

Original Research Article

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Assessment of Rapeseed-Mustard Varieties for Resistance against White Rust (*Albugo candida*) Infection

ABSTRACT

Rapeseed-Mustard plays a crucial role in India's oilseed production, facing challenges from various factors affecting yield and seed quality. Notably, white rust stands out as a significant biotic stressor, causing substantial losses in both yield and seed quality, particularly in oil content. To address this, utilizing resistant varieties is the most cost-effective and environmentally friendly approach for disease management. However, the available resistant sources are limited. In a recent study during the Rabi seasons of 2021-22 and 2022-23, thirty rapeseed mustard genotypes were assessed under white rust prevalent conditions. Out of 30 genotypes tested, none were found to be free of white rust disease infection and none were classified as resistant. However, 11 genotypes were registered under moderately resistance while, 19 genotypes susceptible category. Promisingly, some genotypes exhibited resistance to white rust, offering potential for developing superior cultivars to effectively manage the disease in regions where mustard cultivation is prevalent.

Keywords: Rapeseed-mustard, white rust, genotypes, resistance

1. INTRODUCTION

India stands as one of the world's leading producers of edible oils, with rapeseed-mustard being a significant annual oilseed crop, contributing around 25% to the nation's total oilseed production. Spanning 8.06 million hectares, it yields 11.75 million tonnes at a productivity rate of 1458 kg/ha (Anonymous, 2022). Ranking second in area after soybean, rapeseed-mustard is crucial due to its higher oil content (39-44%).

This agriculturally important crop has a rich cultivation history in India, China, and is gaining prominence in Australia. Predominantly grown during the Rabi season, its cultivation is concentrated in states such as Rajasthan, Madhya Pradesh, Uttar Pradesh, Haryana, West Bengal, Assam, Jharkhand, Gujarat, North Eastern States, and Bihar, collectively representing 96% of the crop's area and production. Despite the substantial oilseed cultivation, India remains a major importer of edible oil (Singh *et al.*, 2022). The mustard sector in India experiences fluctuations in area, production, and yield due to various biotic and abiotic stresses, as highlighted by Singh *et al.* (2022).

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33 Prominent biotic stresses leading to yield reduction in rapeseed-mustard include *Alternaria*
34 blight (*Alternaria brassicae*), white rust (*Albugo candida*), *Sclerotinia* stem rot (*Sclerotinia*
35 *sclerotiorum*), and downy mildew (*Hyaloperonospora parasitica*). Severe infections, particularly by
36 *Alternaria* blight, white rust, and downy mildew, can result in economically significant yield losses
37 ranging from 20% to 60% (Bisht *et al.*, 1994).

38 When occurring together, white rust and downy mildew can cause a substantial impact, leading to a
39 37-47% reduction in pod formation and a 17-54.5% decrease in seed yield (Mukherjee *et al.*, 2001).
40 Effective management strategies include the utilization of resistant varieties, acknowledged for being
41 cost-effective and environmentally friendly. Notably, germplasm screening has identified varying levels
42 of host resistance against white rust, providing a promising avenue for mitigating these challenges
43 (Ahmad *et al.*, 2014; Dharavath *et al.*, 2017).

44 The resistance to diseases, particularly through the presence of a single gene (R-gene) or multiple
45 genes with modest effects, significantly influences crop production, quality assurance, environmental
46 safety, and overall yield. Within Brassica species, considerable genotypic variability is observed
47 concerning white rust resistance. Notably, *Brassica juncea*, a member of the Indian gene pool,
48 displays high susceptibility to white rust, while the germplasm from East European origins exhibits
49 varying degrees of resistance, ranging from highly resistant to moderately resistant (Chand *et al.*,
50 2022).

51 To ensure consistent and stable crop performance, the development of varieties with inherent
52 resistance or tolerance to diseases, including white rust, emerges as an economically viable,
53 environmentally safe, and sustainable strategy. The research focus is directed towards evaluating
54 rapeseed-mustard genotypes for their resistance to white rust under field conditions, with a particular
55 emphasis on the cost-effectiveness and environmental advantages associated with the cultivation of
56 resistant varieties.

57 **2. MATERIALS AND METHODS:**

58 This study involved the examination of 30 locally collected rapeseed-mustard cultivars. The cultivars
59 were cultivated in a randomized block design with three replications during the Rabi seasons of 2021-
60 22 and 2022-23 at the experimental field of the Department of Plant Pathology, College of Agriculture,
61 BUAT, Banda (U.P.). The planting arrangement followed an augmented design with two rows for each
62 genotype, each row extending 3 meters. The spacing was maintained at 30 cm between rows and 15
63 cm between individual plants. The incidence of white rust disease was systematically observed and
64 recorded throughout the experimental period.

