

# **Effect of plant growth regulators on nutrient and quality changes in *Zingiber officinale* Rosc.**

## **ABSTRACT**

An experiment was conducted during 2020-2022 to study the effect of different plant growth regulators (PGRs) on nutrient and quality parameters in ginger variety IISR Varada. Foliar spray of PGRs with 6-benzyl adenine purine (6-BAP), cycocel (Chlormequat chloride), gibberellic acid (GA) and paclobutrazol (PBZ) with five concentrations viz., 50ppm, 100ppm, 150ppm and 200ppm were applied 4th month after planting and water spray treated as control. Nutrients composition and biochemical components were observed on 5th month after planting and oleoresin was observed in the rhizomes after harvest. Results of pooled data over two years revealed that the total chlorophyll content of leaves was maximum in plants treated with GA at 100ppm. The quality parameter essential oil was shown an increased among all the PGRs and the maximum was noticed in Paclobutrazol at 100ppm concentration (82% increase from normal value), but there is a reduction in the oleoresin content from the normal standard value. The protein content in the rhizomes was estimated to be higher in the GA treated plants at 100ppm.

*Keywords: PGRs, ginger, nutrients, oleoresin.*

## **1. INTRODUCTION**

Ginger (*Zingiber officinale* Rosc.) belongs to the Zingiberaceae family is mainly valued as a spice, medicine and as vegetable since the ancient days. Ginger is one of the oldest and renowned commercial spices well known for its flavour, aroma and pungency and ginger is an essential part in flavouring the food, production of alcoholic beverages in foreign countries, in perfume industry, pharmaceutical and also for the industrial use. Ginger rhizome has an important antioxidative, antitumorigenic, antimicrobial and antiviral agent [1,2,3]. Even though, India is the major producer of ginger its production falls behind demand due to the market's high demand for its nutritional and therapeutic properties. To withstand the situation, augmented production with higher quality is necessary. One of the profitable methods getting popularized nowadays is the use of suitable plant growth regulators.

Berova and Zlatev [5] studied the effects of plant growth retardant Paclobutrazol, on the physiology and yield of tomato. The effect of GA on the photosynthetic performance, growth and yield of mustard and *Nigella* was reported [5,6]. Benzyl amino purine and gibberellin maintain the production of chlorophyll content and antioxidant enzymes under inundation conditions in soyabean [7]. Application of cycocel, potassium chloride, salicylic acid improved the quality of ginger rhizome [8]. Cycocel 100, 500, 1000 ppm, and Ethrel 50,100,200 ppm were effective when applied three times at 15-day intervals, beginning at 70 DAP in ginger [9]. Augmentation of nutraceuticals, productivity and quality of ginger (*Zingiber officinale* Rosc.) through triacontanol application was reported [10]. Foliar application of CCC at 500 ppm showed the highest yield with good quality ginger rhizomes under Tamil Nadu conditions. Besides the suitable PGRs, since the PGRs perform in a concentration dependent manner, the concentration of PGRs also needed to be standardized. Keeping this

view, the research experiment was designed with four different plant growth regulators at four different concentrations and its effect on the nutrient, biochemical and quality of produce.

## 2. MATERIAL AND METHODS

### 2.1 Plant Materials and treatments

The experiment was conducted during 2020-2022 at the ICAR-Indian Institute of Spices Research, Kozhikode, Kerala, India (Longitude 75.780411° E, latitude 11.258753°N). The experiment was set up in polyhouse conditions with average temperature, light intensity, and the humidity 20-30 °C, 200 to 800 k lux and 60 -70g/m<sup>3</sup>. The variety IISR Varada was used in this experiment which is a high yielding and good quality variety with an average yield of 22.6 tonnes per hectare and also tolerant to diseases. Poly bags size of 40x40x10 cm were filled with potting mixture consisting of soil, sand and farmyard manure in 2:1:1 proportion and planted the ginger rhizomes having the average weight of 25g in these bags. Nutrients were provided as per package of practice recommended by ICAR-IISR, Kozhikode. The experiment was laid out in factorial CRD with three replications. Physicochemical properties of the soil were: texture-sandy loam, pH-4.48, E.C-506.3 µs/cm, available N, P and K 170.8 ppm, 4.11ppm and 245ppm, respectively. The following four PGRs viz., 6 –benzyl adenine purine, cycocel (chlormequat chloride), gibberlic acid and paclobutrazol at four different concentrations 50ppm, 100ppm, 150ppm and 200ppm in PGRs were sprayed at 90 and 120 days after planting. Spraying of water was considered as control. Each treatment was replicated three times. The plants were kept free from weeds and watered when required.

