

Enhancing Seed Quality of Chickpea (*Cicer arietinum* L.) Through Foliar Application of Plant Growth Regulators and Nutrients under Water Deficit Conditions

Abstract

Chickpea cultivation in rainfed regions, such as Madhya Pradesh, faces challenges due to water stress, impacting crop yield and quality. The study conducted assesses the impact of foliar spray with plant growth regulators and nutrients on the germination percentage, seedling length, and seed vigor indices of chickpea varieties (JG 36 and JG 14) under water deficit conditions. Cytokinin analogues, known for breaking dormancy, and other growth regulators and nutrients were applied during the flowering stage. The results revealed significant differences in germination percentage, seedling length, seed vigour index I, and seed vigour index II among irrigation levels, varieties, and foliar spray of plant growth regulators and nutrients. Under different irrigation levels, D₁ (Irrigation at 30 DAS and at flower initiation) recorded the highest germination percentage (97.24%), seedling length (31.41 cm), seed vigour index I (3054.81), and seed vigour index II (52.77) compared to control D₂ (Drought stress at Flowering up to physiological maturity). JG 14 (V1) performed well, achieving the highest germination % of 96.99%. Among the treatments, foliar application of Benzyladenine (BA) at 40 ppm (T₃) resulted in the highest germination percentage (98.17%) and seed vigour index II (60.33), whereas T₈ (BA 40ppm + ZnSO₄ 1%) exhibited the highest seedling length (31.83 cm) and seed vigor index I (3099.85). This study demonstrates that foliar spray with plant growth regulators and nutrients, particularly Benzyladenine at 40 ppm, significantly improves germination percentage, seedling length, and seed vigor index, offering potential solutions for enhancing chickpea productivity under water deficit conditions.

Keywords: Seed quality, Benzyladenine, Water deficit stress, Plant growth regulators

Introduction

Globally, Chickpea is prevalent in regions with temperate and subtropical climates (FAOSTAT, 2011). In Madhya Pradesh, chickpea is often grown as a rainfed crop, utilizing the monsoon rainfall during the Rabi season (Pande *et al.*, 2012). The crop's resilience to withstand water stress and its ability to perform nitrogen fixation are crucial in rainfed agriculture, particularly in regions with limited irrigation facilities (Islam *et al.*, 2021). In India, drought is one of the predominant constraints on production within semi-arid ecosystems. Salinity, high temperatures, and low temperatures, water stress also negatively impact plant growth and agricultural productivity in these regions (Golla, 2021). Among the various abiotic stresses, drought is identified as the most severe, significantly restricting crop production and yield, especially in arid and semi-arid areas (Araujo *et al.*, 2015). Severe water deficit

condition leads to reduction in crop growth, physiology and yield of the crop (Sabaghpour, 2003).

Drought stress results in a decrease in both germination percentage and seedling growth (Ashraf *et al.*, 2006). In the grain-filling stage of the crop, a water deficit becomes a crucial factor, restricting the rate of seed filling and various processes associated with seed quality. The productivity of chickpea, particularly in semi-arid regions, is connected to factors such as insufficient photosynthates in pods and seed setting. The application of macro and micro nutrients through foliar means has been suggested as a better solution (Parimala *et al.*, 2013). Water stress is identified as the primary abiotic stress that hampers crop growth, development, and overall production on a globally (Shahbaz *et al.*, 2011).

The application of bioregulators from external sources has emerged as an innovative technology for enhancing stress tolerance in crop plants (Vineeth *et al.*, 2017). The importance of combination of bio-regulators and nutrients in enhancing crop yields has long been recognized, and this cost-effective technology has now demonstrated its value in substantially increasing agricultural productivity. The challenge of moisture stress during the flowering and seed-setting stages, leads to decline in seed yield (Anbessa and Bejiga, 2002). Thiourea is recognized for its ability to break dormancy and promote germination (Germchiet *et al.*, 2011). In potato tuber, application of thiourea at optimal concentration enhances the germination process and the development of multiple sprouts (Rehman *et al.*, 2002).

