

Review Article

REPRODUCTION IN INSECTS

Abstract

Reproduction is a fundamental process in all life forms, including insects, in which offspring are produced from the parent organisms. Insects reproduce through both sexual and asexual reproduction, ensuring rapid population increase. Sexual reproduction involves the use gametes from both male and female insects, whilst asexual reproduction permits solitary creatures to lead to genetically indistinguishable progeny. Reproductive ecology deals with the study of how physiological characteristics, behavioral, and environmental elements impact on insect reproduction. The complexity of insect reproduction is focused by important processes known as vitellogenesis, which is required for egg development and transgenerational immunity. The i5k initiative planned to sequence more than 5,000 insect genomes to increase our apprehension and understanding, yet problems still exist because of genome assembly issues and budgetary limitations. Understanding the reproductive morphology and methods of insects, as well as oviposition-related genes, is critical for effective pest management and biological control measures. Insect reproductive research contributes immensely to evolutionary biology, conservation, and agricultural productivity by understanding life cycles, reproductive behaviors, and pollination roles. Insect neurobiology, microbiome and environmental entomology are three most important study fields that will come up with new insights into basic biological processes as well as anticipated pest management and conservation measures in the face of environmental changes in the coming future.

Keywords: Reproduction, Ecology, Vitellogenesis, Oviposition, Evolutionary, Neurobiology, Microbiome.

1.0 INTRODUCTION

Reproduction is the basic feature of all known life of organisms and the natural process by which offspring are produced from their parents [62]. Generally, there are two forms of reproduction: sexual and asexual reproduction. In sexual reproduction, two reproductive cells, the male and female organisms called gametes, which contain half the number of chromosomes of normal cells, are created by meiosis, and the male gamete fertilizes the female gamete of the same species to create a fertilized zygote. This produces an offspring whose genetic makeup is derived from those of the two parental organisms. In asexual reproduction, an organism can reproduce without the involvement of another organism, creating a genetically similar or identical copy of itself.

In similar research, it was stated that sexual and asexual modes of reproduction are carried out in insects, thus ensuring their rapid increase in numbers. Reproductive ecology deals with the organism's physiology, behaviors, and the abiotic and biotic environments in which it lives to determine how it reproduces [62]. The female reproductive functions, physiological traits, and egg characteristics determine the percentage number of offspring a species can produce. Numerous interconnected factors **inflict restrictions on** this reproductive outcome, which can be categorized as limitations arising from the female's physical condition, the abiotic conditions faced by the female and her eggs, and the presence of natural predators [68].

The process of female femaleinsect reproduction starts with vitellogenesis, a process of accumulation of yolk protein during **the formation of oocyte**. The fusion of vitellogenesis has been reported in follicle cells [26], hemocytes [4][37], nurse cells [51][48], and in **a number of** insect species. Furthermore, in honey bees, vitellogenesis acts as a pathogen receptor and transfers pathogen-derived fragments into the offspring to attain trans-generational immunity [23][65][82]. In the small brown planthopper, vitellogenesis integrated by hemocytes can expedite the upright transmission of **the** rice stripe virus [37].

The number of vitellogenesis genes between different insect species ranges from one to three in general [11].

Vitellogenesis is important for egg development during adulthood and embryonic development after oviposition. Vitellogenesis also acts to safeguard the queen and the worker bees from oxidative tension and extends the lifespan of honey bees [66][12][38]. Moreover, vitellogenesis plays an important role in recognizing fat body sugars and gustatory insight in the worker bees [77][76][54]. In this review, we will be exploring reproduction strategies, the reproductive organs, genomes, and their functions in insects as this will help to design effective biological control measures in further research programs because understanding reproduction in insects is key to the successful understanding of oviposition. Oviposition comprises all physical and biological processes from egg laying to egg hatch, which is essential to the survival and fitness of all insect species.

1.1 IMPORTANCE OF INSECT REPRODUCTION

Ecosystems rely on the essential functioning of insects as the most important and major herbivores of most terrestrial communities in the world. An important pillar of the food web is supported by insects, which transfer plant energy further up the food chain. Many species of fish, birds, mammals, amphibians, reptiles, and birds rely mostly on insects for food and sustenance. During the breeding season, birds rely on insects to provide food for their young [61], to maintain a species' separate identity while combining to form new individuals in a population. This leads to the asexual replication of genetic copies of organisms. The emergence of new species promotes the evolution of organisms, which is essential for their survival due to the constant change in their environment [61]. To produce species variations since no two people are alike and because each person inherits certain genetic traits from both parents, resulting in a somewhat different version of themselves and over hundreds of years, these minute differences add together to generate new species [52].

1.2 SEXUAL AND ASEXUAL REPRODUCTION IN INSECTS

Asexual reproduction consists of several types in the animal kingdom, these include budding, fission, fragmentation, and parthenogenesis. Parthenogenesis is the commonest form in insects and it occurs when an unfertilized egg develops into an offspring. Depending on the species, an organism produced through asexual reproduction can be diploid or haploid. Sexual reproduction is defined as the type of

production that occurs when the female insect produces eggs, which are fertilized by the male sperm cell, and then the eggs are usually placed near the required food for nutrition.

2.0 INSECT REPRODUCTIVE ANATOMY

2.1 OVERVIEW AND FUNCTIONS OF THE MALE REPRODUCTIVE PARTS

The most important part of the male reproductive system is a pair of testes, which is mostly located closer to the back of the abdomen. Testis development, including spermatogenesis, is affected by developmental stages, temperatures, nutritional conditions, and hormones [16][19]. In the yellow dung fly (*Scathophagastercoraria*), the testes atrophy after mating [78]. Similarly, in *Meign* (*Drosophila melanogaster*), there was a significant reduction in the testes size after five successive matings [46]. In general, testis development is closely related to spermatogenesis [17] and testis size usually increases as spermatogenesis becomes more active [44]. However, the relationship between testis size and spermatogenesis depends on the species and developmental stage. The shape of the testis is circular, oval, or elongated, perhaps corresponding to sperm length, probably because sperm develop greatly during spermiogenesis [39]. Sperm is produced at the follicles, which is an example of the functional unit, located within the testis. An insect testis consists of hundreds of follicles, usually arranged parallel to each other. Near the distal end of each follicle, there are a group of germ cells known as spermatogonia that divide by mitosis and increase in size to form spermatocytes. These spermatocytes migrate toward the basal end of the follicle, pushed along by continued cell division of the spermatogonia [52].

Each spermatocyte undergoes meiosis, resulting in four haploid spermatids that undergo further development into mature spermatozoa within the follicle. The mature sperm then exit the testes through short ducts known as vasa differentia and accumulate in storage chambers called seminal vesicles, which are typically enlarged sections of the vasa. These seminal vesicles are connected to similar ducts known as vasa deferentia, which converge near the body's midline to form a single ejaculatory duct that

facilitates the release of sperm through the male's copulatory organ, known as an aedeagus. Additionally, the male reproductive system is often accompanied by one or more pairs of accessory glands, which **are glands that** produce seminal fluid to nourish sperm and spermatophores, protective protein structures that encase sperm and aid in their transfer to the female during copulation [9].

