

Original Research Article

Development of Pink Pigmented Facultative Methylo trophs' (PPFMs) Consortium formulation and its efficacy on Chilli (*Capsicum annuum*)

ABSTRACT

Consortium from Pink Pigmented Facultative Methylo trophs (PPFMs) isolates (*Methylobacterium populi* AAU PPFM C-7, *Methylobacterium radiotolerans* AAU PPFM C-17, *M. populi* AAU PPFM C-19, *M. populi* AAU PPFM T-2 and *M. radiotolerans* B-2) were prepared and pot house experiments were laid out in completely randomized design with four replications on Chilli crop. The results of the study indicated significant effect of PPFMs consortium on growth parameters of chilli (*viz.* shoot length, root length, shoot weight, root weight, number of branches per plant) that could be result of the plant growth hormones produced by PPFMs (*i.e.*, IAA, ACC deaminase and Siderophore production).

Keywords: PPFMs, Foliar spray, Phyllosphere, Solanaceous crops, Plant growth promotion, Chilli

1. INTRODUCTION

The plant canopy is a habitat to a wide range of microorganisms that have beneficial, harmful or neutralistic effects. Large fraction of the earth's surface is covered with plants, that leaf surfaces often represent a substantial multiple of the soil surface area. The terrestrial leaf surface area is over 6.4×10^8 km² that might be colonized by microbes to harbor bacterial population of about 10^{26} cells. As an ecological niche, the plant phyllosphere supports highly abundant methylo trophic species ranging from 10^4 to 10^7 colony forming unit (CFU) per leaflet (Mizuno., 2012). The phyllosphere represents the largest biological surface, estimated to be two times the surface of the earth and is involved in many processes required for the ecosystem including primary biomass production (food and feed), oxygen release and carbon-dioxide fixation (Delmotte *et al.*, 2009). This surface is considered a hostile environment for bacterial epiphytes. Morphological feature of leaf surface and its surroundings both make phyllosphere ecosystem crucial than another habitat. Healthy leaf epidermal cells constantly secrete organic and inorganic molecules, such as amino acids, sugars (including sucrose, glucose and fructose), methanol and various salts to their surfaces (Corpe and Rheem, 1989; Fiala *et al.*, 1990).

Bacteria are the dominant inhabitants of crop phyllosphere and mainly represent on leaves in a large number. Pink Pigmented Facultative Methylophs (PPFMs) are heterotrophic microflora of the surface of young leaves and are group of Prokaryotic aerobic eubacteria. They are the endophytic prime inhabitants of wide variety of plant species. Their distinctive pink pigmentation is due to carotenoids; render them to be tolerant to extreme light condition and reactions. These features could explain their occurrence in diverse ecological system such as soil, plants, air and water. Most common niche for synergism between *Methylobacterium* and plant is the phyllosphere, especially leaf surface. The genus *Methylobacterium* is one of the dominant genera, which act as a symbiont with plants by consuming methanol and in turn providing vitamin B₁₂, auxins and cytokinin useful for the plant growth and systemic resistance (Madhaiyan *et al.*, 2007). Their capacity for adapting to changing environmental conditions, growing at high rates on methanol and a variety of C₁-C_n compounds (facultative) reflects their metabolic potentiality. Thus, broadly the *Methylobacterium* species are non-pathogenic and ubiquitous, found in variety of habitats on earth (Green, 2006).

Chilli is an important vegetable and spice crop grown throughout the world. In India, it occupies the area of 287.05 thousand of ha with production of 3406.03 thousand ton. Foliar spraying for crop nutrition has gained the attention of researchers and farmers due to their superior effect. Plants are able to absorb essential elements through their leaves in foliar feeding. The absorption takes place through their stomata and also through their epidermis. Transport is usually faster through the stomata, but total absorption may be as great through the epidermis. Foliar feeding of PGPR and their metabolites has become standard practice now a day in smart agriculture for cultivation of vegetables and fruits and that's why we attempted study to role of native pink pigmented facultative methylophic (PPFMs) foliar inhabitant bacterial isolates as plant growth promoting probiotics was studied.

