

Original Research Article

Effect of restraint stress and cadmium chloride administration on cerebral antioxidants in female Wistar rats.

ABSTRACT

Cadmium is a toxic heavy metal which promotes oxidative stress in various organs including the brain. Restraint stress is a model of stress which involves physical constraint to induce homeostatic imbalance. This study aimed to evaluate the interaction of restraint stress and cadmium toxicity on cerebral antioxidants in female Wistar rats. 24 female Wistar rats (180-220g) were randomly divided into 4 groups (n=6 each): Control (CTL), Restraint stress alone (RSS), Cadmium alone (CCC), Cadmium + Restraint stress (RSC). The experimental groups were subjected to cadmium chloride 100mg/kg b.w. orally and restraint stress for 30 minutes using wire mesh. 24 hours post last cadmium administration and restraint stress exposure, all animals were anaesthetized and sacrificed. The brain was excised, homogenized and analyzed for antioxidant enzymes (superoxide dismutase, catalase, glutathione peroxidase), lipid peroxidation (Malondialdehyde). Results showed that there was significant ($p < 0.05$) decrease in superoxide dismutase in cadmium alone and restraint stress alone groups when compared to control. In the restraint stress alone and cadmium alone groups, cerebral catalase was significantly ($p < 0.05$) decreased when compared to Control. Additionally, a marked ($p < 0.05$) decrease in catalase activity was observed in cadmium + restraint group when compared to restraint stress alone and Cadmium alone

groups. Furthermore, cadmium exposure led to a notable ($p < 0.05$) decrease in cerebral glutathione peroxidase when compared to control. The combined exposure to cadmium and restraint stress significantly ($p < 0.05$) decreased cerebral glutathione peroxidase when compared to restraint stress and Cadmium alone groups. In the cadmium alone group, there was a pronounced ($p < 0.05$) increase in Malondialdehyde when compared to the control group. In conclusion, this present study have shown that combined exposure to cadmium and restraint stress increased free radical production and significantly decreased antioxidant system indicating that combined mechanism of both factors adversely induced neurological damage.

Keywords: Cadmium chloride, restraint stress, oxidative stress, antioxidant enzymes, lipid peroxidation.

INTRODUCTION

Stress is any imbalance in homeostasis caused by either extrinsic or intrinsic stressor [1]. Stress is regulated by the coordinated interaction between the nervous, endocrine and immune system through the hypothalamic-pituitary-adrenal (HPA) axis and the sympathetic adreno-medullary (SAM) axis [2]. Restraint stress is an experimental model that mimics human stress experience in animals [3]. Rats are exploratory animals therefore confining and immobilizing them can induce both physical and psychological stress [4]. Research has established a link between repetitive stress, oxidative stress and inflammation dysregulation [5].

One of the major causes of environmental toxicity globally is heavy metal; this is due to increase in urbanization, industrialization and indiscriminate disposal of waste products containing heavy metals [6]. Cadmium (Cd) is a heavy metal which occurs naturally in the earth crust. There is increased bioavailability of Cd in the environment due to increase anthropogenic activities such as electroplating, battery production and sludge fertilization [7]. Cadmium is known to exert toxicological effect on various organs system including the kidney, liver, heart, and brain [8]. Humans are exposed to cadmium either through ingestion of contaminated food and water or inhalation of contaminated air. Cadmium gains entry into the nervous system via the blood brain barrier accumulating in the brain, resulting in alteration in several neurological functions [9]. Cadmium exposure has also been linked to mitochondrial dysfunction

in cerebral neurons resulting in oxidative imbalance [10]. Hence, this study seeks to evaluate the effect of restraint stress and cadmium chloride administration on cerebral antioxidant in female Wistar rats.

2.0 Material and methods

2.1 Chemical and compounds

Cadmium chloride was **manufactured by** Kermel, China. Chloroform, Normal Saline, distilled water, Buffered formalin, phosphate buffer saline were purchased from department of science laboratory, LAUTECH, Oyo, Nigeria.