Thiourea, also recognized as thiocarbamide due to its nitrogen (-NH₂) and sulfur (-SH) elements, plays a role in influencing plant growth under stressful conditions (Garg *et al.*, 2006). Additionally, it has been observed to facilitate seed germination, promote growth, and alleviate stress in plants (Wahid *et al.*, 2017). The effectiveness of thiourea extends to overcoming both environmentally induced and inherent seed dormancy by fostering seed germination (El-Keblawy and Gairola, 2017). Thiourea increased seed germination, foliar application led to enhanced gas exchange properties, and supplementing the medium supported improved root growth and proliferation (Wahid *et al.*, 2017).

According to Kulsumbiet *al.* (2020), the application of Thiourea at a 1% concentration resulted significantly superior seed quality parameters compared to the control, including speed of germination, shoot length, root length, seedling dry weight, and seedling vigor index. Yasar *et al.* (2016) found that germination percent

was more on 50 μ M BAP than 10 μ M Thidiazuron (TDZ) in pea seeds. TDZ leads to a reduction in the percentage of seed germination (Kim *et al.*, 2019). This inhibitory effect of higher TDZ levels on seed germination has been documented in various plant species, including instances in *Carica papaya* and *Lotus corniculatus* (Murthy *et al.*, 1998; Bhattacharya and Khuspe, 2001; Patil *et al.*, 2012). Hence, this study was taken up to evaluate the efficiency of foliar application of macro and micro nutrients on yield and seedling quality parameters of chickpea.

Material and Methods

Chickpea varieties JG 36 and JG 14 was sown in Experimental Research Farm, Seed technology Research Unit, JNKVV, Jabalpur during *Rabi* 2021-2022 and 2022-2023 in split-split plot design with three replications. Using soil moisture content data and soil water potential data, the water deficit conditions of the crop were assessed. During the water deficit condition, the foliar spray of plant growth regulators and nutrients was applied at the 50% flowering stage for both D₁- Control (Irrigation at 30 DAS and at flower initiation) and D₂-Drought (Drought stress at Flowering upto physiological maturity). During flowering stage, irrigation was provided only to D₁. Water stress was imposed in D₂ during the reproductive stage.

In both the conditions, different plant growth regulators and nutrients were applied at 50% flowering stage viz., T₁-control (no spray), T₂-Thiourea(TU) @ 1000 ppm, T₃ – Benzyladenine (BA) @ 40 ppm, T₄ – Thidiazuron (TDZ) @ 10 ppm, T₅ – 1%ZnSO₄, T₆ – 1% KCl, T₇ – TU @ 1000 ppm + 1% ZnSO₄, T₈ - BA 40 @ ppm + 1% ZnSO₄ , T₉ - TDZ @ 10 ppm + 1% ZnSO₄ , T₁₀ - TU @ 1000 ppm + 1% KCl, T₁₁- BA @ 40 ppm+ 1% KCl, T₁₂- TDZ @ 10 ppm + 1% KCl. After the harvest, the seeds were kept for germination test was conducted as per the International Seed Testing Association (ISTA) procedure by adopting between paper method in a germinator.

Procedure

Three replications of 100 seeds each from their respective treatments were taken. The paper towel was moistened with distilled water, ensuring it was damp but not dripping wet. The moistened paper towel was placed on a flat surface. On one half of the paper towel, seeds were aligned evenly, maintaining consistent spacing between them. The other half of the moistened paper towel was folded over the seeds to fully cover them. An additional layer of butter paper was used to fold the paper towel, creating a humid microenvironment conducive to germination. To further support this environment, the paper towel was kept in a tray filled with water. The

samples were then placed in a germinator at a temperature of 25 ± 2 °C for 10 days, maintaining a relative humidity of 90% (Anonymous, 1999). The tray's water level was refilled as needed. After a 10-day period, the germinated seeds were categorized as normal seedlings, abnormal seedlings, hard seeds, or dead seeds. The germination percentage was then calculated exclusively based on the count of normal seedlings.

$$\text{Germination percentage} = \frac{\text{Number of seeds germinated}}{\text{Total number of seeds}} \times 100$$

Seedling length (cm)

