

1 Evaluation of Tomato (*Solanumlycopersicum*L.) Accessions Resistance against Fusarium Wilt
2 (*Fusariumoxysporum*).
3
4

5 **ABSTACT**

6 **Aim:** Fusarium wilt (*Fusariumoxysporum*) is one of the major biotic factors limiting tomato production
7 globally. Research was conducted in three environments to screen and identify Nigeria tomato
8 accessions that are resistant to fusarium wilt. **Methodology:** Twelve (12) prominent tomato accessions
9 were collected and screened against *Fusariumoxysporum* by artificial inoculation under field conditions in
10 threediverse environments. The experiments were laid out in a Randomized Complete Block Design with
11 three replications. Data were collected on plant height (cm), number of leaves per plant, number of fruits
12 per plant, and fresh fruits yield per plant (kg) and analyzed using IRRRI STAR software. Disease scoring
13 was done using a 0-9 point rating scale in the evaluation trials. **Result:** The results show some
14 significance levels ($P<0.05$) in the traits studied within the accessions. The screening result revealed that
15 none of the twelve prominent accessions is highly resistant to the pathogen. One accession was
16 resistant. Three were moderately resistant; three were moderately susceptible and five were highly
17 susceptible. **Conclusion:** The accessions that were resistant and moderately resistant could be used as
18 a gene donor for breeding for tomato cultivars resistant to *Fusariumoxysporum*. Moreover, the accessions
19 that have been identified to be resistant and moderately resistant with high yield should be cultivated
20 before improved cultivars that will be resistant to *Fusariumoxysporum*will be readily available and
21 accessible in rainforest and derived guinea savanna agro-ecological zones of Nigeria.
22

23 **Keywords:** *Fusariumoxysporum*; Resistance; Screen; Susceptible; Tomato.
24

25 1. INTRODUCTION

26 Tomato (*Solanumlycopersicum L.*) is one of the most significant vegetable crops globally. It is widely
27 cultivated for its nutritional and economic values [1]. It can be grown in diverse agro-ecological regions
28 throughout the year provided irrigation facilities are available. Africa's total tomato production was
29 approximately 24 million metric tons in 2023, with Egypt, Nigeria, and Morocco being the leading
30 producers [2]. It is rich in vitamins and minerals [3]. It also contains phytochemicals that are excellent
31 antioxidants that can reduce the risk of heart disease [4]. Tomato fruit quality relies on the cultivar,
32 cultural practices adopted, harvest time and method, storage, and handling procedures [5]. The fruits can
33 be processed industrially or domestically. The crop's economic importance cannot be neglected due to its
34 substantial contribution to food security and income for millions of farmers [6].
35

36 Despite tomato significance, its production is limited by various biotic factors, including Fusarium wilt,
37 caused by the soil-borne pathogen *Fusariumoxysporum f. sp. lycopersici (Fol)*. Fusarium wilt is one of the
38 most significant diseases affecting tomato production globally. The incidence is more pronounced in
39 acidic soils and tropical regions where the environment favoured it [7]. Fusarium wilt is a vascular disease
40 that significantly impedes tomato productivity by colonizing the xylem vessels leading to leaf yellowing,
41 stunted growth, wilting, and plant death [8]. According to Jarvis [9], economic losses due to Fusarium wilt
42 are huge, affecting production in the field and greenhouse production systems.

43
44 Given the persistent threat posed by Fusarium wilt, the development and deployment of resistant tomato
45 varieties are critical for the sustainable management of the disease. Resistant cultivars provide the most
46 cost-effective and environmentally friendly means of controlling Fusarium wilt [10]. However, the
47 effectiveness of these cultivars is often compromised by the emergence of new Fol races and pathogen
48 variants. Therefore, continuous evaluation of tomato accessions for resistance to Fusarium wilt is
49 essential to identify and develop cultivars that can withstand current and emerging strains of the pathogen
50 [11]. The research aims at screening and identifying Nigeria tomato accession (s) that are resistant to
51 Fusarium Wilt.

52

53 **2. MATERIALS AND METHODS**

54

55 **2.1 The study locations**

56 The experiment was conducted in three field environments; Biological Garden of the University of Medical
57 Sciences Ondo-City, Teaching and Research Centre, Ekiti State University, Ado-Ekiti and Oke-Ako/Irele
58 Farm Settlement, Oke-AkoEkiti, all in Nigeria during the early cropping season of 2024 and at the
59 Department of Crop, Horticulture and Landscape Design Laboratory, Ekiti State University, Ado-Ekiti
60 Nigeria. Ondo and Ado-Ekiti lie in the rainforest agroecological zone, while OkeAko-Ekiti lies in the
61 derived guinea savanna.