The stock solution of each plant growth regulator was prepared by initially dissolving in a surfactant and then diluting it to appropriate concentrations. Each growth regulator has particular surfactant such as BAP in 1N NaOH (freshly prepared), Cycocel in water, Paclobutrazol in ethanol or methyl alcohol and GA in alcohol. The prepared solution in various concentrations was sprayed to the plant manually using a spray machine in the morning.

### 2.2 Estimation of chlorophyll content

Chlorophyll 'a' and 'b' as well as total chlorophyll content from selected leaves (1 g) were extracted with 80% acetone and quantified according to Arnon's method [11]. Spectrum absorption was measured at 645, 663 and 652 nm and the chlorophyll contents were expressed as mg g<sup>-1</sup> of fresh.

The formula for calculation of total chlorophyll content was

$$\text{Total chlorophyll} = \frac{\text{OD}_{652} \times 1000}{34.5} \times \frac{V}{1000 \times W}$$

Where,

V=final volume of the chlorophyll extract

W= fresh weight of the tissue extracted

### 2.3 Estimation of nutrient content in leaves and rhizomes

For the nutrient uptake studies, leaves and rhizomes were oven-dried at 60°C and powdered using mixer grinder. The nitrogen (N) uptake was assessed using the Kjeldahl method [12]. For the estimation of the P, one-gram powdered plant sample were digested using a mixture of nitric acid (HNO<sub>3</sub>) and hydrochloric acid (HCl 60%) with a ratio of 9:4 (v: v) and assessed using spectrophotometer at 660nm [13]. Potassium (K) was estimated using an atomic absorption spectrophotometer (Varian AA 240FS) [14].

## 2.4 Estimation of protein content in rhizome

Total soluble protein in leaves and rhizomes were estimated by using the method of Lowry. [15] by using BSA as standard. Fresh leaves (100 mg) were added in test tubes having a 10 ml phosphate buffer. The content was centrifuged at 3000 rpm for 10 minutes and the supernatant was collected and made up to 10 ml. 1 ml of the supernatant was pipette out to a test tube and 5 ml of alkaline copper tartrate reagent and 0.5 ml of folin reagent were added. The color intensity was measured at 660 nm in spectrophotometer and the amount of soluble protein present in the sample was calculated by using bovine serum albumin as standard and expressed as mg g<sup>-1</sup> fresh weight.

## 2.5 Estimation of oleoresin content

Quality parameters such as essential oil, oleoresin content, fiber content and dry recovery percentage were estimated using standard procedures. Rhizome samples were dried and powdered and oleoresin estimation was done using ASTA [16] method. Ten gram of sample was weighed and packed in cotton wool and placed in glass column (18 × 500 mm) with stopcock. To this, 50 ml of acetone was added and kept to stand overnight. The filtrate extracted through the non-absorbent cotton was collected in a pre-weighed 100ml beaker and column was washed with acetone. The extracts in the beaker were evaporated to dryness and weighed to determine the percentage of oleoresin. The amount of oleoresin was estimated gravimetrically.