Five normal seedlings from the germination test were randomly selected for measurement of seedling length on tenth day. The seedling length was measured from the collar region to the tip of the primary leaf. The mean seedling length was expressed in centimetres. To calculate the mean (average) seedling length, the lengths of all the measured seedlings were summed, and then this sum was divided by the total number of seedlings that were measured.

$$\text{Mean Seedling length} = \frac{\text{Sum of seedling lengths}}{\text{Total number of seedlings}}$$

Seed vigour index-I

Seed vigour Index is a measure used to assess the overall vigour or health of seedlings. Seed vigor Index (VI) was calculated using the formula given by Abdul-Baki and Anderson (1973)

$$\text{Vigour index I} = [\text{Root length (cm)} + \text{Shoot length (cm)}] \times \text{Germination \%}$$

Seed vigour index – II

The weight of 10 seedlings (grams), excluding the cotyledons, was measured on the 10th day after drying them at 60-80°C in an oven for 24 hours. The lot of seedlings that displayed the highest dry weight was considered to be the most vigorous. This method helps assess the seedlings' overall health and growth potential based on their accumulated dry weight, excluding the energy reserves stored in the cotyledons.

$$\text{Vigour index I} = \text{Seedling dry weight (g)} \times \text{Germination \%}$$

Statistical analysis

The statistical analysis, namely two-way ANOVA, and Tukey's Honest Significant Difference (HSD) test at a 5% level of significance, was conducted using

R 4.2.2 statistical software. All results were expressed as the average of three replications. Treatment effects were determined through analyses of variance using the Split-Split plot design (Gomez and Gomez, 1984).

Result and Discussion

The analysis of variance conducted on data of two consecutive years (2021-2022 and 2022-2023), as well as the pooled data revealed a significant variation in germination percentage, seedling length, seed vigor index I, and seed vigour index II (Table 2).

i) Effect of foliar spray of plant growth regulators and nutrients on Germination percentage and seedling length of chickpea under water deficit condition

Seed quality is influenced by various factors including environmental conditions, genetic factors, and the moisture and fertility of the soil (Soleimani *et al.*, 2000). Seed germination stands out as an important and highly sensitive stage in the plant life cycle (Ashraf and Mehmood *et al.*, 1990). Seed germination and seedling growth significantly contributes for determining the yield of the crop (Rauf *et al.*, 2007). The results from the pooled analysis of two consecutive years (Table 1 and Figure 1) indicates the range of germination % was found to be 95.58 % to 98.17%. Under the irrigation level, germination % was found to be significant. The highest (97.24%) germination % was found in D₁ (Irrigation at 30 DAS and at flower initiation) followed by 96.33 % in D₂ (Drought stress at Flowering upto physiological maturity). Among the varieties, V₁ (JG36) recorded highest (96.99%) germination % followed by 96.58% in V₂ (JG 14). Among the treatments, it was observed that foliar application of Benzyladenine (BA) at 40 ppm (T₃) resulted in the highest germination percentage (98.17%) which is at par with all other treatments. In contrast, the control treatment (T₁) recorded the lowest (95.58%) germination percentage which is on par with treatment T₂ (96.21%).

Our result is in similarity with Morsy *et al.*, (2018) who stated that seed germination (%) decreased with increase in water deficit condition. Seeds produced under moisture deficit conditions exhibited delayed and uneven germination, along with reduced seedling growth and storage viability (Awosanmiet *et al.*, 2022). Water deficit stress during the vegetative stage has adverse effects on both seed germination and plant establishment. However, legume plants exhibit drought

tolerance during the late vegetative stage of the crop (Subbaramamma *et al.*, 2017). Okcu *et al.* (2005) found that the primary effect of drought is impaired germination and early seedling growth, ultimately impacting the growth and productivity of legume (Hossain *et al.*, 2010).

