

# Antiviral Efficacy of Neem, Garlic, and Ginger Extracts on Newcastle Disease Virus in Poultry: Impact of Concentration and Heat Treatment

## ABSTRACT

**Aims:** Newcastle disease virus (NDV) is a paramyxovirus that causes significant mortality rates in poultry, often known as Newcastle disease or Ranikhet. This virus has the potential to inflict serious economic losses on farmers. As there is no effective therapy for NDV infection, the current research investigated the efficiency of medicinal plant extracts against this virus.

**Place and Duration of Study:** The experiment was conducted at the Department of Physiology, Biochemistry, and Pharmacology and the Department of Microbiology and Veterinary Public Health, Chittagong Veterinary and Animal Sciences University (CVASU), Bangladesh, during a period from January to April 2017.

**Methodology:** Plant samples (neem bark, neem leaf, garlic, and ginger) were collected from the medicinal garden of CVASU. Samples were ground, and 300 g of ground samples were mixed with distilled water in a 1:5 ratio. Mixtures were stirred for 1 hour by an electric stirrer (1000 rpm) and left overnight. All the mixtures were filtrated through Whatmanno. 1 filter paper separately. Finally, aqueous extracts were prepared from the filtrated sample with the help of a round bottom flask of a rotator vacuum evaporator. Then, the extracts were categorized into different groups to determine the possible effects of extract concentration and heat treatment on the antiviral potential of plant extracts. The plant extract was mixed with viable NDV (8 HA; field strain) and kept at 27.3 °C for 30 minutes. To determine virus inactivation, a 0.2-ml mixture was inoculated into nine-day-old embryonated chicken eggs and incubated. After 48 hours, the allantoic fluid was harvested, and a hemagglutination (HA) assay was performed to determine the virus HA titer.

**Results:** The antiviral effect of plant extract is described in terms of HA titers, specifically the geometric mean titer (GMT). The lower GMT titer value of the plant extract showed higher antiviral activity. However, Neem is more efficient against NDV than other extracts (garlic and ginger). The antiviral activities of these extracts can vary due to the concentration and heat treatment (autoclave) of the extracts. The antiviral potency of all plant extracts declines with decreasing concentration. Heat treatment significantly ( $p = 0.02$ ) decreases the plant's extract antiviral efficacy.

**Conclusion:** This study suggests the potential use of common local medicinal plants to treat Newcastle disease in poultry, although active compounds of those plants have not yet been studied. Finally, these plants can be a promising source for developing antiviral drugs against Newcastle disease.

**Keywords:** Newcastle disease, Anti-viral medicine, Medicinal plants, Aqueous extracts, Hemagglutination test.

## 1. INTRODUCTION

Newcastle disease virus (NDV) is a negative-sense single-stranded RNA virus under the family Paramyxoviridae (Mayo, 2002). It causes a severe disease in poultry named Newcastle disease, or Ranikhet. This disease remains a serious economic challenge to all segments of the poultry industry because of its contagious and mortality (0–100%) records (Chollom et al., 2012; Sulaiman et al., 2013), although mortality varies depending on the

18 pathotype of the virus. Medicinal plants are a rich source of natural compounds such as  
19 tannins, polyphenols, proanthocyanidins, sulfonamides, anti-adhesives, etc. that exhibit  
20 antiviral (Mukhtar et al., 2008; Gupta et al., 2014) and anti-inflammatory (Gupta et al., 2014)  
21 activities. Many reports showed that many indigenous communities used their herbal  
22 preparations for veterinary use (Madadgar et al., 2013). Like many other medicinal plants,  
23 Neem (*Azadirachta indica*) has been used in Ayurvedic medicine for more than 2000 years,  
24 and now it is being used in modern medicine, cosmetics, and pharmaceuticals as the global  
25 scenario is changing towards the use of nontoxic plant products. Its medicinal values come  
26 from the fruits, seeds, leaves, roots, and bark (Biswas et al., 2002). Various preparations of  
27 neem obtained from its different parts have been found to exert anti-bacterial, anti-viral, anti-  
28 malarial, anti-oxidant, anti-fungal, anti-mutagenic, anti-carcinogenic, contraceptive, and  
29 antiulcer activity (Subapryya and Nagini, 2005; Bonsu et al., 2012). Garlic has been an  
30 interesting plant for centuries as a medicinal panacea. A broad range of pathogenic  
31 organisms, including bacteria, fungi, protozoa, and viruses, are sensitive to fresh, crushed  
32 garlic (Mehrbood et al., 2009). *Zingiber officinale* (family Zingiberaceae), commonly known as  
33 ginger, is commonly used as an effective medicine against coughs and colds. It was also  
34 found effective against Newcastle disease virus during an *in vitro* experiment (Kikuzaki et al.,  
35 1991; Sharma and Gupta, 1998; Untari et al., 2022). However, since there is a lack of  
36 sufficient information about the antiviral potential (and factors affecting this potential) of  
37 neem leaves, neem bark, garlic, and ginger against the Newcastle disease virus, this study  
38 focused on finding out the effects of concentration and heat treatment (during autoclaving)  
39 on the anti-viral activity of these plant extracts. These plants had been used as traditional  
40 medicines by the natives for many years due to their antibacterial, antifungal, anti-allergic,  
41 anti-viral, and other important medicinal properties.