62

63 **2.2 Experimental materials**

64 The experimental materials used for the work comprised 12 Nigeria tomato accessions obtained in
65 different agroecological zones of Nigeria (Table 1). The tomato accessions were named after the place of
66 their collection for easy identification. The pathogen, *Fusariumoxysporium* was isolated from infected
67 tomato **plant leaves and** roots showing typical symptoms of the pathogen from the tomato field of the
68 Teaching and Research Farm of Ekiti State University, Ado-Ekiti.

69

70

71

72

73 **Table 1:** List of 12 tomato plant accessions screened for resistance to *Fusariumoxysporium* and their
 74 place of collection.

Accession Code	Source of collection	of Agro-Ecological Zone	Accession Code	Source of collection	of Agro-Ecological Zone
Acc. 1	Ikole-Ekiti,	FZ	Acc. 7	Otun-Ekiti	DS
Acc. 2	Calabar	FZ	Acc. 8	Kabba	DS
Acc. 3	Okitipupa	FZ	Acc. 9	Ilorin	SGS
Acc. 4	Ilesa	FZ	Acc. 10	Lafia	SGS
Acc. 5	Akpoka	DS	Acc. 11	Zaria	NGS
Acc. 6	Oro	DS	Acc. 12	Kano	NGS

75 Note: NGS: Northern Guinea Savanna; SGS: Southern Guinea Savanna; DS: Derived Savanna; FZ:
 76 Forest Zone

77

78 **2.3 Isolation, purification and its pathogenicity test of the *Fusariumoxysporium* inoculum**

79 Infected tomato **plant leaves and** roots showing typical symptoms of *Fusariumoxysporium* were chopped
 80 into pieces with a sterilized chopper and surface sterilized with 0.1 per cent mercuric chloride for 35
 81 seconds and washed with sterilized water. The water was drained properly and transferred into potato
 82 dextrose agar (PDA) medium in Petri dishes aseptically. 40 µg l⁻¹ of streptomycin was added to the Petri
 83 dishes to avoid bacterial contamination before pouring the potato dextrose agar. The PDA Petri dishes
 84 were incubated for 120 hours at 28 ± 2 °C. After three days of inoculation, radiating mycelia growth was
 85 seen at the edges of the infected bits and the edge of the fungal colonies were carefully transferred to
 86 PDA medium slants in a refrigerator at 10°C and were periodical sub-culturing for the studies. The
 87 pathogen was identified as *Fusariumoxysporium* based on its morphological and cultural characteristics
 88 [12-13]. The pathogenicity of the isolates was established using Koch's postulates. The agar slants and
 89 Petri plates containing *Fusariumoxysporium* inoculums were stored according to Harlapure *et al.* [14].

90

91 **2.4 Field Experimental design and cultivation condition**

92 The experiment was laid out in a Randomized Complete Block Design (RCBD) with three replicates
 93 across three environments. The seeds of different tomato accessions across different agroecological
 94 zones of Nigeria were nursed at the standard nursery of Ekiti State University Teaching and Research
 95 Farm, Ado-Ekiti and transported to different experimented sites for transplanting. The seedlings were
 96 transplanted on a well-prepared plot at 18:00 of the day. Each of the accessions was raised in 8 m² plots.
 97 Good agronomic practices were adopted to obtain perfect plants.

98

99

100

101 **2.5 Inoculum preparation and inoculation**

102 The isolated and purified *Fusariumoxysporium* inoculum stored at 50C was re-cultured in a PDA medium.
103 Miura et al [15] procedure was adopted in the preparation of conidial suspension. Tween 20 was added to
104 gelatin (0.02% Tween 20 in 0.25% gelatin) to the prepared suspension to enhance a proper adherence of
105 conidia to the tomato aerial parts [16]. The tomato plant was inoculated 15 days after transplanting by
106 spraying the prepared 1 x 10⁶ spores per ml of conidial suspension containing 0.02% Tween 20 in 0.25%
107 gelatin per plot using a knapsack sprayer. The inoculum was sprayed around 18:00 hours of the day and
108 ensured that the entire plant surface became wet with conidial suspension and left overnight. Water was
109 sprayed with a knapsack sprayer on the tomato plants six times at two-hour intervals after 12 hours of
110 inoculation.