65 To analysed the percent disease index (PDI), observations on white rust occurrence were recorded
66 every 10 days from 10 randomly selected plants of each row, both during the vegetative and true leaf
67 stage (42nd days after sowing) under natural epiphytotic conditions.

68 Observations recorded on ten randomly selected plants from each row of each tagged genotype. The
 69 observation on disease severity of white rust disease was recorded at 10 days interval using 0-9
 70 rating scale (Conn *et al*, 1990) (Table 1).

71 The intensity was calculated with formulae = $\frac{\text{Sum of all disease rating}}{\text{No.of leaves observed} \times \text{Max. disease rating}} \times 100$

72 The Area Under Disease Progress Curve (AUDPC) was calculated by the formula as under (Shaner
 73 and Finney 1977)

$$\text{AUDPC} = \sum_{i=1}^{n-1} \left[\frac{y_i + y_{i+1}}{2} \right] (t_{i+1} - t_i)$$

74 Where,

75 Y_i = White rust severity (%) at the 1st observation

76 $t_{(i+1)} - t_i$ = Time (days) between two disease scores

77 n = Total number of observations

78 Infection rate

79 Apparent infection rates per unit days helps estimated the progress of the disease in the field on two
 80 different time points. Formula for weekly interval given by Vanderplank in 1963 was used to calculate
 81 the Logarithmic infection rates.

$$\text{Infection rate}(r) = \frac{2.3}{t_2 - t_1} \log_e \frac{x_2(1 - x_1)}{x_1(1 - x_2)}$$

82 **Where**

83 (r) = Apparent infection rate

84 t_1 = time during first observation

85 t_2 = time (days) during second observation

86 $t_2 - t_1$ = time intervals between two observation

87 x_1 = per-cent disease intensity value in decimal at corresponding t_1 time

88 x_2 = per-cent disease intensity value in decimal at corresponding t_2 time

89 Log e = natural log

90

91 **Table-1: Disease rating scale for evaluation of genotypes reaction:**

Rating score	Leaf area covered (%)	Disease reaction
0	No symptoms	Immune (I)
1	< 5	Highly resistant (HR)
3	5-10	Resistant (R)
5	11-25	Moderately resistant (MR)
7	26-50	Susceptible (S)

9	>50	Highly susceptible (HS)
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93 **3. RESULTS AND DISCUSSION:**

94 Data presented in table-2 & Fig-1 evident that combined mean of percent disease severity in different
 95 genotypes were ranged between 18.49 to 30.51% during crop session 2022 & 2023. Data are in
 96 accordance with that minimum percent disease severity was recorded in genotypes Giriraj (18.49%)
 97 differ statistically with Tejashwani (20.40%) followed by RH746 (20.66%), GSC7 (21.34%), Krishna
 98 (21.35%), PC6 (21.53) and NRCHB101 (22.18%). The next group of genotypes that showed
 99 promising were super express, RH406, DHM44, and PM29, which obtained respective results of
 100 22.30, 23.45, and 24.02% which, during crop sessions 2022 and 2023, shown non-significant
 101 differences in their effectiveness against the rapeseed-mustard white-rust.

102 For the genotypes KMH8765, PHR126, PM30, RLC3, KMH721, Basanti, and RH749, the
 103 reported percentages of disease severity (25.49, 25.65, 25.77, 25.83, 26.43, 26.47, and 27.44%)
 104 were statistically equivalent to each other. Subsequently, genotypes of Radha, Dhakar, Peelasona,
 105 GSC6, and DRMR116540 were registered as promising; their respective genotypes were stated as
 106 27.57, 27.68, 28.18, 28.59, and 28.66, however they were revealed to be comparable. A total of 56-92
 107 (30.51%), RCH (31.47%), Ganga (30.28), PGSH1707 (2.68%), Nirmlabold (29.45), Lahar (29.25), and
 108 Kalasona (29.24) had the highest percentage of disease severity. Similar to present findings Li *et al.*,
 109 (2007) were screened out 44 genotypes of *Brassica juncea* against white rust and concluded that
 110 most of the genotypes of Indian gene pool showed moderate to susceptible reaction against white
 111 rust disease. Previous researchers identified certain *Brassica* germplasms as resistant, but in the
 112 current study, the screened Brassica lines differed from those earlier findings. Some lines were
 113 identified as susceptible, while others fell into the moderately resistant category in the present
 114 research. The data presented in the table-2 indicates clear that the range of the pooled AUDPC of
 115 white rot in several cultivars of rapeseed-rapeseed-mustard was 244.60 to 493.78. RCH1 recorded
 116 the highest pooled AUDPC (493.78), which was followed by 56-92 (482.05), Ganga (466.53),
 117 PGSH1707 (456.26), Nirmla bold (437.94), Lahar (426.33), Kalasona (412.03), DRMR_116540
 118 (407.56), GSC6 (406.95), Peelasona (394.92), Dhakar (384.98), Radha (381.75), RH 749 (370.75),
 119 KMH 721 (363.50), Basanti (362.94), RLC3 (352.53), PM 30 (346.58), PHR 126 (345.79), KMH 8765
 120 (-339.13), PM 29 (-317.21), DHM 44 (-311.44), RH 406 (297.58), Super express (298.42), NRCHB
 121 101 (292.15), PC6 (279.40), Krishna (-273.26), and GSC7 (273.22). Giriraj (244.60) genotypes had
 122 the lowest pooled AUDPC, followed by Tejashwani (256.50) and RH 746 (261.38).

