

Effect of biofertilizers on soil quality parameters in gerbera (*gerbera jamesonii* bolus) cultivation

Abstract: A field experiment was carried out at the Experimental Farm, Department of Horticulture, Assam Agricultural University, Jorhat, during the periods of 2015-2016 and 2016-2017. The experiment was set in Randomized Block Design (RBD) consisting of three replications and eight treatments. The Treatments comprised of T_0 (Control), T_1 {*Bacillus subtilis* (4% solution)}, T_2 {*Microbacterium laevaniformans* (4% solution)}, T_3 NPK (@15:10:20 g m⁻²), T_4 Vermicompost (5 kg per plot), T_5 (½ NPK + ½ Vermicompost + *Bacillus subtilis*), T_6 (½ NPK + ½ Vermicompost + *Microbacterium laevaniformans*), T_7 (½ NPK + ½ Vermicompost + Consortium) and T_8 (Consortium). The results revealed that the soil parameters were significantly influenced by the treatments. Studies revealed that soil pH (5.26), Organic Carbon (0.86%), N (276.9 kg ha⁻¹), K (135.1 kg ha⁻¹), Microbial Biomass Carbon (379.23 µg g⁻¹ soil 24 hour⁻¹) were found highest in T_4 . For P (44.6 kg ha⁻¹) and Electrical Conductivity (0.19 dS/m) was recorded highest in treatment T_7 .

Keywords: Influence, Biofertilizers, Vermicompost, Soil fertility and sustainability.

1. INTRODUCTION

In recent years, floriculture is fast emerging as highly competitive, commercial and economic activity with potential for earning valuable foreign exchange. The flower markets both at national and international levels are very much quality oriented. The overall quality of the cut flowers is governed by the cultivars grown, growing conditions and pre and post harvest management practices. Performance of each gerbera cultivar varies with region, season and other growing conditions.

“Today, agrochemicals are being used excessively in crop production due to high trend in industrialization and population explosion in the world. Their continuous application has introduced major challenges for farmers in the form of soil infertility, nutrient imbalance, accumulation of toxic chemicals in the soil which have an adverse effect on the soil productivity, ecosystem destruction, environmental degradation and also affecting the yield and quality of the product” (Eman *et al*, 2008). “Steady decline in soil organic matter levels due to continuous cropping with injudicious applications of chemical fertilizers has led to negative nutrient balances in Indian agriculture, impaired soil health and weaken factor productivity” (Rao, 2007).

“Soil quality and its degradation depend on a large number of physical, chemical, biological, microbiological and biochemical properties, the last two being the most sensitive since, they respond rapidly to changes. The microbiological activity of a soil directly influences ecosystem stability and fertility and it is widely accepted that a good level of microbiological activity is essential for maintaining soil quality. The microbiological activity *viz.*, the enzymatic activities play a key role in soil nutrient cycling, its activity is essential in both the mineralisation and transformation of organic matters and plant nutrients in soil ecosystem” (Dick and Tabatabai, 1993). “Soil enzyme activities are very sensitive to both natural and anthropogenic disturbances and show a quick response to induced changes” (Dick, 1997).

To cope with all these problems a cheaper, better and safer way is necessary in order to improve the soil fertility status, maximize the agricultural productivity with minimum Eco hazards. All these criteria can be achieved through application of bio-fertilizers which is known as "microbial inoculants", these are the products containing the living cells (Mainly bacteria & fungi) that naturally activate the microorganisms found in the soil, restoring the soil fertility and improve physico-chemical and biological properties of soil.

2. MATERIALS AND METHODS

The experiment was undertaken in the Experimental Farm of Department of Horticulture, Assam Agricultural University, Jorhat-785013 during the year 2015-17. The experimental soil was sandy loam in texture, well drained and having pH 5.1. The experiment was set out in a Randomized Block Design (RBD) which was replicated thrice. Cultivar Indukumari having uniform vigour and age were selected and planted on 15th of October in both the years of the study at a spacing of 30 cm x 30 cm. The area of the experimental plot was 134.25 sq.m. and that of the individual bed was 1.5m x 1.5m (2.25 sq.m.) were raised to 25 cm from the ground level to avoid water stagnation. The crops were raised by following nine treatments in both the years.