Swain *et al.* (2014) have noted that early stage of the crop particularly seed germination and initial seedling growth, are critical stages that are adversely affected by water stress. The results from the pooled analysis of two consecutive years (Table 1) indicate the range of seedling length was found to be 26 cm to 31.83 cm. With respect to irrigation levels, seedling length was found to highest (31.41cm) in D₁ (Irrigation at 30 DAS and at flower initiation) followed by (28.49 cm) D₂ (Drought stress at Flowering upto physiological maturity). Among the varieties, highest (30.30 cm) seedling length was found to be observed in V₂ (JG 14) followed by V₁ (JG 36) with 29.6cm. Among the treatments, maximum (31.83 cm) seedling length was observed for treatment T₈ - BA 40 @ ppm + 1% ZnSO₄ which is on par with all other treatments, whereas minimum seedling length was observed for control T₁ (26.00 cm) which is on par with T₂ (28.64cm), T₆ (29.77cm), T₇ (28.29cm), T₉ (28.98cm) and T₁₀ (30.21cm) respectively. Our result is in similarity with Soleymani *et al.* (2018) who reported that there was a significant reduction in germination percentage, seedling growth as the stress level increases. Mohammadkhani and Heidari (2008) also observed a reduction in root and shoot length, as well as the fresh and dry weight of maize seedlings with an increase in stress levels.

Figure 1. Effect of Germination % on foliar spray of plant growth regulators and nutrients in chickpea under water deficit conditions

