

Original Research Article

GC-MS Analysis and Biological Roles of Phytochemical Compounds in n-Hexane Extract of *Durio zibethinus* Murr. Seeds

ABSTRACT

Aim: The current study was to assess the phytochemical compounds in *Durio zibethinus* seeds that can be used for medicinal and industrial purposes.

Methodology: The n-hexane extract of the seeds was analyzed using Gas Chromatography–Mass Spectrometry. The Gas Chromatography–Mass Spectrometry (GC-MS) analysis of n-hexane extract from *D. zibethinus* seeds revealed the presence of twelve compounds.

Results: The chemical constituents identified are beta-Guaiene (0.85 %), Hexadecanoic acid, methyl ester (3.82 %), 9,12-Octadecadienoic acid, methyl ester (1.26 %), Phytol (1.36 %), Methyl stearate (2.08 %), Lupeol (1.97 %), n-Tetracosanol-1 (17.34 %), Squalene (32.75 %), Triacontanediol (34.47 %), 1-Heptacosanol (1.89 %), Eicosane (0.88 %) and Hexadeca-2,6,10,14-tetraen-1-ol, 3,7,11,16-tetramethyl- (29.34 %).

Conclusion: The presence of these chemical constituents in the seed extract has authenticated the scientific evidences for its physiological properties relating to human health.

Keywords: *Durio zibethinus* Murr.; Phytochemicals; GC-MS analysis; Biological activities

1. INTRODUCTION

The adoption of medicinal plants as a source of possible medications has become incredibly successful. Most plants are extremely helpful in the drug development process since they naturally create chemical compounds or molecules with pharmacological or bioactive qualities [1]. The broad category of "medicinal plants" includes a wide range of botanical species that produce compounds that are beneficial to human fitness and wellbeing. In the past, the creation of pharmaceutical medications for diseases including cancer, infections, digestive problems, diabetes, and hypertension has benefited significantly from using medicinal plants [2]. Different parts of plants, such as leaves, fruits, roots and seeds have been recognized for their medicinal activities [3].

A tropical fruit plant called durian (*Durio zibethinus* Murr.; Family Bombacaceae) is grown in Malaysia and other Southeast Asian countries. Based on its appearance like the thorny thrones of Asian kings and its high nutritional status, it is referred to as the "King of Tropical Fruit" [4]. Durian is said to have anti-oxidant, anti-cancer, anti-heart disease, anti-diabetic, and anti-obesity properties [5]. The capacity to enhance the immune system is one of the purported medicinal and therapeutic benefits of durian fruit. Its fruit pulp maybe a great source of nutrients like proteins, carbohydrates, fibers and dietary fat [6]. It is also disclosed

that durian seed, pulp and peel flour possess nutritional, structural, antioxidant and anti-inflammatory properties [7]-[8].

Natural chemical substances (phytochemicals) are essential for promoting health benefits. [9]. Considering their roles in plant metabolism, phytochemicals can be categorized as primary or secondary metabolites. For plants to grow and develop, primary metabolites such as sugars, amino acids, proteins, nucleic acids' purines and pyrimidines, and chlorophylls are essential. Moreover, in addition to safeguarding plants from environmental hazards including pollution, stress, UV exposure, and illnesses, they also contribute to the colour, aroma, and flavour of plants [10]. As opposed to that, secondary metabolites such as carotenoids, phytosterols, alkaloids, terpenes and polyphenols (including flavonoids, phenolic acids, tannins, stilbenes, coumarins, and lignans) have different roles in a plant's environment, including luring pollinators and functioning as natural defenses against pathogens and predators [11]-[12]. Therefore, this study uses Gas Chromatography-Mass Spectrometry (GC-MS) to thoroughly investigate the bioactive compounds found in the n-hexane extract of *Durio zibethinus* Murr. seeds, helping to further our understanding of the chemical makeup of this less-studied area of the *Durio zibethinus* Murr. plant. There are attempts to study the biological activities of the detected bioactive compounds present in the n-hexane extract of *Durio zibethinus* Murr. seeds.

2.MATERIALS AND METHODS

2.1Sample Collection and Authentication

The plant materials, *Durio zibethinus* Murr. seeds were collected at Crown Estate of Igbinedion University, Okada, Edo State, Nigeria in the month of April, 2023. At the Taxonomy section, Biological Sciences Department, Igbinedion University, Okada, Nigeria, the seeds samples were identified and authenticated. The Voucher No. IUOH/001/1371 was assigned.

2.2Preparation and Extraction of Plant Materials

The *Durio zibethinus* Murr. seeds were washed and air-dried, the seed coats were carefully peeled off manually from the seeds. These seeds were appropriately ground into little particles using a mortar and pestle and stored in a polythene bag. By soaking 2.0 kg of the crushed seeds in 10.0 L of methanol at ambient temperature for 72 hours, the extract was obtained. The extracts were filtered with Whatmann filter paper. The extract was separated from the solvent using a Soxhlet apparatus. The resulting extract was stored in an air-tight container in a refrigerator until it was required for GC-MS analysis [13].

2.3GC-MS Analysis

Instruments and Chromatographic Conditions

The GC-MS analysis was performed on an Agilent Technologies interfaced [Model: 7890A (GC)] with Mass Selective Detector model: 5975C (MSD). The electron ionization was maintained at 7 eV with an ion source temperature stationed at 250 °C. Highly pure helium gas (99.9 % purity) was utilized as carrier gas, while HP-5ms (30mm X 0.25mm X 0.320µm) was utilized as the stationary phase. The oven temperature was programmed to begin with a temperature of 140 °C, held for 5 minutes at 4 °C per minute and allowed to reach a

maximum temperature of 240 °C for 15 minutes, thereafter the temperature was maintained for another 6 minutes at the rate of 3.5 °C/minute.