111

112 **2.6 Disease assessment, data collection and analysis**

113 Disease scoring of the inoculated tomato plants was done 50 days after inoculation according to
114 Challagulla et al.[17]. Data were collected plant height (cm), number of leaves per plant, number of fruits
115 per plant and fresh fruit yield per plant (Kg). The data collected was analyzed using IRR I STAR software
116 (18). Means were separated by Duncan's multiple range test (DMRT) (P = 0.05).

117

118 **3.RESULTS AND DISCUSSION**

119 **Table 3** presents the mean values for fresh fruit yield per plant and some other yield components of
120 twelve (12) Nigeria tomato plant accessions screened against fusarium wilt across three environments.
121 The parameters studied in the experiment show some significant levels (P>0.05) among the twelve
122 accessions. The level of significance indicated the extent of genetic variability among the accessions.
123 This indicated that genetic improvement can be made within these accessions [19].The result shows that
124 accession from Zaria (127.33) had the highest mean value for plant height, followed by Ilesa and Otun-
125 Ekiti with 126.00 and 121.00 respectively. Okitipupa (35.00) recorded the highest value for the number of
126 fruits per plant followed by Kabba (30.33). Vegetatively, Ilesa (72.33) had the highest number of leaves
127 per while Kano (26.00) had the least number of leaves per plant. Zaria (1.84) had the highest fresh fruit
128 yield per plant followed by Kabba (1.54). Accession from Otun-Ekiti and Oro dies off during the flowering
129 stage. They couldn't produce fruits. The variances observed in these traits among twelve accessions
130 could be attributed to different genetic make-up of the accessions [20].

131

132

133

134

135

136 Table 2: Scale for rating of disease in tomato plant

Grade	Disease severity	Host response
0	No lesion or wilt observed	Highly Resistant
1	Small brown specks of pin point size	Resistant
2	Small roundish to slightly elongated, necrotic gray spots, about 1-2 mm in diameter, with a distinct brown margin. Lesions are mostly found on the lower leaves	Moderately Resistant
3	Lesion type same as in 2, but significant number of lesions on the upper leaves	Moderately Resistant
4	Typical susceptible Fusarium lesions, 3 mm or longer infecting less than 4% of leaf area	Moderately Susceptible
5	Typical susceptible Fusarium lesions of 3mm or longer infecting 4-10% of the leaf area.	Moderately Susceptible
6	Typical susceptible Fusarium lesions of 3 mm or longer infecting 11-25% of the leaf area.	Susceptible
7	Typical susceptible Fusarium lesions of 3 mm or longer infecting 26-50% of the leaf area.	Susceptible
8	Typical susceptible Fusarium lesions of 3 mm or longer infecting 51-75% of the leaf area. Many leaves are dead.	Highly Susceptible
9	Typical susceptible Fusarium lesions of 3 mm or longer infecting more than 75% leaf area affected. The plant dies-off.	Highly Susceptible

137
138 **Table 4** presents the summary of tomato accessions wilt severity recorded across the three
139 environments. The result shows that none of the accessions evaluated was highly resistant to fusarium
140 wilt disease. Accession from Kano was resistant to fusarium wilt. Accessions from Ikole, Calabar, and
141 Zaria were moderately resistant. Accessions from Okitipupa, Akpoka, and Kabba were moderately
142 susceptible while accession from Ilesa, Ilorin, Lafia, Oro and Otun-Ekiti were highly susceptible. This work
143 is in agreement with the findings of Sultan et al., [21] who reported different host responses in some
144 hybrid tomato plants to fusarium wilt. The disparity in host response to fusarium wilt within the accessions
145 could be a result of the differences in their genetics [20]. The resistant and moderately resistant
146 accessions identified can be exploited in a breeding programme for the development of fusarium wilt
147 resistant for commercial cultivars. Accession from Kano recorded a poor yield but is resistant to fusarium
148 wilt.

149 150 **4. Conclusion**

151 This work revealed that none of the tomato plant accessions grown in the prominent tomato growing
152 areas in different parts of Nigeria were highly resistant to fusarium wilt under field artificial inoculation.