Yield of oleoresin on dry weight basis was calculated using the formula.

$$\text{Oleoresin (\%)} = [\text{Weight of residue (g)} / \text{weight of sample (g)}] * 100$$

## 2.6 Estimation of Essential oil content

Essential oil content on fresh weight basis was determined by steam distillation of freshly harvested rhizomes using Clevenger apparatus, Clevenger's [17] method. The prepared sample was accurately weighed and transfers in to a flask. Then about 500ml of water was added and the flask was assembled in a Clevenger trap. The flask was heated with stirring and the distillation rate was maintained. Distill until two consecutive readings taken at one hour intervals show no change of oil volume in the trap. Cool to the room temperature, allow the stand until the oil layer is clear and read the volume of oil collected.

$$\text{Volume of oil (ml)} \\ \text{Volume of the oil \%} = \frac{\text{Volume of oil (ml)}}{\text{Weight of the sample (g)}} \times 100$$

## 2.7 Statistical analysis

The experiments were performed in a factorial completely randomized design with three replications. The SAS 9.3 statistical analysis package's General Linear Models (GLM) tool was used to do an analysis of variance on all data (Cary, North Carolina-based SAS Institute) Duncan's new multi-product line.

### 3. RESULTS AND DISCUSSION

In the present study, foliar application of growth regulators, it significantly affected the nutrient, biochemical and quality traits of ginger.

#### 3.1 Leaf Chlorophyll content

The foliar spray of growth regulator gibberellic acid significantly increased the chlorophyll content in leaves at 180 DAP than other PGRs (Fig.1). Among the interaction mean, Gibberellic acid (GA) at 100ppm showed 1.40 mean values for total chlorophyll content in the pooled year data followed by Cycocel at 150ppm. The minimum chlorophyll content was recorded in control treatment. GA has reported to increase the leaf chlorophyll content in *Gladiolus* (El-Naggar [18]). GA significantly increased the total chlorophyll contents in *Mentha piperita*. Gibberellic acid could induce the cell division among the leaves and thereby increase the surface area and increased its total gibberellin content in potato plants [18]. GA increased chlorophyll concentration per leaf while also increasing leaf area, resulting in a drop in chlorophyll per unit area and darker leaves than GA untreated leaves [20]. The treatment of GA in soyabean leads to increase in chlorophyll content and thereby increase the rate of photosynthesis [10]. The increased chlorophyll content, net photosynthetic rate may also be due to the increased potassium content in the leaves since the potassium can affect the respiration, photosynthesis, leaf NPK content, chlorophyll development, water content of leaves, carbon dioxide (CO<sub>2</sub>) assimilation and carbon movement [21]. In present study, GA influenced the synthesis of chlorophyll irrespective of all the concentrations.

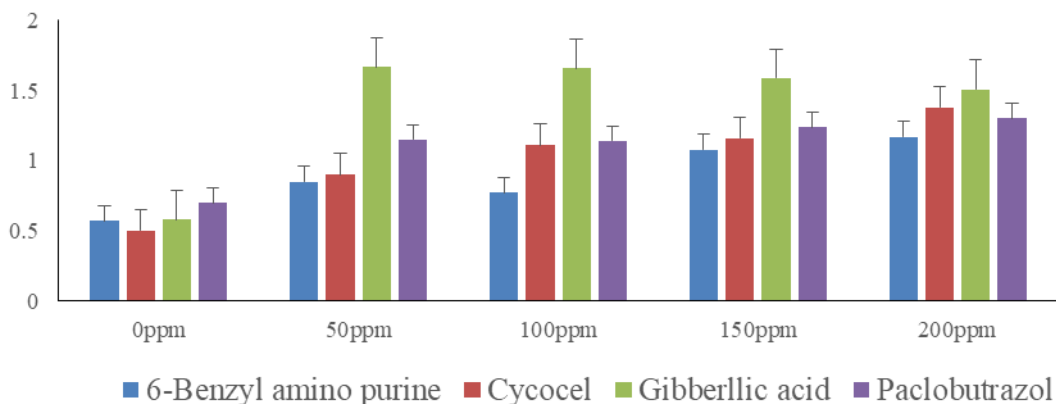


Fig. 1 Effect of PGRs on the chlorophyll content of the leaves

#### 3.2 N, P and K content of leaf and rhizomes

Foliar application of the different PGRs significantly affected the N, P, K content of leaf and rhizome. From the results, higher N was reported in Cycocel at 100ppm (Table 1), and maximum K was reported in GA at 100ppm (Table 2), but the P content in the leaves was found to be insignificant (Data not shown). In rhizomes, higher K content observed in GA treated plants at 100ppm (Table 4), higher N content in GA at 150ppm (Table 3) and higher