## 42 43 **2. MATERIAL AND METHODS**

### 44 45 **2.1 Sample collection**

46 The experiment was conducted at the Department of Physiology, Biochemistry, and  
47 Pharmacology and the Department of Microbiology and Veterinary Public Health, Chittagong  
48 Veterinary and Animal Sciences University (CVASU), Bangladesh, from January 2017 to  
49 April 2017.

### 50 51 **2.2 Virus sample**

52 A previously isolated and stored (-80 °C) field strain of the Newcastle disease virus was  
53 collected from a repository of the Department of Microbiology and Veterinary Public Health,  
54 CVASU. After thawing, the virus sample was treated with an antibiotic (Gentamicin) to  
55 prevent bacterial contamination. Then the virus samples were subjected to a  
56 hemagglutination (HA) test to confirm the viability and concentration of the virus.

### 57 58 **2.3 Plant samples**

59 The bark and leaf of Neem (*Azadirachta indica*), Garlic (*Allium sativum*), and Ginger  
60 (*Zingiber officinale*) from the medicinal plant garden of Shahedul Alam Qudary Teaching  
61 Veterinary Hospital, CVASU, Bangladesh.

### 62 63 **2.4 Plant extract preparation**

64 Collected neem leaves, garlic, and ginger were thoroughly cleaned with water to remove dirt  
65 and unwanted materials. Then 300-gram (g) samples were ground using an electrical  
66 grinder. The ground samples (300 g) were mixed with distilled water at the ratio of 1:5  
67 (sample: water) for neem leaf, garlic, and ginger (Iwalokun et al., 2004; Kwawukume et al.,  
68 2013; Gupta and Chaphalkar, 2015). In the case of neem bark, the collected barks were cut  
69 into small pieces and dried at room temperature (27°C) for several weeks. Then dried barks  
70 were pulverized using an electric grinder. Finally, 300 g of powder was mixed with distilled

71 water in a 1:10 ratio (Mahmood et al., 2017). Sample mixtures were stirred for 1 hour by an  
72 electric stirrer (1000 rpm) and left overnight. All the mixtures were filtrated through  
73 Whatmanno. 1 filter paper separately. Then 300 ml of filtrates from different samples were  
74 taken into a round bottom flask of a rotator vacuum evaporator and condensed by the  
75 evaporation of solvent from the filtrate in a water bath at 56.7°C for 3-5 hours. After the  
76 evaporation of solvent from the filtrate, the condensed extracts were preserved in a tightly  
77 corked-labeled bottle and stored at 4 °C.

## 78 79 **2.5 Categorization of plant extracts**

80 Each plant extract was broadly categorized into two groups: Group-I: extract without heat  
81 treatment, and Group-II: extract treated with heat at 121°C for 15 min. Then every group was  
82 further divided into three sub-groups by mixing PBS with concentrated plant extract, such as  
83 sub-group I: 100% extract (concentrated extract); sub-group II: 75% extract (three parts  
84 extract and one part PBS); and sub-group III: 50% extract (one part extract and one part  
85 PBS).

## 86 87 **2.6 Toxicity test of samples**

88 All types of plant extract were checked for any possible toxicity to chicken embryos by an  
89 embryonated egg inoculation assay. 0.2 ml of aqueous plant extract was inoculated in nine-  
90 day-old embryonated chicken eggs (5 eggs per concentration) collected from the Regional  
91 Poultry Farm (RPF), Chittagong. Then all eggs were incubated for 48 hours to check for  
92 embryo mortality.

