

Phyto-extracts for managing *Alternaria* leaf spot on mung bean

ABSTRACT

The present study was investigated to evaluate the effect of selected phyto-extracts for the management of *Alternaria* leaf spot on mung bean caused by *Alternaria alternata* (Fr.) Keissler, under field conditions. Three replications of mung bean were planted in a randomized block design. At the research plot of the Central Research Field, Department of Plant Pathology, Sam Higginbottom University of Agriculture, Technology and Sciences, Prayagraj during *Zaid* season of 2023. The effect of neem leaf extract, onion bulb extract, ginger rhizome extract, garlic bulb extract, turmeric rhizome extract and Mancozeb were compared and tested the plant height (cm), pod length (cm), disease intensity (%) and B:C ratio recorded at 45, 60 and 75 DAS interval. The effect of foliar spray of T₄-garlic bulb extract @ 10% was found the best for managing *Alternaria* leaf spot of mung bean under field conditions as minimum disease intensity(34.4%), maximum plant height (65.4cm), pod length significantly increased (8.4cm)and highest B:C ratio(1:2.05).Followed by minimum disease intensity(35.5%),plant height (60.9cm), pod length (7.8 cm) and highest B:C ratio(1:1.97) was recorded inT₁-neem leaf extract @ 10%followed by T₅-turmeric rhizome extract @ 10% when compared to (Treated check) T₆-Mancozeb @0.2% and (Untreated check)T₀-control. Overall results revealed that T₄-garlic bulb extract @10% is significantly effective against *A. alternata* (Fr.) Kessler.

Keywords: *Alternaria alternata* (Fr.) Keissler, garlic bulb extract, neem leaf extract, turmeric rhizome extract, ginger rhizome extract, onion bulb extract.

1. INTRODUCTION

Mung bean [*Vigna radiata* (L.) Wilczek] is an important summer food legume crop of short duration and cultivated in the tropical and sub-tropical countries of the world. In India, this crop is cultivated in all three seasons viz., *Kharif*, *Rabi* and *Zaid*. Its seed are a good source of dietary proteins, vitamins and minerals for humans especially for marginal and vegetarian people. It is an ideal crop for small holding farmers due to its low input requirement, nitrogen fixation capability and short duration. These qualities make it highly suitable for crop rotation [1,2]. Worldwide, the total cultivated area and production of mung bean are 7 million ha and 5 million tonnes respectively. The yield potential of this crop ranges from 2.5 to 3.0 t ha⁻¹; however, the global average productivity is 0.5 t ha⁻¹. South Asia contributes 90% of total mung bean production worldwide, and India is the largest producer in South Asia with production of about 1.5 to 2.0 million tonnes from about 3 to 4 million hectare annually, with average productivity of 0.5 t ha⁻¹ [3,4].

Abiotic and biotic stresses caused significant decline in legume yield in South Asia and South East Asia. Among biotic stresses, fungal diseases are responsible for reducing yield up to 40–60% in mung bean [5]. Fungal pathogens can infect mung bean plants at different stages, such as during emergence, seedling, vegetative and reproductive stages and cause substantial damage leading to yield loss or complete failure of production. Species of the genera *Fusarium* (wilt), *Rhizoctonia* (wet root rot), and *Macrophomina* (dry root rot) infect mung bean plants during seed/seedlings stages (seed-borne or soil borne), while species of

36 the genera *Colletotrichum* (anthracnose), *Alternaria* and *Cercospora* (leaf spot),
37 *Erysiphe/Podosphaera* (*Sphaerotheca*) (powdery mildew) affect plants during vegetative and
38 reproductive stages [6].[7]reviewed the status of web blight in mungbean and
39 recently,[8]published a review on yellow mosaic in mungbean and urd bean from India.