2.4 Identification of Phytochemical Compounds

Identification of phytochemical compounds and interpretation of mass spectrum GC-MS was conducted using the National Institute of Standard and Technology (NIST) database which contain more than 62,000 patterns. This was followed by comparing the spectrum of unknown component with the spectrum of the known component using computer searches on a NIST Ver.2.1 MS data library. Hence process establishes the name, structure and molecular weight of the components of the test materials

3. RESULTS

Table 1: GC-MS analysis of the n-hexane extract from *D. zibethinus* seeds

Number	Retention Time (min)	Name of the Compound	Molecular Formula	Composition (%)	Mass Spectral Data
1	114.684	Beta-Guaiene	C ₁₅ H ₂₄	0.85	119,133,147, 161 ,175
2	17.498	Hexadecanoic acid, methyl ester	C ₁₇ H ₃₄ O ₂	3.82	15,27,41,57, 74
3	19.88	19,12-Octadecadienoic acid (ZZ), methyl ester	C ₁₉ H ₃₄ O ₂	1.26	25,27,41,55, 67
4	20.262	Phytol	C ₂₀ H ₄₀ O	1.36	27,41,57, 71 ,95
5	20.382	Methyl stearate [Octadecanoic acid, methyl ester]	C ₁₉ H ₃₈ O ₂	2.08	25,41,43,57, 74
6	26.535	Lupeol	C ₃₀ H ₅₀ O	1.97	41, 43 ,68,81,95
7	27.437	n-Tetracosanol-1	C ₂₄ H ₅₀ O	17.34	27,41, 55 ,83,97
8	29.351	Squalene	C ₃₀ H ₅₀	32.75	27,41,55, 69 ,81
9	30.234	51,30-Triacontanediol	C ₃₀ H ₆₂ O ₂	34.47	25,41, 55 ,69,82
10	30.348	1-Heptacosanol	C ₂₇ H ₅₆ O	1.89	27,41, 55 ,83,97
11	30.635	Eicosane	C ₂₀ H ₄₂	0.88	27,41,43, 57 ,71
12	31.054	Hexadeca-2,6,10,14-tetraen-1-ol, 3,7,11,16-tetramethyl-	C ₂₀ H ₃₄ O	1.34	14,27, 41 ,55,69

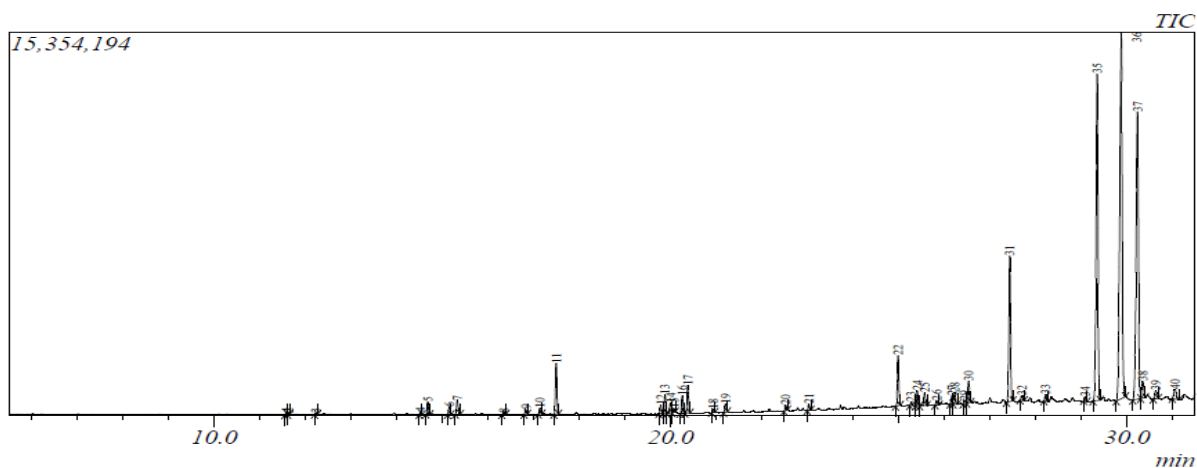


Figure1: GC-MS Chromatogram of n-hexane extract of *D. zibethinus* seeds

The mass spectra of the compounds with the highest percentage peak areas were also represented in *Figures2-13* respectively.

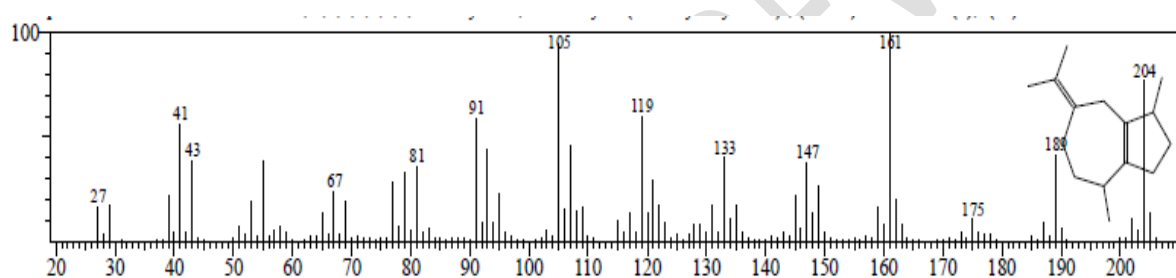


Figure2: Mass spectrum of Beta-Guaiene (RT: 14.684, Area % = 0.85) from *D. zibethinus* seeds

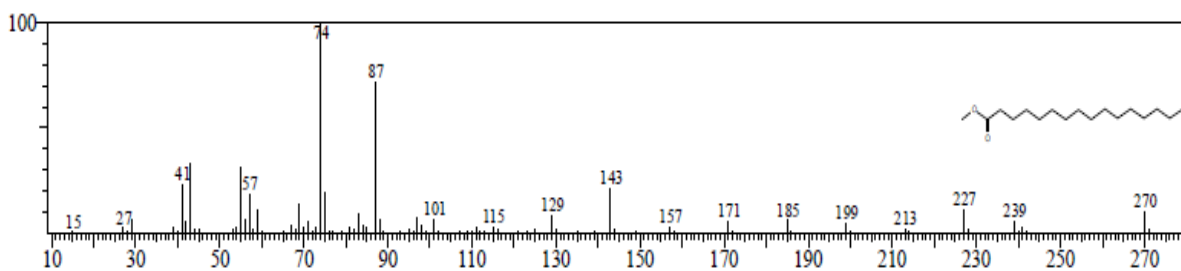


Figure3: Mass spectrum of Hexadecanoic acid, methyl ester (RT: 17.498, Area % = 3.82) from *D. zibethinus* seeds