## 93 94 **2.7 Preparation of 8 HA unit virus**

95 A virus sample (0.2 ml) was propagated in embryonated chicken eggs. After propagation,  
96 allantoic fluid, collected after 48 hours of incubation of eggs, was subjected to an HA test to  
97 determine 1 HA unit. Based on the Hemagglutination (HA) test result, an 8 HA unit virus  
98 concentration was prepared by mixing 15 ml of sterile PBS with 1 ml of allantoic fluid.

## 99 100 **2.8 Embryonated eggs inoculation**

101 Inoculums were prepared by properly mixing the plant extracts from all sub-groups with 8 HA  
102 units of the virus at a 1:1 (500 µl extract with 500 µl virus) ratio in an Eppendorf tube and  
103 were incubated at room temperature for 30 minutes. Five nine-day-old embryonated chicken  
104 eggs per concentration were inoculated with 0.2 ml of inoculum following standard  
105 procedure. The inoculated eggs were incubated for 48 hours. The eggs were candled after  
106 24 hours of inoculation to check embryo mortality. After 48 hours of incubation, eggs were  
107 transferred to a chilling temperature and kept for 24 hours. About 10% control for both  
108 viruses and extracts was maintained during the whole procedure. After 24 hours of chilling,  
109 allantoic fluids were harvested from each egg using a sterile syringe. The fluid was taken  
110 into the falcon tube. Then the HA test was carried out using 1% chicken RBC, collected from  
111 a specific pathogen-free flock, to determine the virus titer in the allantoic fluid.

## 112 113 **2.9 Statistical analysis**

114 All data were entered into a spreadsheet in MS Excel 2010. The data were sorted, cleaned,  
115 and coded using the Excel program before exporting for the analysis of the Geometric mean  
116 titer (GMT). Finally, the GMT of HA was used to determine reduced HA titer (%) and the  
117 effect of two key factors-concentration and heat treatment, on the antiviral potential of plant  
118 extract.

# 119 120 **3. RESULTS**

## 121 122 **3.1 Toxicity of Extract**

123 In the toxicity testing assay, no plant extract caused mortality in chicken embryos within 48  
 124 hours of incubation (Table 1).

125 **Table 1: Determination of toxicity of extracts in embryonated eggs**

126

Plant extracts	Concentration (%)	Total no of eggs inoculated	Death observed in embryonated eggs			
			24 hours		48 hours	
			without heat-treated plant extracts used	heat treated Plant extracts used	24 hours	48 hours
Neem Leaf	100	20	A (0/5)	A(0/5)	A (0/5)	A(0/5)
	50	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
	33	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
Neem Bark	100	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
	50	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
	33	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
Garlic	100	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
	50	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
	33	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
Ginger	100	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
	50	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)
	33	20	A (0/5)	A (0/5)	A (0/5)	A (0/5)

127

A= Indicates all alive eggs after 48 hr inoculation of extract

128

### 3.2 Overall antiviral potential of extracts

129 The anti-viral effect (reduced HA) of plant extract was found to be variable when compared  
 130 with the virus control group (Table2). For the NDV field strain, the geometric mean of HA titer  
 131 (GMT) of the virus control group was 128 HA units. 100%, 50%, and 33% of aqueous extracts  
 132 of neem bark from group-I (without autoclave) exhibited 0 HA titer (GMT), which indicates  
 133 complete (100%) inactivation of the virus in the embryonated chicken egg assay. Group II  
 134 (using an autoclave) had GMT values of 16, 64, and 84, which correspond to decreases  
 135 in viral titer of 87.5%, 50%, and 34.3% at 100%, 50%, and 33% concentrations, respectively.  
 136 On the other hand, ginger extract was found to be the least effective medicinal plant against  
 137 the ND virus. It showed high GMT values (101.59, 111.43, and 111.43 for group-I, and 97,  
 138 111.43, and 128 for group-II) that reflect the weak anti-viral potential (20.6%, 12.9%, and  
 139 12.9% GMT reduction by sub-groups of groups-I; 24.21%, 12.9%, and 0% GMT reduction by  
 140 sub-groups of group-II) of ginger against NDV. All neem leaf extract sub-groups of groups I  
 141 showed similar anti-viral activity (GMT 64 and 50% GMT reduction) despite being different in  
 142 concentration, although sub-groups of group II showed gradually decreased anti-viral activity  
 143 due to dilution. The GMT values were 64, 84, and 111.43, reflecting 50%, 34.3%, and 12.9%  
 144 virus titer reduction, respectively, for three sub-groups of neem leaf extract. Garlic extracts  
 145 are good enough to reduce ND virus titers if they are not autoclaved. During this study, non-  
 146 autoclaved extracts reduced 58% to 71.2% of the virus titers, whereas **heat-treated**  
 147 **(autoclaved)** extracts reduced 0% to 43.3% only.