40

41 Mung bean is an important pulse crop in many Asian countries including Pakistan, where the
42 diet is mostly cereal based. Mung bean similar to other pulses, is a good source of protein,
43 vitamins, minerals and calories [9].A wide variety of fungus isolates has been identified that
44 infect the mung bean crop causing dry root rot, charcoal rot and cercospora leaf spot [1].
45 The most prominent fungal disease of mung bean is the leaf spot disease. The symptoms
46 include the formation of spots of different kind and shape on leaves (sometimes on stems
47 and fruits; depending upon the type of the fungus). The genus *Alternaria* has its various
48 species that are known to cause a significant damage of crops in the field as well as at late
49 harvesting stage. It is well known pathogen affecting pulses, cereals fruits and vegetables
50 that result in severe agricultural losses [10,26,27,28].

51

52 A wide variety of research based on epidemiological studies have revealed that *Alternaria*
53 *alternata* spores are the most commonly recognized fungal spore in the environment
54 [11].*Alternaria* black rot, *Alternaria* leaf spot and *Alternaria* brown spot are the most
55 significant phenotypes of plant diseases caused by *A. alternata* spores [12]. *Alternaria* leaf
56 spot damage is more prominent on aerial parts and approximately upto 80% losses were
57 recorded.Natural plant extracts modulate plant growth and are involved in plant defense
58 responses, including limiting pathogen development [13]. The action of natural compounds
59 such as terpenoids, phenolics, and alkaloids are not specific, and their effects on pathogens
60 are versatile[14]. Thus the present study aimed to evaluate the effect of selected phyto-
61 extracts for the management of *Alternaria* leaf spot on mung bean. Keeping the above in
62 view the study was undertaken during *Zaid* season 2023, at the Central Research Field,
63 Department of Plant Pathology, Sam Higginbottom University of Agriculture, Technology
64 And Sciences, Prayagraj.

65

66 2. MATERIAL AND METHODS

67

68 The experiment was carried out at the Central Research Field, Department of Plant
69 Pathology, Sam Higginbottom University of Agriculture, Technology And Sciences, Prayagraj
70 during *Zaid* season 2023.The study was laid-out with Randomized Block Design (RBD) with
71 three replications.Three sprays of all treatments were given at an interval of 15 days.
72 Treatments were imposed after appearance of the first disease symptoms. Observations on
73 plant height (cm) and disease intensity (%) of *Alternaria* leaf spot of mung bean were
74 recorded at 15 days interval, pod length (cm) and B:C ratiodata were obtained after the
75 harvest on physiological maturity.The treatments comprised of application of selected phyto-
76 extracts *viz.*neem, garlic, ginger, onion, turmeric @10% andmancozeb @0.2% (treated
77 check) and control (untreated). The crop was sprayed three times at 45, 60, and 75 DAS of
78 interval. The disease intensity of *Alternaria* leaf spot was recorded after ten days of spray.
79 Plant disease intensity (PDI)was calculated after each spray by using 0-5 disease rating
80 scale on the basis of percentage area of foliage infected by the pathogen.

81

82 2.1 Isolation and identification of the pathogen

83

84 Diseased samples collected from different areas during the season and isolation of
85 pathogenwas carried out in the laboratory. Firstly collected diseased samples washed
86 thoroughly under the tap water and then cut into small pieces 2-4 mm in size with the help of
87 a sterilized blade in such a way that the sample contained a 50 per cent healthy portion as
88 well as a 50 per cent diseasedportion. The surface of the pieces was sterilized by using 1

89 per cent Sodium hypochlorite solution for 30 seconds to 1 minute, then finally wash well with
 90 the three changes of sterilized distilled water and to remove excess water then pieces were
 91 placed on blotter paper. With the help of a sterilized inoculating needle place the sample
 92 pieces on petri plates containing potato dextrose agar medium under the aseptic conditions
 93 in the laminar airflow chamber. Five pieces on PDA media on each plate. Inoculated petri
 94 plates kept in an incubator at 25±2°C and examine at frequent intervals to check the growth
 95 of the target fungal pathogen.
 96 Identification of the target pathogen (*Alternaria alternata*). The fungal colony culture was
 97 initially white, cottony, and abundant in aerial mycelium, but it progressively turned grey.
 98 With aerial mycelium, the old culture appeared entirely grey, and clear concentric rings
 99 formed on the PDA medium. Conidiophores ranged simple to branched, and appeared singly.
 100 Conidia were seen in lengthy chains on conidiophores, with strong walls, beaked edges, and
 101 transverse and longitudinal septation and golden to brown in appearance. The fungus was
 102 identified as *A. alternata* based on colony characteristics and morphological characteristics of
 103 conidiophores and conidia [15,16].