2.2 FUNCTIONS OF THE FEMALE REPRODUCTIVE PARTS

The female reproductive system is present in the abdominal cavity of female insects. The female reproductive system of insects consists of the following organs or parts: ovaries, lateral oviduct, median oviduct or vagina, spermatheca, accessory glands, and gonopophysis. The following are the functions of the various female reproductive parts:

Ovaries: An insect's intestine **is located** above its anterior abdominal chamber, which houses a single pair of ovaries. It is situated **on both sides of the female insect's abdomen, inside the tergum**. The insect's two ovaries, which are mesodermal in origin, generate eggs. Many ovarioles, or functional units, make up each ovary. Every ovariole is muscular, elongated, and coated in an epithelial layer [73]. Each ovariole of an insect is composed of many pieces, which means pedicel and terminal filament germarium oocytes. A long, tubular **structure called the** terminal filament is located in the ovariole's anterior region. Germarium apical consists of oogonia [30]. The outer sheath of the ovariole covers the basal vitellarium. It includes a variety of oocyte types, including primary, secondary, and tertiary oocytes. Vitellin, sometimes known as the yolk, is produced in the adipose tissue, released into the hemolymph, and then absorbed by the oocytes via endocytosis. The terms panoistic type and meroistic type refer to two kinds of ovarioles that are primarily seen in insects. **Panoistic type ovariole: this type is the most basic and has fewer cells. It is primarily seen in** orthopterans and beetles (coleopterans). Lepidoptera and Hymenoptera insects are the primary hosts of meroistic type ovarioles. A nursing cell is present in this form of ovariole to nourish the oocytes [53].

Lateral Oviduct: The pedicel of an insect's ovariole forms one pair of lateral oviducts. Every ovariole's pedicel opens into the lateral oviduct. **Different types of eggs are communicated by lateral oviducts.**

Every lateral oviduct culminates in the formation of a median oviduct below the colon after extending postero-ventrally along the lateral margin of the proctodeal. The lateral oviducts originate from the ectoderm and are muscle-derived.

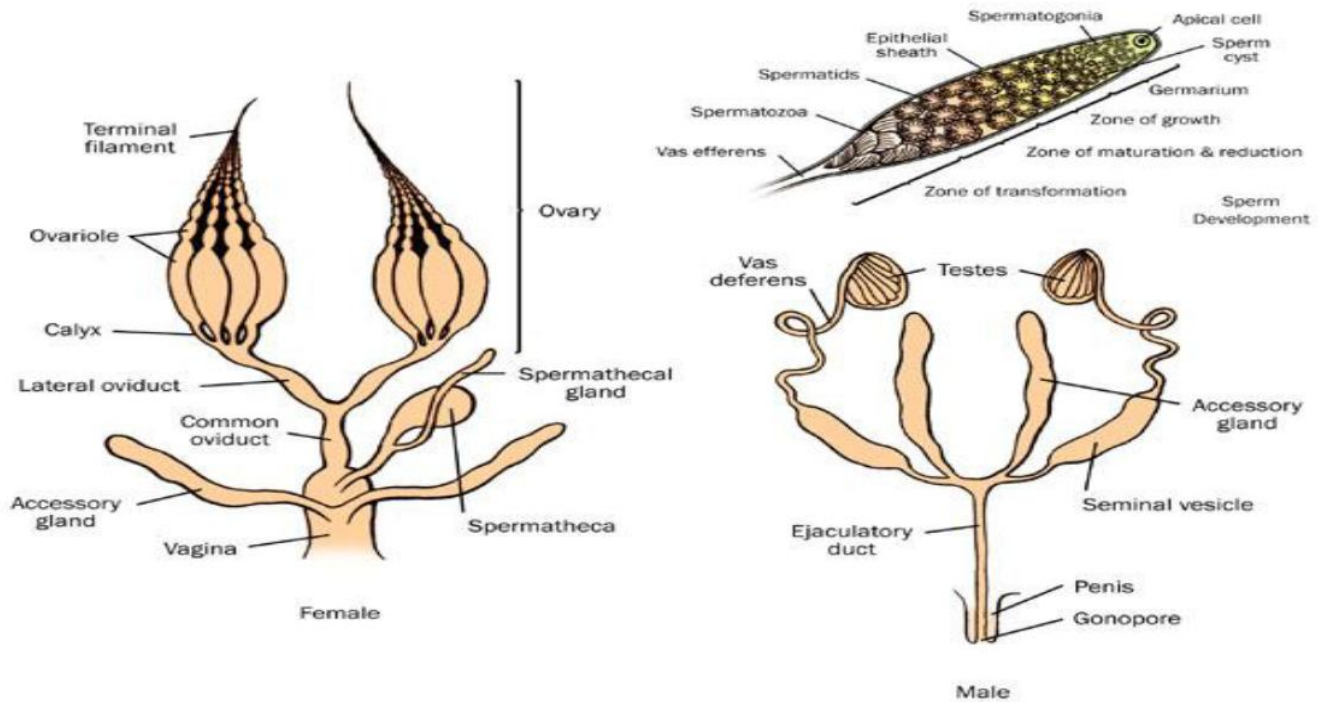
Median Oviduct or Vagina: A common oviduct, median oviduct, or vagina is formed posteriorly by the posterior union of the two lateral oviducts. As a result, the two lateral oviducts meet at a unique "Y" shape. In comparison to the lateral ducts, the median tube is relatively wider. Because of its ectodermal origin, it is muscular and has a modified cuticle lining it. It moves posteriorly and opens ventrally between the plates of the ovipositor. It is also called Barsa copulatrix in moths [30].

Spermatheca: The term refers to a sac-like tubular structure located at the posterior end of the common oviduct. The spermatheca, which consists of a long, coiled duct and a circular chamber, is located in the abdominal cavity near its posterior end, in the midline, below the hindgut. It opens into the genital chamber and has a creamy white color. Spermatheca serves as an insect's storage chamber, holding sperm throughout the process of copulation. Within the common oviduct, the egg is fertilized by these sperm. Insect spermatheca also functions as glands that secrete spermathecal fluid, which feeds sperm [9].

Accessory Glands: A single pair of accessory glands is usually observed in insects, although some insects possess two pairs. These glands are typically located at the apex of the common oviduct. They are responsible for secreting adhesive material that aids in attaching the egg to the substrate. In certain insects, such as grasshoppers, the secretion from the accessory glands contributes to the formation of egg pods. Moreover, in numerous aquatic insects, these glands produce gelatinous fluids that serve to protect the eggs. Wasp species utilize the secretion from their accessory glands to immobilize their prey.

Gonopophysis: The posterior part of the female insect's abdomen features an opening known as the gonopophysis. This structure is also referred to as the ovipositor of the female insect. Enclosed by a chitinous framework, the gonopophysis plays a crucial role in the process of egg deposition and the creation of egg pods [73].

Fig .1 Diagram of the reproductive parts of male and female insects [30].