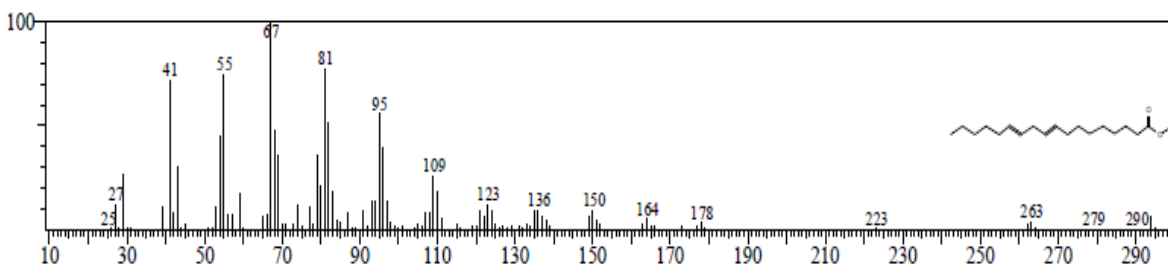


Figure4: Mass spectrum of 9,12-Octadecadienoic acid, methyl ester (RT: 19.881, Area % = 1.26) from *D. zibethinus* seeds

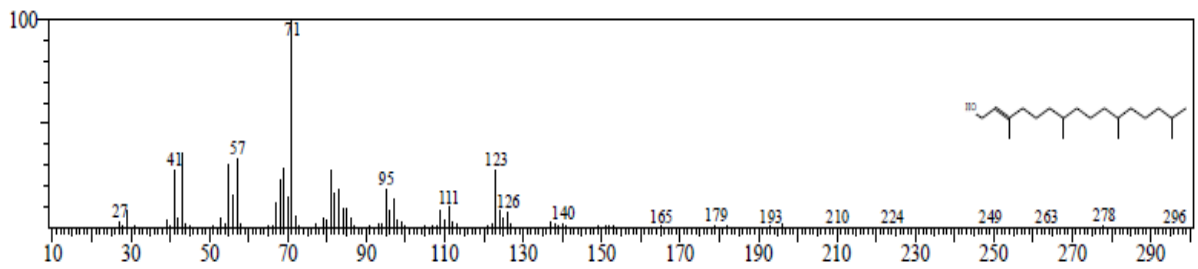


Figure5: Mass spectrum of Phytol (RT: 20.262, Area % = 1.36) from *D. zibethinus* seeds

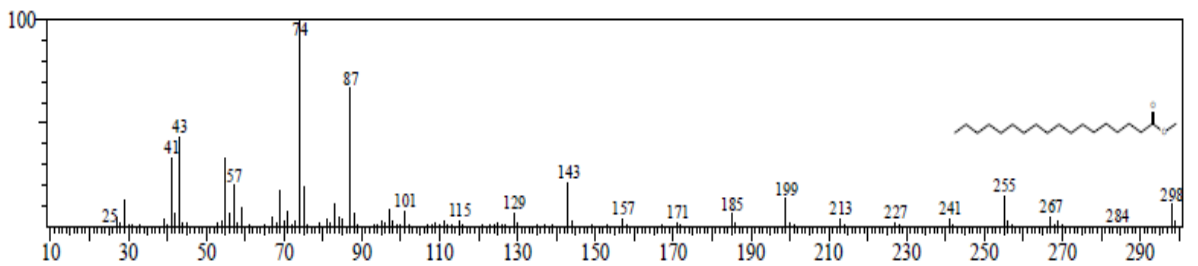


Figure6: Mass spectrum of Methyl stearate (RT: 20.382, Area % = 2.08) from *D. zibethinus* seeds

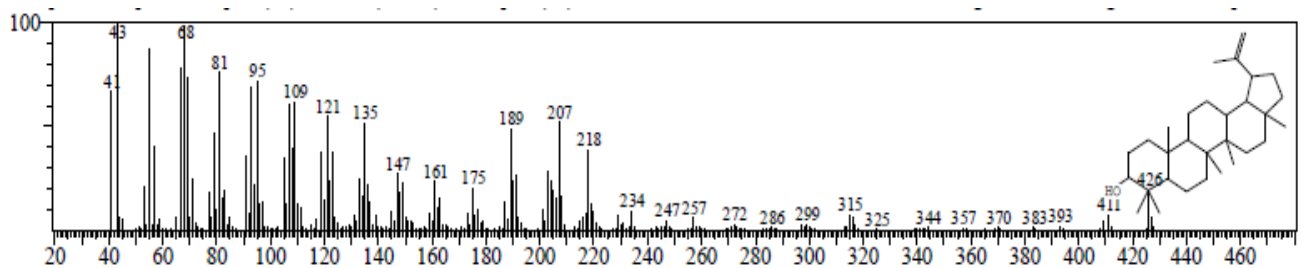


Figure7: Mass spectrum of Lupeol (RT: 26.535, Area % = 1.97) from *D. zibethinus* seeds

