

## **Cytokines Levels and Anti-tuberculosis drug resistance among HIV Seropositive Patients in Ibadan, Southwest, Nigeria.**

### **Abstract**

Failure of effective solutions to tuberculosis are caused by human immunodeficiency virus infection, delay in diagnosis and or proper treatments, mostly lead to resistance to certain drugs. These are said to be either by mutation or through horizontal transfer where organism obtain resistance from other microorganisms. More so, susceptibility to TB disease rises on mutations in cytokine receptors which ultimately lead to defective signaling and alter the immune surveillance and bacterial killing. This study observed 217 tuberculosis patients living with human immunodeficiency virus at University College Hospital, Ibadan. The sputum of known HIV positive patients were collected in standard screw-capped leak-proof sputum container with specific clinic identification number. Each sample was divided into two parts. One part decontaminated with N-acetyl L- Cystine (NaOH-NALC) and used for culture, while the second part was processed without decontamination and used to perform GeneXpert MTB/RIF assay using ultra MTB/RIF cartridge. 10mls of blood was collected from each eligible and consented client. Out of which 5mls were added to K<sub>3</sub> EDTA bottles for HIV confirmation test and cytokines levels estimation. There were 105(48.4%) males and 112(51.6%) females with a mean age of  $42.95 \pm 8.286$  years and a range of 28-65 years. Higher *Mycobacterium.tuberculosis* isolation rate was found among male than females and among those aged  $\geq 40$ . Similarly, *M. bovis* recorded higher isolation rate among male than female and among age group 40 and above. There was no significant difference between culture positivity and sex, as well as age of the subjects. There was no significant association between cytokines (IL-10 and IFN- $\gamma$ ) and sex of the subjects ( $P > 0.05$ ). However, females had higher IFN- $\gamma$  level of  $231.06 \pm 61.81$  IU/ml when compared to  $227.53 \pm 57.92$  IU/ml obtained from males. There was no significant association between cytokines (IL-10 and IFN- $\gamma$ ) and sex, but age group  $> 40$  years had higher IL-2 and IFN-G level of  $226.53 \pm 52.39$  IU/ml and  $232.41 \pm 66.63$  IU/ml respectively. There was significant association between culture positivity (TB) and cytokines (IL-10 and IFN- $\gamma$ ), and geneXpert positivity (TB) is significantly associated with high cytokines levels ( $F = 68.729$ ,  $p = 0.000$ ) ( $P < 0.05$ ). There was an association between high cytokines levels and anti-TB drug resistance

( $P < 0.05$ ). Highest cytokines levels; IL-10 ( $401.00 \pm 7.07$  IU/ml) and IFN- $\gamma$  ( $401.00 \pm 7.07$  IU/ml) were recorded in Levofloxacin resistance. There was an association between high cytokines levels and genotypic anti- drug resistance ( $P < 0.05$ ). with highest cytokines levels; IL-10 ( $400.12 \pm 12.37$  IU/ml) and IFN- $\gamma$  ( $472.87 \pm 31.93$  IU/ml) recorded in Rifampicin resistance. This entire study revealed that there is high prevalence of MDR-TB among PLHIV, with Rifampicin monoresistance having the highest resistance rate among the drugs tested, and pro and inflammatory cytokines (IL-10 and IFN- $\gamma$ ) play significant role in the development of Multi Drug Resistant-Tuberculosis among people living with HIV.

**Keywords: Tuberculosis, Cytokines levels, anti-tuberculosis drug resistance.**

## **Introduction**

The major causes of failure to effective solutions to tuberculosis (TB) are human immunodeficiency virus (HIV) infection, epidemics of TB and delay in diagnosis and proper treatment (Naidoo et al., 2019). In 2022, the estimated number of incident cases of TB were about 10.6 million, deaths from TB among HIV-negative patients were 1.13 million and deaths from HIV-positive TB were about 0.17 million (WHO, 2023a).

Drug-resistant TB may arise due to the failure in implementing proper TB control programs and properly managing TB cases (WHO, 2022). As a result, more than 206,030 MDR-TB cases are estimated to occur worldwide each year, of which about 6% takes place in Nigeria (Dayyab et al., 2022). World Health Organization (WHO) pointed out in a survey recently that about 20% of MDR-TB isolates were also extensively drug-resistant (XDR) i.e. resistant to INH and RIF and to at least one of the most efficient second-line drugs (WHO, 2022). Bacteria may become antibiotic resistant due to two reasons: one mutation in some drug target genes and second through horizontal transfer in which an organism obtain drug resistance genes from other microorganisms (Peraman *et al.*, 2021). Most of the microorganisms acquire resistance in the environment through horizontal transfer because mutations occur at low frequency.

Cytokines have a protective and pathological responses with the network of pro-inflammatory, T1 and T2 cytokines through reactions with complex mycobacterial antigens and RD regions.