2. MATERIAL AND METHODS

Development of phyllospheric PPFMs bacterial consortium and their plant growth promoting (PGP) activity

(A) Consortium development

I. Sources of native PPFMs bacterial isolates

The PPFMs were isolated from the phyllosphere of solanaceous crops (*i.e.*, chilli, tomato, brinjal, tobacco and potato) from research farms of Anand Agricultural University farms by leaf imprinting method (Savitha *et al.*, 2015). They were further screened for their methanol consuming and plant growth promoting activities. From the thirty isolates, five potent PPFMs isolates C-7, C-17, C-19, T-2 and B-2 were selected for consortium preparation and to study its efficacy on chilli crop in pot study.

II. Compatibility test of PPFM isolates

In vitro plate bioassay was carried out to determine compatibility of the native phyllospheric PPFM isolates. Each phyllospheric PPFM bacterial isolate was grown in NMS broth for 5-6 days. On Nutrient agar plate, one isolate (100 µl) was spreaded and the well was made by cork borer, in the well of the plate 50 µl of the other phyllospheric PPFM isolates were filled. The plates were incubated at 30±2°C for 5 days and observed every 24 h for inhibition of the test cultures if any (Sateesh and Sivasakthivelan, 2013).

III. Consortia preparation

PPFM bacterial isolates were grown in broth (Nutrient broth supplemented with 0.5% methanol) to ensure maximum number. Determination of population density of each isolate in broth was done by Neubauer hemocytometer. Individual culture in specific proportion was mixed to reach population density of 5 x 10⁸ in final product and used for experiment.

(B) Efficacy of PPFM in Consortium on Chilli cv. GVC-111 under pot house condition during Kharif 2017-18

Experimental site

Pot experiment was conducted in pot house during *kharif* season of the year 2017-18 at Department of Agricultural Microbiology, AAU, Anand. The details of the experiment described below.

Table 1. Treatment details

Treatment details	
T ₁	Treated check: Chemical foliar fertigation (19:19:19) @ 2 g/lit
T ₂	Indigenous Cow urine @ 2% v/v
T ₃	Vermiwash @ 10% v/v

T ₄	Phyllospheric PPFMs Consortium (5 × 10 ⁸ CFU/ml)
T ₅	AAU Methylo trophic bacterial consortium (phyllospheric and rhizospheric) for Rice (5 × 10 ⁸ CFU/ml)
T ₆	Absolute control (Water spray)

Treatment Application:

Treated check T₁, T₂ and T₃ foliar spray at 30 and 45 days after transplanting (DATP).
Microbial Foliar spray T₄ and T₅ at 30 and 45 DATP @ 1 lit/ha.

Experimental details under pot study:

No. of treatments: 6

No. of replication: 4

Design: Completely Randomized Design (CRD)

Pot size/capacity: 10 kg soil/pot

Crop: Chilli (*Capsicum annuum* L.)

Variety: Gujarat Vegetable Chilli – 111 (GVC-111)

Recommended Fertilizer dose (kg/ha): 100-50-50 (NPK)

3. RESULTS AND DISCUSSION

EFFICACY OF PPFM BACTERIAL CONSORTIUM ON CHILLI cv. GVC-111 IN POT HOUSE DURING KHARIF 2017-18 Effect of PPFM Consortium on Growth Attributing Characters

Shoot length

Table 2. Effect of PPFMs consortium on shoot length

Treatments	30 DATP	45 DATP	90 DATP
T ₁	22.45	36.00 ^b	70.25 ^e
T ₂	22.08	36.65 ^b	76.25 ^d
T ₃	21.50	36.57 ^b	79.33 ^c
T ₄	22.58	38.93 ^a	88.42 ^a
T ₅	22.43	36.86 ^b	83.17 ^b
T ₆	22.51	35.44 ^b	66.50 ^f
CD			
C.V. %	3.30	2.73	1.85
S.Em. ±	0.37	0.50	0.72

Note: Treatment means with the letter/letters in common are not significant by Duncan's New Multiple Range Test at 5% level of significance

