

Antibacterial activity and biotechnological application of Endophytic Bacteria isolated from seaweed

ABSTRACT:

Seaweeds are multicellular, photosynthetic organisms. Seaweeds are a type of algae that mainly grows in the ocean, which is also known as sea vegetable. Seaweeds come in various colors like green, red, brown, and black, and they are known for their high nutritional value. Many seaweed species are edible and offer a range of culinary possibilities. Some even have commercial importance, used in various products like sushi, nori sheets, and even cosmetics and fertilizers. Seaweeds are often considered superfoods due to their high nutritional value, rich in vitamins and minerals like iodine, calcium, magnesium, and iron. Endophytic bacteria are significant contributors to the plant microbiome and are essential for the survival, development, and growth of the plant. Hormones, nutrients, secondary metabolites, and other tiny molecules are among the substances secreted by endophytic bacteria that have a substantial impact on plant growth and development. Auxins, cytokinin, and gibberellins, for instance, are phytohormones produced by some endophytic bacteria that can encourage cell division and elongation, resulting in greater plant development. Polyhydroxybutyrate (PHB) is a biodegradable and bio-based thermoplastic polyester. PHB is synthesized as an intracellular storage compound by various microorganisms, including certain bacteria like *Cupriavidus necator*, *Ralstonia eutropha*, *Bacillus megaterium*, and some archaea. It is stored in the form of dense granules within the cells. Proteases play a vital role in catalyzing the breakdown of peptide bonds in proteins. This enzymatic function is crucial not only for digestion in the human body but also in biotechnological and industrial applications. In industries like food processing, pharmaceuticals, and laundry detergents, protease activity is harnessed to break down proteins.

KEYWORDS: Polyhydroxybutyrate (PHB), Seaweed, secondary metabolites, thermoplastic polyester

INTRODUCTIONS

Seaweeds, also known as sea vegetables, are indeed fascinating marine plants. They thrive in shallow waters and along rocky shorelines, forming distinct zones based on water depth. Kelp is a notable example, often found in waters up to 50 meters deep. These plants serve as a crucial food source for marine life. Seaweeds anchor themselves to the seafloor through holdfasts, but unlike typical plants, they don't extract nutrients through their roots. Instead, they absorb nutrients directly from the surrounding water. Seaweeds come in various colors like green, red, brown, and black, and they are known for their high nutritional value. Many seaweed species are edible and offer a range of culinary possibilities. Some even have commercial importance, used in various products like sushi, nori sheets, and even cosmetics and fertilizers. Their versatility and nutritional benefits make them a valuable resource both in the ocean ecosystem and for human consumption.

Seaweeds are indeed fascinating and diverse organisms found in coastal areas around the world. They play a significant role in marine ecosystems and have various uses for both humans and other marine life. Red algae, scientifically known as Rhodophyta, are particularly noteworthy due to their vibrant red hue, which is the result of pigments like chlorophyll A, phycoerythrin, and phycocyanin. These red algae are not only edible and important for human consumption but also serve as a vital source of nutrition for many marine creatures. They contribute to the food web by providing sustenance to fishes and shellfish. Seaweeds, as a group of multicellular, photosynthetic organisms, are integral to marine ecosystems and have diverse applications, from food and medicine to industrial purposes. They are categorized into three main groups based on coloration: red, green, and brown seaweeds. Each group has its unique characteristics and uses. Seaweeds are often considered superfoods due to their high nutritional value, rich in vitamins and minerals like iodine, calcium, magnesium, and iron.

Endophytic bacteria are fascinating microorganisms that have a significant impact on plant growth and health. They secrete a wide range of substances, including hormones, nutrients, and secondary

metabolites, which play important roles in development of plant. Phytohormones like auxins, cytokines, and gibberellins produced by certain endophytic bacteria can stimulate cell division and elongation in plants, leading to enhanced growth. Nitrogen fixation is another crucial service provided by some endophytic bacteria, making nitrogen more readily available to the host plant, which is essential for its growth and development.

The research into the antibacterial activity of endophytic microorganisms has indeed advanced significantly in recent years, and various methods have been employed to study their potential in combating bacterial infections. Here's a brief overview of the methods mentioned:

Agar Diffusion Assay: This method involves embedding the test bacteria in an agar medium and then applying an endophytic bacterial extract or a purified compound onto the medium. The appearance of a clear zone around the application site indicates the presence of antibacterial compounds, as the compounds inhibit bacterial growth.

Minimum Inhibitory Concentration (MIC) Assay: In this assay, the target bacteria are exposed to a range of concentrations of the endophytic bacterial extract or purified compounds. The goal is to determine the lowest concentration that effectively inhibits bacterial growth. This provides a quantitative measure of the antibacterial potency of the tested substances.