123 The data in the table revealed the apparent white rust disease infection rate in several rapeseed
 124 rapeseed-mustard cultivars during the 2022–2023 crop season. From the data, the genotypes Kala
 125 Sona (0.526) had the highest observed r value, followed by RCH-1 (0.515), GSC-6 (0.510), KMH-721
 126 (0.497), 56-92 (0.493), Peela Sona (0.491), Lahar (0.490), RLC-3 (0.489), Dhakar, Radha, & NRCHB-
 127 101 (0.479), Ganga & PGSH-1707 (0.478), Basanti (0.473), PM-29 (0.472), PM-30 & PC-6 (0.470),
 128 PHR-126 (0.466), and Krishna (0.463). KMH-8765 (0.453), RH-749 & Nirmala Bold & DRMR-116540

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129 (0.460), RH-406 (0.457), GSC-7 (0.484), and RH-746 (0.446). The genotypes Giriraj & Tejshwani had
 130 the lowest apparent infection rate (0.444), followed by DHM-44 & Super express (0.430).

131 The findings in the table-3 showed that all rapeseed-mustard genotypes were
 132 infected with white rust disease, however the severity differed across genotypes. The data on per-cent
 133 disease severity were classified into several categories (based on the disease rating scale) to
 134 determine the reaction of genotypes to white rust disease. Out of 30 genotypes tested, none were
 135 found to be free of white rust disease infection and none were classified as resistant. However, 11
 136 genotypes namely, Giriraj, RH-746, RH-406, NRCHB-101, PM-29, PC-6, GSC-7, Tejashwani, DHM-
 137 44, Krishna and Super express were registered under moderately resistance while, 19 genotypes
 138 namely, PM-30, GSC-6, RLC-3, RCH-1, PGSH-1707, PHR-126, RH-749, Nirmala Bold, Basanti,
 139 Kalasona, Peelasona, Radha, KMH-721, 56-92, Dhakar, KMH-8765, Ganga, Lahar and DRMR-
 140 116540 were registered moderately susceptible category. In the current study, none of the assessed
 141 genotypes showed complete resistance to infection, aligning with Awasthi *et al.* (2012) findings that
 142 emphasized the susceptibility of key *B. juncea* varieties in India to white rust. The diverse responses
 143 of different genotypes to pathogens, varying in susceptibility, may stem from the complex interplay of
 144 resistance gene expression and the genetic background affecting genotype-pathogen interactions, as
 145 emphasized by Singh *et al.* (2021). The dynamics of how hosts and pathogens interact are
 146 significantly shaped by both macro and micro environmental factors, ultimately influencing the
 147 disease's severity, as discussed by Tamang *et al.* (2022).

148 **Table-2: Pooled data of rapeseed mustard cultivars against white rust:**

Pooled data of rapeseed-mustard cultivars against white rust 2022 and 23				
S. No.	Name of genotypes	Percent disease severity	Pooled AUDPC	Pooled r value
1	Giriraj	18.49 (25.45)	244.60	0.444
2	RH746	20.66 (27.01)	261.38	0.446
3	RH406	23.45 (28.94)	297.58	0.457
4	NRCHB101	22.18 (28.07)	292.15	0.479
5	PM29	24.02 (29.33)	317.21	0.472
6	PM30	25.77 (30.49)	346.58	0.470
7	PC6	21.53 (27.63)	279.40	0.470
8	GSC6	28.59 (32.30)	406.95	0.510
9	GSC7	21.34(27.49)	273.22	0.484
10	RLC3	25.83 (30.52)	352.53	0.489
11	RCH1	31.47 (34.10)	493.78	0.515
12	PGSH1707	29.68 (32.98)	456.26	0.478
13	PHR126	25.65 (30.39)	345.79	0.466
14	TEJASHWANI	20.40 (26.83)	256.50	0.444
15	DHM44	23.50 (28.98)	311.44	0.430
16	KRISHNA	21.35 (27.50)	273.26	0.463
17	RH749	27.44 (31.56)	370.75	0.463
18	NIRMLA BOLD	29.45 (32.84)	437.94	0.460
19	BASANTI	26.47 (30.94)	362.94	0.473
20	KALASONA	29.24 (33.45)	412.03	0.526
21	GANGA	30.28 (33.18)	466.53	0.478
22	PEELASONA	28.18 (32.05)	394.92	0.491
23	RADHA	27.57 (31.66)	381.75	0.479
24	KMH721	26.43 (30.92)	363.50	0.497