Additional role of pro-inflammatory cytokines as biomarker signature for disease severity, bacterial load, culture conversion and treatment outcome have been discovered in previous studies. Mycobacterial growth prevention is achieved by the joint role of cytokines and chemokines as they coordinate the initiation, expansion, inflammation, recruitment, differentiation, activation and localization of mononuclear cells. Array of cytokines such as IL-12, IL-18, IFN- $\gamma$ , TNF- $\alpha$ , and IL-6 play early protection roles against both MDR-TB and DS-TB. Mycobacterium tuberculosis (MTB) clearance in granuloma of non-human primate model rendered through the T cells with balanced secretion of pro and anti-inflammatory cytokines. However, in the chronic infection setting, the balance between protection and pathology is lost due to altered expression of cytokines that accompany delayed resolution of inflammation and tissue repair. The susceptibility to TB disease rises on mutations in cytokine receptors which ultimately lead to defective signaling and aborts the immune surveillance and bacterial killing (Pavithra et al, 2023).

There is limited studies on cytokine signatures for DR-TB and latent TB. Hence, this study aimed to determine the relationship of cytokines levels and drug resistance among TB/HIV co-infected clients in Ibadan.

## **Materials and Methods**

### **Study area**

The study was carried out at the University College Hospital (UCH), Ibadan. The hospital serves as the central hub for healthcare services, medical education, and research in the southwest region of Nigeria. It is located at longitude 7.3569°N and latitude 3.8743°E.

### **Study Population**

The study included 217 TB patients attending University College Hospital, living with HIV, as well as those without HIV infection who served as the control group.

## **Sample Collection**

### **Collection of sputum sample**

Sputum samples from a deep cough were collected among known HIV positive patients on ART, attending HIV clinic at UCH, Ibadan. Each patient was instructed to produce and submit 2 sputum specimens within two consecutive days. The specimens were collected in standard screw-capped leak-proof sputum container with specific clinic identification number. The first sputum specimen was obtained on the first contact with the centre (spot specimen) while the second specimen was an early-morning specimen produced at home after cleaning the mouth with water. The two sputum specimens were processed at the same time.

### **Collection of samples for biochemical and haematological assays**

About 10mls of blood was collected from each eligible and consented client. Out of which 5mls were added to K<sub>3</sub> EDTA bottles for HIV confirmation test, viral load determination, CD4+ cell count, cytokines, and full blood count estimation. The remaining 5mls blood was added to lithium heparin bottles for electrolyte panel analysis, liver function tests, and lipid profile estimation.

### **Analysis of sputum specimens**

Each sputum sample was divided into two parts. One part was decontaminated with N-acetyl L-Cystine (NaOH-NALC). The second part was processed without decontamination and used to perform GeneXpert MTB/RIF assay using ultra MTB/RIF cartridge.

### **Inoculation and Culture Method**

The sediment pellet of decontaminated sputum was resuspended in deionized water by vortexing to make homogeneous inoculum; 50 microlitres of this inoculum was inoculated on Lowenstein Jensen(LJ) slants, Ogawa slants as well as blood agar slants in triplicate. These bottles were sealed with parafilm to prevent contamination and dessication.

The inoculated slants were incubated in MSE incubator model 4032 at 37<sup>0</sup>C for eight weeks. These were observed after 48 hours and thereafter weekly for appearance of macroscopic growth. The growth on LJ, Ogawa as well as blood agar slants were observed after keeping the slants in diffuse sunlight for about one hour for pigmentation (Murli *et al*, 2009). The methods for the preparation of the three media used as shown in the appendix.

### **Estimation of cytokine IL10**

The estimation of the interleukin 10 was done using the NWLSS Human IL-10 ELISA kit, which is able to recognize native and recombinant human IL-10.

### **Estimation of cytokine IFN- $\gamma$**

The estimation of the IFN- $\gamma$ , or type II interferon was done using the Cohesion Bioscience Human IFN gamma ELISA kit, which employs an antibody specific for Human IFN-gamma coated on a 96-well plate.

### **Data analysis:**

Analyses of all obtained data were performed by using STATA/IC version 23.0. The  $\chi^2$  test was used to calculate p value when appropriate. P values <0.05 was considered statistically significant.

## **Results**

Table 1 showed that 105(48.4%) were male, while 112(51.6%) were females. Moreover, 26(52.0%) of the control subjects were male, while 24(48.0%) were females. The mean age with SD was  $42.95 \pm 8.286$  years with a range of 28-65 years, while the control group had a mean age with SD of  $42.24 \pm 4.745$  years with a range of 35-50 years.

Table 2 showed that male subjects had higher *M.tuberculosis* isolation rate of 19.0% compared to 13.4% for females. Similarly, higher *M.bovis* isolation rate of 4.7% was found in males while females had 2.7%. NTM had more occurrences in males (3.8%), compared to females (1.8%).

Table 3. Showed that significantly higher *M.tuberculosis* isolation rate of 21.4% was found among people aged  $\geq 40$  years compared to 10.5% among people  $< 40$  years at ( $P < 0.05$ ). Age group  $\geq 40$  years had higher isolation rate (4.5%) of *M.bovis* compared to those  $< 40$  (2.9% ), though, not statistically significant at ( $P > 0.05$ ). NTM had a non-significant higher isolation rate of 3.6% among age group  $\geq 40$  years compared to 1.9% for age group  $< 40$  years ( $P > 0.05$ ).