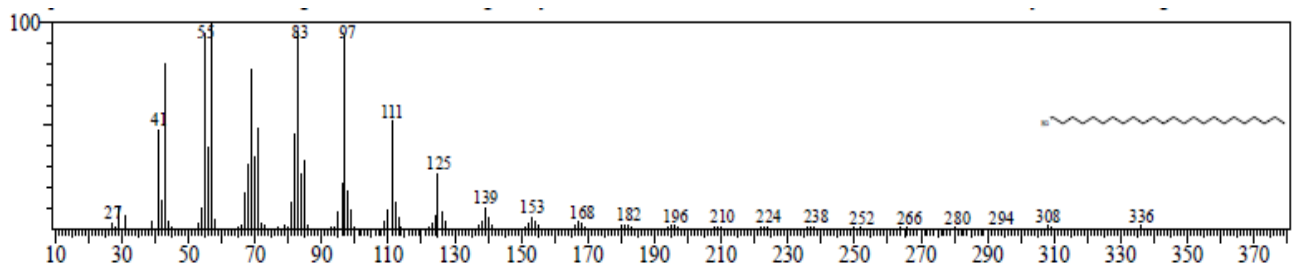


Figure8: Mass spectrum of n-Tetracosanol-1 (RT: 27.437, Area % = 17.34) from *D. zibethinus* seeds

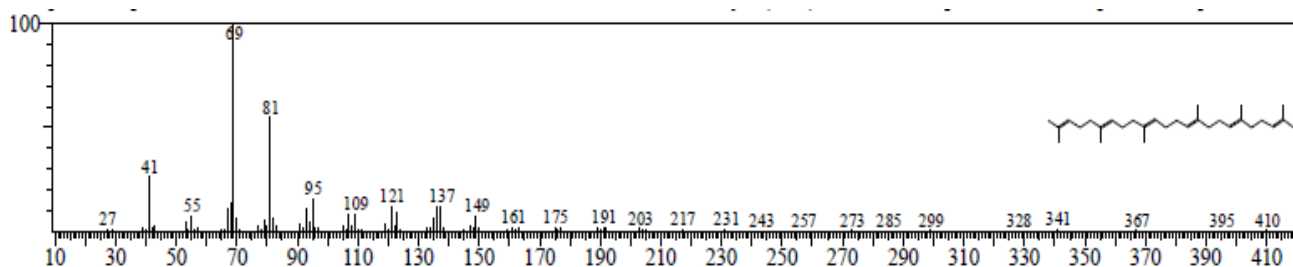


Figure9: Mass spectrum of Squalene (RT: 29.351, Area % = 32.75) from *D. zibethinus* seeds

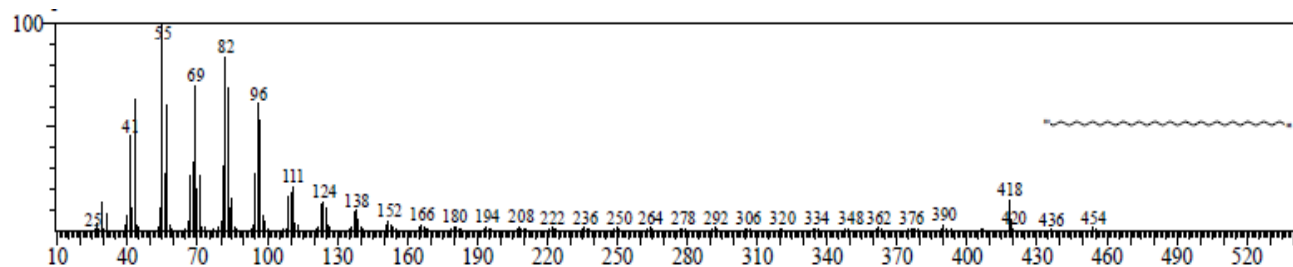


Figure10: Mass spectrum of 1,30-Triacontanediol (RT: 30.234, Area % = 34.47) from *D. zibethinus* seeds

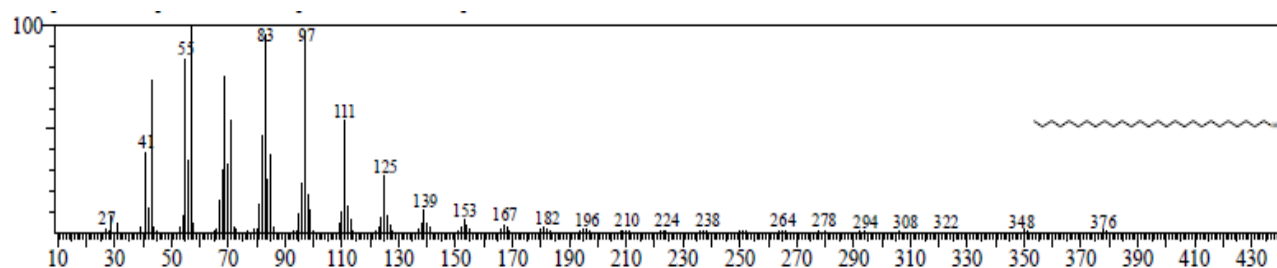


Figure11: Mass spectrum of 1-Heptacosanol (RT: 30.348, Area % = 1.89) from *D. zibethinus* seeds

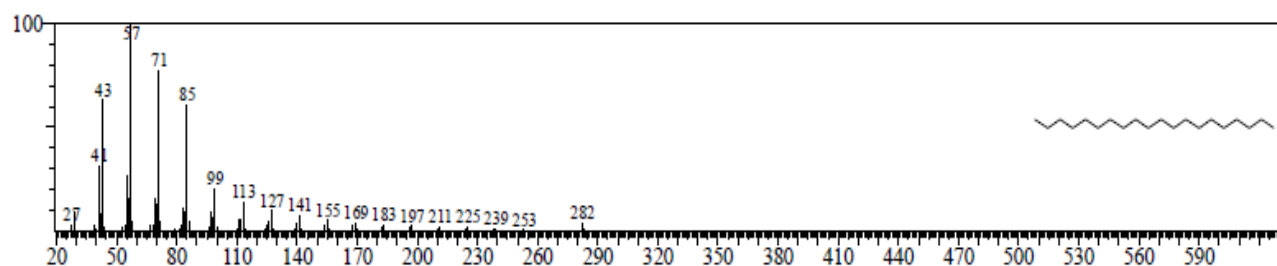


Figure12: Mass spectrum of Eicosane (RT: 30.635, Area % = 0.88) from *D. zibethinus* seeds