25	56-92	30.51 (33.51)	482.05	0.493
26	DHAKAR	27.68 (31.73)	384.98	0.479
27	KMH8765	25.49 (30.31)	339.13	0.453
28	LAHAR	29.25 (32.72)	426.33	0.490
29	DRMR116540	28.66 (32.35)	407.56	0.460
30	SUPER EXPRESS	22.30 (28.16)	298.42	0.430
	C.D.		1.322	
	SE(m)		0.466	
	SE(d)		0.659	
	C.V.		2.649	

*Figure in parenthesis are angular transformed value

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150 **Table-3: Response of different genotypes against white rust of crucifers**

151	Rating scale	Reaction	No. of genotypes	Name of genotypes
152	5	Moderately Resistant	11	Giriraj, RH-746, RH406, NRCHB-101, PM29, PC-6, GSC-7, Tejashwani, DHM- 44, Krishna, Super express.
157	7	Susceptible (S)	19	PM30, GSC-6, RLC-3, RCH-1, PGSH1707, PHR-126, RH-749, Nirmla Bold, Basanti, Kalasona, Peelasona, Radha, KMH-721, 56-92, Dhakar, KMH8765, Ganga, Lahar, DRMR116540

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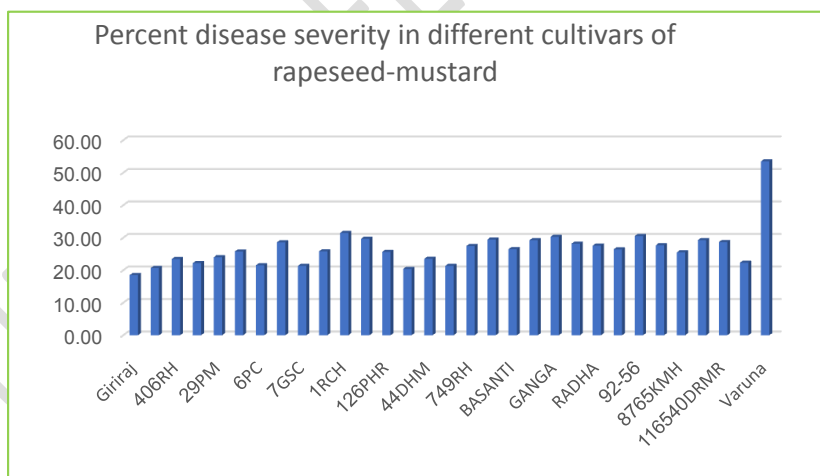
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179 **Fig-1: Percent disease severity in different cultivars of rapeseed-mustard**

181 4. CONCLUSION:

182 Germplasm lines of rapeseed-mustard have showed moderately resistant response in field
 183 screening trials against white rust disease while some of lines observed as a susceptible. These lines
 184 may be valuable in future breeding programs to develop resistant cultivars for commercial cultivation

185 in farmer's fields. To address evolving diseases, identifying various resistance genes in crop species
186 is crucial. India has recently made significant efforts to expand mustard cultivation in non-traditional
187 locations for diversifying the cropping system. Thus, maintaining access to donor parents with high
188 resistance against white rust disease is essential.

189 To achieve the goal, perform controlled lab tests on diverse genotypes for the disease. Confirm
190 resistance in a controlled laboratory with artificial inoculation and molecular markers tied to resistant
191 genes. Field testing is essential due to occasional disease escape. Hosts with more resistance genes
192 prevent new pathogen races, as the pathogen requires virulent genes to overcome host resistance.

