

Occurrence of potentially pathogenic bacteria in commercially sold seafood from Misamis Occidental, Philippines

ABSTRACT

Seafood is a nutritious food that is an enviable component of a healthy diet. Nevertheless, there are health risks associated with the consumption of seafood.

This study aimed to investigate the occurrence of potentially pathogenic *Vibrio* spp., *Staphylococcus aureus*, and *Salmonella* spp. in 200 seafood samples purchased from several retail outlets in Ozamiz City, Philippines. Different samples analyzed using standard microbiological techniques were clam, shrimp, squid, and fish (anchovy and oil sardines). The *Vibrio* species identified and isolated were *V. parahaemolyticus* (47%), *V. cholerae* (16%) and *V. alginolyticus* (9%). Thirty percent (30%) of the samples analyzed were positive for *S. aureus* and 9% of the samples were contaminated with *Salmonella* sp. The total aerobic plate count mean log₁₀ cfu/g of clams, shrimps, head/gills, intestine of fish (oil sardine, anchovy), and squids were considered as medium to high risk of transmitting pathogenic bacteria. Moreover, the clams and head/gills of oil sardines were considered to be at medium to high risk of transmitting pathogenic *V. parahaemolyticus*. The findings revealed that seafood samples analyzed microbiologically were loaded with potentially pathogenic bacteria. This result is of public health concern because these organisms are documented as causes of food-borne diseases.

Keywords: pathogenic bacteria, food-borne pathogen, shrimp, squid, marine environment

1. INTRODUCTION

“Seafood a vital food component for a large portion of the world’s population”[1]. “However, there are health risks linked to the ingestion of seafood. One of the main risks involves the consumption of improperly cooked or raw seafood that might be contaminated by foodborne pathogens in the marine environment”[2]. “Various outbreaks of bacterial diseases associated with the consumption have been reported in both developed and developing countries” [3]. “From these seafood-borne bacteria, *Vibrio* spp. *S. aureus*, and *Salmonella* spp. comprise an important group of pathogenic bacteria that frequently cause human disease and even death when the contaminated food is consumed raw or improperly cooked”[4].

“In the Asian region, *Vibriosp.* has been recognized as the leading cause of foodborne outbreaks in various countries, including Japan, India, China, Korea, and Malaysia”[5] [6]. “Investigations have shown that many outbreaks may lead to acute gastroenteritis, including diarrhea, headache, vomiting, nausea, and fever, and are caused by ingestion of infected seafood”[7]. “For food samples, temperature abuse may be instigated by improper storage or a long holding time on the exhibit rack at the retail level without proper temperature control”[8] [9]. “Among more than 20 *Vibrio* species identified to be linked with human disease, *V. cholerae*, *V. parahaemolyticus*, and *V. vulnificus* are the pathogenic species of *Vibrio* that pose the greatest threat to human health”[10]. “Most of these *Vibrio* species secrete enterotoxins in water, foods, or in the gastrointestinal tract”[11].

“Aside from *Vibriosp.*, several bacteria in seafood may cause infection in humans. Examples of these bacteria that may cause foodborne illness are *Salmonella*, *Staphylococcus*, *Shigella*, and *Listeria*”[12]. “*S. aureus* is one of the most popular sources of gastroenteritis worldwide, which is caused by the ingestion of food that contains preformed toxins. Humans are

common carriers of *S. aureus* in the nose, throat, and skin infections. Therefore, the organism can be just transmitted to seafood during handling”[13].

“Seafood is not the usual habitat of *Salmonella*, but still, the occurrence of *Salmonella* in seafood is escalating. Hence, fishery products have been recognized as **major spreaders** of foodborne pathogens”[14][15]. “*Salmonella* is associated with fish and fishery products, thereby causing a public health problem. Various authors have reported the incidence of enterotoxigenic *S. aureus* and *Salmonella* in seafood”. [16] Infection with these bacteria may affect only the intestines or may spread from the intestinal tract to the blood vessels and then to other body sites.