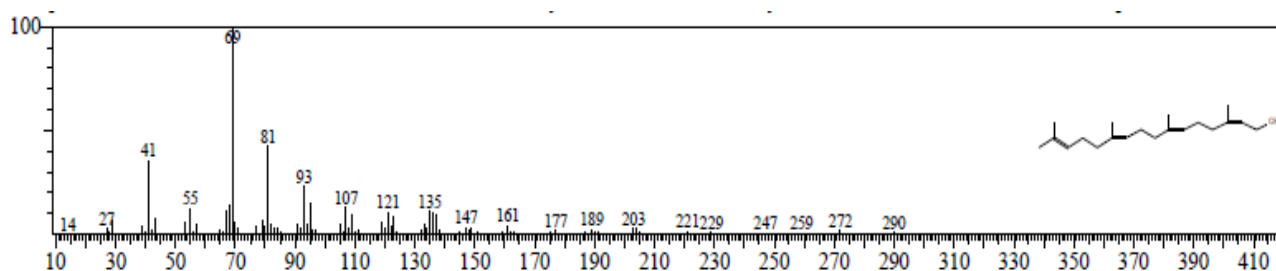


Figure13: Mass spectrum of Hexadeca-2,6,10,14-tetraen-1-ol, 3,7,11,16-tetramethyl- (RT: 31.054, Area % = 1.34) from *D. zibethinus* seeds

4. DISCUSSION

4.1 GC-MS Analysis

GC-MS is one of the leading techniques to identify the constituents of volatile matter, long chain, branched chain hydrocarbons, alcohols, acids, esters etc. Twelve compounds (phytochemical constituents) were identified in the n-hexane extract of *D. zibethinus* seeds through GC-MS analysis. The chromatogram peaks were integrated and thereafter compared with the database of spectrum of known compounds stored in the GC-MS NIST library. The chemical constituents identified are beta-Guaiene (0.85%), Hexadecanoic acid, methyl ester (3.82%), 9,12-Octadecadienoic acid, methyl ester (1.26 %), Phytol (1.36 %), Methyl stearate (2.08 %), Lupeol (1.97 %), n-Tetracosanol-1 (17.34 %), Squalene (32.75 %), Triacantanediol (34.47 %), 1-Heptacosanol (1.89 %), Eicosane (0.88 %) and Hexadeca-2,6,10,14-tetraen-1-ol, 3,7,11,16-tetramethyl- (29.34 %).

Figure 1 shows the GC-MS chromatogram of the peak of the compounds detected. The fragmentation pattern and molecular formula were used to confirm the identification of the phytochemical compounds. The retention time (RT), percentage composition and mass spectral data of active principles are presented in *Table 1*.

4.2 Biological Activities of the Phytochemicals Detected through GC-MS Analysis

The biological roles of phytochemicals in human metabolism has long been established. Phytochemicals are very active constituents and are abundant in nature. They are grouped and have significant roles in preventing various diseases. The use of these phytochemicals is done in a combination of multiple phytochemicals and other drugs as well [14, 15].

Phytochemicals exhibit a wide range of therapeutic roles, including antioxidant, anti-inflammatory, anti-diabetic, analgesic, anti-cancer, neuroprotective, and anti-microbial activities [16,17,18,19]. Most phytochemical constituents identified in *D. zibethinus* seeds are terpenoids, fatty acid esters, terpene alcohols, triterpenoids, and fatty alcohols that contribute to the antibacterial, antifungal, antioxidant, anti-inflammatory activities etc. Among the phytochemicals identified in *D. zibethinus* seed oil are as follows:

Beta-Guaiene (**1**) is a sesquiterpenoid. It has a bicyclic structure with a cyclohexene ring and a cyclopropane ring. Beta-Guaiene has been shown to exhibit various biological activities, including antimicrobial, anti-inflammatory, and anticancer activities [20].

Hexadecenoic acid methyl ester (**2**) is a saturated fatty acid methyl ester that shows antioxidant, antitumor, immunostimulant, chemopreventive and lipoxygenase inhibitory, antimicrobial activities and it is a potent mosquito larvicide [21]. 9,12-octadecadienoic acid

(Z,Z)-, methyl ester (**3**) is also a saturated fatty acid methyl ester that shows antioxidant [22] and anti-inflammatory activities [23,24]. Methyl stearate or Octadecanoic acid, methyl ester (**5**) is another saturated fatty acid methyl ester identified in *D. zibethinus* seed oil. Mazumder et al. [25] have reported that methyl stearate or octadecanoic acid, methyl ester has anti-inflammatory, intestinal lipid metabolism regulation, nematocidal, antinociceptive, antioxidant, antimicrobial and antifungal activities.

Phytol (**4**) is a diterpene member of the long-chain unsaturated acyclic alcohols and exert a wide range of biological effects. It has been reported that phytol is a potential candidate for a broad range of applications in the pharmaceutical and biotechnological industry. Recent investigations with phytol demonstrated anxiolytic, metabolism-modulating, cytotoxic, antioxidant, autophagy- and apoptosis-inducing, antinociceptive, anti-inflammatory, anti-cancer, immune-modulating, and antimicrobial effects [26,27,28]. Another diterpenoid alcohol detected in *D. zibethinus* seed oil was Hexadeca-2,6,10,14-tetraen-1-ol, 3,7,11,16-tetramethyl- (**12**). It has been reported that Hexadeca-2,6,10,14-tetraen-1-ol, 3,7,11,16-tetramethyl- shows antimicrobial, diuretic and anti-inflammatory activities [29,30].

Lupeol is a pentacyclic triterpenoid that shows anti-malarial, anti-microbial, anti-angiogenic, as well as anti-inflammatory and antiarthritic activities. Lupeol has also been reported as an effective higher blood-brain barrier permeability [31]. Lupeol inhibits neuroinflammation in cerebellar cultures and induces neuroprotection associated with the modulation of astrocyte response and expression of neurotrophic and inflammatory factors [32].