Time-Kill Assay: This assay monitors the growth of the test bacteria over time in the presence of the endophytic bacterial extract or compounds. It helps in understanding the kinetics of antibacterial activity, including the rate and extent of bacterial inhibition.

Polyhydroxybutyrate (PHB) is indeed a fascinating biodegradable thermoplastic polyester with a range of promising applications. Here are some key points about PHB:

Microbial Synthesis: PHB is synthesized as an intracellular storage compound by various microorganisms, including certain bacteria like *Cupriavidus necator*, *Ralstonia eutropha*, *Bacillus megaterium*, and some archaea. It is stored in the form of dense granules within the cells.

Biodegradability: One of the primary advantages of PHB is its biodegradability. Unlike traditional petroleum-based plastics, PHB can be broken down by natural processes, making it a more environmentally friendly option.

Biocompatibility: PHB is biocompatible, meaning it is well-tolerated by living organisms. This property makes it suitable for various applications in medicine and biomedicine.

PHB finds applications in multiple fields such as Agriculture, medicine and biomedical.

Proteases play a vital role in catalyzing the breakdown of peptide bonds in proteins, leading to the formation of smaller peptide chains or amino acids. This enzymatic function is crucial not only for

digestion in the human body but also in biotechnological and industrial applications. In industries like food processing, pharmaceuticals, and laundry detergents, protease activity is harnessed to break down proteins. For example, in laundry detergents, protease activity is essential for effectively removing stains containing protein-based substances like blood or food. This demonstrates the versatility of proteases in improving product performance and cleanliness in various sectors.

AIM

The study of the antibacterial activity and biotechnological application of Endophytic Bacteria isolated from seaweed

OBJECTIVES

- Collect and identify seaweed specimens from the coastal region of Tamil Nadu.
- Isolate and characterize endophytic bacteria residing within the seaweed.
- Evaluate the antibacterial potential of endophytic bacteria against UTI pathogens.
- Produce and characterize Polyhydroxybutyrate (PHB) using endophytic bacteria.
- Screen endophytic bacteria for enzymatic activity.

MATERIALS AND METHODS

COLLECTION OF SAMPLES

Seaweed samples were gathered from Sathyabama University in Tamil Nadu. Initially, the seaweeds were observed and selected based on variations in their morphology and color. After selection, the collected seaweed samples were carefully placed in Polythene zip-lock sample covers. They were then promptly transported to a laboratory and rinsed gently with sterile water. To eliminate excess moisture from the seaweed samples, they were gently pressed with sterile tissue paper. Further sterilization of the seaweed samples was performed to remove external microorganisms, facilitating the isolation of endophytic bacteria.

BACTERIAL ISOLATION AND IDENTIFICATION

The process involves isolating endophytic bacteria from seaweed. The selected segment of thallus are immersed in 95% ethanol for 30 sec and 4% sodium hypochlorite solution for 3 mins. Rinse the thallus in sterile distilled water three times and dry the surface area under appropriate condition. After drying cut the thallus segment approximately of 0.5cm and place it in the petri

plate containing tryptic soy agar medium(TSA) supplemented with ketoconazole(100mg/L) to suppress the fungal growth. It includes surface sterilization of seaweed segments, followed by their placement on tryptic soy agar medium supplemented with ketoconazole to suppress fungal growth. After incubation at 37°C for 24 hours, endophytic bacterial colonies are observed and transferred to fresh TSA plates to obtain pure colonies. This method is commonly used for studying endophytic bacteria associated with seaweed.

MORPHOLOGICAL CHARACTERISTICS

The colony characteristics like size, shape, color, elevation, texture, margin, and opacity when studying bacterial isolates. All actinobacterial morphotypes were grown in LB broth and incubated at 28±2 °C at 120 rpm. Additionally, observing the growth cycle of individual isolates using a light microscope and performing Gram staining every 24 hours for up to 10 days provides valuable information about their growth patterns and Gram stain reactions. This comprehensive approach helps in characterizing and understanding the isolated endophytic bacteria.

MICROSCOPIC EXAMINATION

GRAM STAINING:

A loopful of culture was smeared on the sterile glass slide and air dried. The smear was subsequently flooded with crystal violet, left for 1 minute, washed, and then flooded with Gram's iodine for another minute. After this, decolorizing agents (90% alcohol) were introduced, followed by a thorough wash. Finally, the counterstain safranin was applied for 20 seconds, followed by washing. The slide was then examined under a microscope, and the results were recorded.