3.0 TYPES OF REPRODUCTION IN INSECTS

There are generally seven types of reproduction through which insects produce their young. These are listed and explained below:

Viviparity: Hagan [33] strictly defined viviparity in insects as the process in which they “give birth to living offspring which have hatched from the egg within the mother’s body.” The author, however, recognized the fact that some species can deposit eggs with embryos in different stages of development and considered this behavior a different phenomenon, called “ovoviviparity.” Later, Hagan [34] presented a new classification where the viviparity of insects is classified into four groups, based on the modifications related to the strategies utilized to nourish the embryo during development. These are:

- **Ovoviviparity:** The egg contains enough yolk to nourish the embryo till hatching and there are no special structures associated with embryo nourishment.
- **Adrenotropic viviparity:** The egg contains enough yolk to nourish the embryo till hatching and there are special structures associated with embryo nourishment.

- **Haemocoelous viviparity:** The development of the embryo occurs in the haemocoel, not in genital ducts, and embryonic nourishment is derived from maternal tissue.
- **Pseudoplacental viviparity:** The embryo obtains at least part of its nourishment using a pseudoplacenta.

Adenoparous: This strategy occurs when eggs have adequate yolk, and young offspring are fed from milk glands after their release, also, pupating instantly without feeding. A typical example is the (*Glossina pupipara*) of Diptera [50]. This process is further divided into two groups:

- **Pseudoplacental:** This process occurs when eggs with little yolk feeding and nourishment are through a pseudo placenta. An example are psocoptera, dermapteran, and aphids.
- **Haemocoelous:** This strategy occurs when nourishment of young offspring occurs in the hemolymph of the parent mother. Young offspring are born either by the rupture in the walls of the parent or by the genital canal. A typical example is strepsipterous and larvae of cecidomyids [50].

Parthenogenesis: The third most important type of production in insects is parthenogenesis, which is described as the ability of female insects to reproduce without fertilization. This process occurs due to genetic factors, heredity, climatic factors, unsuccessful finding of a mate, and hormonal imbalance within the body of the insect. Parthenogenesis is subdivided into:

- **Sporadic:** Occurs occasionally. An example is the silkworm.
- **Constant:** Occurs regularly or frequently. An example is thrips.
- **Cyclic:** This is an alternation of generations. A typical example is aphids.

Based on the sexes of the offspring produced, parthenogenesis can be classified as:

- **Arrhenotoky:** Only males are produced. An example is Hymenoptera.
- **Thelytoky:** Only females are produced. An example is acridids.
- **Amphytoky:** Occurs when both females and males are produced. An example is hymenopterans.

- **Paedogenesis/Neoteny:** This strategy of parthenogenesis occurs when immature insects or stages give birth to young ones, usually due to hormonal changes. A typical example is cecidomyids [32].

Polyembryony: This phenomenon occurs when insects reproduce by giving birth to two or more offspring instead of a single one, as two or more embryos are generated from a single egg. An example is the endoparasitic Hymenoptera-like platygaster [75].

Hermaphroditism: This type of reproductive phenomenon occurs when both male and female gonads exist in the same individual, which may be functional, as in *Icerya purchasi*, or nonfunctional, as in stonefly (*Perla marginata*) [7][70].

Castration: Castration in individual insects occurs mainly because of the development of the reproductive organs. An example is well-developed ovaries developing in female insects, similarly, well-developed testes develop in male insects. The third is insects with underdeveloped ovaries in worker honey bee insects.

Alternation of Generation: This is another strategy employed by insects through parthenogenesis and bisexual reproduction. For example, aphids exhibit this phenomenon of reproduction by parthenogenesis in summer and sexual reproduction.

4.0 REPRODUCTIVE STRATEGIES IN INSECTS

Semelparity and Iteroparity: Insects demonstrate two reproductive systems, namely semelparity and iteroparity. Semelparity encompasses reproducing once in an organism's lifetime, resulting in a larger number of offspring before the death of the organism. A typical example is cicadas, which endure underground for extended periods before emerging to mate, lay eggs, and die. In iteroparity, an organism reproduces several times in its lifespan, producing smaller numbers of offspring at each reproductive stage. Butterflies are a typical example of iteroparity because they mate and lay eggs several times throughout their life cycle [22].

Drifting Behavior: It has been observed that certain insects, such as wasps, ants, and honeybees, use a distinct reproductive strategy called drifting behavior. These insects are classified as social insects because one or a small number of females are in charge of laying eggs while other **members of the colony** search for food. **In order to** mate and procreate, some colony affiliates leave their original nest and attach to a new one. A study on the bumblebee species *Bombus terrestris* was carried out [8]. The study found that fertile workers use a unique **strategy called drifting behavior** to avoid reproductive rivalry in their nests and instead mate and reproduce in different colonies. Workers who are fertile or infertile exhibit this fluctuating behavior, even if their propensity to immigrate to other colonies and procreate within their new colonies is mostly determined by their level of fertility.

5.0 REPRODUCTIVE AND MATING BEHAVIOR IN INSECTS

The term used to describe the appropriation of energy and resources towards reproduction in insects is known as reproductive effort or behavior. Insects manifest a wide variety of reproductive strategies, and the stage of reproductive effort can differ significantly among species. Some insects reveal a high reproductive effort, where **a major portion** of their energy and resources are committed to reproduction. A typical example is the female green-veined white butterflies (*Pieris napi*), which boost their reproductive effort when they acquire large male donations, but speckled wood butterflies (*Parargeaegeria*) do not. The green-veined white butterflies produced more eggs but did not invest more resources per egg. This behavior was important because donating capacity in green-veined white butterflies is heritable, which leads to greater fecundity and high-donating sons [79].

Egg Size: Egg size is vital in insect reproductive strategies, with remarkable involvement in offspring success. Insects manifest a different scale of egg sizes, which can be linked to different reproductive strategies. In the burying beetle (*Nicrophorus vespilloides*), the body mass of the offspring has a positive effect on egg size when care after hatching is absent, **but this** action dissipates when there is post-hatching care. In the European earwig (*Forficula auricularia*), the quantity of pre-hatching care has consequences **on the egg size of offspring** number, with a positive correlation only in clutches, getting high ranges of pre-hatching care.

Parent-Offspring Conflict and Sibling-Sibling Conflict (Sibling Rivalry): Another important aspect of insect reproduction is the parent-offspring conflict, guided by genetic relatedness and maximum investment approaches. Insects usually engage in sibling-sibling conflict, competing for sometimes scarce resources, for example, parental care, space, and food, which affects reproductive success.

Courtship Behavior: Insects have gone through a series of courtship behaviors to attract and mate with their partners. The obvious example occurs when the insect employs the act of calling and courtship songs; this strategy is used by fruit flies, crickets, and mosquitoes to lure and persuade their mates to mate with them. A different behavior exhibited is dancing and foreplay, which is observed in male flies, dung beetles, springtails, and apterygotes. Nuptial gifts are another strategy used by some male insects, where food or indigestible tokens are handed over to females during courtship, as observed in hangingflies, katydids, and balloon flies. Aphrodisiac strategies are also employed by some insects; a typical example is the dust produced by male butterflies. Lastly, visual signals are also utilized by some insects to attract their mates or partners, as seen in fireflies and butterfly species. By employing these different plans and strategies, insects can discover and copulate with their partners, ensuring the continuation of their species to avoid extinction [55].