P in GA treated plants at 50ppm (Table 5). The control plants significantly lower content of N, P and K was observed both in rhizomes and leaves. Nitrogen content in the leaves was found to be increased with the treatment of Cycocel and found that the Cycocel treated plants contained more N, P, Ca and Mg but less K in tomato plants. The maximum K content in leaves due to the application of GA alone or in combination with potassium nitrate in *Solanum lycopersicum* [22]. Foliar application of GA improved the leaf potassium content by 17.65% in Indian Mustard [23]. Leaf nitrogen and phosphorus content was found to increase due to the application of GA<sub>3</sub> and KNO<sub>3</sub> was reported in *Cucumis sativus* plants [24]. The maximum N, P and K content in the rhizomes in GA treated plants influences the protein content in the rhizomes and increasing its nutritive [25]. In other report phosphorus was increased in the sweet pepper fruit due to the application of GA [26]. In present study also showed the positive relation in the application of exogenous growth regulators in the content of major nutrient in the plant leaves and rhizomes.

### **3.3 Protein content in rhizome**

Among the four PGRs applied the GA at 100ppm (69.04) sprayed plant rhizome registered significantly higher protein. The increased protein content in rhizome might be due to the increased Nitrogen content in the rhizomes leading to the production of maximum amino acids biosynthesis and thereby overall increase in the protein content [27,28,29,30]. Enhancing effect of the protein content in the rhizomes may be due to the direct role of Nitrogen in their biosynthesis [31]. Biosynthesis of certain hormones (gibberellins, auxins, and cytokinins) involved in protein synthesis, and the formation of the ribosome structure is influenced by the N content [25]. In present study, all PGRs showed significant influence on rhizome protein content (Fig. 2).

### **3.4 Oleoresin and essential oil content**

The essential and oleoresin content in ginger determines the quality of a ginger. The treatments with the different growth regulators could influence the quality of ginger, since it acts on the biochemical pathways. Polled data showed that the maximum quantity of essential oil was reported in paclobutrazol (2.03) at a concentration of 100ppm (2.14) followed by GA treated plants. The oleoresin content in the ginger found to decrease due to the PGRs application. There was 82% increase from the normal value in the essential oil content. According to Farooqi [32] 200ppm of kinetin resulted in the biomass production and thereby rises in essential oil in Mint (*Mentha arvensis*). The obtained results substantiate that by paclobutrazol (growth retardant) reduce the vegetative growth and thus leads to the major portioning of biomass. As a result of increase in the number of leaves with the application of 100 mg/ L of gibberlic acid (GA) higher essential oil content was reported in sage (*Salvia officinalis*) [33]. Maximum oleoresin content in the GA applied plants may be due to the increase in the number of leaves. Both the quality and quantity of the essential oil and oleoresin are influenced by the application of plant growth regulators. The increased nitrogen content in leaves could enhance oil content and yield in aromatic plants as the amount of biomass yields per unit area, leaf area development and the photosynthetic rate by the high nitrogen content [34].

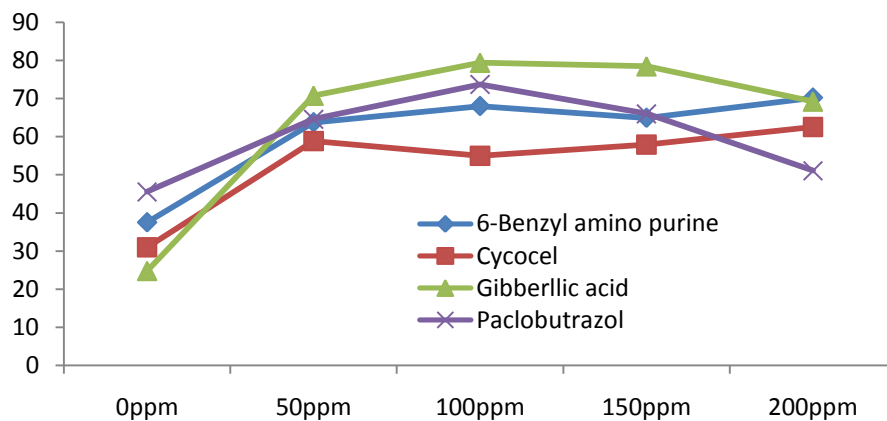


Fig. 2 Effect of PGRs on the protein content of the rhizomes

Table 1. Effect of different concentrations of plant growth regulators on the N content of leaves