193 REFERENCES

- 194 1. Ahmad, HN, Perveen, R, Chohan S, Yasmeen G, Mehmood MA, & Hussain W. Screening of
195 canola germplasm against *Albugo candida* and its epidemiological studies. *Pakistan Journal of*
196 *Phytopathology*. 2014; **26**(2): 169-173.
- 197 2. Anonymous. Directorate of Economics and Statistics, Ministry of Agriculture and Farmers
198 welfare. 2022; pp.260.
- 199 3. Awasthi RP, Nashaat NI, Kolte SJ, Tewari AK, Meena PD, Bhatt R. Screening of putative
200 resistant sources against Indian and exotic isolates of *Albugo candida* inciting white rust in
201 rapeseed-mustard. *J. Oilseed Brassica*. 2012; **1**: 27–37.
- 202 4. Bisht IS, Agrawal RC, Singh R. White rust (*Albugo candida*) severity in mustard (*Brassica juncea*)
203 varieties and its effects on seed yields. *Plant Varieties & Seeds*. 1994; **7**(2): 85-89.
- 204 5. Chand S, Singh N, Prasad L, Nanjundan J, Meena VK, Chaudhary R. Inheritance and allelic
205 relationship among Gene-gene (s) for white rust resistance in Indian Mustard-mustard [*Brassica*
206 *juncea* (L.) Czern & Coss]. *Sustainability*. 2022; **14**(18): 11620.
- 207 6. Conn KL, Tewari JP, Awasthi RP. A disease assessment key for Alternaria black spot in rapeseed
208 and mustard. *Canadian Plant Disease Survey*. 1990; **70**:19-22.
- 209 7. Dharavath N, Mehera, B, Nath S, Patra SS, Rout S. Effect of different sowing dates and
210 application of pesticides on growth and yield of mustard crop (*Brassica juncea*). *Int. J. Pure App.*
211 *Biosci*. 2017; **5**(1): 178-187.
- 212 8. Li CX, Sivasithamparam K, Walton G, Salisbury P, Burton W, Banga Surinder, Banga SS, Shashi
213 Chattopadhyay C, Kumar A, Singh R, Singh D, Agnohotri A, Liu SY, Li YC, Fu TD, Wang YF,
214 Barbetti MJ. Expression and relationships of resistance to white rust (*Albugo candida*) at
215 cotyledonary, seedling, and flowering stages in *Brassica juncea* germplasm from Australia,
216 China, and India. *Australian Journal of Agricultural Research*. 2007; **58**(3): 259–264.
- 217 9. Mukherjee AK, Mohapatra T, Varshney A, Sharma R and Sharma RP. Molecular mapping of a
218 locus controlling resistance to A-*Albugo candida* in Indian mustard. *Plant Breed* 2001; **120**: 483-
219 487.
- 220 10. Shaner G, Finney RE. The effect of nitrogen fertilization on the expression of slow mildewing
221 resistance in Knox wheat. *Phytopathology*. 1977; **67**:1051-1056.
- 222 11. Singh K, Akhtar J, Shekhawat N, Meena VS, Gupta A, Meena BR, Gupta V. Identification of new
223 source of resistance against white rust. *Journal of Oilseed Brassica*. 2022; **13**(2): 100-104.

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- 224 | 12. Singh OW, Singh N, Kamil D, Singh VK, Devi TP, Prasad L. Morpho-~~Molecular~~ ~~molecular~~
225 | ~~Variability~~ ~~variability~~ and ~~Host~~ ~~host~~ ~~Reactivity~~ ~~reactivity~~ of *Albugo candida* ~~Isolates~~ ~~isolates~~
226 | ~~Infecting~~ ~~infecting~~ *Brassica juncea* ~~Genotypes~~ ~~genotypes~~ in India. J. Plant Pathol. 2021: **103**:
227 | 139–153.
- 228 | 13. Tamang S, Saha P, Bhattacharya S, Das A. Unveiling ~~Genotype~~ ~~genotype~~ × ~~Environment~~
229 | ~~environment~~ ~~Interactions~~ ~~interactions~~ towards ~~Identification~~ ~~identification~~ of ~~Stable~~ ~~stable~~ ~~Sources~~
230 | ~~sources~~ of ~~Resistance~~ ~~resistance~~ in ~~Chickpea~~ ~~chickpea~~ ~~Collar~~ ~~collar~~ ~~Rot~~ ~~rot~~ ~~Pathosystem~~
231 | ~~pathosystem~~ ~~Exploiting~~ ~~exploiting~~ GGE ~~Biplot~~ ~~biplot~~ ~~Technique~~ ~~technique~~. Australas. Plant Pathol.
232 | 2022: **51**: 47–58.
- 233 | 14. Vander-plank ~~J.P.J.E.~~ Plant disease: Epidemics and controls. Academic Press, New York, 349 p.
234 | 1963.

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