6.0 THE COMPLEXITY OF INSECT SEMINAL FLUID

Female insects are modified internally through mating. These female insects are changed in behaviors, physiologically and even anatomically. These mated female insects exhibit a complete variation dramatically from their virgin self. These variances are caused by proteins and other molecules transmitted in the male's ejaculate. The ejaculate is comprised of sperm and seminal fluid, the term proteins surrounding a rich diversity of proteins, lipids, nucleic acids, carbohydrates, hormones, mucus, vitamins, water, vesicles, microbes, and in some species, glandular cells [2][14][36][35]. In several insects, the protein constituent of the ejaculate, acquired mainly from the male accessory glands, is especially varied. Recently identified and analyzed in the accessory gland in the malaria vector is amounted to 121 [64][14][6], while almost 100 seminal fluid proteins (SFPs) are transferred to females in the process of mating in a different mosquito, that is the yellow fever vector *Aedes aegypti* [71]. In insects such as *Drosophila*, seminal fluid proteins' amino acid variation is further added by a diversity of

post-translational modifications, for example, glycosylation, which may widen their effect on their roles [28]. Insect ejaculates are normally classified into two main types [80]. Firstly, in *Drosophila melanogaster*, the male ejaculate sperm may be free swimming within the seminal fluid system. On the other hand, the male ejaculate may be transmitted as a spermatophore, a classical example is the springtails and butterflies where all the non-sperm and sperm are wrapped within a proteinaceous capsule [80].

7.0 GENERAL FUNCTIONS OF MALE INSECT REPRODUCTIVE GLAND PRODUCTS

Seminal fluid constituents have been involved in the induction of different post-mating phenotypes of females [49][35][58][60][2]. The seminal fluid proteins have been reported to be very important in steering these changes, a typical example is *Drosophila melanogaster*. The performance of SFPs in female insects can be grouped into those that have effects on behavior, physiology, and anatomy. Other seminal fluid proteins have a crucial role within the male insect, by processing other seminal proteins as they go through the male's reproductive tract to the female [43][41][42]. Some other functions include the promotion of activities of the sperm in the female reproductive tract, an example is the successful release of sperm from storage [2].

8.0 INSECT PEST GENOMES ON DIAMONDBACK MOTH

Agricultural pest management is an important interest in entomology, but genome sequencing of these important pests has fallen behind that of other insects. The 'i5k' ambition was launched by Robinson and colleagues in 2011, with the objective to sequence over 5000 insects and arthropods with desirable biological research significance before 2017. The aims and objectives of the 'i5k' ambition are still not realized because of two main challenges: problems in assembling genomes of insects and financial constraints [63]. National Center for Biotechnology Information (NCBI) has registered 1,219 insect genome-sequencing programs. Currently, complete genomes of only 28 species of agricultural pests have been sequenced but part of these genomes have low scaffold N50 values, low assembly quality, and low gene integrity. These problems may be due to the high heterozygosity of the majority of insect

pests. Diamondback moth (*Plutellaxylostella*) is a dominant pest, attacking and causing severe damage to cruciferous crops, and rapidly developing unimaginable resistance to pesticides. The genome of the diamondback moth is only 343 Mb, but its high degree of heterozygosity brings about considerable challenges in genome assembly. The diamondback moth genome was sequenced in 2013 and reported to be the first insect genome to be sequenced with a high degree of heterozygosity. In the process of sequencing the diamondback moth genome, You et al. [80] adopted the fosmid-to-fosmid sequencing strategy to obtain 1,819 scaffolds. The genome annotation found 18,071 genes and 781 noncoding RNAs, and the N50 of this genome assembly is 737 kb. The insect has 1,412 specific genes, and the amplification of gene families involved in detoxification metabolism and participating in protection was done.

9.0 GENES ASSOCIATED WITH INSECT PRODUCTION

9.1 Oviposition-Related Genes

Oviposition-related genes have consistently been a key focus in insect molecular biology. Research over time has enhanced our understanding of these genes, including those associated with oviposition glands, oogenesis, oviposition site selection, and ovulation and hatching. Female insects, especially certain pest species, have an incredibly high reproductive capacity, laying hundreds to thousands of eggs each. The fecundity of these females and the role of oviposition-related genes are crucial for sustaining insect populations [47].

Insect oviposition is governed by a genetic pathway consisting of various oviposition-related genes, which are mainly categorized into four groups: genes controlling oviposition glands, genes related to the egg surface, genes involved in oviposition site selection, and genes associated with ovulation and hatching. These genes can directly or indirectly influence insect oviposition [18][31][57]. Modern techniques such as RNA interference (RNAi) and CRISPR/Cas9 have been employed to study these genes in detail. The combination of RNAi with other methods to control plant diseases and pests has begun to see some commercial applications [40][81][13].

9.2 Oviposition-Related Gland Genes

The physiological and biochemical activities in female insects, including those related to oviposition, are controlled by specific genes. For instance, the vitellogenin gene is highly expressed in the ovaries and salivary glands of insects [54]. During the transition from larva or nymph to adult female, various physiological and biochemical processes support the development of organs involved in oviposition. In insects, especially in species like *Apis mellifera*, the vitellogenin gene is crucial for regulating immunity, stress tolerance, lifespan, feeding, and oviposition behaviors. Interfering with the expression of this gene can lead to lipid loss in females and lipid accumulation in ovaries, impacting ovary maturity [25][54].

9.3 Oogenesis-Related Genes

During ovarian development, oocytes continuously acquire nutrients to form ova. Previous research has mapped the dynamic gene expression landscape during oogenesis, identifying 1,932 genes that are differentially expressed at various stages, particularly from late vitellogenesis to early choriogenesis. Vitellogenesis and choriogenesis are key processes in oocyte development and maturation, regulated by factors involved in hormonal control of oogenesis and transgenerational hormonal effects. The vitellogenin gene, for example, plays a significant role in determining the nutritional status of oocytes and their capacity for continuous division. During oogenesis, vitellogenin protein is synthesized in the fat body, released into the hemolymph, and incorporated into maturing oocytes through vitellogenin receptor-mediated endocytosis.

9.4 Oviposition-Site-Selection-Related Genes

To ensure the next generation of larvae or nymphs survives after the female's ovaries have fully matured, identifying oviposition sites with adequate food is crucial [67]. Females rely heavily on the sensilla on their antennae to locate these sites, as these sensilla express a variety of olfactory genes. These genes encode proteins responsible for the sense of smell, which play a vital role in female mating and selecting suitable oviposition locations [1]. Research indicates that a female's sense of smell is essential for recognizing hosts, mating, and oviposition. Key genes linked to olfactory proteins and receptors in female insects include those for odorant-degrading enzymes, odorant receptors, ionotropic receptors,

and sensory neuron membrane proteins. These genes encode the main proteins involved in chemical signaling [29][45][83].