BACTERIAL CONFIRMATION TEST

VITEK® MSPRIME is a valuable tool for bacterial confirmation tests, offering automated identification with a diverse and reliable database for rapid and certain microbial identification.

ANTIBACTERIAL ANALYSIS

Preparation of broth cultures:

Two UTI pathogens (Escherichia coli, Staphylococcus aureus) were inoculated in Nutrient broth and incubated at 37°C for 24 hours, it was used in antibacterial analysis.

UNDER PEER REVIEW

38 grams of MHA (Muller Hinton Agar) was dissolved in 1000 ml of distilled water and then autoclaved for 15 minutes at 121°C. Once the medium was about 45°-50°C, it was poured into sterile petri dishes. Then it was allowed to set completely.

ANTIBACTERIAL ACTIVITY

In this experiment, you swabbed an overnight broth culture onto MHA plates, created wells, added spice extracts, and used Streptomycin as a positive control. After incubation, you measured the diameter of the zone of inhibition, and the result is 4.8 mm.

PHB PRODUCTION

Materials:

The endophytic bacterial culture was isolated and inoculated into the culture to produce the PHB.

Culture conditions:

A mineral salt medium used for PHA (Polyhydroxyalkanoate) production. The composition of the medium includes:

2.5g/L K_2HPO_4

2.5g/L KH_2PO_4

5g/L glucose (added separately)

2g/L yeast extract

0.2g/L $MgSO_4 \cdot 7H_2O$

0.01g/L $FeSO_4 \cdot 7MnSO_4$

0.007g/L $H_2O_7H_2O$

The pH of the medium was adjusted to 7.5, and sterilization of the medium without glucose was performed for 20 minutes at 121°C. Glucose was sterilized separately and then added to the medium. This process is likely for the cultivation of microorganisms to produce PHA.

Sudan Black B staining:

The Sudan Black B staining method is commonly used to detect the presence of

polyhydroxybutyrate (PHB) granules in bacterial cells. Here's a summary of the process:

Preparation of Sudan Black B Solution:

Mix 0.3 mg of solid Sudan Black B dye with 100 mL of 70% ethanol to prepare the staining solution.

Smear Preparation:

Smears of the positive bacteria are prepared and heat-fixed onto glass slides.

Staining with Sudan Black B:

The prepared smears are stained with the Sudan Black B solution for 15 minutes. This staining step helps to visualize the PHB granules.

Wash with Xylene:

After staining, the smears are washed with xylene. This step is likely performed to remove excess stain and enhance contrast.

Counter-Staining with Safranin:

The smears are counter-stained with Safranin (5% in distilled water) for 1 minute. Safranin is a red stain and is used to provide contrast to the blue-black PHB granules stained by Sudan Black B.

Microscopic Observation:

The stained smears are then observed under a microscope with immersion oil at a magnification of 100×. Cells containing blue-black cytoplasmic granules are considered PHB producers.

Preservation:

PHB-producing cells are preserved in 2% glycerol, likely for further analysis or long-term storage. Stained slides were observed to determine the cellular characteristics of the isolates at the microscope with immersion oil at 100X.

Extraction:

Bacterial Culture: Bacterial cells are grown and incubated for 48 hours.

Harvesting: After incubation, the bacterial cells are harvested by centrifugation at 5,000 rpm for 10 minutes. This separates the cells from the culture medium.

Drying: The harvested bacterial cells are dried to determine the Cell Dry Weight (CDW), which serves as an index of biomass.

PHA and Lipid Extraction: The dried cells are treated with boiling chloroform to dissolve both

UNDER PEER REVIEW

the PHA and lipids present in the cells.

Filtration: The chloroform-treated solution is then filtered using Whatman No. 1 filter paper to remove the remaining cell mass. This step isolates the dissolved PHA and lipids.

Precipitation of PHA: Methanol is added to the filtrate, which selectively precipitates the PHA from the solution.

Centrifugation: The solution containing the precipitated PHA is subjected to centrifugation at 12,000 rpm for 10 minutes. This step separates the PHA from the rest of the solution.

Drying: The isolated PHA is air-dried to remove any remaining solvent.

Weighing and Storage: The dried PHA is weighed to determine its yield and is then stored for further studies or applications.

FTIR Analysis:

FTIR (Fourier Transform Infrared) spectroscopy is a common technique used to analyze the chemical composition of substances by measuring their infrared absorption spectra

- Chloroform was used as a medium for your sample.
- The sample was applied as a smear over a NaCl (sodium chloride) block, which is often used as a sample holder for FTIR analysis.
- The PerkinElmer RX1 FTIR spectrophotometer was used for the analysis.
- The FTIR spectrum of the sample was collected in the wavelength range of 450-4000 cm^{-1} .