Although several technological alternatives that could augment the quality of the food supply have been **implemented**, instances of food-borne diseases caused by consumption of seafood still emerge. In Iligan City, Philippines [17], an outbreak of cholera killed two people. Health authorities have begun a series of **tests** of water samples in the villages. Other reports said some of the infected individuals were associated with the consumption of improperly cooked **seafood**, particularly **oysters**.

In the provinces of Zamboanga del Sur, Lanao del Norte, and Misamis Occidental Philippines, massive **mortality** were observed among pond-cultured shrimp due to *Vibrio harvey* **which** was isolated purely from the lymphoid organs and hepatopancreas of affected shrimp, **and** histopathological observations indicated a severe bacterial infection [18]. Disease problems have **been recorded** in shrimp-finish polyculture in Misamis Occidental due to bacteria [19]. Nevertheless, studies on the presence of potentially pathogenic bacteria **in seafood** are scarce. This study aimed to examine the occurrence of potentially pathogenic *Vibrio* sp., *S. aureus*, and *Salmonella* sp. **in** fresh seafood sold from several **retail** outlets in the province of Misamis Occidental, Philippines, **and** to highlight that consumption of raw and ingestion of improperly cooked sea foods may pose serious health **hazards**.

2. MATERIAL AND **METHODS**

2.1 Sample collection and processing

Simple random sampling of two hundred (200) samples of commercially sold seafood such as clams (40), shrimps (40), squids (20), **and** fish [anchovy (50) and oil sardines (50)] were purchased from **10 randomly selected** retail outlets in the province of Misamis Occidental, Philippines. Samples were placed in sterile, labeled, sealed plastic bags and were transported to the laboratory under **refrigerated conditions** for analysis. The samples were aseptically removed from the container. After the skin ranging, **sterile forceps** and knife appropriate for opening the shell to remove the intestine and gills were used. For shrimp samples, the shell was peeled and detached from the **fresh** part and intestine. For fishes, samples of head/gills, intestines, and flesh were analyzed microbiologically, **whereas** only the flesh of squid was examined using standard microbiological techniques. For the clams, the flesh and intervalve water were used for bacterial analyses. **Approximately** 1g from each sample was weighed and dispensed into **9ml** of sterile distilled water and shaken vigorously to mix properly. Ten-fold dilutions were made, and dilutions (10^{-6} and 10^{-7}) were selected for microbial enumeration using the standard pour plate method by Fawole and Oso (2001)[20].

2.2 Enrichment procedures

Five (5) grams of each shellfish flesh **was** incised using a sterile scalpel after removal of the carapace. These 5 g meat samples were homogenized in 45 ml of 3% NaCl containing 1% alkaline peptone water (APW, pH: 8.6) **using** a sterile blender. The shellfish homogenates were incubated at 37°C for 18 h [21].

2.3 Isolation procedures and identification of **the bacterial colonies**

Analysis of *Vibriospp.* in seafood samples was performed in **triplicate**. Standard procedures of the U.S. Food and Drug Administration (FDA) **was** used to detect the contamination of potentially pathogenic *Vibrio* species [22] with some modifications. Following incubation, shellfish homogenates were inoculated using an inoculating loop on **thiosulfate citrate bile salts** sucrose agar media (TCBS, Hi-Media, India) and incubated at 37°C for 18 h [23]. For each seafood sample, 25g were homogenized in **225ml** sterile 0.1% peptone water using a sterile blender. A **10-fold** serial dilution was prepared in sterile 0.1% peptone water as described in the Bacteriological Analytical Manual [24]. **The spread** plate method was carried out using 10 and ten dilutions on TCBS agar. The TCBS plates were incubated at 37°C for **18-24 h**, and counts were made for each colony type. Discrete colonies were aseptically **subcultured** into fresh agar plates to **obtain pure** colonies.