The presence of important alcoholic compound detected in *D. zibethinus* seed oil was n-Tetracosanol-1 (**7**). This is a fatty acid derivative of lignoceric acid that shows antimicrobial, antioxidant, anticancer, antimutagenic and nematocidal activities [33]. It also lowers cholesterol and enhances immune functions [34]. n-Tetracosanol-1 is one of the constituents of policosanol which modifies several cardiovascular disease risk factors by reducing LDL oxidation, platelet aggregation and endothelial cell damage [35]. Another alcoholic compound detected in *D. zibethinus* seed oil was 1-Heptacosanol (**10**). Researchers have observed that 1-heptacosanol has antimicrobial and antioxidant activities [36,37].

Squalene (**8**) belongs to the class of triterpene hydrocarbon, and is known to be a biochemical precursor for all steroids in plants and animals. It has several applications in the food, pharmaceutical, and medical sectors. It is essentially used as a dietary supplement, vaccine adjuvant, moisturizer, cardio-protective agent, anti-tumor agent and antioxidant [38].

Squalene has been reported to help in the synthesis of cholesterol, effective as skin emollient, scavenging free radicals, and inhibit the fungal growth [39].

Eicosane (**11**) is a long-chain hydrocarbon and has been reported for its antibacterial, anti-inflammatory, analgesic, and antipyretic activity [40,41,42,43]. In addition, eicosane may promote wound healing due to their potent free radical scavenging, hydroxyproline and glutathione action (antioxidant) [44,45]. From GC-MS Analysis results, it is obvious that *D. zibethinus* seed accumulates essential phytochemicals, hence, the essential oil extracted from the seed of *D. zibethinus* Murr. can be used in developing new leads of therapeutic interest.

5. CONCLUSION

In conclusion, the bioactive phytochemicals found in *D. zibethinus* Murr. seeds are abundant and essential for maintaining human health and wellbeing. According to these studies, the

essential oil extracted from the seeds of *D. zibethinus* Murr. may be useful in the formulation of nutritional and/or therapeutic products. This research has contributed to the global effort to encourage the usage of herbal drugs from natural sources which has been generally accepted as safer drugs to synthetic ones.

REFERENCES

- 1 Kaloudas D. and Penchovsky R. Plant-Derived Compounds and Their Potential Role in Drug Development. *Research Anthology on Recent Advancements in Ethnopharmacology and Nutraceuticals*. 2022: 502–517.
DOI: 10.4018/978-1-6684-3546-5.ch026.
- 2 Priya S. and Satheeshkumar PK. Natural Products from Plants: Recent Developments in Phytochemicals, Phytopharmaceuticals, and Plant-Based Nutraceuticals as Anticancer Agents. *Functional and Preservative Properties of Phytochemicals*. 2020: 145–163.
<https://doi.org/10.1016/B978-0-12-818593-3.00005-1>
- 3 Dzobo K. The Role of Natural Products as Sources of Therapeutic Agents for Innovative Drug Discovery. *Comprehensive Pharmacology*. 2022: 2022: 408–422.
doi: [10.1016/B978-0-12-820472-6.00041-4](https://doi.org/10.1016/B978-0-12-820472-6.00041-4)
- 4 Husin NA, Bhore SJ, Rahman S. and Karunakaran R. A Review on the Nutritional, Medicinal, and Genome Attributes of Durian (*Durio zibethinus* L.), the King of Fruits in Malaysia. *Bioinformation*. 2018;14(6): 265-270. DOI: [10.6026/97320630014265](https://doi.org/10.6026/97320630014265)
- 5 Siburian R, Aruan DGR, Barus T, Haro G. and Simanjuntak P. Phytochemical Screening and Antidiabetic Activity of N-hexane, Ethyl acetate and Water Extract from Durian Leaves (*Durio zibethinus* L.). *Oriental Journal of Chemistry*. 2019;35(1):487-490. <http://dx.doi.org/10.13005/ojc/350166>
- 6 Adeniyi SA, Oke VJ. and Olatunji GA. GC-MS Analysis of Phytochemical Constituents in Methanol Extract of Wood Bark from *Durio zibethinus* Murr. *International Journal of Medicinal Plants and Natural Products*. 2019;5(3):1-11.
<http://dx.doi.org/10.20431/2454-7999.0503001>
- 7 Charoenphun N. and Klangbud WK. Antioxidant and Anti-inflammatory Activities of Durian (*Durio zibethinus* Murr.) Seed, Pulp and Peel Flour. *PeerJ*. 2022;10: e12933.
<https://doi.org/10.7717/peerj.12933>
- 8 Permatasari ND, Witoyo JE, Masruri M, Yuwono SS. and Widjanarko SB. Nutritional and Structural Properties of Durian Seed (*Durio Zibethinus* Murr.) Flour Originated from West Kalimantan, Indonesia. *IOP Conference Series: Earth and Environmental Science*. 2022: 1012: 012038, DOI 10.1088/1755-1315/1012/1/012038
- 9 Begum VSM, Tariq NPM, Tariq MJH. and Shariq KM. Plants Secondary Metabolites as Medicines: A Review. *International Journal of Zoological Investigations*. 2022;08(01): 490–493. <https://doi.org/10.33745/ijzi.2022.v08i01.056>
- 10 Sokan-Adeaga AA, Sokan-Adeaga MA, Sokan-Adeaga ED, Oparaji AN, Edris H, Tella EO, Balogun FA, Aledeh M. and Amubieya OE. Environmental Toxicants and Health Adversities: A Review on Interventions of Phytochemicals. *Journal of Public Health Research*. 2023;12(2):22799036231181226. doi: [10.1177/22799036231181226](https://doi.org/10.1177/22799036231181226)
- 11 Nedjimi B. Phytoremediation: A Sustainable Environmental Technology for Heavy