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151 **Table 2: HA activity of Plant extract against NDV in embryonated eggs**

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Concentration	Plants	No of	HA titer of virus treated	Control	Reduced HA (%)
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	Extract	eggs (n)	with extracts Geometric mean-titer (GMT)		GMT	=(*C-*E)/*C	
			Without heat treated	Heat treated		Without heat treated	Heat treated
<b>100% (1:1)</b>	Neem leaf	10	64	64	128	50	50
	Neem Bark	10	0	16		100	87.5
	Garlic	10	36.75	84		71.2	34.3
	Ginger	10	101.59	97		20.6	24.21
<b>50% (1:2)</b>	Neem leaf	10	64	84		50	34.3
	Neem Bark	10	0	64		100	50
	Garlic	10	45.25	97		64.6	24.2
	Ginger	10	111.43	111.43		12.9	12.9
<b>33% (1:3)</b>	Neem leaf	10	64	111.43		50	12.9
	Neem Bark	10	0	84		100	34.3
	Garlic	10	53.81	128		58	0
	Ginger	10	111.43	128		12.9	0

163 Haemagglutination titer of virus control group,

164 Haemagglutination titer of extracts treated embryonated group eggs

155

156 However, the antiviral activity of garlic also varied due to dilution (GMT 36.5, 45.25 and

157 53.81 in group-I; 84, 97 and 128 in group-II).

158

### 159 3.3 Effect of concentration and heat treatment on the antiviral potential of

### 160 plant extract

161 There was a favorable correlation found between plant extract concentration and antiviral

162 activity.

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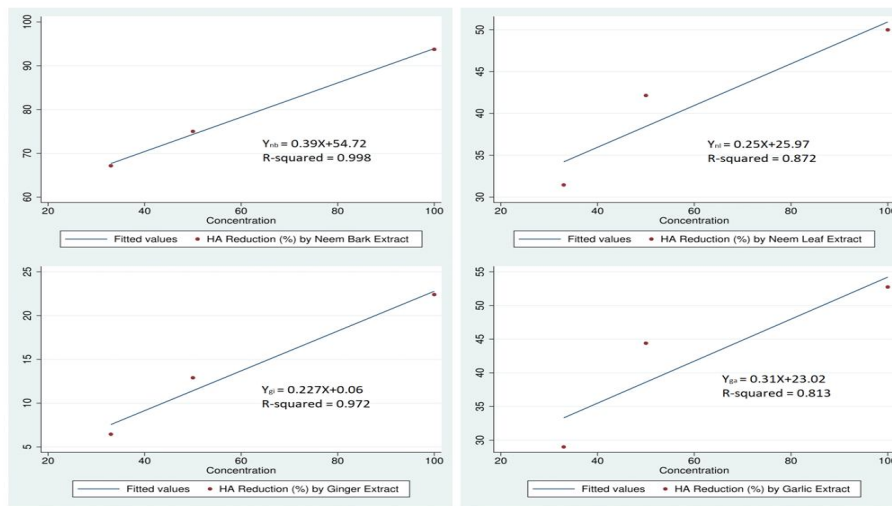
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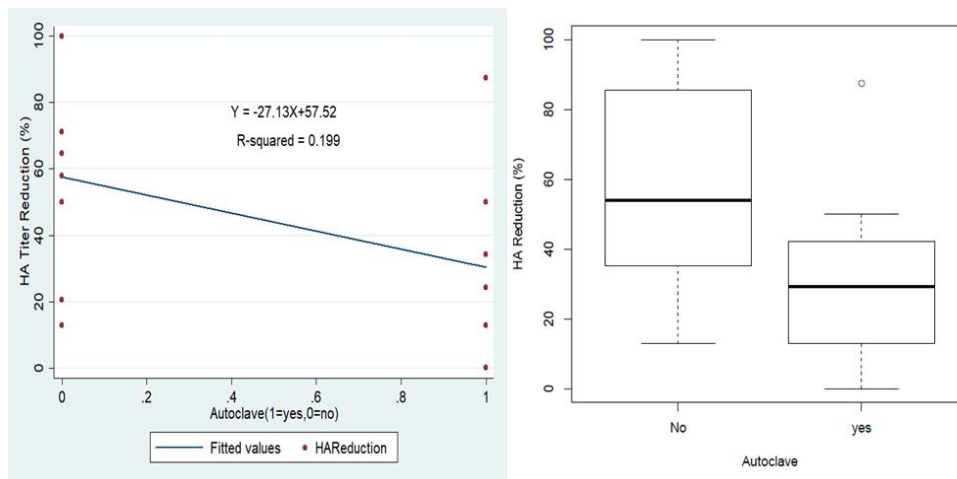