Colonies recognized as discrete were carefully examined macroscopically for cultural characteristics. Pure isolated colonies were subjected to Gram staining and **grown** at different salt concentrations **by placing** the colonies into tubes **containing** peptone water and 0%, 3%, 6%, **and** 10% NaCl, and these tubes were incubated at 37°C for 24 h [25]. **In addition**, all bacterial colonies from different samples **grown** on TCBS plates were chosen to be streaked on the surface of **trypticase soya agar** slants (TSA; Oxoid, UK) added with 2% NaCl and incubated for 24 **h** at 37°C. Further identification of

Vibrio sp. was performed using physiological, morphological, and different biochemical tests available in the laboratory. The isolates were identified by comparing their characteristics with those of recognized taxa, as illustrated and described by [26] [27] [28] [29]. To isolate and identify *S. aureus*, 10 g of each seafood sample was homogenized with 90 ml of a sterile 0.1% peptone water solution. Serial dilutions of the homogenate were performed using sterile peptone water and plated in triplicates on Baird Parker agar (Merck 1.05406). After incubation at 35 °C for 48 h, the distinctive black colonies with apparent halo of *S. aureus* were subjected to Gram staining to clarify bacterial morphology. Staphylococci are spherical Gram-positive bacteria that occur in microscopic cluster. *S. aureus* forms a relatively large yellow colony on rich medium, whereas *S. epidermidis* has a comparatively small white colony [30].

For the detection of *Salmonella* sp., the EN ISO 6579 reference method was used. This process currently involves non-selective pre-enrichment in buffered peptone water followed by selective enrichment in Rappaport–Vassiliadis soya broth and Muller–Kauffmann tetrathionate/novobiocin broth followed by selective plating of these on Salmonella–Shigella agar. Incubation was made at 35°C, and colonies resembling *Salmonella* sp. or *Shigella* sp. *Salmonella* will produce hydrogen sulfide (H₂S) gas but will not ferment lactose. The resulting bacterial colonies will appear colorless with black centers. Any suspect salmonella strains that are isolated are then confirmed by biochemical and serological test methods.

2.4 Bacterial counts and data analysis

Following proper incubation, dilutions of 30–300 colonies were selected and counted. The number of colony-forming units per g (cfu/g) of seafood was computed by multiplying the number of bacteria by the dilution. Total aerobic plate count (TAPC), *Vibrio* sp., *S. aureus*, and *Salmonella* sp. counts that were attained for cfu/g of food were converted into log₁₀ values. Seafood was classified as having a no-to-low risk of transmitting pathogenic bacteria if the TAPC was 5.0 log₁₀ cfu/g and if the counts of *Vibrio* spp., *S. aureus*, and *Salmonella* sp. were 3.0 log₁₀ cfu/g. Furthermore, if the value is at least 5.0 log₁₀ cfu/g for the total bacterial counts and at least 3.0 log₁₀ cfu/g for *Vibrio* sp., *S. aureus*, and *Salmonella* sp., the seafood is classified as medium-to-high risk of transmitting pathogen [31][32].

3. RESULTS AND DISCUSSION

3.1 The Occurrence of *Vibrio* spp.

Seafood samples analyzed microbiologically in this study showed varying degrees of bacterial contamination. The results on the contamination of *Vibrio* spp. are summarized in Tables 1a and 1b. Of the 200 seafood samples tested, 65.5% were contaminated with potentially pathogenic *Vibrio* spp. The high incidence of *Vibrio* spp. could be due to the nature of these bacteria, which are known as halophilic waterborne and commonly dwell in environmental water sources worldwide. brackish water, marine environments, and freshwater may support the growth of these organisms, which are pathogenic to humans [33].

Table 1a Distribution and frequency of occurrence of *Vibrio* spp. isolates from fresh seafood samples.