ENZYMATIC ACTIVITY

AMYLOLYTIC ACTIVITY

The starch hydrolysis test is a common method to assess a bacterial isolate's ability to produce amylase, an enzyme that breaks down starch. Here are the key steps of the procedure you described:

Preparation of Starch Hydrolysis Agar: Starch hydrolysis agar, which is a selective medium containing starch, is prepared and sterilized by autoclaving at 121°C and 15 lb pressure for 15 minutes to ensure it's free from contaminants.

Pouring the Medium: After sterilization, the agar medium is carefully poured into petri plates in a laminar flow hood to maintain sterility.

Cooling to Room Temperature: The petri plates with the agar medium are allowed to cool to room temperature within the laminar flow hood to prevent contamination.

Inoculation: Sterile inoculation loops or swabs are used to transfer bacterial isolates onto the surface of the agar in the petri plates.

Incubation: The inoculated petri plates are then incubated at 37°C for 24 hours to allow bacterial growth and potential amylase production.

Iodine Test: After incubation, an iodine solution is directly added to the bacterial colonies on the agar surface. The iodine solution will turn blue-black in the presence of starch, but if amylase is produced and has hydrolyzed the starch, there will be a clear zone around the bacterial colonies where the iodine solution remains yellow-brown.

PROTEOLYTIC ACTIVITY

On blood agar media, the activity of proteolytic bacteria was qualitatively tested. Clear zones around the colonies show that the bacteria are able to degrade protein.

RESULT AND DISCUSSION

IDENTIFICATION OF SEAWEED:

The identified seaweed is the *Sargassum* species. They have branched thallus with hollow berry-like floats (pneumatocysts). The fronds are generally small and have serrated edges. Sexual reproduction takes place in most species but the pelagic species reproduce by fragmentation. The largest members can reach several meters in length.



Figure 5 : Antibacterial activity of isolated endophytic bacteria against *Staphylococcus aureus*

S.N O	ORGANISMS	ZONE OF INHIBITION (mm)			
		20µl	50µl	75µl	Positive control (streptomycin)
1.	<i>E.coli</i>	9mm	13mm	17mm	14mm
		10mm	12mm	15mm	15mm
2.	<i>Staphylococcus aureus</i>	10mm	12mm	15mm	15mm

Table 2 : Antibacterial assay of isolated endophytic bacteria against *E.coli* and *Staphylococcus aureus*

UNDER PELL

PHB PRODUCTION

SUDAN BLACK B STAINING:

Results from the initial Sudan Black B staining on test isolate cultured in petri dishes showed that colonies' colors altered to a dark greenish-blue hue when the dye was poured into the dishes. This symptom is taken to indicate that test isolates have PHB accumulations.

UNDER PEER REVIEW

PHB POWDER Bioplastics are more importance

nowadays

due to their biocompatible and biodegradable

nature which is an alternative for petroleum based plastics. *Bacillus cereus* is the bacterium that was isolated from the seaweed and was found to produce PHB upto 900 mg of its dry cell weight.

FTIR RESULT:

The PHB extraction powder was analyzed by Fourier transform infrared (FTIR). The PHB powder functional group was observed by FTIR peaks. The alkanes (C-H) group were located in 2924.09. They show alkanes consist of single-bonded carbon and hydrogen atoms. In 1743.65 peak shows the ketones (C=O) group were located and carbonyl groups are composed of a carbon atom double-bonded to an oxygen atom. Alkenes groups were located in 1643.35. In alkene, carbon carbon double bond (C=C) is the functional group. Nitro Compounds also have been presented. NO₂ is strongly present in the extract polymer. In 1442.75 the aromatic compounds (C=C) were observed. The functional group that contains only carbon and hydrogen is an aromatic ring which is a six-carbon ring with alternating double bonds. Alkyl & Aryl Halides groups also presented. When a mixture of alkyl halide and aryl halide is treated with sodium in dry ether alkyl arene is formed.