2.2 Experimental Protocol and Animals

Twenty-four (24) female rats (180-220g) were used for the study. The rats were kept in a **well-ventilated** animal house (12/12 hour light and dark cycle). The rats were acclimatized for two weeks and had unrestricted access to clean water and feed before the experiment. All protocols and treatment procedures were done according to the Institutional Animal Care and Use Committee (IACUC) guidelines, in strict compliance with the National Institutes of Health (NIH) guideline for the care and use of laboratory animals. After acclimatization, the rats were divided randomly into four groups with six (6) rats in each group and the experiment lasted for 21 days.

Group I = Control group (CTL), were given feed and water *ad libitum*.

Group II= Restraint Stress Alone (RSS), were subjected to restraint stress using wire mesh for 30 minutes daily.

Group III= Cadmium Alone (CCC), were administered cadmium chloride (100mg/kg b.w) orally and daily **[11]**.

Group IV= Cadmium+ Restraint stress (RSC) received cadmium chloride (100mg/kg b.w) daily and were subjected to restraint stress for 30 minutes daily.

Animals subjected to restraint stress were placed in a prone position within a wire mesh setup, carefully designed to avoid applying pressure to the head and neck while keeping the rats completely immobile.

The setup ensured that the animals were well-ventilated throughout the procedure and not in pain. Pain is detected in rats by their squeaks [12].

2.3 Collection and Preparation of samples

Twenty four hours post administration; rats were anesthetized by placing each rat in a dessicator containing cotton wools soaked with chloroform. Brain organ was excised and divided into two parts. The first part was homogenized using mortar and pestle on cold ice. Then, it was centrifuged at 5000rpm for ten minutes. The homogenate supernactant was assayed for biochemical analysis. The second part of the brain was used for histological examination.

2.4 Biochemical Tests

Glutathione perioxide (GPx), catalase (CAT), malondialdehyde (MDA), and superoxide dismutase (SOD) activities in brain tissues were evaluated spectrophotometrically in accordance with the enclosed pamphlets attached to commercial kits purchased from Biodiagnostic (Cairo, Egypt).

2.5 Histology Evaluation

Cerebral specimens were collected in a small, labeled test tube containing 10% buffered formalin for light microscopy examination. Twenty-four (24) after specimen collection, tissues were dehydrated in ascending ethanol concentrations (70% 24 h, 90% 1 h, and 100% 1 h). After this, the tissues were cleared by Xylene, embedded in paraffin wax, and cut into small sections (5 μ M) using Reichert's Rotatory Microtome. All sections were stained by Harris Hematoxylin and Eosin (H&E) stain as described by Horobin. (2013). Then the slides were examined under a light microscope.

2.6 Statistical Analysis

SPSS (version 16.0) was used for all statistical analyses. All results obtained are expressed as Mean \pm Standard Error of the Mean (SEM). Data were analyzed using one-way ANOVA and Duncan's *posthoc* test for multiple comparisons. P value < 0.05 was considered to be statistically significant.

3.0 Results and Discussion

Results

Effects of restraint stress and cadmium chloride administration on Cerebral Antioxidant System in female Wistar rats in both control and experimental groups

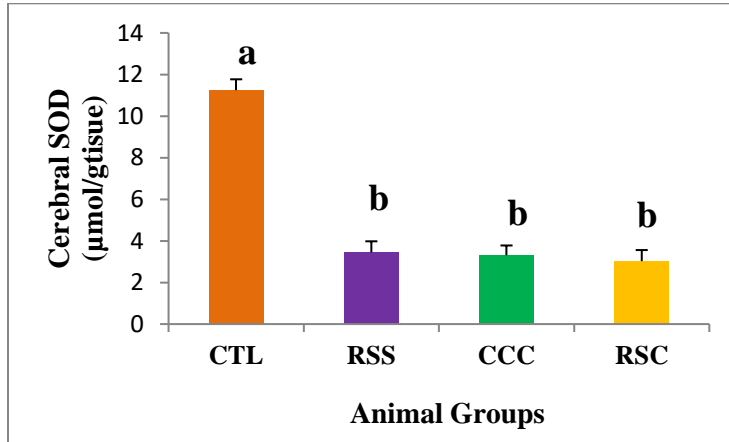


Figure 1: Effect of restraint stress and cadmium chloride administration on cerebral superoxide dismutase in female Wistar rats.