170 **Figure 1:** Regression lines reflecting effects of concentration of plant extracts on their anti-viral

171 potential

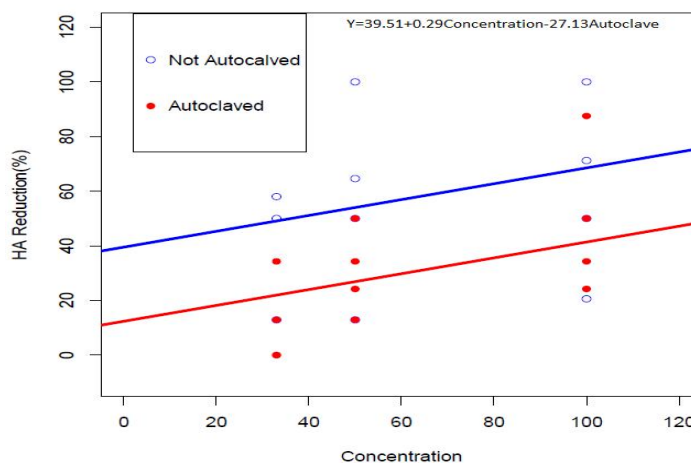
172

173 The antiviral potential of all plant extracts decreases if the concentration of extract  
 174 decreases. The regression analysis shows that concentration adversely affects the antiviral  
 175 activity of neem bark ( $R^2=0.998$ ; slope=0.39), Neem leaf ( $R^2=0.872$ ; slope=0.25), Garlic  
 176 ( $R^2=0.813$ ; slope=0.31) and Ginger ( $R^2=0.972$ ; slope=0.23) extracts (Figure 1). On the other  
 177 hand, heat treatment has a negative effect on antiviral potential of plant extract (Figure  
 178 2). Due to autoclaving, the antiviral properties of plants decrease significantly ( $p=0.02$ ).  
 179



180 **Figure 2:** Regression line and box plot reflecting the effect of heat treatment (during  
 181 autoclaving) on the antiviral activity of studied medicinal plant extracts  
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183 However, the effect of concentration on the virus titer reduction ability of plant extract is not  
 184 modified by heat treatment (Figure 3).  
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 201 **Figure 3:** Combined effect of concentration and heat treatment on anti-viral activity of  
 202 studied medicinal plants  
 203

204 **4. DISCUSSION**

205 Medicinal plants are the ultimate source for treating living organisms in humans and animals  
 206 suffering from infectious and non-infectious diseases (Arunkumar and Muthuselvan, 2009).  
 207 In this study, four aqueous extracts of three plant species, neem (*Azadirachta indica*), garlic  
 208 (*Allium sativum*), and ginger (*Zingiber officinale*), were prepared and tested against NDV,