9.5 Genes Related to Oviposition and Hatching

Oviposition, which encompasses egg laying and hatching, is a crucial phase in insect reproduction. The field of research on ovulation-associated genes is still in its infancy, particularly regarding the roles of individual genes. For instance, studies have shown that the recombinant Homeobox protein gene reduces the number of eggs laid by *Dermanyssusgallinae*. Additionally, applying recombinant Degakr protein from the Akr gene to the adult *Dermanyssusgallinae* impacts egg-laying, resulting in a 42% decrease in oviposition [58]. Numerous genes regulate the physiological and behavioral processes of females during oviposition, including ovulation and the maintenance of the internal egg environment and surface humidity [77].

DOPA decarboxylase, vital for corneous tanning, affects molting, survival, and reproduction in *R. prolixus*. Stern et al. [74] discovered that knocking out the Aromatic amino acid decarboxylase 2 (AADC2) gene led to delayed ovulation, lower hatchability, and higher mortality in hatched nymphs. Current research focuses on genes related to environmental stability and hatching. For example, RNAi-mediated knockout of the Transformer-2 homologue gene reduced reproduction in *Diaphorinacitri*, with silenced females laying fewer eggs and a 29.86% reduction in hatching rates compared to controls [81]. Another study identified that the myxoid protein is essential for oviposition in *Nilaparvatalugens*, with Mucin (Muc) genes encoding proteins necessary for normal eggshell formation and oviposition.

10.0 CHALLENGES IN INSECT REPRODUCTION

Predation Risk: Because predators serve as crucial regulators of prey populations, predator threat is one of the most significant processes in ecosystem operation [21]. This selection pressure can influence demographic features, geographical distribution, population structure, and the evolution of physical characteristics, life history traits, and behavioral tactics in both prey and predators [20]. Attack rates provide the most precise assessment of the influence of predation on prey populations; nevertheless, in natural systems, they are notoriously difficult to quantify due to the infrequent nature of predation events

and predators' fast consumption or disposal of prey carcasses. Seasonality and the complexity of the habitat's structural structure are two factors that can affect the risk of predation [69].

Competition for Mates: Darwin [56] described sexual selection as the differential reproductive success resulting from **competition for mates**. Instead of mating with multiple females, males might find it more advantageous to fertilize as many eggs as possible. Parker [56] **was the pioneer in** recognizing that male competition could extend within the female, a phenomenon he termed "sperm competition." Parker defined sperm competition as the rivalry between the sperm of multiple males within a single female for egg fertilization. This competition extends sexual selection beyond mating, persisting until conception. There are two types of post-mating sexual selection: male competition (sperm competition) and female choice (cryptic female choice) [72][74].

11.0 SIGNIFICANCE OF INSECT REPRODUCTIVE STUDIES

Insect reproductive studies are significant for several reasons:

- a. **Understanding Life Cycles:** Insect reproductive research helps us understand the life cycles of various insect species. This information is essential for understanding their population dynamics, behavior, and ecology.
- b. **Pest Management:** Insect reproductive studies provide information on pest insects' reproductive practices. This knowledge can be utilized to build efficient pest management techniques, such as controlling insects at various periods of their life cycle.
- c. **Evolutionary Biology:** Studying insect reproduction helps us understand evolutionary processes. Insects use a variety of reproductive techniques, such as sexual and asexual reproduction, multiple mating, and intricate courtship activities. Scientists can learn more about evolutionary mechanisms by researching these strategies.
- d. **Conservation:** Insect reproductive studies are essential **for the conservation of endangered insects. Understanding their reproductive biology aids in the development of conservation strategies aimed at preserving their breeding sites and guaranteeing successful reproduction.**

e. **Agricultural Productivity:** Insects **play an important part** in pollination, which is required for agricultural productivity. Insect reproduction research contributes to a better knowledge of their role as pollinators and how to improve their reproductive behavior for agricultural production.

In conclusion, insect reproductive research has important implications for many domains, including ecology, pest management, evolutionary biology, conservation, and agriculture.

12.0 DISCUSSIONS AND CONCLUSIONS

Reproduction is the basic feature of all known life of organisms and the natural process by which offspring are produced from their parents. Insects reproduce both sexually and asexually, enabling their rapid population growth. Reproductive ecology is concerned with the organism's physiology, habits, and the abiotic and biotic settings in which it lives to determine how it reproduces. Recent research into oviposition-related genes in insects has improved significantly. However, the complexities of ovarian growth, the physiological processes of oviposition, and the involvement of antennae in selecting oviposition sites during ovulation and egg hatching are more complex than previously imagined. **It is evident that** no single gene can fully regulate the oviposition behavior of females; instead, a network of genes works in concert to govern this behavior comprehensively. **When examining insect reproductive strategies, several key examples include parent-offspring conflict, sibling-sibling conflict, egg size, semelparity, iteroparity, drifting behavior, courtship behavior, and reproductive behavior.** These strategies provide insights into how insects achieve successful reproduction within ecosystems. Regarding the complexity of insect seminal fluid, mating induces internal modifications in female insects, altering their behavior, physiology, and anatomy. These changes result in a dramatic transformation from their virgin state, driven by proteins and other molecules in the male's ejaculate. The products of male insect reproductive glands, particularly seminal fluid constituents, play a role in inducing various post-mating phenotypes in females. The effects of seminal fluid proteins on female insects can be categorized into changes in behavior, physiology, and anatomy.

Insect reproduction can be classified into seven main types: oviparity, viviparity, parthenogenesis, polyembryony, hermaphroditism, castration, and alternation of generation. Genome sequencing of

significant pests has lagged compared to other insects. To date, the complete genomes of only 28 agricultural pest species have been sequenced, with many exhibiting low scaffold N50 values, poor assembly quality, and low gene integrity, likely due to high heterozygosity in most insect pests. Research in insect molecular biology has consistently focused on genes associated with oviposition. Advances have been made in understanding the mechanisms of oviposition-related genes, including those related to the oviposition gland, oogenesis, oviposition-site selection, and genes involved in ovulation and hatching.

13.0 FUTURE RESEARCH DIRECTIONS

With **an adequate** understanding of insect biology and reproductive system, newly discovered disciplines of study within entomology are emerging. A typical example is insect neurobiology, which involves understanding the neural techniques that emphasize insect behavior. This education can provide insights into basic neurological functions and can be applied to robotics and artificial intelligence. Secondly, another interesting **important field of study is insect microbiomes, which comprise microbial communities that live in symbiosis with insects [15]**. These microbes **play an important role** in insect physiology, digestion, immunity, and reproduction. Understanding these **associated** links can lead to innovative pest management tactics and a better understanding of basic biological processes. Another developing area, environmental entomology, which studies the connections between insects and their habitats, is gaining traction in the face of climate change and habitat loss. The goal of this study is to predict how environmental changes would affect insect populations and to identify potential conservation strategies.