Source/ concentration	0	50ppm	100ppm	150ppm	200ppm	Mean
BAP	1.27	2.34	2.41	2.55	2.11	2.13
Cycocel	1.18	2.49	2.97	3.01	2.36	2.40
GA	1.09	2.43	2.57	2.19	2.05	2.06
PCA	1.15	2.36	2.63	2.15	2.51	2.16
Mean	1.17	2.40	2.64	2.47	2.25	
(P = .05)						
Hormone (H)	0.06					
Concentration (C)	0.08					
H*C	0.22					

Table 2. Effect of different concentrations of plant growth regulators on the K content of leaves

Source/ concentration	0	50ppm	100ppm	150ppm	200ppm	Mean of hormones
BAP	1.20	1.90	2.10	1.97	1.70	1.77
Cycocel	1.21	1.62	1.80	1.59	1.68	1.58
GA	1.32	2.22	1.91	1.79	1.90	1.82
PCA	1.41	1.70	1.72	1.99	1.82	1.72
Mean	1.31	1.86	1.88	1.83	1.77	
(P = .05)						
Hormone (H)	0.02					
Concentration (C)	0.04					

H\*C 0.08

Table 3. Effect of different concentrations of plant growth regulators on the N content of rhizomes

Source/ concentration	0	50ppm	100ppm	150ppm	200ppm	Mean
BAP	0.21	1.44	1.52	2.55	2.11	1.56
Cycocel	0.14	1.55	2.57	2.19	2.05	1.70
GA	0.32	1.38	2.97	3.01	2.36	2.00
PCA	0.25	1.34	2.63	2.15	2.51	1.77
Mean	0.23	1.42	2.42	2.47	2.25	
( <i>P</i> = .05)						
Hormone (H)	0.02					
Concentration (C)	0.13					
H*C	0.23					

Table 4. Effect of different concentrations of plant growth regulators on the K content of rhizomes

Source/ concentration	0	50ppm	100ppm	150ppm	200ppm	Mean
BAP	1.25	2.06	1.85	2.07	1.94	1.83
Cycocel	1.02	1.89	1.72	1.89	2.32	1.76
GA	1.09	2.31	2.51	1.92	1.97	1.96
PCA	1.13	1.98	2.36	1.87	1.94	1.85
Mean	1.12	2.06	2.11	1.93	2.04	
( <i>P</i> = .05)						
Hormone (H)	0.03					
Concentration (C)	0.02					
H*C	0.05					

Table 5. Effect of different concentrations of plant growth regulators on the P content of rhizomes

Source/ concentration	0	50ppm	100ppm	150ppm	200ppm	Mean
BAP	0.14	0.32	0.30	0.31	0.28	0.27
Cycocel	0.08	0.30	0.29	0.29	0.29	0.25
GA	0.07	1.13	0.96	1.12	0.26	0.70
PCA	0.11	0.31	0.32	0.32	0.32	0.27
Mean	0.1	0.51	0.46	0.51	0.28	
( <i>P</i> = .05)						
Hormone (H)	0.01					
Concentration	0.01					

#### 4. CONCLUSION

It is obvious from the experiment that the application of 6-benzyl adenine purine (6-BAP), cycocel (Chlormequat chloride), gibberellic acid (GA) and paclobutrazol (PBZ), at four concentrations 50ppm, 100ppm,150ppm and 200ppm leads to various changes in the nutrient, biochemical content in leaves as well as in quality of rhizomes . From the overall observations application of Gibberellic acid at 100ppm at 4 months after the planting is significantly influenced the nutrient and biochemical changes in ginger.

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