209 which causes economic losses to the poultry industry across the world. In this study, the  
210 antiviral impact of plant extract is described in terms of HA titers, especially the geometric  
211 mean titer (GMT). The lower the GMT titer value, the stronger the antiviral effect of plant  
212 extract, which was also described in another study by Sulaiman et al. (2011), Ong et al.  
213 (2014), and Mahmood et al. (2017). In this research, all the studied plants were found to be  
214 non-toxic to the chicken embryo, and their antiviral activities against NDV can vary due to  
215 differences in species, concentration, and heat treatment. These findings prove the previous  
216 research findings (Keqiang and van Bruggen, 2001; Mahmood et al., 2017). Neem is  
217 comparatively more effective against NDV compared to other plant extracts; this may be due  
218 to the difference in their phytochemical properties (Song et al., 1997; Parida et al., 2002).  
219 Without autoclaving, the use of neem extracts (both leaf and bark) shows almost similar  
220 effectiveness against NDV, irrespective of extract concentration. But after autoclaving, the  
221 antiviral activities of neem extract decrease with the decrease in extract concentration  
222 (Sulaiman et al., 2011; Ong et al., 2014; Mahmood et al., 2017). This phenomenon indicates  
223 the negative effect of heat treatment on the antiviral potential of neem plant extracts, which  
224 may happen due to the breakdown or degeneration of biochemical compounds (such as  
225 nimbidin, sodium nimbidate, epicatechin, catechin, etc.) that trigger antiviral activity (Biswas  
226 et al., 2002). Despite being affected by heat and concentration, the neem plant can be an  
227 incredible source of anti-viral therapeutics against NDV, as it also plays an important role in  
228 strengthening the immune system and inactivating viruses effectively (Sadekar et al., 1998;  
229 Awolu et al., 2013; Elbasuni et al., 2023). Garlic is a rich source of allicin and ajoene (Ankri  
230 and Mirel, 1999). Previously, garlic was found effective against infectious bronchitis virus  
231 during propagation in embryonated chicken eggs (Shojai et al., 2016). Besides, it can  
232 significantly increase the antibody titer against the ND and avian influenza viruses in poultry  
233 (Eid and Iraqi, 2014). This study found that garlic can also inactivate NDV; however, the  
234 extract's concentration and heat treatment have a major impact on its antiviral activity.  
235 Ginger, a source of different phenolic derivatives such as gingerol, paradole, bisabolene,  
236 zingerone, etc., has been used as an anti-viral agent in humans and animals for many years.  
237 Gupta and Chalkar (2015) described the anti-inflammatory and anti-viral potential against  
238 NDV of garlic. During this research, garlic was found to be able to inactivate NDV, but to a  
239 narrow extent. However, interestingly, the anti-viral activity of undiluted extract may be  
240 increased due to heat treatment. Due to the presence of gingerol, paradole, bisabolene,  
241 zingerone, zingiberol, and 6-shogaol, ginger (*Zingiber officinale*) can act as an anti-  
242 inflammatory and anti-viral agent. In previous research, ginger extracts were found effective  
243 against NDV (Mishra et al., 2012; Gupta and Chaphalkar, 2015), but in this study, ginger  
244 showed a lower antiviral effect against NDV, which may be due to the species differences of  
245 ginger and/or strain differences of virus used in these studies. Furthermore, the antiviral  
246 activity of ginger is conversely related to the dilution and autoclaving of the extract.

247

## 248 5. CONCLUSION

249 The aqueous extracts of all studied plants are effective against the Newcastle disease virus  
250 of poultry, although neems-bark extract is likely the most effective among the plant extracts.  
251 Heat treatment can affect the antiviral potential of medicinal plant extracts, which is worse in  
252 the case of garlic extract. Besides, the concentration of extract also has a positive  
253 relationship with the antiviral activities of studied plants. Although the present study has not  
254 studied details, especially active compounds, of the studied plants, it unveils the possibility of  
255 using common local medicinal plants to treat Newcastle disease in poultry.

256

## 257 STUDY LIMITATIONS AND FUTURE DIRECTIONS

258

259 This study focused solely on the antiviral effectiveness of neem, garlic, and ginger aqueous  
260 extracts in embryonic eggs against NDV. The current study did not investigate details,  
261 particularly the active components of the studied plants. So further study is needed to

262 identify the active elements of these plants and their potential impacts on live birds. It will aid  
263 with the development of novel antiviral medications for Newcastle disease.

264

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266

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270

## 271 **COMPETING INTERESTS**

272

273 The authors declare that they have no known competing financial interests or personal  
274 relationships that could have appeared to influence the work reported in this paper.

275

## 276 **AUTHORS' CONTRIBUTIONS**

277

278 Mohammad Mahbub Hasan - Conceptualization, Methodology, Data curation, Formal  
279 analysis, writing, editing, Abdul Ahad- Conceptualization, Supervision, administration.

280

## 281 **ETHICAL APPROVAL**

282

283 Ethical Approval (CVASU/Dir(R&E) EC/2022/435(1)/7) was taken from the Ethical Approval  
284 committee of the Director of Research of Extension, Chittagong Veterinary & Animal  
285 Sciences  
286 University.

287

## 288 **Disclaimer (Artificial intelligence)**

289

290 Author(s) hereby declare that NO generative AI technologies such as Large Language  
291 Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the  
292 writing or editing of this manuscript.

293

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