Values are expressed as mean \pm SEM (n= 6). Bars with superscript of different letters are significantly ($p < 0.05$) different from each other. Bars with superscript of same letters are not significantly different from each other.

There was significant ($p < 0.05$) decrease in SOD levels in RSS, CCC, and RSC group when compared to CTL. However, there was no significant difference in RSC when compared to RSS and CCC.

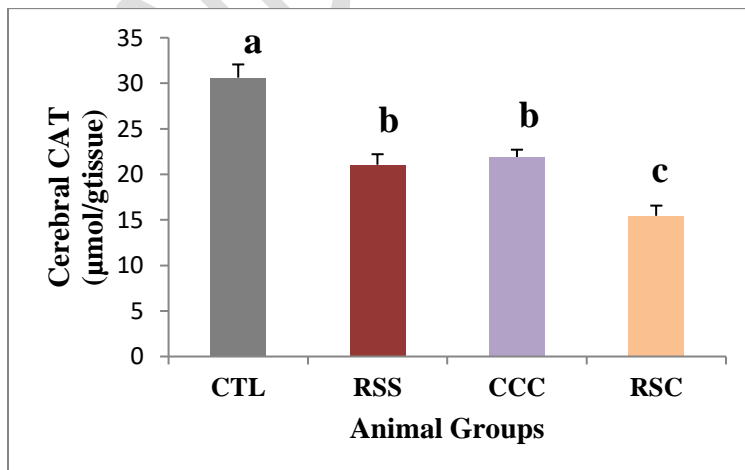


Figure 2: Effect of restraint stress and cadmium chloride administration on cerebral catalase in female Wistar rats.

Values are expressed as mean \pm SEM (n= 6). Bars with superscript of different letters are significantly ($p < 0.05$) different from each other. Bars with superscript of same letters are not significantly different from each other.

There was a significant ($p < 0.05$) decrease in cerebral catalase in both RSS and CCC groups when compared to CTL. Additionally, a significant ($p < 0.05$) decrease in catalase activity was observed in RSC when compared to RSS and CCC groups.

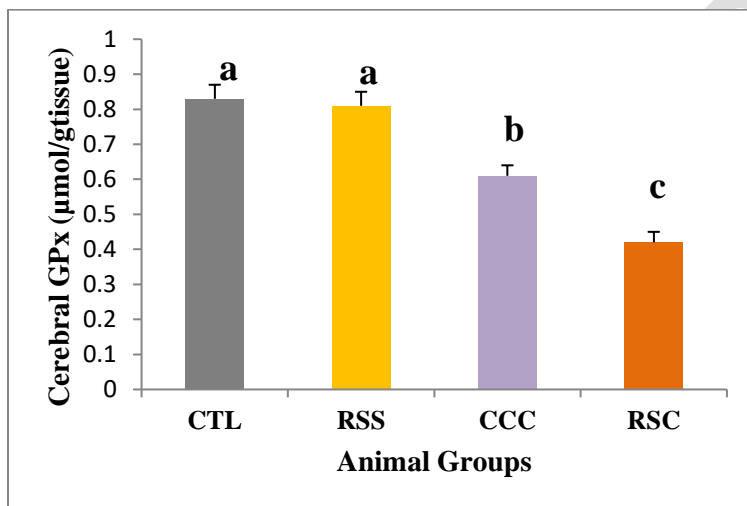


Figure 3: Effect of restraint stress and cadmium chloride administration on cerebral glutathione peroxidase (GPx) in female Wistar rats.