153 Only one accession of the twelve accessions screened was found to be resistant. However, the only
 154 resistant varieties had poor fruit yield. The moderately resistant accessions could be exploited in disease
 155 resistance breeding programs using plant breeding techniques for the development of varieties Farmers
 156 are advised to grow the accessions that are moderately resistant in Southwestern Nigeria to avert fruit
 157 yield loss due to fusarium wilt pending the time tomato cultivars that are resistance to fusarium will readily
 158 available and affordable.

159
 160 **Table 3:** Mean performance of parameters taken from 12 Nigeria Tomato accessions screened against
 161 Fusarium wilt disease across the three environments.

Acc.	PH (cm)	NLP-1	NFP-1	FFYP-1 (kg)
Ikole-Ekiti	90.00 ^{ab}	53.67 ^{abc}	24.00 ^{abc}	1.26 ^{bc}
Calabar	79.67 ^{ab}	72.33 ^f	12.33 ^{de}	1.73 ^a
Okitipupa	60.00 ^b	48.00 ^a	35.00 ^e	1.52 ^{abc}
Ilesa	126.00 ^a	72.33 ^{bcd}	7.67 ^{bcd}	0.74 ^{de}
Akpoka	75.67 ^{ab}	51.67 ^a	20.67 ^{cde}	1.52 ^{abc}
Zaria	127.33 ^a	61.67 ^{bcd}	24.33 ^a	1.84 ^{cd}
Otun-Ekiti	121.00 ^a	55.00 ^{ab}	0.00 ^{de}	0.00 ^f
Kabba	90.00 ^{ab}	66.67 ^{abc}	30.33 ^{bc}	1.54 ^{abc}
Ilorin	66.00 ^b	41.33 ^{ab}	17.67 ^{abc}	0.72 ^{de}
Lafia	66.67 ^b	34.67 ^{def}	7.67 ^e	0.43 ^{ef}
Oro	53.69 ^b	32.00 ^{ef}	0.00 ^{ab}	0.00 ^f
Kano	43.68 ^c	26.00 ^f	18.00 ^{cd}	0.34 ^{ab}

162 Means with the same letter(s) in each Column are not significantly different (P<0.05) according to
 163 Duncan's Multiple Range Test (DMRT). Note: ACC= Accessions, PH= Plant Height, NLP= Number of leaf
 164 per plant, NFP= Number of fruits per plant, FFYP= Fresh fruits yield per plant.

165 **Table 4:** Summary of Tomato accessions wilt severity recorded in across the three environments.

Accessions	Host Response	Accessions	Host Response
Ikole-Ekiti,	Moderately Resistant	Kabba	Moderately Susceptible
Calabar	Moderately Resistant	Ilorin	Highly Susceptible
Okitipupa	Moderately Susceptible	Lafia	Highly Susceptible
Ilesa	Highly Susceptible	Oro	Highly Susceptible
Akpoka	Moderately Susceptible	Kano	Resistant
Zaria	Moderately Resistant	Otun-Ekiti	Highly Susceptible