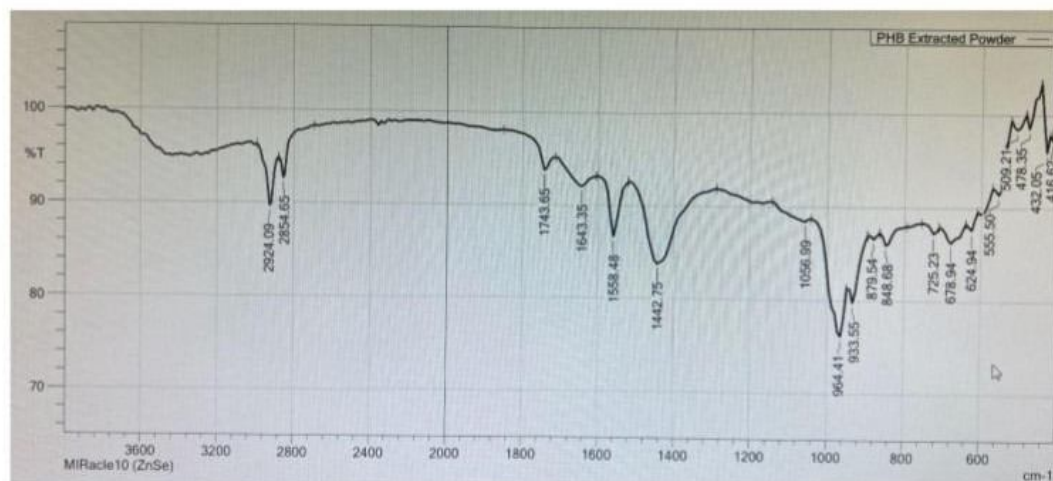


Figure 8 : FTIR report of PHB powder

AMYLOLYTICACTIVITY:

The isolated bacteria showed prominent growth on starch hydrolyzing, a zone of Inhibition was observed after the addition of iodine.



Figure 9: Amyolytic activity of isolated endophytic bacteria on Starch agar

PROTEOLYTICACTIVITY:

The isolate bacteria showed prominent growth on blood agar by the formation of zone of clearance after incubation



Figure 10 : Proteolytic activity of isolated endophytic bacteria on blood agar

REFERENCE

1. Amiluddin N. M., 2007, Study of Growth and Carragenan Content of Seaweed *Kappaphycusalvarezii*(Lin.,1758),Affected by Ice ice in the Water of Puri island, Seribu Islands, Thesis for M.S, Graduate Scholl of Bogor Agricultural University, Bogor, pp.1-78
2. Aris M, 2011, Identification, Pathogenicity of Bacteria and the Use of Gene 16S rRNA for Ice-ice Detection on Seaweed Aquaculture ,*Kappaphycusalvarezii*(Linn.,1758), Thesis for M.S,Graduate School of Bogor Agricultural University (IPB), Bogor, pp.1-127
3. Austin B., and Austin D.A., (eds). 1993, Bacterial fish pathogen disease in farmed and will fish, Second Edition, Ellis Horwood Limited. Department of Biological Sciences, Heriot-Watt University, England, pp.553
4. Buchanan R.E., and Gibbons N.E., 1974 Bergey's Manual of Determinative Bacteriology, 8th ed. USA, The Williams & Wilkins Co, Inc, pp.1272
5. Flewelling AJ, Johnson JA, Gray CA Isolation and bioassay screening of fungal endophytes from North Atlantic marine macroalgae. *Bot Mar*, 2013; R6: 287–297.
6. Furbino LE, Godinho VM, Santiago IF, Pellizzari FM, Alves TMA, Zani CL, Junior PAS, Romanha AJ, CarvalhoAJO, Gil LHVG, Rosa CA, Minnis AM, Rosa LH. Diversity patterns; 27 ecology and biological activities of fungal communities associated with the endemic macroalgae across theAntarctic peninsula. *MicrobEcol*, 2014; 67: 775–787.
7. Godinho VM, Furbino LE, Santiago IF, Pellizzari FM, Yokoya NS, PupoD, Alves TM, Junior PA, Romanha AJ, Zani CL, Cantrell CL, Rosa CA, Rosa LH. Diversity and bioprospecting of fungal communities associated with endemic and cold adapted macroalgae in Antarctica. *ISME J*, 2013;7: 1434–1451.
8. Khan MS, Zaidi A, Wani PA. Role of phosphate solubilizing microorganisms in sustainable agriculture A review. *Agronomy and Sustainable Development*, 2007; 27: 29-43.
9. Loque CP, Medeiros AO, Pellizzari FM, Oliveira EC, Rosa CA, Rosa LH. Fungal community associated with marine macroalgae from Antarctica. *Polar Biol*, 2010; 33: 641–648.
10. Dawes EA, Senior PJ. The role and regulation of energy reserve polymers in microorganisms. *Adv MicrobPhysiol*1973;10:135-266.
11. Pringsheim EG, Wiessner W. Minimum requirements for heterotrophic growth and reserve substance in *Beggiatoa*. *Nat*1963;197:02.
12. Kannan LV, Rehacek Z. Formation of Poly- β -hydroxybutyrate by Actinomycetes. *Indian J Biochem*1970;7:126-9.