- Metals Decontamination. *SN Applied Sciences*. 2021; 3: 286.
<https://doi.org/10.1007/s42452-021-04301-4>
- 12 Agidew MG. Phytochemical Analysis of Some Selected Traditional Medicinal Plants in Ethiopia. *Bull. Nat. Res. Centre*. 2022;46(1): 87.
<https://doi.org/10.1186/s42269-022-00770-8>
- 13 Nwafor MN, Anosike AC, Onodugo CA, Ya'u M. and Babandi A. Antioxidant, Phytochemical Composition and Proximate Analysis of *Mangifera haden* Seeds. *International Journal of Scholarly Research in Chemistry and Pharmacy*. 2022;01(01): 011-023. <https://doi.org/10.56781/ijsrcp.2022.1.1.0022>
- 14 Ding Y, Hou R, Yu J, Xing C, Zhuang C. and Qu Z. Dietary Phytochemicals as Potential Chemopreventive Agents against Tobacco-Induced Lung Carcinogenesis. *Nutrients*. 2023;15: 491. doi: 10.3390/nu15030491.
- 15 Mandal MK, Mohammad M, Parvin SI, Islam MM, Gazi HAR, Alberto AKM, da Costa MJ. and Carvalho JCT. A Short Review on Anticancer Phytochemicals. *Pharmacognosy Reviews*. 2023;17:11–23. doi: 10.5530/097627870236.
- 16 Anwar S, Shamsi A, Shahbaaz M, Queen A, Khan P, Hasan GM, Islam A, Alajmi M.F, Hussain A, Ahmad F. and Hassan MI. Rosmarinic acid Exhibits Anticancer Effects via MARK₄ Inhibition. *Scientific Reports*. 2020;10:1–13.
doi: 10.1038/s41598-020-65648-z.
- 17 Anwar S, Khan S, Anjum F, Shamsi A, Khan P, Fatima H, Shafie A, Islam A. and Hassan MI. Myricetin Inhibits Breast and Lung Cancer Cells Proliferation via Inhibiting MARK₄. *Journal of Cellular Biochemistry*. 2022;123:359–374.
doi: 10.1002/jcb.30176.
- 18 Divekar PA, Narayana S, Divekar BA, Kumar R, Gadratagi BG, Ray A, Singh AK, Rani V, Singh V, Singh AK, Kumar A, Singh RP, Meena RS and Behera TK. Plant Secondary Metabolites as Defense Tools Against Herbivores for Sustainable Crop Protection. *International Journal of Molecular Sciences*. 2022;23:2690.
doi: 10.3390/ijms23052690.
- 19 Yeshi K, Crayn D, Ritmejeriyè E. and Wangchuk P. Plant Secondary Metabolites Produced in Response to Abiotic Stresses has Potential Application in Pharmaceutical Product Development. *Molecules*. 2022;27: 313. doi: 10.3390/molecules27010313.
- 20 Safriansyah W, Sinaga SE, Supratman U. and Harneti D. Phytochemistry and Biological Activities of *Guarea* Genus (Meliaceae). *Molecules*. 2022; 27: 8758.
<https://doi.org/10.3390/molecules27248758>
- 21 Shaaban MT, Ghaly MF. and Fahmi SM. Antibacterial Activities of Hexadecanoic acid methyl ester and Green-Synthesized Silver Nanoparticles Against Multidrug-Resistant Bacteria. *Journal of Basic Microbiology*. 2021;61(6): 557-568.
- 22 Elwekeel A, Hassan MHA, Almutairi E, Alhammad M, Alwhbi F, Abdel-Bakky MS, Amin E. and Mohamed EIA. Anti-Inflammatory, Anti-Oxidant, GC-MS Profiling and Molecular Docking Analyses of Non-Polar Extracts from Five *Salsola* Species. *Separations*. 2023; 10: 72-86. <https://doi.org/10.3390/separations10020072>
- 23 Janbaz K, Aslam N, Imran I. and Jabeen Q. Evaluation of Anti-inflammatory, Analgesic and Antipyretic Activities of *Salsola imbricata* Forssk in Rats. *Journal of Animal and Plant Sciences*. 2021; 31: 862–867.

- 24 Mohammed HA, Al-Omar MS, Mohammed SA, Alhowail AH, Eldeeb HM, Sajid MS, Abd-Elmoniem EM, Alghulayqeh OA, Kandil YI. and Khan RA. Phytochemical Analysis, Pharmacological and Safety Evaluations of Halophytic Plant, *Salsola Cyclophylla*. *Molecules*. 2021; 26: 2384.
- 25 Mazumder K, Nabila A, Aktar A. and Farahnaky A. Bioactive Variability and *In Vitro* and *In Vivo* Antioxidant Activity of Unprocessed and Processed Flour of Nine Cultivars of Australian *lupin* Species: A Comprehensive Substantiation. *Antioxidants*. 2020; 9(4): 282. <https://doi.org/10.3390/antiox9040282>
- 26 Islam MT, Ali ES, Uddin SJ, Shaw S, Islam MA, Ahmed MI, Chandra Shill M, Karmakar UK, Yarla NS, Khan IN, Billah MM, Pieczynska MD, Zengin G, Malainer C, Nicoletti F, Gulei D, Berindan-Neagoe I, Apostolov A, Banach M, Yeung AWK, El-Demerdash A, Xiao J, Dey P, Yele S, Jóźwik A, Strzałkowska N, Marchewka J, Rengasamy KRR, Horbańczuk J, Kamal MA, Mubarak MS, Mishra SK, Shilpi JA. and Atanasov AG. Phytol: A Review of Biomedical Activities. *Food and Chemical Toxicology*. 2018; 121: 82-94.
doi: 10.1016/j.fct.2018.08.032. Epub 2018 Aug 18. PMID: 30130593.
- 27 Bobe G, Zhang Z, Kopp R, Garzotto M, Shannon J. and Takata Y. Phytol and its Metabolites Phytanic and Pristanic acids for Risk of Cancer: Current Evidence and Future Directions. *European Journal of Cancer Prevention*. 2020; 29(2): 191-200.
doi: 10.1097/CEJ.0000000000000534. PMID: 31436750; PMCID: PMC7012361.
- 28 Anoor PK, Yadav AN, Rajkumar K, Kande R, Tripura C, Naik KS. and Burgula S. Methanol Extraction Revealed Anticancer Compounds Quinic Acid, 2(5H)-Furanone and Phytol in *Andrographis paniculate*, *Molecular and Clinical Oncology*. 2022; 17(151): 13. <https://doi.org/10.3892/mco.2022.2584>
- 29 Erdoğan MK, Geçibesler İH. and Behçet L. Chemical Constituents, Antioxidant, Antiproliferative and Apoptotic Effects of a New Endemic *Boraginaceae* species: *Paracaryumbingoelianum*. *Results in Chemistry*. 2: 100032.
<https://doi.org/10.1016/j.rechem.2020.100032>
- 30 Sadeghi H, Rostamzadeh D, Kokhdan EP, Asfaram A, Doustimotlagh AH, Hamidi N. and Hossein S. *Stachys pilifera* Benth: A Review of Its Botany, Phytochemistry, Therapeutic Potential, and Toxicology. *Evidence-Based Complementary and Alternative Medicine*. 2022; 2022: 9. <https://doi.org/10.1155/2022/7621599>
- 31 Park JS, Rehman IU, Choe K, Ahmad R, Lee HJ. and Kim MO. A Triterpenoid Lupeol as an Antioxidant and Anti-Neuroinflammatory Agent: Impacts on Oxidative Stress in Alzheimer's Disease. *Nutrients*. 2023; 15(13): 3059.
doi: 10.3390/nu15133059. PMID: 37447385; PMCID: PMC10347110.
- 32 Oliveira-Junior MS, Pereira EP, de Amorim VCM, Reis LTC, do Nascimento RP, da Silva VDA, Costa SL. Lupeol Inhibits LPS-Induced Neuroinflammation in Cerebellar Cultures and Induces Neuroprotection Associated to the Modulation of Astrocyte Response and Expression of Neurotrophic and Inflammatory Factors. *International Immunopharmacology*. 2019; 70: 302-312.
<https://doi.org/10.1016/j.intimp.2019.02.055>
- 33 Kumari N. and Menghani E. Evaluation of Antibacterial Activity and Identification of