104
 105 **2.1.1 Purification and maintenance of the pathogen**

106
 107 The culture obtained was purified once after the pathogen identity confirmed. Periodic sub
 108 culturing on PDA slants was done to keep the pure culture. Throughout the investigation, this
 109 pure culture employed. The cultures of the fungus were sub-cultured on PDA slants and kept
 110 in laboratory at 28±1 °C for 15 days. Further, these cultures were sub-cultured once in a
 111 month and used for future purpose [17].

112
 113 **2.2 Preparation of Phyto-extracts**

114
 115 Phyto-extract was prepared from, leaves of neem (*Azadirachta indica*), bulb of garlic
 116 (*Allium sativum*), rhizome of ginger (*Zingiber officinale*), bulb of onion (*Allium cepa*), rhizome of
 117 turmeric (*Curcuma longa*) washing with running tap water followed by sterile distilled water,
 118 air dry at 27°C and ground to obtain extracts of each plant species the extraction was
 119 done by means of pestle and mortar. Water extract was obtained by adding one gm of tissue
 120 in one ml of water (1:1w/v) and filtered through double layers of muslin cloth. This forms the
 121 standard solution (100%). The phyto-extracts were sprayed at the rate of 10% prepared from
 122 standard solution [18]. All the treatments were given as foliar sprays. Phyto-extracts were
 123 sprayed @ 10 ml/liter of water, mancozeb @ 2 ml/ liter of water.

124
 125 **2.3 Recording the disease intensity (%)**

126
 127 After germination, five plants per treatment per replication were randomly selected.
 128 Regularly watched for first appearance of disease. The observation on disease intensity was
 129 recorded using a progressive 0-5 scale, as shown in (Table 1 and Plate 1). Numerical
 130 rating grade was given on the basis of percentage of area infected by pathogen on the
 131 leaves [19] as described below:

132
 133 **Table 1: Disease rating scale**

134

Grade	Leaf area covered
0	No symptoms of disease on leaves (Immune)
1	Spots size covering 1-10% of leaf area (Resistant)
2	Spots size covering 11-25% of leaf area (Moderately resistant)

3	Spots size covering 26-50% of leaf area (Moderately susceptible)
4	Spots coalesce to form big patches covering 51-75 % of leaf area (Susceptible)
5	Big spots size covering >75 % of leaf area (Highly susceptible)

135
136
137
138
139
140
141
142
143
144
145
146
147
148
149
150
151
152
153
154
155
156
157
158
159
160
161
162
163
164
165
166
167
168
169
170
171
172
173
174
175
176
177
178
179
180
181
182

2.3.1 Per cent disease intensity (PDI)

$$\text{PDI} = \frac{\text{Sum of all numerical ratings}}{(\text{Total number of leaves rating} \times \text{Maximum disease grade})} \times 100$$



Plate 1: Disease rating scale

2.4 Economics analysis

Cost of cultivation, gross return, net return and benefit cost ratio was worked out to evaluate the economics of each treatment, based on the existing market prices of input and output [20].

2.4.1 Cost of cultivation (ha⁻¹)

The cost of cultivation for each treatment was work out separately, taking into consideration all the cultural practices followed and costs of inputs used in the cultivation.

2.4.2 Gross return (ha⁻¹)

The gross return from each treatment was calculated by using the following formula:

$$\text{Gross return (ha}^{-1}\text{)} = \text{Yield (q/ha)} \times \text{Price (Rs./q)}.$$

2.4.3 Net return (ha⁻¹)

The net profit from each treatment was calculated separately by using the following formula:

$$\text{Net return} = \text{Gross return (ha}^{-1}\text{)} - \text{Cost of cultivation (ha}^{-1}\text{)}.$$

2.4.4 Benefit Cost ratio

The benefit cost ratio was calculated by using the following formula:

$$\text{Benefit Cost ratio} = \frac{\text{Net return (ha}^{-1}\text{)}}{\text{Total cost of cultivation (ha}^{-1}\text{)}}$$

2.5 Statistical analysis

183 The data obtained were statistically analyzed by the methods suggested by [21].The
184 standard error and critical difference (C. D.) was calculated at 5% level
185 of significance for comparing treatment means.