REFERENCES

1. Alonso, D. P., M. Campos, H. Troca, R. Kunii, F. Tripet, and P. E. M. Ribolla. (2019). Gene expression profile of *Aedes aegypti* females in courtship and mating. *Sci. Rep.* 9: 15492.
2. Avila, F. W., Sirot, L. K., LaFlamme, B. A., Rubinstein, C. D., Wolfner, M. F., Avila, F. W., Wolfner, M. F., & LaFlamme, B. A. Insect Seminal Fluid Proteins: Identification and Function. *Annual Review of Entomology.* 2011. 56(1), 21–40.
3. Avila, FW., Sirot, LK., LaFlamme, BA., Rubinstein, CD., Wolfner, MF., Avila, F. W., Wolfner, M. F., & LaFlamme, B. A. (2011). Insect Seminal Fluid Proteins: Identification and Function. *Annual Review of Entomology*, 56(1), 21–40. 2011. <https://doi.org/10.1146/annurev-ento-120709-144823>.
4. Bai, H., and Palli, SR. Identification of G protein-coupled receptors required for vitellogenin uptake into the oocytes of the red flour beetle, *Tribolium castaneum*. *Sci. Rep.* 2016. 6:27648. doi: 10.1038/srep27648.
5. Bai, H., Qiao, H., Li, F., Fu, H., Sun, S., Zhang, W., Et al. Molecular characterization and developmental expression of vitellogenin in the oriental river prawn *Macrobrachium nipponense* and the effects of RNA interference and eyestalk ablation on ovarian maturation. 2015. *Gene* 562, 22–31. doi: 10.1016/j.gene.2014.12.008
6. Baldini, F., Gabrieli, P., Rogers, D. W., & Catteruccia, F. Function and composition of male accessory gland secretions in *Anopheles gambiae*: a comparison with other insect vectors of infectious diseases. *Pathogens and Global Health.* 2012. 106(2), 82–93. <https://doi.org/10.1179/2047773212Y.0000000016>.
7. Beck, H., Übereinen weiteren Fall von lateralem Hermaphroditismus beider, Ameise *Polyergus rufescens* Latr. *Zool Anz.* 1958. 161, 243–249.
8. Châline, N., Blacher, P., Yagound, B., Lecoutey, E., Devienne, P., and Chaméron, S. Drifting behavior as an alternative reproductive strategy. 2013.
9. Chapman, R. F. (2013). *The Insects: Structure and Function.* Cambridge University Press.

10. Constantini, D., Bruner, E., Fanfani, A. & Dell 'Omo, (2007). G. Male-biased predation of western green lizards by Eurasian kestrels. *Naturwissenschaften* 94. 2007 1015–1020. <https://doi.org/10.1007/s00114-007-0284-5>.
11. Corona, M., Libbrecht, R., Wurm, Y., Riba-Grognuz, O., Studer, R. A., and Keller. Vitellogenin underwent sub functionalization to acquire caste and behavioral specific expression in the harvester ant *Pogonomyrmex barbatus*. 2013. *PLoS Genet.* 9: e1003730. doi: 10.1371/journal.pgen.1003730.
12. Corona, M., Velarde, R. A., Remolina, S., Moran-Lauter, A., Wang, Y., Hughes, K. A., Et al. Vitellogenin, juvenile hormone, insulin signaling, and queen honey bee longevity. *Proc. Natl. Acad. Sci. U. S. A.* 2007. 104, 7128–7133. doi: 10.1073/pnas.0701909104.
13. Dao, YX., and NK. Illiny. Effect of parental RNA interference of a transformer-2 homologue on female reproduction and offspring sex determination in Asian citrus psyllid. *Physiol. Entomol.* 2018. 43: 42–50.
14. Dottorini, T., Nicolaides, L., Ranson, H., Rogers, D. W., Crisanti, A., & Catteruccia, F. (2007). A genome-wide analysis in *Anopheles gambiae* mosquitoes reveals 46 male accessory gland genes, possible modulators of female behavior. *Proceedings of the National Academy of Sciences of the United States of America.* 2007. 104(41), 16215–16220. 724 <https://doi.org/10.1073/pnas.0703904104>.
15. Douglas AE. *Insects and their beneficial microbes.* Princeton University Press.
16. Droney DC., The influence of the nutritional content of the adult male diet on testis mass, body condition, and courtship vigour in a Hawaiian *Drosophila*. *Functional Ecology.* 1998. 12:920-928.
17. Du Q, Wen L, Zheng SC, Bi HL, Huang YP, Feng QL, Liu L. Identification and functional characterization of *double sex* gene in the testis of (*Spodoptera litura*). *Insect Science.* 2019. 26:1000-1010.
18. Du, L., M. M. Wang, J. L. Li, S. Y. He, J. X. Huang, and J. Wu. Characterization of a vitellogenin receptor in the bumblebee, *Bombus anthogenesis* (Hymenoptera, Apidae). *Insects.* 2019. 10: 1–14

19. Dumser JB., The regulation of spermatogenesis in insects. *Annual Review of Entomology*.1980. 25:341-369.
20. Eberhard, W. G. *Female control: sexual selection by cryptic female choice*. — Princeton University Press, Princeton. 1996. 501 pp.
21. Endler, JA. Defense against predators in Predator-prey relationships, Feder, M. E. & Lauder, GV. (Eds). 1986. (Texas University of Chicago Press, 1986). 7.
22. Fritz, R. S., Stamp, N. E., and Halverson, T. G. Iteroparity and semelparity in insects. *The American Naturalist*, .1982. 120(2), 264-268.
23. Garcia, J., Munro, ES., Monte, MM., Fourrier, MC., Whitelaw, J., Smail, DA., Et al... Atlantic salmon (*Salmo salar* L.) serum vitellogenesis neutralizes infectivity of infectious pancreatic necrosis virus (IPNV). *Fish Shellfish Immunol*. 2010. 29, 293–297. doi: 10.1016/j.fsi.2010.04.010.
24. Genome 10K Community of Scientists. Genome 10K: a proposal to obtain whole-genome sequence for 10,000 vertebrate species. *The Journal of Heredity*. 2009. 100, 659–674.
25. Ghosh, SKB., W. B. Hunter, AL. Park, and DE. Gundersen-Rindal. Double-stranded RNA oral delivery methods to induce RNA interference in phloem and plant-sap-feeding Hemipteran insects. *Jove-J. Vis. Exp*. 2018. 135: 57390.
26. Gilbert, LI., Serafin, RB., Watkins, NL., and Richard, DS. Ecdysteroids regulate yolk protein uptake by *Drosophila melanogaster* oocytes. *J. Insect Physiol*.1998. 44, 637–644. doi: 10.1016/S0022-1910(98)00020-1
27. Gillott, C. (2003). Male accessory gland secretions: modulators of female reproductive physiology and behavior. *Annual Review of Entomology*, 48(1), 163–184. <https://doi.org/10.1146/annurev.ento.48.091801.112657>.
28. Gligorov, D., Sitnik, JL., Maeda, RK., Wolfner, MF., & Karch, FA. Novel Function for the Hox Gene *Abd-B* in the Male Accessory Gland Regulates the Long-Term Female Post-Mating Response in *Drosophila*. *Plos Genetics*,9(3). 2013.e1003395. <https://doi.org/10.1371/journal.pgen.1003395>.
29. González, A. G., M. E. R. Meléndez, G. I. Ballesteros, C. C. Ramírez, and R. P. Millanao.