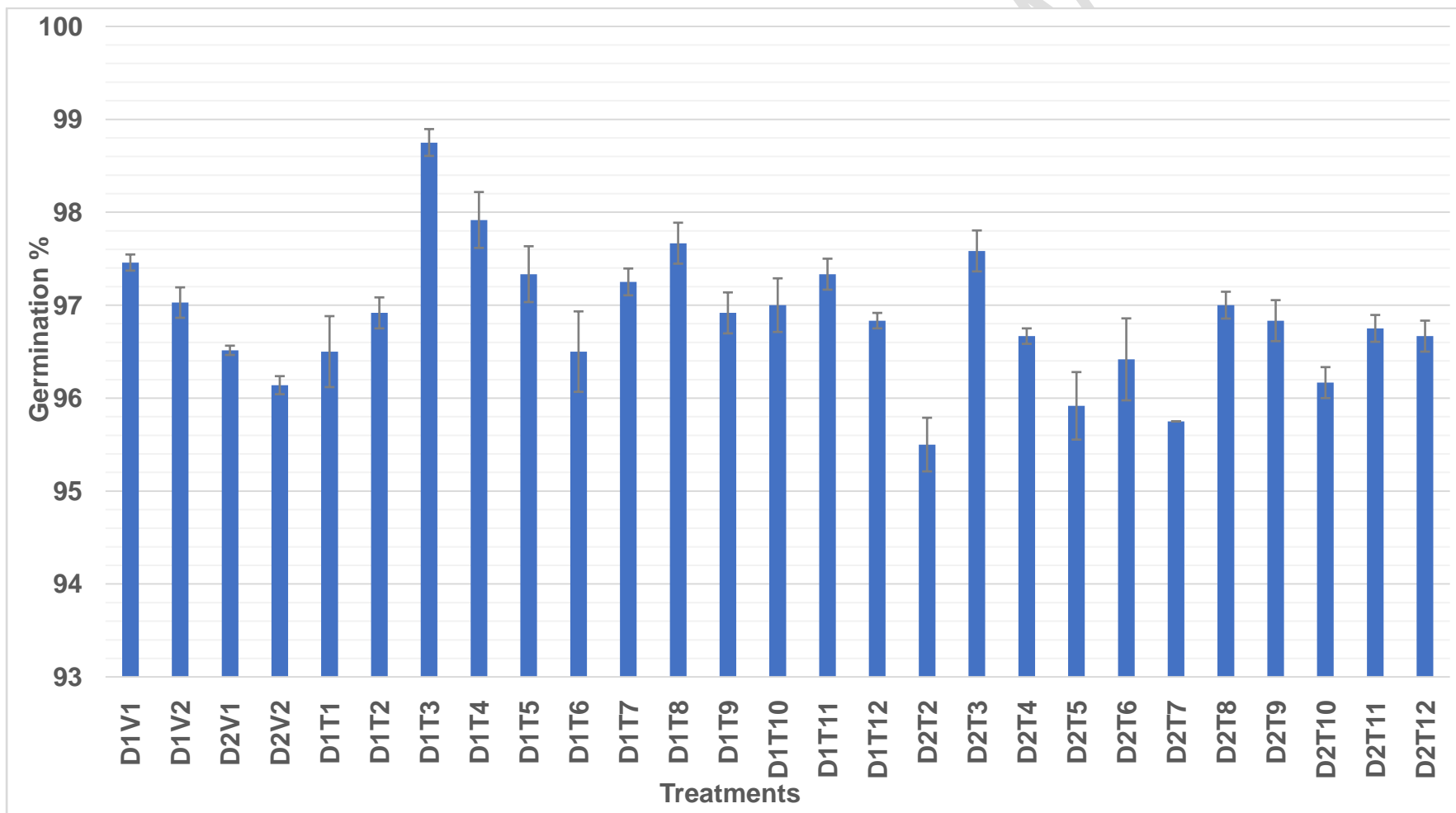


Table 1: Germination % and Seedling length as affected by irrigation level, varieties, plant growth regulators and nutrients of chickpea.

Treatments	Germination %			Seedling length (cm)		
	Main plot: Irrigation (D)					
	2021-2022	2022-2023	Pooled	2021-2022	2022-2023	Pooled
D ₁	97.39 ^a	97.10 ^a	97.24 ^a	31.80 ^a	31.02 ^a	31.41 ^a
D ₂	96.36 ^b	96.29 ^a	96.33 ^b	28.41 ^b	28.57 ^b	28.49 ^b
SEm±	0.10	0.16	0.12	0.07	0.10	0.06
Subplot: Varieties (V)						
V ₁	97.21 ^a	96.76 ^a	96.99 ^a	30.17 ^a	29.04 ^b	29.60 ^b
V ₂	96.54 ^b	96.62 ^a	96.58 ^b	30.05 ^a	30.55 ^a	30.30 ^a
SEm±	0.06	0.08	0.03	0.05	0.11	0.04
Sub-sub plot: Treatments (T)						
T ₁	95.83 ^d	95.33 ^e	95.58 ^f	26.50 ^d	25.51 ^c	26.00 ^b
T ₂	96.25 ^{cd}	96.17 ^{de}	96.21 ^{ef}	30.44 ^{abcd}	26.83 ^{bc}	28.64 ^{ab}
T ₃	98.25 ^a	98.08 ^a	98.17 ^a	34.53 ^a	28.43 ^{abc}	31.48 ^a
T ₄	97.25 ^b	97.33 ^{abc}	97.29 ^{bc}	32.84 ^{ab}	29.71 ^{abc}	31.28 ^a
T ₅	96.75 ^{bc}	96.50 ^{bcd}	96.63 ^{bcd}	31.78 ^{ab}	31.40 ^a	31.59 ^a
T ₆	96.50 ^{bcd}	96.42 ^{cde}	96.46 ^{de}	29.44 ^{bcd}	30.09 ^{ab}	29.77 ^{ab}
T ₇	96.50 ^{bcd}	96.50 ^{bcd}	96.50 ^b	27.13 ^{cd}	29.46 ^{abc}	28.29 ^{ab}
T ₈	97.08 ^{bc}	97.58 ^{ab}	97.33 ^{bcd}	31.11 ^{abc}	32.56 ^a	31.83 ^a
T ₉	96.83 ^{bc}	96.92 ^{bcd}	96.88 ^{cde}	28.74 ^{bcd}	29.22 ^{abc}	28.98 ^{ab}
T ₁₀	96.83 ^{bc}	96.33 ^{cde}	96.58 ^{bcd}	29.06 ^{bcd}	31.37 ^{ab}	30.21 ^{ab}
T ₁₁	97.17 ^b	96.92 ^{bcd}	97.04 ^{bcd}	28.94 ^{bcd}	32.33 ^a	30.64 ^a
T ₁₂	97.25 ^b	96.25 ^{cde}	96.75 ^{bcd}	30.78 ^{abcd}	30.62 ^{ab}	30.70 ^a
SEm±	0.19	0.23	0.15	0.95	0.96	0.93

The values with same letter cases are not significantly different at p<0.05 level. *, ** and *** denotes values are significant at p<0.05, 0.01 and 0.001 levels, respectively.