The Treatments were T₀ (Control), T₁ {*Bacillus subtilis* (4% solution)}, T₂ {*Microbacterium laevaniformans* (4% solution)}, T₃ NPK (@15:10:20 g m⁻²), T₄ Vermicompost (5 kg per plot), T₅ (½ NPK + ½ Vermicompost + *Bacillus subtilis*), T₆ (½ NPK + ½ Vermicompost + *Microbacterium laevaniformans*), T₇ (½ NPK + ½ Vermicompost + Consortium) and T₈ (Consortium). Where, as per recommendation package and practices fertilizers was applied at the time of field preparation. N, P₂O₅ and K₂O respectively which were applied NPK @15:10:20g m⁻². All the fertilizers were applied four days ahead of planting. Half of urea, full dose of SSP, MOP was applied at the time of basal dose. The second dose of N was applied at 30 days after planting. Consortium used in the experiment was the mixture of *Bacillus subtilis* and *Microbacterium laevaniformans*. Depending upon the nature of substrate vermicompost contains 1.5-2.5% N, 0.9-1.7% P, 1.5-2.4% K, 0.5-1.0% Ca, 0.2-0.3% Mg, 0.4-0.5% S and other micronutrients with vitamins, enzymes and hormones.

Observations on soil parameters

Soil samples collected from each plot after the harvest of the crop were air dried, ground and sieved through 2mm diameter and stored in butter paper bags with proper tagging and used for various analysis.

2.1 Soil pH

Soil pH was determined before and at harvest by glass electrode method (Jackson, 1973).

2.2 Organic carbon(OC) by Wet Digestion (%)

Organic Carbon was calculated with the Wet digestion method described by Walkley, (1934).

2.3 Soil Electrical Conductivity(ds/m)

The electrical conductivity (EC) indicates the amount of soluble (salt) ions in soil. The determination of electrical conductivity (EC) is made with a conductivity cell by measuring the electrical resistance of a 1:5 soil:water suspension. We use KCl solution as a reference solution of 0.01M concentration. This reference solution has an electrical conductivity of 1.413 ds/m at 25 °C (Rayment and Higginson, 1992).

2.4 Available nitrogen (kg ha⁻¹)

Available N of the soil sample was estimated by modified Kjeldahl's method as described by Jackson (1973) and expressed as kg ha⁻¹.

2.5 Available phosphorous (kg ha⁻¹)

Available P in soil sample was extracted by Bray's method as outlined by Jackson (1973) and expressed as available P₂O₅ (kg ha⁻¹).

2.6 Available potassium (kg ha⁻¹)

Available K content of the soil sample was extracted with neutral normal ammonium acetate as outlined by Jackson (1973). The potassium content was determined with the help of Flame Photometer and expressed as available K₂O (kg ha⁻¹).

2.7 Soil microbial biomass carbon (µg g⁻¹)

Microbial biomass carbon was determined by chloroform fumigation- extraction technique following the method of Vance *et al.* (1987).

Statistical analysis

The experimental data obtained from various observations were analysed statistically by using Fisher's method of analysis of variance in Randomized Block Design as described by Panse and Sukhatme (1978). Significance or non significance of the variance due to various treatments effect was determined by calculating respective 'F' values.

The standard error of the differences was calculated by using the formula:

$$S.Ed = \sqrt{\frac{2 \times \text{Error meansquare}}{\text{Number of replication}}}$$

The critical differences (C.D.) were calculated to find out the significance and non-significance of mean differences between the treatments. CD was calculated by using the following expression:

$$C.D = S.Ed \times t_{0.05}$$

Where, $t_{0.05}$ = tabulate value of 't' at 5 per cent level of probability for appropriate degree of freedom

3. RESULT AND DISCUSSION

Influence of biofertilizers on soil parameters:

3.1 Soil pH

"Data recorded on soil pH initially was 5.1. There was a significant increase in soil pH (5.26) in treatment T₄(Table.1). Higher pH in the organic treatments might be due to the deactivation of Al³⁺ and concomitant release of basic cations due to addition of organic matter"(Gogoi *et al.*, 2015). "However, application of different organic sources did not effect the soil pH much perhaps due to great buffering action of organic matter present in the organic manures"(Rajkonwar, 2012). Similar results were reported by Prakash *et al.* (2002) and Sushma *et al.* (2007). Also, "higher pH might be due to the increase in microbial activities in the root zone which decomposes organic manures and also fix unavailable form of mineral nutrient into available forms in soil thereby substantiates crop requirement and improve organic carbon level and stabilize soil pH". Similar result was also reported by Tekasangla *et al.*(2015) in cauliflower.

3.2 Organic carbon(OC) by Wet Digestion (%)

The initial organic carbon was to be recorded as 0.58% which was significantly influenced by different treatments. There was a significant increase in organic carbon (0.86%) in treatment T₄(Table.1). "It might be due to relatively higher carbon content in the organic manure compared to other organic and inorganic treatments. Similar findings were reported by various workers"(Gupta *et al.*, 2000; Bullock, 2002; Garcia-Ruiz, 2008 and Chanu, 2015). Organic carbon of soil acts as a sink and source of nutrients for microbial population, which regulates the availability of different nutrients through microbial transformation. The net increase in organic carbon was much higher with organic manures with microbial consortium. It is probably due to application of organic inputs and their releasing behaviour of different acids.