Values are expressed as mean \pm SEM (n= 6). Bars with superscript of different letters are significantly ($p < 0.05$) different from each other. Bars with superscript of same letters are not significantly different from each other.

There was significant ($p < 0.05$) decrease in cerebral glutathione peroxidase in CCC group when compared to CTL. In the RSS group, no significant difference was observed when compared to CTL. However, there was significant ($p < 0.05$) decrease in RSC when compared to RSS and CCC groups.

Table 1: Effect of restraint stress and cadmium chloride administration on cerebral malondialdehyde in female Wistar rats.

Cerebral MDA (nmol/gtissue)	CTL	RSS	CCC	RSC
	25.38±1.11 ^a	26.24±1.18 ^a	32.59±1.64 ^b	33.49±1.99 ^b

Values are expressed as mean \pm SEM ($n = 6$). Mean values with superscript of different letters are significantly ($p < 0.05$) different from each other. Groups with superscript of same letters are not significantly different from each other.

In female Wistar rats exposed to CCC group there was significant ($p < 0.05$) increase in MDA level when compared to CTL. However, there was no significant difference in RSS group when compared to CTL. In the RSC group there was no significant difference when compared to the RSS and CCC groups.

Histological findings

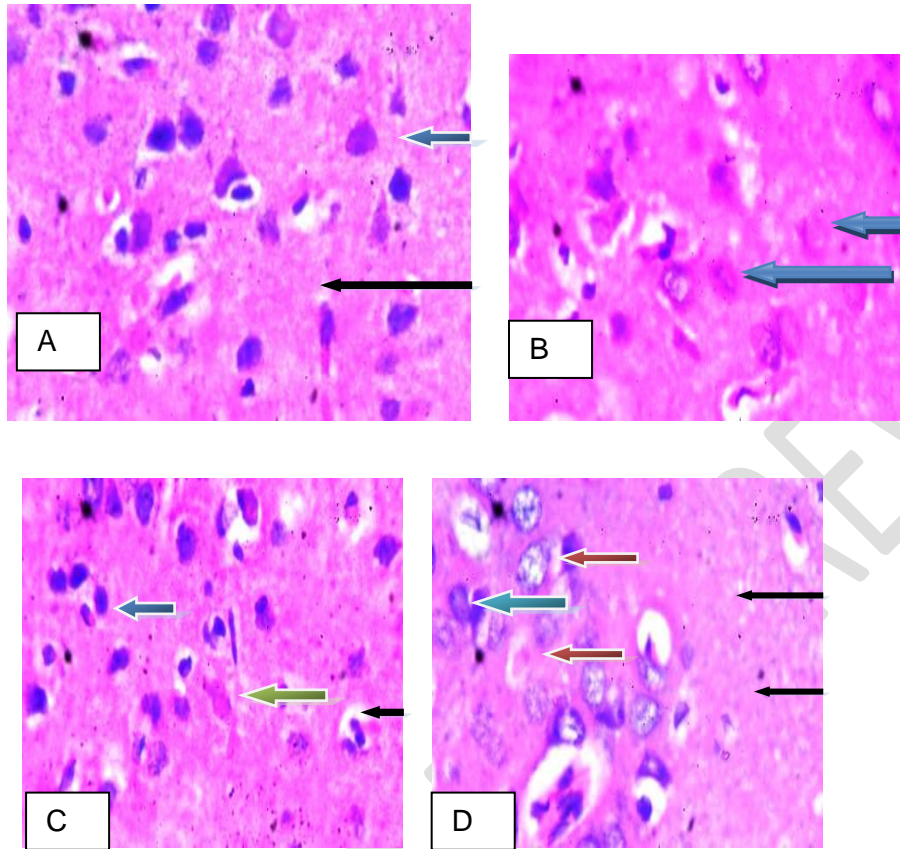


Figure 4: Effect of cadmium chloride administration and restraint stress on cerebral histology in female Wistar rats.