Table 2: Results of the two-way ANOVA and Tukey multiple range tests for the comparative effects of plant growth regulators and nutrients on the seed germination %, seedling length(cm), seed vigour index I and Seed vigour index II of Chickpea under water deficit condition.

Treatments	Germination %			Seedling length (cm)			Seed Vigour Index I			Seed Vigour Index II		
	2021-2022	2022-2023	Pooled	2021-2022	2022-2023	Pooled	2021-2022	2022-2023	Pooled	2021-2022	2022-2023	Pooled
D	0.61 ^a	0.96ns	0.73*	0.39***	0.62**	0.37***	43.65***	41.17**	32.58***	1.31**	0.46**	0.89**
V	0.23**	0.32ns	0.13***	0.20ns	0.48***	0.16***	13.52**	35.05***	13.31***	0.63***	0.40**	0.35***
T	0.53***	0.66***	0.44***	2.60***	2.69***	2.60***	252.41***	263.09***	253.36***	4.46***	4.55***	4.46***
D x V	0.45ns	0.70ns	0.49ns	0.32*	0.58*	0.29ns	31.24**	43.98*	24.88ns	1.06ns	0.50***	0.67***
D x T	0.85*	1.05**	0.79***	3.49ns	3.62*	3.50ns	337.65ns	351.83*	338.66ns	5.99***	6.09ns	6.00ns
D x V x T	1.09**	1.37ns	0.95ns	4.92***	5.11ns	4.94ns	477.19***	497.82ns	478.81ns	8.46***	8.61***	8.45*

^a F-values. ns: not significant F ratio ($p < 0.05$); *, ** and *** indicate significant at $P < 0.05$, 0.01 and 0.001, respectively.

ii)Effect of foliar spray of plant growth regulators and nutrients on Seed Vigour index I and seed vigour index II of chickpea seeds under water deficit condition

The results from the pooled analysis of two consecutive years (Table 3) indicate the range of seed vigour index I was found to be 2487.11 to 3099.85. Seed vigour index I was found to be highest (3054.81) in D₁ (Irrigation at 30 DAS and at flower initiation) followed by D₂ (2745.37) (Drought stress at Flowering upto physiological maturity). With respect to varieties, seed vigour index I was found be higher (2927.7) in V₂ (JG 14) followed by 2872.48 in V₁ (JG 36). With respect to the foliar application of plant growth regulators and nutrients, highest (3099.85) seed vigour index I was found to be in treatment T₈(BA 40 @ ppm + 1% ZnSO₄) which is in on par with all other treatments except T₁. Seed vigour index I was found to be lowest (2487.11) in control T₁ which is on par with T₂ (2756.39), T₆ (271.78), T₇ (2731.74) and T₉ (2807.85), respectively. According to Dhandas *et al.* (2004), the traits most susceptible to drought stress are seed vigor index and plumule length.

The results from the pooled analysis of two consecutive years (Table 3) indicate the range of seed vigour index II was found to be 46.81 to 60.33. Under the irrigation level, seed vigour index II was found to be significant. The highest (52.77) seed vigour index II was found in D₁ (Irrigation at 30 DAS and at flower initiation) followed by 49.91 in D₂ (Drought stress at Flowering upto physiological maturity). Among the varieties, V₁ (JG36) recorded highest (53.13) seed vigour index II followed by 49.57 in V₂ (JG 14). Among the treatments, it was observed that foliar application of Benzyladenine (BA) @ 40 ppm (T₃) resulted in the highest seed vigour index II (60.33) which is on par with T₄ (53.16). In contrast, the control treatment (T₁) recorded the lowest seed vigour index II 46.81 which is at par with treatment T₃ (60.33).Our result is in conformity with Abdoli *et al.* (2012) found that drought stress led to a decrease in seedling vigor index. Movahhedy-Dehnavy *et al.* (2007) reported that the foliar application of nutrients increased the germination percentage, germination rate, seedling dry weight, and seed vigor index (El-Beltagiet *al.*, 2022). Uniform germination rate and seedling emergence are important factors for establishing a good crop stand (Murunguet *al.*, 2003).