Sex-and tissue-specific expression of odorant- binding proteins and chemosensory proteins in adults of the scarab beetle *Hylamorpha elegans* (Burmeister) (Coleoptera: Scarabaeidae). Peer J. 2019. 14: 1–21.

30. Gullan, P. J., & Cranston, P. S. (2014). *The Insects: An Outline of Entomology*. Wiley-Blackwell.

31. Guo, W., J. Lü, M. J. Guo, S. M. Chen, B. L. Qiu, W. Sang, C. X. Yang, Y. J. Zhang, and H. P. Pan. De novo transcriptome analysis reveals abundant gonad-specific genes in the ovary and testis of (*Henosepilachnavigintioctopunctata*.) 2019. Int. J. Mol. Sci. 20: 1–17.

32. H.M. Parker., The relationship of parthenogenesis in virgin Chinese painted quail (*Coturnix chinensis*) hens with embryonic mortality and hatchability following mating Poult Sci.2012.

33. Hagan, H. 1948. A Brief Analysis of Viviparity in Insects. Journal of the New York Entomological Society.2022. 56(1): 63-68.

34. Hagan, H. Embryology of the viviparous insects. New York, Ronald Press. Zilberman, B. et al.: Viviparity in Staphylinidae and reproductive behavior of *Corotoca* Pap. Avulses Zool., v.59: e20195919 9/10 Hölldobler, B.; Kwapich, C.L. & Haight, K.L. 2018. Behavior and exocrine glands in the myrmecophilous beetle *Lomechusoidesstrumosus* (Fabricius, 1775) (formerly called *Lomechusastrumosa*) (Coleoptera: Staphylinidae: Aleocharinae). 2019. 1954. Plos One, 13(7): e0200309.

35. Hopkins, B. R., Sepil, I., &Wigby, S. Seminal fluid. Current Biology,27(11), R404–82 R405. 2017. <https://doi.org/10.1016/j.cub.2017.03.063>

36. Hopkins, B. R., Sepil, I., &Wigby, S. Seminal fluid. Current Biology,27(11), R404824R405<https://doi.org/10.1016/j.cub.2017.03.06>.

37. Huo, Y., Yu, Y., Chen, L., Li, Q., Zhang, M., Song, Z., Et al. Insect tissue-specific vitellogenin facilitates transmission of plant virus. PlosPathog. 2018. 14: e1006909. doi: 10.1371/journal.ppat.1006909.

38. Ihle, KE., Fondrk, MK., Page, RE., and Amdam, GV. Genotype effect on lifespan following vitellogenin knockdown. 2015. Exp.Gerontol.61,113–122. doi: 10.1016/j.exger.2014.12.007

39. Joly D. Bressac C., Sperm length in Drosophilidae (Diptera): Estimation by testis and receptacle lengths. *International Journal of Insect Morphology & Embryology*. 1994. 23:85-92.
40. Killiny, N., and A. Kishk. 2017. Delivery of dsRNA through topical feeding for RNA interference in the citrus sap piercing-sucking hemipteran, *Diaphorinacitri*. *Insect Biochem. Phys.* 2017. 95: 1–13.
41. Laflamme, B. A., & Wolfner, M. F. Identification and function of proteolysis regulators in seminal fluid. *Molecular Reproduction and Development*. 2013. 80(2), 80–101. <https://doi.org/10.1002/mrd.22130>.
42. LaFlamme, B. A., Ravi Ram, K., & Wolfner, M. F. The *Drosophila melanogaster* seminal fluid protease “Seminase” regulates proteolytic and post-mating reproductive processes. *PLoS Genetics*. 2012. 8(1), 30–32. <https://doi.org/10.1371/journal.pgen.1002435>.
43. LaFlamme, BA., Avila, FW., Michalski, K., & Wolfner, MFA. *Drosophila* protease cascade member, seminal Metalloprotease-1, is activated stepwise by male factors and requires female factors for full activity. *Genetics*,196(4),11171129. 2014. <https://doi.org/10.1534/genetics.113.160101>.
44. Laranjo LT, Haifig I, Costa Leonardo AM. Morphology of the male reproductive system during postembryonic development of the termite (*Silvestri Termes euamignathus*) (Isoptera: Termitidae). *Zoologist Anzeiger*. 2018. 272:20-28.
45. Li, H., Gu. TZ, Chen, CY., Huang, KR, Chen. RX and Hao.DJ. Identification and expression patterns of chemosensory proteins in the black-back prominent moth, (*Closterarestitura*)(Lepidoptera: Notodontidae). *Eur. J. Entomol*. 2019.116: 372–391.
46. Linklater JR, Wertheim B, Wigby S, Chapman T., Ejaculate depletion patterns evolve in response to experimental manipulation of sex ratio in *Drosophila melanogaster*. *Evolution*. 2007. 61:2027-2034.
47. Lu, W., Q. Wang, M. Y. Tian, J. Xu, J. Lv, S. G. Wei, and A. Z. Qin. Reproductive traits of *Glenea cantor* (Coleoptera: Cerambycid: Lamiinae). *J. Econ. Entomol*. 2013. 106: 215–220.
48. Matsumoto, T., Yamano, K., Kitamura, M., and Hara, A. Ovarian follicle cells are the site of

- vitellogenin synthesis in the Pacific abalone *Haliotis discus hannai*. *Comp. Biochem. Physiol. Part A: Mol. Integr. Physiol.* 2008. 149, 293–298. doi: 10.1016/j.cbpa.2008.01.003.
49. McGraw, LA., Suarez, SS., & Wolfner, MF. On a matter of seminal importance. *Bio Essays*.2015. 37(2), 142–147. <https://doi.org/10.1002/bies.201400117>.
50. Meier R, Kotrba M, Ferrar P. Ovoviviparity and viviparity in the Diptera. *Biol. Rev. Camb. Philos.Soc.* 1999; 74:199–258. Reviews fly reproductive mechanisms.
51. Melo, AC., Valle, D., Machado, EA., Salerno, AP., Paiva-Silva, GO., Cunha, ES., et al. (2000). Synthesis of vitellogenin by the follicle cells of (*Rhodniusprolixus*). *Insect Biochem. Mol. Biol.* 30, 549–557. 2000. doi: 10.1016/S0965-1748(00)00023-0.
52. Meyer JR. Survival Strategies. *General Entomology*. [Internet] 2014 [Accessed 17 December). Available <https://www.cals.ncsu.edu/course/ent425/library/tutorials/ecology/survival.html>. genome provides insights into herbivory and detoxification. *Nature Genetics*.2015. 45, 220–225.
53. N.C. State General Entomology. [https://genent.cals.ncsu.edu/bug bytes/reproductive system/females.2015](https://genent.cals.ncsu.edu/bug%20bytes/reproductive%20system/females.2015).
54. Nunes,FM.,K.E.Ihle,NS.Mutti,ZL.Simões,andGV.Amdam.The gene *vitellogenin* affects microRNA regulation in honey bee (*Apis mellifera*) fat body and brain. *J. Exp. Biol.* 2013. 216: 3724–3732.
55. O'woma, O. O., Chigozirim, U. P., Emmanuel, O., and Chukwuebuka, E. M. Reproductive and survival strategies utilized by insect. A review. *American Journal of Zoological Research*. 2016. 4(1), 1-6.
56. Parker, GA. Sperm competition and its evolutionary consequences in the insects. *Biol. Rev. Camb. Philos. Soc.*1970. 45: 525–567.
57. Peng, L., Q. Wang, M. M. Zou, Y. D. Qin, L. Vasseur, L. N. Chu, Y. L. Zhai, S. J. Dong, L. L. Liu, W. Y. Hn , et al. 2020. CRISPR/Cas9-mediated vitellogenin receptor knockout leads to functional deficiency in the reproductive development of (*Plutellaxylostella*). *Front Physiol.*2020. 10: 1–13.