186

187 3. RESULTS AND DISCUSSION

188

189 Under field conditions, three sprays of all the treatments were taken up at 45, 60 and 75
190 DAS. The results are presented on Table 2; Fig. 1, 2 and plate 2.

191

192 3.1 Disease intensity (%) of Alternaria leaf spot on mung bean

193

194 The data presented in the Table 2 and depicted in Fig. 1 reveals that disease intensity (%) of
195 mung bean significantly decreased in treatment T₄ – Garlic bulb extract (34.4 %) followed by
196 T₁ – Neem leaf extract (35.5 %), T₅ – Turmeric rhizome extract (36.7 %), T₃ – Ginger
197 rhizome extract (38.0 %) and T₂ - Onion bulb extract (39.0 %) as compared to (Treated
198 check) T₆ – Mancozeb (32.8 %) and (Untreated check) T₀ – Control (40.9 %).

199 Comparing the treatments with CD value (1.07), all the treatments (T₁, T₂, T₃, T₄, T₅, T₆)
200 were found significant over (Untreated check) T₀ – Control. Among the treatments (T₂, T₃)
201 were found non-significant to each other. (T₁ and T₄) were found to be significant over all the
202 other treatments.

203

204 3.2 Plant height (cm) on mung bean

205

206 The data presented in the table 2 and depicted in Fig. 1 reveals that plant height (cm)
207 of mung bean significantly increased in treatment T₄ – Garlic bulb extract (65.4 cm) followed
208 by T₁ – Neem leaf extract (60.9 cm), T₅ – Turmeric rhizome extract (58.8 cm), T₃ –
209 Ginger rhizome extract (57.0 cm) and T₂ - Onion bulb extract (55.2 cm) as compared to
210 (Treated check) T₆ – Mancozeb (69.7 cm) and (Untreated check) T₀ – Control (48.1 cm).

211 Comparing the treatments with CD value (2.06), all the treatments (T₁, T₂, T₃, T₄, T₅, T₆)
212 were found significant over (Untreated check) T₀ – Control. Among the treatments (T₂, T₃);
213 (T₃, T₅) were found non-significant to each other. (T₁, T₂ and T₄) were found to be significant
214 over all the other treatments.

215

216 3.3 Pod length (cm) on mung bean

217

218 The data presented in the table 2, Fig. 1 and depicted in plate 2 reveals that pod length (cm)
219 of mung bean significantly increased in treatment T₄ – Garlic bulb extract (8.4 cm) followed
220 by T₁ – Neem leaf extract (7.8 cm), T₅ – Turmeric rhizome extract (7.4 cm), T₃ – Ginger
221 rhizome extract (7.0 cm) and T₂ - Onion bulb extract (6.8 cm) as compared to (Treated
222 check) T₆ – Mancozeb (9.4 cm) and (Untreated check) T₀ – Control (6.2 cm).

223 Comparing the treatments with CD value (0.53), all the treatments (T₁, T₂, T₃, T₄, T₅, T₆)
224 were found significant over (Untreated check) T₀ – Control. Among the treatments (T₂, T₃);
225 (T₃, T₅) and (T₅, T₁) were found non-significant to each other. (T₂ and T₄) were found to be
226 significant over all the other treatments.