The observation on shoot length of the plant were measured at 30, 45 and 90 DATP and analyzed that showed no significant difference in plant height at 30 DATP, but at 45 DATP and 90 DATP it showed significant effect as T₄ was had the highest shoot length (38.93 cm) which was found significantly superior over T₆ (35.44 cm). Similarly at 90 DATP T₄ showed the highest shoot length (88.42 cm) which was found significantly superior over T₆ showing (66.50 cm), Which was followed by T₅ (83.17 cm), T₃ (79.33 cm), T₂ (76.25 cm) and T₁ (70.25 cm). T₄ was statistically superior over T₁. The increased shoot length might be the result of IAA production as it has important role in stimulation of cell elongation and division that aids to longer and PPFMs have been reported to produce IAA, Similar findings were reported by Subhaswaraj *et al.* (2017) as they studied the impact of seed bacterization with *M. extorquens* MM2 on seed germination, root and shoot length where they found that *M. extorquens* MM2 has capacity to produce IAA (6.16 µg/ml) which resulted in increased seed vigour index and its foliar application contributed a substantial impact to growth of tomato plant. Sentilkumar and Krishnamurthy (2017) also reported that application of *Methylobacterium* by 1% foliar spray resulted in significant increase in plant height compared to control.

Table 3. Effect of PPFMs consortium on chlorophyll content (mg/cm²)

Treatments	30 DATP	45 DATP	90 DATP
T ₁	46.73	40.48 ^b	51.17 ^{cd}

T ₂	45.37	41.36 ^{ab}	52.08 ^{bcd}
T ₃	45.53	41.76 ^{ab}	53.46 ^b
T ₄	45.99	42.82 ^a	57.26 ^a
T ₅	45.65	41.43 ^{ab}	53.23 ^{bc}
T ₆	45.53	38.38 ^c	50.64 ^d
S.Em. ±	0.33	0.70	0.72
C.V. %	1.42	3.43	2.70

Note: Treatment means with the letter/letters in common are not significant by Duncan's New Multiple Range Test at 5% level of significance

Effect of PPFMs consortium on chlorophyll content of Chilli cv. GVC-111

The observation on chlorophyll content of leaves were measured at 30, 45 and 90 DATP that showed no significant difference at 30 DATP but at 45 and 90 DATP, the difference were significant. At 45 DATP, T₄ (42.82 mg/cm²) was found significantly superior over T₆ (38.38 mg/cm²) and T₅ (41.43 mg/cm²), T₃ (41.76 mg/cm²) and T₂ (41.36 mg/cm²) found at par with the T₄. Similarly at 90 DATP T₄ showed the highest chlorophyll content (57.26 mg/cm²) which was found significantly superior over T₆ (50.64 mg/cm²), which was followed by T₃ (53.46 mg/cm²) and T₅ (53.23 mg/cm²). The siderophore production by the PPFMs may have contributed to the increase in chlorophyll content as iron is a crucial component of chlorophyll molecules, so improved iron uptake leads to higher chlorophyll synthesis. PPFMs also interact with plant hormones like auxins and cytokinins. These hormones can stimulate chlorophyll biosynthesis by upregulating the expression of relevant genes. Many other workers have also reported the effect of PPFMs on chlorophyll content. Methylobacterium application has showed increased in photosynthetic activity by increasing the stomatal count, chlorophyll content and malic acid content in crops (Madhaiyan *et al.*, 2004). Awasthy *et al.* (2020) reported that 2% foliar spray of PPFM at 30 and 50 days increased chlorophyll content in Paddy. Satyan *et al.* (2018) reported increase the chlorophyll stability index of small cardamom due to PPFM inoculation. Our study also has a similar trend as evidenced by the higher chlorophyll content in the treatments compared to the controls.

Effect of PPFMs consortium on phyllosphere microbial population of chilli cv. GVC-111

The PPFMs bacterial population were measured on 30, 45 and 90 DATP at initial and three days after application of respective treatment. Results of the same showed no significant difference 30 DATP initial, but the observation after three days of treatment application revealed that T₄ was having highest PPFMs population (5.52 log cfu/g of leaves) which was found significantly higher and T₅ (5.47 log cfu/g) was at par with T₄. Similarly at 45 DATP T₄ was having the significantly higher PPFMs population both at initial (4.51 log cfu/g) and three days after treatment application (5.50 log cfu/g) which were at par with T₅ initially (4.49 log cfu/g) and three days after treatment application (5.49 log cfu/g). At 90 DATP the PPFMs population data showed that T₄ having was significantly higher (5.52 log cfu/g) population which was at par with T₅ having (5.48 log cfu/g). Higher bacterial populations in of T₄ (PPFMs consortium) and T₅ (Rhizospheric and phyllospheric methylootrophs) after foliar spray shows their survival and proliferation in phyllosphere which is believed as their habitat.