Haematoxylin and Eosin stained photomicrographs of the cerebrum of control and experimental rats. Histological sections of CTL rats (A) and RSS rats (B) showed a normal neuronal (blue arrow), capillaries (red arrow), and stroma (black arrow) while in the CCC (C) rats the cerebrum showed several normal neuronal cells (blue arrow), few neuronal cells with moderate necrosis (green arrow), mildly dilated capillaries (black arrow), and the stroma appear normal (black arrow). In the RSC (D), cerebrum with normal neuronal cells (blue arrow), few depleted neuronal cells (red arrow), capillaries and stroma appear normal (black arrow) was seen (H & E, $\times 400$).

Discussion

Various researches have established that cadmium toxicity induced oxidative stress in animals and human [14, 15, 6]. Chronic stress has been linked to oxidative stress [16]. The brain is particularly vulnerable to oxidative stress, any form of disruption in this organ have a serious impact on the entire body. Antioxidant enzymes such as superoxide dismutase and Catalase protect cells against oxidative damage caused by reactive oxygen species (ROS) [17]. In fig 1 and 2, results observed in the restraint stress alone and cadmium alone groups is in-line with the previous studies of Zafir and Banu, [18].; Alnahdi and Sharaf, [19] where there was significant ($p < 0.05$) decrease in cerebral superoxide dismutase (SOD) and catalase (CAT) levels following cadmium intoxication and stress induction when compared to control. Previous studies have shown that cadmium and restraint stress independently can increase the production of reactive oxygen species (ROS) resulting in the depletion of antioxidant system. Cadmium disrupts cellular defense system by depleting antioxidant levels and increasing accumulation of ROS which can result in lipid peroxidation [20]. Additionally, Cd can replace some essential elements like zinc and iron which are necessary for SOD and CAT activities and thus, decrease their antioxidant capacity [9]. Exposure to repetitive stress leads to hyperactivation of the hypothalamic pituitary adrenal axis and sludge in catecholamine release, this increase in glucocorticoid and catecholamine hormones promotes oxidative metabolism thereby increasing ROS production and decreasing the SOD and CAT levels [21]. The combination of cadmium and restraint stress when compared to cadmium alone and restraint stress alone groups showed no significant difference in SOD level suggest that the level of SOD might have been reduced to a threshold by either cadmium alone or stress alone. However, the combined effect of cadmium and restraint stress further decreased the Catalase level suggesting both cadmium and stress exacerbate oxidative stress.

Glutathione peroxidase (GPx) is an enzymatic anti-oxidant that functions to protect cells from oxidative stress by catalyzing the reduction of hydrogen peroxide (H_2O_2) and lipid peroxide into oxygen and water [22]. Result observed in fig 3 is consistent with the previous research of Shagirtha et al. [23], where cadmium alone group showed a significant ($p < 0.05$) decrease in cerebral glutathione peroxidase when compared to control. This indicates that overproduction of free radicals in cadmium-exposed rats caused oxidative damage to membrane lipid and protein, which led to a decrease in antioxidant enzymes

glutathione peroxidase [23]. The combined exposure to cadmium and restraint stress led to a further reduction in glutathione peroxidase activity, indicating that these factors intensified oxidative stress.

Malondialdehyde is the byproduct of lipid peroxidation, this occurs when oxidative stress damages cell membranes [24]. In table 1, elevated level of MDA observed in the cadmium alone group is consistent with the findings of Ojo et al. [25] where cadmium alone group was significantly ($p < 0.05$) increased when compared to control. This is likely due to the production of reactive oxygen species (ROS) such as peroxy, superoxide and hydroxyl radicals. Indirectly, cadmium exposure generates these radicals by disrupting mitochondrial function which stimulates lipid peroxidation by producing endoperoxidases through non-enzymatic cyclization reaction [26]. The combination of restraint stress and cadmium showed no statistical significant difference when compared to the cadmium alone and restraint stress alone groups. This suggests that cadmium alone might have caused maximal effect on lipid peroxidation in which restraint stress may not cause further increase.