227

228

229

230

231

232

233

234

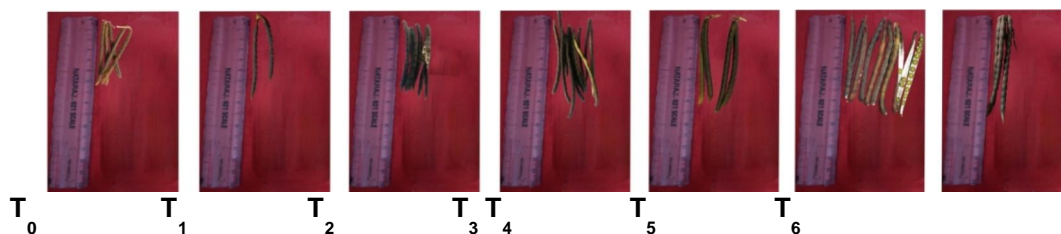


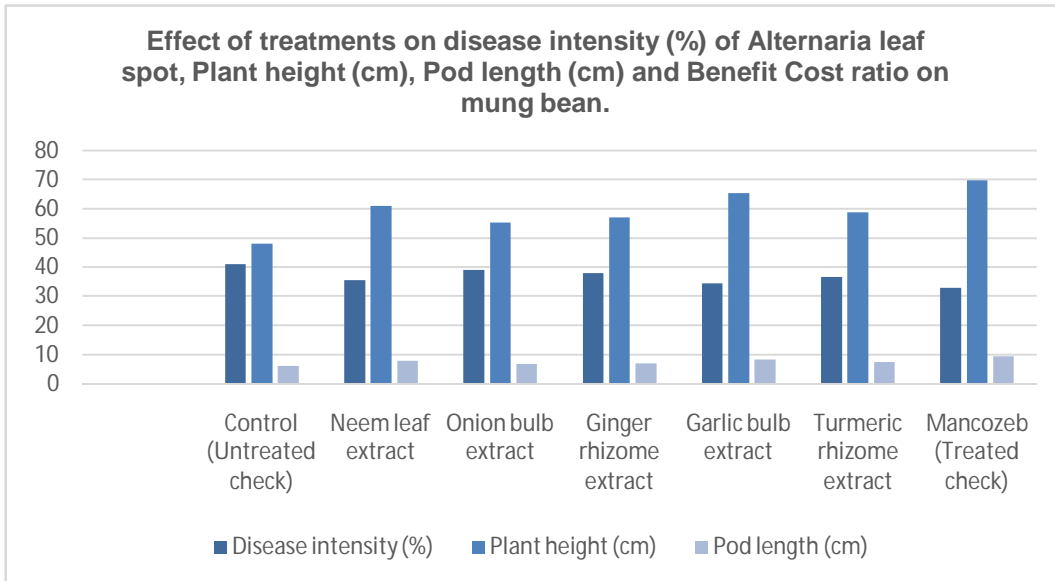
Plate 2 Effect of treatments on pod length (cm) of mung bean after harvest

3.4 Benefit Cost ratio on mung bean

The data presented in the Table 2 and depicted in Fig. 2 reveals that economics analysis of the data over the session that (Treated check) T₆ - Mancozeb recorded higher gross returns Rs. 86,767.00, net returns Rs. 60,587.00 with B:C ratio **1:2.31** followed with T₄ – Garlic bulb extract recorded gross returns Rs. 84,881.00, net returns Rs. 57,051.00 with B:C ratio **1:2.05** as compared to (Untreated check) T₀ – Control gross returns Rs. 64,558.00, net returns Rs. 39,728.00 with B:C ratio **1:1.60**.

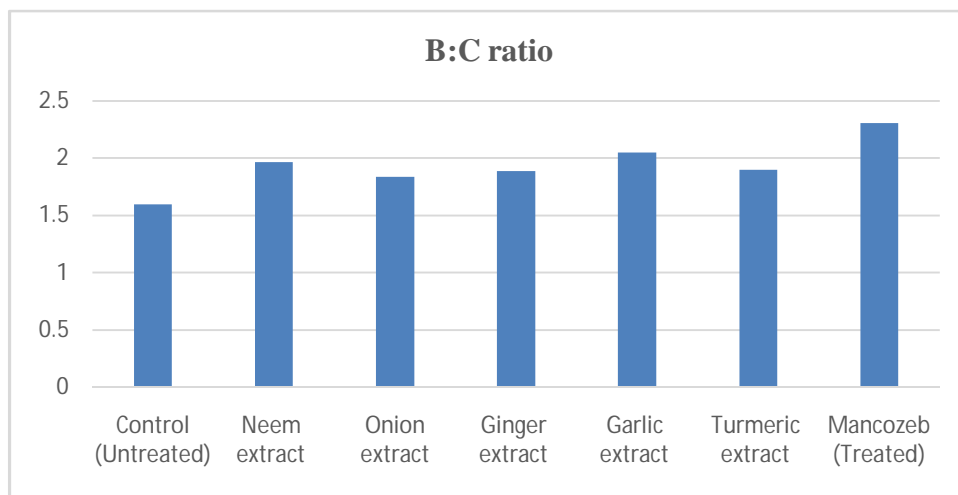
Table 2: Effect of treatments on disease intensity (%) of Alternaria leaf spot, Plant height (cm), Pod length (cm) and Benefit Cost ratio on mung bean.