Table 4. Effect of PPFMs consortium on bacterial population

Treatment	Phyllosphere bacterial population (log CFU/g of leaves)				
	30 DATP		45 DATP		90 DATP
	Initial	After 3 Days of spraying	Initial	After 3 Days of spraying	

T ₁	4.41	4.42 ^b	4.39 ^b	4.39 ^b	4.39 ^b
T ₂	4.40	4.40 ^b	4.37 ^b	4.38 ^b	4.38 ^b
T ₃	4.39	4.41 ^b	4.37 ^b	4.38 ^b	4.41 ^b
T ₄	4.40	5.52 ^a	4.51 ^a	5.50 ^a	5.52 ^a
T ₅	4.41	5.47 ^a	4.49 ^a	5.49 ^a	5.48 ^a
T ₆	4.38	4.39 ^b	4.36 ^b	4.37 ^b	4.36 ^b
S.Em. ±	0.04	0.03	0.03	0.04	0.04
C.V. %	1.68	1.43	1.36	1.52	1.47

Note: Treatment means with the letter/letters in common are not significant by Duncan's New Multiple Range Test at 5% level of significance

Logarithmic transformation given to microbial population values

Effect of PPFMs consortium on root weight, shoot weight, root length, shoot length, number of branches and yield of chilli cv. GVC-111

Table 5. Effect of PPFMs consortium on root weight, shoot weight, root length, shoot length and number of branches chilli cv. GVC-111

Treatment	Root weight (g/plant)	Shoot weight (g/plant)	Root length (cm)	No. of Branch per plant
T ₁	5.37 ^d	70.25 ^d	18.85 ^c	1.72 ^a
T ₂	6.08 ^c	77.65 ^c	19.95 ^{bc}	1.75 ^a
T ₃	6.79 ^b	80.05 ^{bc}	20.51 ^b	1.85 ^a
T ₄	8.26 ^a	91.87 ^a	28.29 ^a	1.88 ^a
T ₅	7.92 ^a	80.78 ^b	21.12 ^b	1.78 ^a
T ₆	5.08 ^d	65.17 ^e	15.16 ^d	1.44 ^b
S.Em. ±	0.24	0.88	0.42	0.06
C.V. %	7.17	2.27	6.82	6.63

Note: Treatment means with the letter/letters in common are not significant by Duncan's New Multiple Range Test at 5% level of significance

Square root transformation was given to No. of branches values

The observation on root weight, shoot weight, root length, shoot length and no. of branches were recorded and analyzed which shows that root weight of T₄ (8.26 g/plant) was significantly superior than T₆ (5.08 g) and T₅ (7.92 g) found at par with T₄. Similarly, T₄ was having root length (28.29 cm) which was significantly higher than T₆ (15.16 cm). The observation on shoot weight also found significant, where T₄ (91.87 g/plant) was significantly superior than T₆ (65.17 g/plant). No. of branches were also significantly higher in T₄ (1.88) compared to T₆ (1.44). Nalayini *et al.* (2014) also reported that application of 75% recommended N and P fertilizers with azophosmet and foliar application of PPFMB either 45 or at 60 DAS resulted in 2.3 q of additional seed cotton than 100% RDF without bioinoculants. Shivakumar *et al.* (2018) reported that foliar application of PPFM has led to increase in root length and root volume on tomato crop under pot experiment. The enhancement PPFM (2%) might be due to involvement in the translocation of photo assimilates from source to sink. Similar results for growth parameters were reported in our study for growth attributing parameters.

4. CONCLUSION

In this study, foliar application of PPFMs consortium has resulted in enhanced growth parameters significantly in chilli under pot house experiment. The shoot length, chlorophyll content, root length, root weight, shoot weight and no. of ranches per plant were found significantly higher in the treatment receiving PPFMs consortium in comparison to control. The Plant growth promoting regulators (IAA, ACC deaminase, siderophore production) has upregulated the plant growth which reflected in overall growth.

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