3.3 Electrical conductivity

The initial soil EC was to be recorded as 0.13 dS/m. Nutrient integrations (both with organic and inorganic sources) showed an increase in EC of soil (Table.1) and data ranged between 0.13 and 0.19 dS m⁻¹. The highest Electrical conductivity of 0.19 dS m⁻¹ was recorded in T₇. More availability of soluble forms of K, Ca, Mg and Na those lead to formation of some salts due to addition of organics, which might be responsible for the higher EC of the soil (Gogoi *et al.*, 2015).

3.4 Available nitrogen (kg ha⁻¹)

The initial available N in the soil was 250.7 kg ha⁻¹. The highest available nitrogen of 276.94 kg ha⁻¹ was recorded under treatment T₄ (Table.2). "Such a build up of available N to fix atmospheric N in the rhizosphere throughout the cropping period might be due to the fact that pH value rises as a result of organic sources and thus lowered the oxidation-reduction process. Organic acid and microbial product of decomposition from organic sources solubilises the insoluble compounds by interacting with their specific binding cations and clay minerals. Therefore, it was seen that application of organic sources was found to be good in enhancing the nitrogen availability in soil" (Workneh *et al.*, 1993). Organically managed soil exhibited great of biological activity of inoculated microorganisms as well as their potential nitrogen fixation (Merelo *et al.*, 2006).

3.5 Available phosphorous (kg ha⁻¹)

The initial available phosphorus content of soil was recorded as 41.1 kg ha⁻¹. The highest available soil phosphorus of 67.84 kg ha⁻¹ was recorded under treatment T₇ (Table.2). Increase in available phosphorus might be due to the application of vermicompost and phosphate solubilising and mineralizing ability of the microorganisms from the soluble form of phosphorus sources (Tao *et al.*, 2008). It is established that application of PSB along the rock phosphate significantly increased the available phosphorus status in soil which could be attributed to the production of organic acids which acts as chelating agent from stable complexes with Fe and Al which are available in acid soil and thereby release phosphorus from clutches of Fe and Al to the soil solution (Biswas, 2008 and Umlong, 2010). Reports are also available on the role of biofertilizers in improving the availability of nutrients (Mishra *et al.*, 1999). Similar results of better availability of Nitrogen and Phosphorus due to biofertilizers and vermicompost were reported in crossandra Cv. Dindigul local (Narasimharaju and Haripriya, 2001), in marigold (Mashaldi, 2000) and in carnation (Gangadharswamy, 2010).

3.6 Available potassium (kg ha⁻¹)

Initially 121.8 kg ha⁻¹ of potassium was recorded in the soil. In case of residual potassium (Table.2), treatment T₄ showed higher potassium content of 135.1 kg ha⁻¹. This might be due to release of potassium from these organic amendments and also due to solubilisation of mineral based potassium or native potassium. The positive influence of organic manure on the available potassium was earlier reported by Srikanth *et al.* (2000). Besides, it could be also due to prevention of leaching loss due to retention of more potassium by organic components while inorganic fertilizers could have released potassium at a faster rate. These results were similar to the findings by Bahadur *et al.*, 2006; Biswas (2008) and Umlong (2010). The positive influence of organic manure on the available potassium in soil might be due to the effect of organic manures on the reduction of potassium fixation; added organic matter interacted with potassium clay to release potassium from the non-exchangeable fraction to the available pool (Reddy, 1998).

3.7 Soil microbial biomass carbon (µg g⁻¹)

The initial soil microbial biomass carbon content of soil was recorded as 228.76 µg g⁻¹ soil 24 hour⁻¹. Significant variation in MBC (Table.3) was observed in the present study. In treatment T₄ resulted in the highest MBC (379.23 µg g⁻¹ soil 24 hour⁻¹). This might be due to the application of organic source of nutrients which improves the microbial and enzymatic activities in soil (Rajkonwar, 2012). The biological properties were higher in the soil might be due to the increase in organic carbon, total N and P

content in the soil with the application of organic inputs specially biofertilizers and vermicompost, which are directly related to the biological properties of the soil.

The response of soil to biofertilizers is well recognized. Though many reports are available from different parts of India on influence of biofertilizer, on soil fertility and sustainability were studied for different locations by various workers. The use of organic amendments has been recognized as an effective means for improving soil aggregation, structure and fertility, increasing microbial diversity and populations, improving the moisture holding capacity of soil and increasing crop yields” (Zink and Allen, 1998). A higher dehydrogenase activity was observed in soil treated with biofertilizers, and a significant correlation was observed between the dehydrogenase activity of microbial populations (Dinesh *et al.*, 2010). Hridya *et al.* (2013) observed that the microbial inoculations significantly increased soil available nutrient contents, enzyme activities such as urease, dehydrogenase and β -glucosidase activity and microbial biomass carbon by reducing the amount of the required fertilizer.