Treatment number	Treatments	Dosage	Disease intensity (%)	Plant height (cm)	Pod length (cm)	Benefit Cost ratio
T ₀	Control (Untreated check)	-	40.9 ^a	48.1 ^a	6.2 ^a	1:1.60
T ₁	Neem leaf extract	10 %	35.5 ^d	60.9 ^d	7.8 ^d	1:1.97
T ₂	Onion bulb extract	10 %	39.0 ^b	55.2 ^b	6.8 ^b	1:1.84
T ₃	Ginger rhizome extract	10 %	38.0 ^b	57.0 ^{bc}	7.0 ^{bc}	1:1.89
T ₄	Garlic bulb extract	10 %	34.4 ^e	65.4 ^e	8.4 ^e	1:2.05
T ₅	Turmeric rhizome extract	10 %	36.7 ^c	58.8 ^c	7.4 ^{cd}	1:1.90
T ₆	Mancozeb (Treated check)	0.2 %	32.8 ^f	69.7 ^f	9.4 ^f	1:2.31
CD at 5%			1.07	2.06	0.53	



264
265
266
267
268

Fig. 1 Effect of treatments on disease intensity (%) of *Alternaria* leaf spot, Plant height (cm), Pod length (cm) and Benefit Cost ratio on mung bean.



269
270
271
272
273
274
275
276
277
278
279
280
281

Fig. 2 Effect of treatments on Benefit Cost ratio of mung bean

3.5 Discussion

The probable reasons for such findings may be due to the antimicrobial properties of garlic extract and neem leaf extract due to the presence of volatile oil which contains diallyl disulphides, diallyl trisulphides and sulphodioxides derived from allicin or ajoene which disrupts the cell membrane of the pathogen and restricts the performance of some enzymes that are important to proliferate fungi in nature and inhibits enzyme formation leading to cell death and loss of vigour, ultimately killing the pathogen by this garlic may indirectly influence the plant growth. The research outcomes align with the similar findings was reported by [22,23,24,25], who revealed that the maximum disease control was found in the

282 garlic treatment, suggesting that the use of garlic extracts could be a viable approach to
283 control *Alternariaalternata* (Fr.) Keissler. But among all the treatments chemical fungicide
284 (Treated check) has shown the minimum disease control has shown with strong fungicidal
285 effect against pathogen. In order to reduce the pathogen, it may produce some toxic
286 chemical residues, they may have potential harmful effects to non-targeted organism. So,
287 considering the ecosystem, phyto-extracts of garlic and neem significantly inhibits the
288 pathogen could lead to better health of the plants this cascade effect may have contributed
289 to the holistic wellbeing of plants, consequently resulting in producing minimum disease
290 intensity, increased plant height, producing maximum pod length growthand leading to a
291 healthier plant.

292

293 **4. CONCLUSION**

294

295 *Alternariaalternata* (Fr.) Keissler was found associated with Alternaria leaf spot disease of
296 mung bean (*Vigna radiata* L.). The disease intensity (%), plant height (cm), pod length (cm)
297 and B:Cratio, overall results revealed that T₄ - Garlic bulb extract @10% is significantly
298 effective against *A. alternata* (Fr.) Kessler as compared to (Treated check) T₆ - Mancozeb @
299 0.2% and (Untreated check) T₀- Control. It is worth mentioning that the conclusions drawn
300 from this study were based on observations carried out under field conditions at Central
301 Research Field, Department of Plant Pathology, Sam Higginbottom University of Agriculture,
302 Technology And Sciences, Prayagraj,during *Zaid* Season 2023, within the agro- climatic
303 conditions of Prayagraj, UP. India.