Samples	No. Tested (%)	No. Positive for <i>Vibrio</i> spp
FISH		
Anchovy	50 (25)	39 (78)
Sardines	50 (25)	42 (84)
SHRIMP	40 (20)	18 (45)
CLAM	40 (20)	22 (55)
SQUID	20 (10)	10 (50)
Total	200 (100)	131 (65.5)

Table 1b. Distribution and frequency of occurrence of *Vibrio* spp. isolates from fresh seafood samples.

Seafood Samples	Number (%) of positive samples			Total Positive for <i>Vibrio</i> (%)
	<i>V. parahaemolyticus</i>	<i>V. cholerae</i>	<i>V. alginolyticus</i>	
FISH				
Anchovy				
Gills	14 (28)	06 (12)	04 (08)	24 (48)
Intestine	05 (16)	04 (08)	02 (04)	11 (22)
Meat	04 (08)	00 (00)	00 (00)	04 (08)
SARDINES				

Gills	16 (40)	04 (08)	02 (04)	22 (44)
Intestine	8 (20)	02 (04)	02 (04)	12 (24)
Meat	06 (12)	02 (04)	00 (00)	08(16)
SHRIMP	10 (25)	06 (15)	02 (05)	18(45)
CLAM	12 (30)	06(15)	04(10)	22(55)
SQUID	06 (30)	02 (10)	02 (10)	10(50)
Total number (%) of positive samples	81 (40.5) /200	32 (16)/200	18 (09) /200	131(65.5)

The samples that harbored most of the *Vibrio* spp. were the fish in which the gills obtained most of the isolates. This could be due to the gill physiology, which serves as a sieve that filters water that comes into the fish from their water ecosystem. Fish could be predisposed to gill disease because the gill is used for the exchange of gasses, and the fish will be hassled because of the intricacy in the exchange due to the heavy load of microbes in water, causing the gill to be infected [34]. Among the *Vibrio* species isolated, *V. parahaemolyticus* was the predominant species (40.5%), followed by *V. cholera* (16%) and *V. alginolyticus* (09%). The findings of this study agree favorably with the results of several authors in a similar study conducted in Brazil, China, India, and Bursa province, Turkey [35][36]. On the other hand, varying rates have been reported in some other studies. In Nigeria, *V. cholerae* was the most frequently isolated *Vibrio* sp. followed by *V. parahaemolyticus*[37].

The predominant *V. parahaemolyticus* is salt tolerant but very sensitive to heat and is destroyed by cooking. However, gastroenteritis is a common cause of foodborne diseases in many Asian countries such as China, Japan, and Taiwan and is recognized in the United States as the leading cause of human gastroenteritis [38]. This species is associated with the consumption of insufficiently or raw cooked seafood, poor handling of seafood during preparation, and improper post-harvest storage conditions [39]. This pathogen is commonly isolated from a variety of seafood including oyster, crab, scallop, lobster, shrimp, and fish [40].

In this study, 16% of the samples tested were positive for *V. cholera*. This bacterium favors alkaline surroundings to thrive well [41]. Perhaps the presence of this bacterium was linked to the low salinity in various locations due to the influx of freshwater into the ocean. This species can cause cholera and is typically a waterborne illness. Food-borne cases, primarily fish outbreaks, are also known carriers of this microorganism.

In this study, the prevalence of *V. alginolyticus* in seafood was 9%. This finding was close to the result of Baffone et al. (2006), who reported 8.16% in clams in Italy. In contrast, Hassanin (2007) reported higher infection rates for *V. alginolyticus* in shrimps in Egypt. This bacterium is one of the most popular *Vibrio* species occurring in marine environments and seafood (Neetoo et al. 2022). *V. alginolyticus* is an opportunistic pathogen, and its pathogenicity is thought to be similar to that of *V. parahaemolyticus*[42]. In India and Taiwan, this *Vibrio* species is linked with a white spot in shrimp while the zoonotic risk of this pathogen has been implicated in soft tissue, wound and ear infections in humans[43]. *Vibrio* spp. and several pathogenic species are commonly associated with outbreaks of *Vibrio* infections due to ingestion of raw or improperly cooked seafood, water contaminated with human feces or sewage, inappropriate post-harvest storage conditions, or poor handling of seafood during preparation.