Table.1 Soil pH and organic carbon (%) and electrical conductivity(dS/m):

Initial pH : 5.1, organic carbon: 0.58% and EC:0.13 dS/m

Treatment	Soil pH			Organic carbon			EC		
	2015-2016	2016-2017	Pooled	2015-2016	2016-2017	Pooled	2015-2016	2016-2017	Pooled
T ₀	5.10	5.09	5.09	0.55	0.57	0.56	0.14	0.14	0.14
T ₁	5.13	5.11	5.12	0.66	0.69	0.68	0.16	0.16	0.16
T ₂	5.15	5.12	5.14	0.67	0.68	0.67	0.16	0.16	0.16
T ₃	4.80	4.49	4.65	0.62	0.64	0.63	0.14	0.16	0.15
T ₄	5.24	5.28	5.26	0.82	0.89	0.86	0.19	0.17	0.18
T ₅	5.19	5.22	5.21	0.73	0.75	0.74	0.18	0.18	0.18
T ₆	5.20	5.23	5.22	0.74	0.78	0.76	0.19	0.18	0.18
T ₇	5.21	5.23	5.22	0.74	0.80	0.77	0.19	0.19	0.19
T ₈	5.11	5.11	5.11	0.67	0.70	0.68	0.16	0.16	0.16
S.Ed (\pm)	0.07	0.06	0.04	0.02	0.04	0.02	0.03	0.04	0.03
CD (5%)	0.16	0.13	0.09	0.04	0.08	0.05	NS	NS	NS

Table.2 Available soil N, P and K (kg ha⁻¹):

Initial N: 250.7 kg ha⁻¹, P₂O₅:41.1kg ha⁻¹ and K₂O : 121.8 kg ha⁻¹

Treatment	Available Nitrogen			Available Phosphorus			Available Potassium		
	2015-2016	2016-2017	Pooled	2015-2016	2016-2017	Pooled	2015-2016	2016-2017	Pooled
T ₀	251.48	251.76	251.62	41.71	42.11	41.91	112.63	112.63	113.59

T₁	259.06	262.99	261.03	48.30	51.37	49.84	118.92	118.92	120.57
T₂	261.18	265.33	263.26	51.33	56.74	54.03	121.67	121.67	123.62
T₃	264.67	266.52	265.59	53.36	60.15	56.76	124.01	124.01	126.66
T₄	275.67	278.22	276.94	59.30	65.34	62.32	134.30	134.30	135.11
T₅	267.67	270.00	268.83	61.00	66.15	63.58	127.04	127.04	127.90
T₆	268.99	269.11	269.05	63.33	68.69	66.01	128.69	128.69	128.34
T₇	270.00	273.30	271.65	64.04	71.64	67.84	131.18	131.18	131.74
T₈	262.78	263.03	262.91	57.09	59.26	58.18	122.80	122.80	123.55
S.Ed (±)	3.48	4.75	3.59	3.46	2.46	1.80	4.26	3.12	2.96
CD (5%)	7.37	10.06	7.61	7.33	5.22	3.82	9.04	6.62	6.28

Table.3 Microbial Biomass Carbon (MBC)

Initial Microbial biomass carbon : 228.76 $\mu\text{g g}^{-1}\text{soil 24 hour}^{-1}$

Treatment	MBC		
	2015-2016	2016-2017	Pooled
T₀	228.48	229.11	228.80
T₁	230.25	235.32	232.79
T₂	233.59	239.33	236.46
T₃	253.33	264.51	258.92
T₄	377.00	381.47	379.23
T₅	321.93	324.92	323.42
T₆	325.33	328.30	326.82
T₇	328.00	331.18	329.59
T₈	241.74	247.75	244.74
S.Ed (±)	3.34	4.65	2.97
CD (5%)	7.07	9.85	6.30

4. CONCLUSION

The investigation revealed that use of organic inputs had significant influence on soil properties at harvest. The highest values of soil pH, soil organic carbon, MBC, and soil available N and K were recorded for the treatments T₇(½ NPK + ½ Vermicompost + Consortium) and T₄(Vermicompost 5 kg plot⁻¹), as regards to EC and P. In recent times, the government has identified the whole N.E. region as an organic zone where most of the cultivated areas have been identified as naturally organic. So, in this

experiment, biofertilizer has been identified as an alternative to chemical fertilizer in order to increase soil fertility and crop production in sustainable farming.

Disclaimer (Artificial intelligence)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc.) and text-to-image generators have been used during the writing or editing of this manuscript.

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