Heavy metals have been associated with significant histological changes in the brain tissues. In this present study, cerebral cortex section examination by light microscope, showed no notable differences among rats in control and restraint alone groups. Cadmium intoxication induced histological changes in the cerebral tissue including mild neuronal necrosis and capillaries dilation, which indicates early stages of neuronal injury driven by cadmium-induced oxidative stress and mitochondrial dysfunction. This is consistent with the findings of Gok *et al.* [27]. Additionally, combined exposure to cadmium and restraint stress led to neuronal degeneration in the cerebrum which could be due to exacerbated disruption of neurovascular integrity. This increased severity in the combined group could be attributed to shared mechanisms such as oxidative stress, neuroinflammation and impaired cellular deoxyribonucleic (DNA) repair capacity.

4.0 CONCLUSION

In conclusion, this present study have shown that combined exposure to cadmium and restraint stress increased free radical production and significantly decreased antioxidant system indicating that combined mechanism of both factors adversely induced neurological damage.

DISCLAMIER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as large language models (Chatgpt, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscript.

INSTITUTIONAL REVIEW BOARD STATEMENT

This study was conducted following the Institutional Animal Care and Use Committee (IACUC) guidelines, in strict compliance with the National Institutes of Health (NIH) guideline for the care and use of laboratory animals.

INFORMED CONSENT STATEMENT

Not applicable

REFERENCE

1. Devi, P.C.B., Reddy, M.A., Zahan, O. and Sharma, J.V.C., 2019. The effect of stress on human life. *Adalya J*, 8, pp.792-811.
2. Mifsud, K.R. and Reul, J.M., 2018. Mineralocorticoid and glucocorticoid receptor-mediated control of genomic responses to stress in the brain. *Stress*, 21(5), pp.389-402.
3. Van Wyk, M., 2022. *Establishing and validating an in vivo rodent model of chronic restraint stress* (Doctoral dissertation, Stellenbosch: Stellenbosch University).
4. Oluwatobi, O.O., Oluwatoyin, O.B., Busuyi, K.D. and Opeyemi, O.G., 2024. Metabolic Effect of Acute Lead and Restraint Stress Exposure on Female Wistar Rats. *Asian Journal of Biochemistry, Genetics and Molecular Biology*, 16(10), pp.17-23.
5. Czarny, P., Wigner, P., Galecki, P. and Sliwinski, T., 2018. The interplay between inflammation, oxidative stress, DNA damage, DNA repair and mitochondrial dysfunction in depression. *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 80, pp.309-321.
6. Genchi, G., Sinicropi, M.S., Lauria, G., Carocci, A. and Catalano, A., 2020. The effects of cadmium toxicity. *International journal of environmental research and public health*, 17(11), p.3782.
7. Hayat, M.T., Nauman, M., Nazir, N., Ali, S. and Bangash, N., 2019. Environmental hazards of cadmium: past, present, and future. In *Cadmium toxicity and tolerance in plants* (pp. 163-183). Academic Press
8. Bhattacharyya, K., Sen, D., Laskar, P., Saha, T., Kundu, G., Ghosh Chaudhuri, A. and Ganguly, S., 2023. Pathophysiological effects of cadmium (II) on human health-a critical review. *Journal of Basic and Clinical Physiology and Pharmacology*, 34(3), pp.249-261.
9. Arruebarrena, M.A., Howe, C.T., Lee, Y.M., and Branco, R.C. (2023). Mechanisms of Cadmium Neurotoxicity. *International Journal Molecular Sciences*, 24, 16558.
10. Li, C.X., Talukder, M., Xu, Y.R., Zhu, S.Y., Wang, Y.X. and Li, J.L., 2024. Cadmium causes cerebral mitochondrial dysfunction through regulating mitochondrial HSF1. *Environmental Pollution*, 360, p.124677.
11. Daverey, J., 2023. The combined effect of radiation and cadmium chloride on the Haematological parameters of mouse. *Journal of science, research and teaching*, 2(8), pp.56-60.