3.2 Occurrence of *S. aureus* and *Salmonella* sp.

The incidence of *S. aureus* in this study was found to be higher than that of *Salmonella* sp. *S. aureus* was detected in 30% of the 200 raw samples of seafood, whereas the incidence of *Salmonella* sp was 9%. The head/gills of fish (oil sardines & anchovy) were heavily loaded with *S. aureus*. Four isolates of *S. aureus* were obtained from shrimp samples, three from clams, and two from squids (Table 2).

It is well documented that freshly caught seafood is free from *S. aureus*, and contamination occurs upon handling [44]. Other conditions such as poor personal hygiene, inadequate refrigeration, and delay in processing are associated with staphylococcal contamination. *S. aureus* is a major cause of food poisoning and a range of extraintestinal infections[45]. Staphylococcal food poisoning is caused by a preformed enterotoxin produced by the growth of bacteria in food that is ingested [46].

The overall prevalence of *Salmonella* sp. in 100 seafood samples tested was in agreement with that reported elsewhere. The findings of the present study showed that the organism was not recovered from the gills and flesh of the fish but was isolated from their intestine only. This result is in agreement with the work of Green [47], in which the organism was never recorded from the flesh of the fish but was documented from the viscera and epithelium. Several scientists believe that fish are possible carriers of *Salmonella*, which accumulate in their intestines for somewhat short periods of time, and some believe that fish become actively infected by *Salmonella*[48][49]. Furthermore, the mollusk and crustacean samples also harbored colonies of *Salmonella*.

The possible cause of *Salmonella* contamination in seafood might be farm runoff and fecal contamination from nearby domesticated or wild animals and poor water quality. In addition to poor distribution, retail marketing, handling, and high stocking, preparation practices, densities, and high water temperature may be responsible for increased *Salmonella* contamination in shrimp [50] (Loest et al. 2022). Other investigations of live bivalve mollusks confirmed that this bacterium could endure coastal water salinity. The bacteria may be found in 0%–2.5% of live shellfish, and intake of these foods

may cause salmonellosis, characterized by enteric fever along with gastroenteritis and diarrhea. Similarly, the United States Food and Drug Administration data revealed that *Salmonella* was the most common contaminant of fish and fishery products [51]. Approximately 12% of the foodborne outbreaks related to the consumption of fish are due to bacteria, including *Salmonella*.

Table 2. Distribution and frequency of *Salmonella* spp. and *S. aureus* isolates from fresh seafood samples.

Seafood Samples	Number (%) of positive samples/number of samples tested	
	<i>Salmonella</i> sp.	<i>S. aureus</i>
FISH		
Anchovy		
Gills	00 (00) /50	08 (16)/50
Intestine	02 (04) /50	06 (12)/50
Meat	00 (00) /50	04 (08)/50
Sardines		
Gills	00 (00) /50	12(24)/50
Intestine	04 (08) /50	04/(08)50
Meat	00 (00) /50	08(16)/50
SHRIMP		
	04 (10) /40	06(15)/40
CLAM		
	06(15)/40	08(20)/40
SQUID		
	02 (10) /20	04(20)/20
Total number (%) of positive samples/total number of samples tested	18 (09) /200	60 (30) /200

3.3 Bacterial counts in seafood

The mean log₁₀ cfu/g of *Vibrio* spp., *S. aureus*, and *Salmonella* sp. and total aerobic plate counts (TAPC) values are presented in Table 3. The clam and shrimp samples obtained the highest TAPC values of 7.61 and 7.48, respectively. These molluscan bivalves and decapod crustaceans are filter feeders, and they may be apt to hoard microbes in nearby waters, which may also contain these microorganisms. They are typically grown and collected in near-shore; shallow estuarine waters [52] and are therefore expected to harbor high concentrations of pathogenic organisms. If this seafood is eaten after a very mild heat treatment or raw, it poses a significant health risk to consumers. The seafood that may be linked with human infection caused by *V. parahaemolyticus* are bivalve mollusks, crustaceans, and finfish, especially those served raw or superficially cooked and exposed to temperature abuse [53]. Of the fishes, the head/gills of oil sardines and anchovy harbored considerable TAPC values of 7.42 and 6.78, respectively.