166
 167
 168

169 **REFERENCES.**

- 170 1. Van Eck J, Kirk DD, Walmsley AM. Tomato (*Lycopersicon esculentum*). In K Wang, ed, Methods
171 in Molecular Biology, Agrobacterium Protocols, Humana Press Inc., Totowa. 2006; 343: 459-473
- 172 2. USDA. USDA FoodData Central: Tomatoes, Red, Raw. Washington, D.C.: USDA 2023.
173 Retrieved from <https://fdc.nal.usda.gov>
- 174 3. FAOSTAT. Food and Agriculture Organization of the United Nations. Global Tomato Production,
175 Rome; 2022. Retrieved from <http://www.fao.org/faostat>
- 176 4. Krishna KR. Effect of chitosan coating on the physicochemical characteristics of guava
177 (*Psidium guajava* L.) fruits during storage at room temperature. Indian J. Sci. Technol. 2014;
178 4:554–558.
- 179 5. Mulat A, Enyew AZ, Dawit F, Ananda MHC. Determination of Heavy Metals in Tomato and its
180 Support Soil Samples from Horticulture and Floriculture Industrial area, Ziway, Ethiopia. Res Dev
181 Material Sci. 1019; 10(1). RDMS.000729.2019. DOI: 10.31031/RDMS.2019.10.000729
- 182 6. Abulosoro PF, Ogunjimi SI, Abulosoro SA. Farmers' perception on the strategies for
183 increasing tomato production in Kabba-Bunu Local Government Area of Kogi State, Nigeria.
184 Agrosearch. 2014;14(2):144-153 <http://dx.doi.org/10.4314/agrosh.v14i2.5>
- 185 7. Srinivas C, Devi DN, Murthy KN, Mohan CD, Lakshmeesha TR, Singh BP, Kalagatur NK,
186 Niranjana SR, Hasheem A, Algarawi AA, Tabassum B, Abd-Allah EF, Nayaka SC, Srivastava
187 RK. *Fusarium oxysporum* f. sp. *lycopersici* causal agent of vascular wilt disease of tomato: Biology
188 to diversity – A review. Saudi Journal of Biological Sciences. 2019; 26; 1315-1324
- 189 8. Hwang J, Ahn J, Choi J, Hwang Y, Jeon H, Kim J. Functional analysis of pathogenesis-related
190 genes involved in *Fusarium* wilt resistance in tomato. Frontiers in Plant Science, 2012; 12,
191 650783. doi:10.3389/fpls.2021.650783
- 192 9. Jarvis, WR. *Fusarium wilt of tomatoes*. St. Paul, MN: American Phytopathological Society. 2018.
- 193 10. Reis A, Boiteux LS, Lopes CA, da Silva JBC. Efficacy of crop rotation and soil solarization for
194 management of *Fusarium* wilt in tomato. Plant Disease, 2019; 103(5), 1012-1018.
195 doi:10.1094/PDIS-10-18-1727-RE
- 196 11. Huang W, Zhao X, Li Y, Liu H, Han L. CRISPR/Cas9-based genome editing for disease resistance
197 in tomato. Plant Biotechnology Journal, 2019; 17(7), 1447-1454. doi:10.1111/pbi.13079
- 198 12. Tuite J. (1969). Plant pathological methods, fungi and bacteria. USA: Burges Publishing
199 Company. 1969.
- 200 13. Namrata RPS, Verma R, Bisen R, Singh R, Teli, B. Inheritance of blast disease resistance in the
201 cross hur 3022 x tetep of rice (*Oryza sativa* L.) Journal of Experimental Biology and Agricultural
202 Sciences, 2019; 7(6), 529 – 535.
- 203 14. Harlapur SI, Kulkarni MS, Wali MC, Kulkarni S. Evaluation of plant extracts, bio-agents and
204 fungicides against *Exserohilum turcicum* (Pass.) Leonard and Suggs. causing turcicum leaf blight
205 of maize. Journal of Agricultural Sciences. 2007;20:541-544.

- 206 15. Miura Y, Ding C, Ozaki R, Hirata M, Fujimori M, Takahashi W, Cai H, Mizuno K. Development of
207 EST-derived CAPS and AFLP markers linked to a gene for resistance to ryegrass blast
208 (Pyricularia sp.) in Italian ryegrass (Lolium multiflorum Lam.). Theoretical and Applied Genetics,
209 2005; 111, 811–818.
- 210 16. Jia Y, Valent B, Lee FN. Determination of host responses to Magnaporthe grisea on detached rice
211 leaves using a spot inoculation method. Plant Disease, 2003; 87, 129– 133.
- 212 17. Challagulla V, Bhattara S, Midmore DJ. Invitro vs in-vivo inoculation: Screening for resistance of
213 Australian rice genotypes against blast fungus. Rice Science, 2015; 22(3), 132-137.
- 214 18. International Rice Research Institute (IRRI). (2013). Standard evaluation system for rice,
215 International Rice Research Institute, Philippines. www.irri.org
- 216 21. Falade MJ, Agbowuro GO. Evaluation of Sweet Potato (Ipomoea batatas) for Early Blast Disease
217 (Alternariasolani). Int. J. Path. Res., 2024; 13(1), 52-60.
- 218 22. Agbowuro GO, Salami AE, Aluko M, Olajide OO. 2021. Phenotypic variability among African Yam
219 Bean landrace accessions from different agro-ecologies of Nigeria Nigerian Agricultural Journal
220 Vol. 52, No. 1
- 221 23. Sultan Z, Kutama AS, Chadi M. Screening of some commonly cultivated tomato varieties
222 fusarium wilt in Jama'are, Bauchi State, Nigeria. Dutse Journal of Pure and Applied Sciences.
223 2024; 10(1a). <https://dx.doi.org/10.4314/dujopas.v10i1a.18>
224