- Bioactive Metabolites by GCMS Technique from *Rhizosphaeractinomycetes*. Indian Journal of Natural Products and Resources. 11(4): 287-294.
- 34Heng YW, Ban JJ, Khoo KS. and Sit NW. Biological Activities and Phytochemical Content of the Rhizome Hairs of *Cibotium barometz* (Cibotiaceae). Industrial Crops and Products. 2020;153: 112612. <https://doi.org/10.1016/j.indcrop.2020.112612>
- 35Godara P, Dulara BK, Barwer N. and Chaudhary NS. Comparative GC–MS Analysis of Bioactive Phytochemicals from Different Plant Parts and Callus of *Leptadenia reticulata* Wight and Arn. Pharmacognosy Journal. 2019;11(1): 129-40.
- 36Vambe M, Aremu AO, Chukwujekwu JC, Gruz J, Luterová A, Finnie JF. and Van Staden J. Antibacterial, Mutagenic Properties and Chemical Characterization of Sugar Bush (*Protea caffra* Meisn.): A South African Native Shrub Species. Plants. 2020; 9(10): 1331. <https://doi.org/10.3390/plants9101331>
- 37Sánchez-Hernández E, Buzón-Durán L, Lorenzo-Vidal B, Martín-Gil J. and Martín-Ramos P. Physicochemical Characterization and Antimicrobial Activity against *Erwinia amylovora*, *Erwinia vitivora*, and *Diplodiaseriata* of a Light Purple *Hibiscus syriacus* L. Cultivar. Plants (Basel). 2021; 10(9):1876. doi: 10.3390/plants10091876.
- 38Paramasivan K. and Mutturi S. Recent Advances in the Microbial Production of Squalene, World Journal of Microbiology and Biotechnology. 2022: 38-91. <https://doi.org/10.1007/s11274-022-03273-w>
- 39Siti FY, Ismail SI, Farah FH. and Mahmud TMM. Phytochemical Composition in Hexane and Methanolic Leaf Extract of *Vernonia amygdalina*. Malaysian Applied Biology. 48(5): 11–17.
- 40Naeim H, El-Hawiet A, Abdel Rahman RA, Hussein A, El Demellawy MA. and Embaby AM. Antibacterial Activity of *Centaurea pumilio* L. Root and Aerial Part Extracts Against Some Multidrug Resistant Bacteria. BMC Complementary Medicine and Therapies. 2020; 20: 79. <https://doi.org/10.1186/s12906-020-2876-y>
- 41Okechukwu PN. “Evaluation of Anti-inflammatory, Analgesic, Antipyretic Effect of Eicosane, Pentadecane, Octacosane, and Heneicosane”. Asian Journal of Pharmaceutical and Clinical Research. 2020; 13(4): 29-35, doi:10.22159/ajpcr.2020.v13i4.36196.
- 42Octarya Z, Novianty R, Suraya N, Saryono. Antimicrobial Activity and GC-MS Analysis of Bioactive Constituents of *Aspergillus fumigatus* 269 Isolated from Sungai Pinang Hot Spring, Riau, Indonesia. Biodiversitas. 2021; 22(4): 1839-1845.
- 43Sharma M. and Mallubhotla S. Diversity, Antimicrobial Activity, and Antibiotic Susceptibility Pattern of Endophytic Bacteria Sourced from *Cordia dichotoma* L. Frontiers in Microbiology. 2022; 13:879386. doi: 10.3389/fmicb.2022.879386
- 44Rhetso T, Shubharani R, Roopa MS. and Sivaram V. Chemical Constituents, Antioxidant, and Antimicrobial Activity of *Allium chinense* G. Don. Future Journal of Pharmaceutical Sciences. 2020;6: 102. <https://doi.org/10.1186/s43094-020-00100-7>
- 45Balachandran A, Choi SB, Beata M-M, Małgorzata J, Froemming GRA, Lavilla CA Jr, Billacura MP, Siyumbwa SN, Okechukwu PN. Antioxidant, Wound Healing Potential and In Silico Assessment of Naringin, Eicosane and Octacosane. Molecules. 2023; 28(3):1043. <https://doi.org/10.3390/molecules28031043>