12. Castelhana-Carlos, M.J. and Baumans, V., 2009. The impact of light, noise, cage cleaning and in-house transport on welfare and stress of laboratory rats. *Laboratory animals*, 43(4), pp.311-327.
13. Horobin, R.W., 2013. How histological stains work. Bancroft's theory and practice of histological techniques. Elsevier, Amsterdam, pp 157–171
14. El-Habit OH, Abdel Moneim AE. Testing the genotoxicity, cytotoxicity, and oxidative stress of cadmium and nickel and their additive effect in male mice. *Biol Trace Elem Res* 2014;159(1–3):364–72.
15. Agnihotri, S.K., Agrawal, U. and Ghosh, I., 2015. Brain most susceptible to cadmium induced oxidative stress in mice. *Journal of Trace Elements in Medicine and Biology*, 30, pp.184-193.
16. Juszczak, G., Mikulska, J., Kasperek, K., Pietrzak, D., Mrozek, W. and Herbet, M., 2021. Chronic stress and oxidative stress as common factors of the pathogenesis of depression and Alzheimer's disease: The role of antioxidants in prevention and treatment. *Antioxidants*, 10(9), p.1439.
17. Ighodaro, O.M. and Akinloye, O.A., 2018. First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defence grid. *Alexandria journal of medicine*, 54(4), pp.287-293.
18. Zafir, A. and Banu, N., 2009. Modulation of in vivo oxidative status by exogenous corticosterone and restraint stress in rats. *Stress*, 12(2), pp.167-177.
19. Alnahdi, H.S. and Sharaf, I.A., 2019. Possible prophylactic effect of omega-3 fatty acids on cadmium-induced neurotoxicity in rats' brains. *Environmental Science and Pollution Research*, 26(30), pp.31254-31262.
20. Unsal, V., Dalkıran, T., Çiçek, M. and Kölükçü, E., 2020. The role of natural antioxidants against reactive oxygen species produced by cadmium toxicity: a review. *Advanced pharmaceutical bulletin*, 10(2), p.184.
21. Chainy, G.B. and Sahoo, D.K., 2020. Hormones and oxidative stress: an overview. *Free Radical Research*, 54(1), pp.1-26.
22. Carmo de Carvalho e Martins, M.D., Martins, da Silva Santos Oliveira, A.S., da Silva, L.A.A., Primo, M.G.S. and de Carvalho Lira, V.B., 2022. Biological indicators of oxidative stress [malondialdehyde, catalase, glutathione peroxidase, and superoxide dismutase] and their application in nutrition. In *Biomarkers in Nutrition* (pp. 1-25). Cham: Springer International Publishing.
23. Shagirtha, K., Bashir, N. and MiltonPrabu, S., 2017. Neuroprotective efficacy of hesperetin against cadmium induced oxidative stress in the brain of rats. *Toxicology and industrial health*, 33(5), pp.454-468.
24. Całyniuk, B., Grochowska-Niedworok, E., Walkiewicz, K.W., Kawecka, S., Popiolek, E. and Fatyga, E., 2016. Malondialdehyde (MDA)—product of lipid peroxidation as marker of homeostasis disorders and aging. In *Annales Academiae Medicae Silesiensis* (No. 70, pp. 224-228). Śląski Uniwersytet Medyczny w Katowicach.
25. Ojo, O.A., Rotimi, D.E., Ojo, A.B, Ogunlakin, A. D., and Ajiboye, B.O. (2023) Gallic acid abates cadmium chloride toxicity via alteration of neurotransmitters and modulation of inflammatory markers in Wistar rats. *Scientific Reports* 13, 1577.
26. Kaur, R., Kaur, J., Mahajan, J., Kumar, R. and Arora, S., 2014. Oxidative stress—implications, source and its prevention. *Environmental science and pollution research*, 21, pp.1599-1613.
27. Gök, E. and Deveci, E., 2022. Histopathological evaluation of IBA-1, GFAP activity in the brain cortex of rats administered cadmium chloride. *Arch. Ital. Biol*, 160(1-2), pp.20-7.