304

305 Disclaimer (Artificial intelligence)

306 Option 1:

307 Author(s) hereby declare that NO generative AI technologies such as Large Language
308 Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during
309 writing or editing of manuscripts.

310 Option 2:

311 Author(s) hereby declare that generative AI technologies such as Large Language Models,
312 etchave been used during writing or editing of manuscripts. This explanation will include the
313 name, version, model, and source of the generative AI technology and as well as all input
314 prompts provided to the generative AI technology

315 Details of the AI usage are given below:

316 1.

317 2.

318 3.

319

320 **ACKNOWLEDGEMENTS**

321

322 The authors express their gratitude to theDepartment of Plant Pathology for providing
323 necessary facilities to carry out the whole experimental research work.

324
325
326
327
328
329
330
331
332
333
334
335
336
337
338
339
340
341
342
343
344
345
346
347
348
349
350
351
352
353
354
355
356
357
358
359
360
361
362
363
364
365
366
367
368
369
370
371
372
373
374
375
376

REFERENCES

1. Nair RM, PandeyAK, WarAR, HanumantharaoB, ShweT, AlamA, et al. Biotic and abiotic constraints in mung bean production-progress in geneticimprovement. *Frontiers in Plant Science*. 2019;10:1340.
2. UdomkunP, TirawattanawanichC, IlukorJ, SridonpaiP, NjukweE, NimbonaP, VanlauweB. Promoting the use of locally produced crops in making cereal-legume-based composite flours: An assessment of nutrient, antinutrient, mineral molar ratios, and aflatoxin content. *Food Chemistry*. 2019;286:651–658.
3. Clarry S. The rise and rise of mung beans in ground cover tm supplement issue 125 November-December. *Grains Research and Development Corporation*. 2016;15.
4. Pratap A, DouglasC, Prajapati U, KumariG, WarAR, TomarR, et al. Breeding progress and future challenges: Biotic stresses. *The mung bean genome. Compendium of plant genomes*. Springer. 2020;55–88.
5. Kaur L, SinghP, Sirari A. Biplot analysis for locating multiple disease resistance diversity in mungbean germ plasm. *Plant Disease Research*. 2011;26(1):55-60.
6. RyleyMJ, TatnellJR. Management of the major foliar diseases of mung beans and peanuts in Australia. In proceedings of the 4th Asian conference on Plant Pathology and the 18th Biennial Australasian Plant Pathology Society Conference. 2011; pp. 61-61.
7. SinghJ, MishraKK, SinghAK. Current status of web blight of mungbean. *An Asian Journal of Soil Science*. 2013;8:495–504.
8. NaimuddinAM, SinghNP. Yellow mosaic of mungbean and urdbean: current status and future strategies. *Journal of Food Legumes*. 2016;29(2):77-93.
9. Khan MN, ShoabM, AshrafMS, QamarR, MahboobA, IjazS. Mung bean (*Vignaradiata*) intercropping enhances productivity of late season irrigated cotton in Punjab. *Asian Journal of Agriculture and Biology*. 2020;8:472-479.
10. Barkai-GolanR, PasterN. *Alternaria* toxins and plant diseases: an overview of origin, occurrence and risks. *World Mycotoxin Journal*. 2008;2:129-140.
11. WoudenbergJHC, Van der MerweNA, JurjevicZ, GroenewaldJZ, CrousPW. Diversity and movement of indoor *Alternaria alternata* across the mainland USA. *Fungal Genetics and Biology*. 2015;81:62-72.
12. LogriecoA, MorettiA, SolfrizzoM. *Alternaria* toxins and plant diseases: an overview of origin, occurrence and risks. *World Mycotoxin Journal*. 2009;2:129-140.
13. Jamiolkowska A. Natural compounds as elicitors of plant resistance against diseases and new. *Biocontrol strategies*. *Agronomy*. 2020;10(2):173.
14. Righini H, Francioso O, Di Foggia M, Quintana AM, Roberti R. Assessing the potential of the terrestrial cyanobacterium *Anabaenaminutissima* for controlling *Botrytis cinerea* on tomato fruits. *Horticulturae*. 2021;7(8):210-216.
15. KeisslerKV. Zur Kenntnis der Pilzflora Krains. *Beihefte Zum Botanischen Centralblatt*. 1912;29:395-440.
16. Hunter BB, BarnettHL. *Deuteromycetes (Fungi Imperfect)*, In: *Handbook of Microbiology: 1, Organismic Microbiology*. Laskin, A. I. and H. A. Lechevalier, Eds. CRC Press, Cleveland, OH; 1973.
17. ToussounTA, NelsonPE. A pictorial guide to the identification of *Fusarium* species. *Australian Plant Pathology Society Newsletter*. 1976;6:11-13.
18. ShekhawatPS, PrasadRL. Antifungal activities of some plant extracts. Inhibition of spore germination. *Indian Phytopathology*. 1971;24:800-802.
19. MckinneyHH. A new system of grading plant diseases. *Journal of Agricultural Research*. 1923;26:195–218.
20. ReddyTY, ReddiGHS. *Principles of Agronomy*. 2nd ed. Kalyani Publishers: Ludhiana; 1995.