58. Perry, J., Sirot, L., & Wigby, S. The seminal symphony: how to compose an ejaculate. *Trends in Ecology Evolution*. 2013.28(7),414–422.
59. Pianka, ER. Niche relations of desert lizards in *Ecology and Evolution of Communities*, Cody, M. L. & Diamond, J. M. (Eds).1975. (Harvard University Press).
60. Poiani, A. Complexity of seminal fluid: A. <https://doi.org/10.1016/j.tree.2013.03.005>. review. *Behavioral Ecology and Sociobiology*, 60(3), 289310. 2006.<https://doi.org/10.1007/s00265-006-0178-0>.
61. Resh VH, Ring TC. *Encyclopedia of Insects* 2nd edition. U.S.A. 2009. Academic Press.
62. Ridley M. *Evolution*, 3rd edition. Blackwell Publishing. 2004. p. 314.
63. Robinson, G.E., Hackett, K.J., Purcell-Miramontes, M., Brown, S.J., Evans, J.D., Goldsmith, M.R. et al. (2011) Creating a buzz about insect genomes. *Science*, 331, 1386.
64. Rogers, D. W., Baldini, F., Battaglia, F., Panico, M., Dell, A., Morris, H R., & Catteruccia, 1047F. Transglutaminase-Mediated Semen Coagulation Controls Sperm Storage in 1048 the Malaria Mosquito. *PLoS Biology*. 2009. 7(12), e1000272. 1049 <https://doi.org/10.1371/journal.pbio.1000272>.
65. Salmela, H., Amdam, GV., and Freitak, D. Transfer of immunity from mother to offspring is mediated via egg-yolk protein vitellogenin. *PLoS Pathog*. 2015. 11: e1005015. doi: 10.1371/journal.ppat.1005015.
66. Seehuus, S. C., Norberg, K., Gimsa, U., Krekling, T., and Amdam, G. V. Reproductive protein protects functionally sterile honey bee workers from oxidative stress. *Proc. Natl. Acad. Sci. U. S. A*. 2006. 103, 962–967. doi: 10.1073/pnas.0502681103.
67. Sétamou, M., J. V. Graça, and J. L. Sandoval. 2016. Suitability of native North American Rosaceae to serve as host plants for the Asian citrus psyllid (Hemiptera: Liviidae). *J. Appl. Entomol.* 140: 645–654.
68. Shashank, DU., Keerthana, M., Singh, MK. and Prasad, S. Reproductive Strategies in Insects: Unravelling the Intricate Reproductive Tactics of Insects. *Vigyan Varta*. 2023. 4(6): 183-186.

69. Shepard, DB. Habitat but not body shape affects predator attack frequency on lizard models in the Brazilian Cerrado. *Herpetologica* 63, 193–202. [https://doi.org/10.1655/0018-0831\(2007\)63\[193:HBNBSA\]2.0.CO;2](https://doi.org/10.1655/0018-0831(2007)63[193:HBNBSA]2.0.CO;2).
70. Sicart, M., Larrouy, G., Hermaphroditism partiel chez un *Culex hortensis*. Bull. soc. hist. nat. Toulouse 97. 1962. 410–412.
71. Sirot, LK., Wolfner, MF., & Wigby, S. Protein-specific manipulation of ejaculate composition in response to female mating status in *Drosophila melanogaster*. Proceedings of the National Academy of Sciences. 2011. 108(24), 9922–9926. <https://doi.org/10.1073/pnas.1100905108>.
72. Smith, RL. (ed). Sperm competition and the evolution of animal mating systems. Academic Press, London. 1984. 687 pp.
73. Snodgrass, R. E. (1935). Principles of Insect Morphology. McGraw-Hill Book Company, Inc.
74. Thornhill, R. Cryptic female choice and its implications in the scorpion fly (*Harpobittacus nigricans*) Am. Nat. 1983. 122: 765–788.
75. Tisserat, B., E.B. Esan and T. Morishige. Somatic embryogenesis in angiosperm. Horticultural Review. 1978. 1: 1–78.
76. Wang, Y., Baker, N., and Amdam, GV. RNAi-mediated double gene knockdown and gustatory perception measurement in honey bees (*Apis mellifera*). J. Vis. Exp. 2013 77:50446. doi: 10.3791/50446
77. Wang, Y., Brent, CS., Fennern, E., and Amdam, G V. Gustatory perception and fat body energy metabolism are jointly affected by vitellogenin and juvenile hormone in honey bees. 2012. PLoS Genet. 8: e1002779. doi: 10.1371/journal.pgen.1002779.
78. Ward PI, Simmons LW., Copula duration and testes size in the yellow dung fly, *Scathophagastercoraria*(L.): the effects of diet, body size, and mating history. Behavioral Ecology and Sociobiology. 1991. 29:77-85.
79. Wedell, N., and Karlsson, B. Paternal investment directly affects female reproductive effort in an insect. Proceedings of the Royal Society of London. Series B: Biological Sciences. 2003. 270(1528), 2065-2071.

80. You, M., Yue, Z., He, W., Yang, X., Yang, G., Xie, M. et al. A heterozygous moth.2013.
81. Yu, XS. Gowda, and N. Killiny. Double-stranded RNA delivery through soaking mediates silencing of the muscle protein 20 and increases mortality to the Asian citrus psyllid, *Diaphorinacitri*. *Pest Manag. Sci.* 2017. 73: 1846–1853.
82. Zhang, S., Dong, Y., and Cui, P. Vitellogenin is an immunocompetent molecule for mother and offspring in fish. *Fish Shellfish Immunol.*2015. 46, 710–715.doi:10.1016/j.fsi.2015.08.011.
83. Zhang, X., S. Yang, Zhang, X. Wang, S. Wang, M. Liu, and J. Xi. Identification and expression analysis of candidate chemosensory receptors based on the antennal transcriptome of *Lissorhoptrusoryzophilus*. *Comp. Biochem. Physiol. Part D. Genomics Proteomics.* 2019. 0: 133–142.