The TAPC value suggests that samples of shrimps, clams, head/gills, and intestines of fishes as well as squids examined in the study were considered as medium to high risk of transmitting pathogenic bacteria (not within acceptable limits, i.e. <5.0 log₁₀ cfu/g for total counts). Moreover, the clams and head/gills of oil sardines were also considered to be at medium to high risk of transmitting pathogenic *V. parahaemolyticus*. From the zoonotic point of view, contamination of potentially pathogenic bacteria in all samples of seafood tested suggests a possible risk to the health of people consuming raw or improperly cooked seafood. Naturally, fish and other seafood have immunity to infections, but their immunity decreases when the concentration of microorganisms is on the increase [54]. Bacteria in seafood improperly cooked could be transmitted to humans as they establish themselves in the intestine, particularly those potentially pathogenic, causing problems, including diseases of various kinds. These bacteria enter the body and then discharge endotoxins that irritate the stomach and bowels. This toxin might lead to stomach cramps, diarrhea, and fever and is called foodborne infection.

Table 3 Total aerobic plate counts, *Vibrio* spp., *S. aureus*, and *Salmonella* spp. counts mean log₁₀ cfu/g (ml) ± standard deviation of seafood in Ozamiz City.

Seafood Item	n	TAPC	<i>Vibrio</i> isolates of <i>S. aureus</i> <i>Salmonella</i>				
			<i>V. parahaemolyticus</i> <i>alginoliticus</i>	<i>V. cholerae</i>	<i>V.</i>		
Fish							
Anchovy	25						
head/gills		6.78±1.7	2.87±1.0	1.23±0.7	1.02±0.6	1.19±0.4	00
intestine		5.60±0.5	1.14±0.9	1.70±0.9	0.37±0.6	1.10±0.5	1.66±1.2
meat		3.64±2.0	1.05±1.1	00	00	1.20±0.1	00
Oil Sardines	25						
head/gills		7.42±2.1	3.04±0.4	1.21±1.0	1.03±0.1	2.19±0.5	00
intestine		5.90±1.9	1.06±0.5	1.07±1.6	1.87±0.6	1.19±0.5	1.15±0.3
meat		3.51±1.0	1.04±0.7	1.02±0.8	00	1.05±0.5	00
Shrimp	20	7.48±1.8	2.69±0.7	1.10±0.7	0.87±0.8	1.73±1.1	1.06±1.8
Clam	20	7.61±1.5	3.14±0.5	1.05±0.4	0.89±0.4	1.93±0.5	1.59±0.2
Squid	10	5.55±0.9	1.20±0.2	1.02±0.2	1.04±0.9	1.14±1.3	1.01±0.9

4. CONCLUSION

Seafood is considered an essential element in the diet and plays a vital role in the development of foodborne diseases. This study revealed that seafood samples analyzed microbiologically could be a source of foodborne bacterial pathogens such as *Vibrio* sp. *S. aureus* and *Salmonella*. The contamination of these bacteria could be attributed to cross-contamination from the surroundings and handling by the sellers and thus constitute a potential health hazard to the public.

Nevertheless, fish and other seafood have to be properly cooked before consumption, and proper quality control measures have to be adopted in the culturing, processing, harvesting, and consumption of seafood. Therefore, monitoring the contamination of potentially pathogenic bacteria in harvested seafood is vital for the sustenance of public health. Further studies have to be conducted to include other seafood products in the microbial analyses and to examine other pathogens probably in seafood.

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