- 377 21. Gomez KA, Gomez AA. Statistical Procedures for Agricultural Research. 2nd ed.
378 John Wiley and Sons: London; 1986.
- 379 22. ChethanaBS, GaneshanG, RaoAS, BellishreeK. *In-vitro* evaluation of plant extracts,
380 bio-agents and fungicides against *Alternariaporri* (Ellis) Cif., causing purple blotch
381 disease of onion. Pest Management in Horticultural Ecosystems. 2012;18(2):194-
382 198.
- 383 23. Kutawa AB, DanladiMD, HarunaA. Antifungal activity of garlic (*Alliumsativum*) extract
384 on some selected fungi. Journal of Medicinal Herbs and Ethnomedicine. 2018;4:12-
385 14.
- 386 24. HassaneinNM, ZeidMA, YoussefAK, MahmoudDA. Efficacy of leaf extracts of neem
387 (*Azadirachtaindica*) and china-berry (*Meliaazedrach*) against early blight and wilt
388 diseases of tomato. Australian Journal of Basic and Applied Sciences.
389 2008;2(3):763-772.
- 390 25. NandagopalV, Ghewande, MP. Use of neem products in groundnut pestmanagement
391 in India. Agricultural and Food Sciences. 2004;3(3):150-155.
- 392
- 393
- 394
- 395 26 SHAGARI, Z. A., M. BELLO, U. K. MOHAMMED, A. I. DABAI, A. MAHMUDA, A.
396 S. BAKI, A. U. IMAM, AND M. GANAU. 2021. "TOXICOLOGICAL ACTIVITY OF
397 THE METHANOLIC LEAF EXTRACT OF SOME MEDICINAL PLANTS USED IN
398 SOKOTO TOWNSHIP AND ENVIRONS". JOURNAL OF ADVANCES IN BIOLOGY
399 & BIOTECHNOLOGY 24 (10):54-62.
400 [HTTPS://DOI.ORG/10.9734/JABB/2021/V24I1030247](https://doi.org/10.9734/JABB/2021/V24I1030247).
- 401 27 SHANKER AK, CERVANTES C, LOZA-TAVERA H, AVUDAINAYAGAM S.
402 CHROMIUM TOXICITY IN PLANTS. ENVIRONMENT INTERNATIONAL. 2005 JUL
403 1;31(5):739-53.
- 404 28 BABULA P, ADAM V, OPATRILOVA R, ZEHNALÉK J, HAVEL L, KIZEK R.
405 UNCOMMON HEAVY METALS, METALLOIDS AND THEIR PLANT TOXICITY: A
406 REVIEW. ORGANIC FARMING, PEST CONTROL AND REMEDIATION OF SOIL
407 POLLUTANTS: ORGANIC FARMING, PEST CONTROL AND REMEDIATION OF
408 SOIL POLLUTANTS. 2010:275-317.