Table 3: Seed vigour Index I and Seed vigour Index II as affected by irrigation level, varieties, plant growth regulators and nutrients of chickpea.

Main plot: Irrigation (D)						
Treatments	Seed Vigour Index I			Seed Vigour Index II		
	2021-2022	2022-2023	Pooled	2021-2022	2022-2023	Pooled
D ₁	3097.90 ^a	3011.72 ^a	3054.81 ^a	52.79 ^a	52.74 ^a	52.77 ^a
D ₂	2738.36 ^b	2752.39 ^b	2745.37 ^b	48.53 ^b	51.32 ^b	49.91 ^b
SEm±	7.17	6.77	5.35	0.22	0.08	0.15
Subplot: Varieties (V)						
V ₁	2934.09 ^a	2810.86 ^b	2872.48 ^b	53.84 ^a	52.41 ^a	53.13 ^a
V ₂	2902.16 ^b	2953.25 ^a	2927.70 ^a	47.48 ^b	51.65 ^b	49.57 ^b
SEm±	3.44	8.93	3.39	0.16	0.10	0.09
Sub-sub plot: Treatments (T)						
T ₁	2538.33 ^d	2435.89 ^c	2487.11 ^b	44.03 ^d	49.58 ^b	46.81 ^b
T ₂	2929.18 ^{bcd}	2583.60 ^{bc}	2756.39 ^{ab}	49.97 ^{bcd}	51.93 ^a b	50.95 ^b
T ₃	3392.71 ^a	2788.13 ^{abc}	3090.42 ^a	65.66 ^a	54.99 ^a b	60.33 ^a
T ₄	3194.68 ^{ab}	2890.88 ^{ab}	3042.78 ^a	53.47 ^{bc}	52.85 ^a b	53.16 ^{ab}
T ₅	3074.93 ^{ab}	3028.25 ^a	3051.59 ^a	45.49 ^d	59.00 ^a	52.24 ^b
T ₆	2841.38 ^{bcd}	2902.18 ^{ab}	2871.78 ^{ab}	45.02 ^d	51.29 ^b	48.15 ^b
T ₇	2618.86 ^{cd}	2844.63 ^{abc}	2731.74 ^{ab}	46.27 ^{cd}	49.89 ^b	48.08 ^b
T ₈	3022.24 ^{abc}	3177.45 ^a	3099.85 ^a	54.09 ^b	49.56 ^b	51.83 ^b
T ₉	2783.28 ^{bcd}	2832.43 ^{abc}	2807.85 ^{ab}	48.58 ^{bcd}	54.64 ^a b	51.61 ^b
T ₁₀	2814.95 ^{bcd}	3021.20 ^{ab}	2918.08 ^a	49.65 ^{bcd}	52.50 ^a b	51.07 ^b
T ₁₁	2812.63 ^{bcd}	3133.39 ^a	2973.01 ^a	51.49 ^{bcd}	47.51 ^b	49.50 ^b
T ₁₂	2994.39 ^{abc}	2946.62 ^{ab}	2970.50 ^a	54.23 ^b	50.62 ^b	52.43 ^b
SEm±	89.81	93.61	90.15	1.59	1.62	1.59

The values with same letter cases are not significantly different at p<0.05 level. *, ** and *** denotes values are significant at p<0.05, 0.01 and 0.001 levels, respectively.

Conclusion

In rainfed chickpea cultivation, water stress poses challenges to yield and seed quality. The germination percentage, seedling length, and seed vigor index were influenced by irrigation levels, varieties, and treatments. Our findings underscore the importance of foliar application of plant growth regulators and nutrients, particularly the application of Benzyladenine at 40 ppm, in mitigating the adverse effects of water deficit conditions on seed quality parameters. The seed

vigor indices I and II reflected the positive impact of foliar applications, with specific treatments, such as T₈ (BA 40 ppm + ZnSO₄ 1%), outperforming others. These results provide practical insights for optimizing chickpea yield and seedling quality under water deficit stress, offering a promising avenue for sustainable crop management.

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