Table 4 showed that there was no significant difference between culture positivity and sex of the subjects ( $p > 0.05$ ). Similarly, there was no significance difference between culture positivity and age of the subjects ( $p > 0.05$ ).

Table 5 showed that there was no significant association between cytokines (IL-10 and IFN- $\gamma$ ) and sex of the subjects ( $P > 0.05$ ). However, females had higher IFN- $\gamma$  level of  $231.06 \pm 61.81$  IU/ml when compared to  $227.53 \pm 57.92$  IU/ml obtained from males.

Table 6, there was no significant association between cytokines (IL-10 and IFN- $\gamma$ ) and sex of the subjects ( $P > 0.05$ ). However, age group  $> 40$  years had higher IL-2 and IFN- $\gamma$  level of  $226.53 \pm 52.39$  IU/ml and  $232.41 \pm 66.63$  IU/ml respectively when compared to  $221.22 \pm 51.20$  IU/ml and  $232.41 \pm 66.63$  obtained from age group  $< 40$  years.

Table 7 showed that there was significant association between culture positivity (TB) and cytokines (IL-10 and IFN- $\gamma$ ) ( $P < 0.05$ )

Table 8 showed that GeneXpert positivity (TB) is significantly associated with high cytokines levels ( $F = 68.729$ ,  $p = 0.000$ ) ( $P < 0.05$ )

Table 9 showed the relationship between Phenotypic DST and Cytokines. There was an association between high cytokines levels and anti-TB drug resistance ( $P < 0.05$ ). Highest cytokines levels; IL-10 ( $401.00 \pm 7.07$  IU/ml) and IFN- $\gamma$  ( $401.00 \pm 7.07$  IU/ml) were recorded in Levofloxacin resistance.

Relationship between LPA and Cytokines as shown in table 10 revealed that there was an association between high cytokines levels and genotypic anti- drug resistance ( $P < 0.05$ ). Highest cytokines levels; IL-10 ( $400.12 \pm 12.37$  IU/ml) and IFN- $\gamma$  ( $472.87 \pm 31.93$  IU/ml) were recorded in Rifamicin resistance.

**Table 1: Demographic characteristics of the participants**

Respondents' demographic Characteristics	Socio-	Cases n(%)	Control n(%)	Total n(%)
<b>Sex</b>				
Male		105(48.4)	26(52.0)	131(49.1)
Female		112(51.6)	24(48.0)	136(50.9)
<b>Age Group (years)</b>				
<40		90(41.5)	17(34.0)	107(40.1)
40-49		83(38.2)	30(60.0)	113(42.3)
50-59		34(15.7)	3(6.0)	37(13.9)
≥60		10(4.6)	0(0.0)	10(3.7)
≥40		127(58.5)	33(66.0)	160(59.9)
Mean (±SD)		42.95 ± 8.286 years	42.24 ± 4.745 years	42.82 ± 7.745 years
Range		28 to 65 years	35 to 50 years	28 to 65 years
<b>Total</b>		<b>217</b>	<b>50</b>	<b>267</b>

**Table 2: Sex and *Mycobacterium* species**

Sex	N	<i>M.tuberculosis</i> n(%)	<i>M.bovis</i>	NTM n(%)
Male	105	20 (19.0)		5 (4.7)
Female	136	4(3.8)		

Female	112	15 (13.4)	3 (2.7)
	2(1.8)		
(P>0.05)			

**Table 3: Age and *Mycobacterium* species**

Age(Years)	N	<i>M.tuberculosis</i>	<i>M.bovis</i>
NTM		n(%)	n(%)
n(%)			
< 40	105	11 (10.5)	3 (2.9)
		2(1.9)	
≥40	112	24 ( 21.4)	5 (4.5)
		4(3.6)	
(P<0.05)			

**Table 4: Patients'' demographic characteristics and Culture results**

Respondents' demographic Characteristics	Socio-	Culture results		Pearson chi-square	p-value
		Positive n(%)	Negative n(%)		
<b>Sex</b>				<b>0.176</b>	<b>0.746</b>
Male		25(23.8)	80(76.2)		

Female	24(21.4)	88(78.6)		
<b>Age Group (years)</b>			<b>2.029</b>	<b>0.188</b>
<40	16(17.8)	74(82.2)		
≥40	33(26.0)	94(74.0)		
<b>Total</b>	<b>49(22.6)</b>	<b>168(77.4)</b>		

\*p<0.05

**Table 5: Gender and Cytokines (IL-2 and IFN-Y)**

Cytokines	Male (n=105)	Female (n=112)	t-test	p-value
IL_2	224.48±48.30	224.18±55.18	0.042	0.966
IFN_Y	227.53±57.92	231.06±61.81	0.434	0.665

\*p<0.05

**Table 6: Age and Cytokines (IL-10 and IFN-y)**

Cytokines	Age Group (years)		t-test	p-value
	<40 years (n=90)	≥40 years (n=127)		
IL_2	221.22±51.20	226.53±52.39	0.742	0.459
IFN_G	225.04±48.73	232.41±66.63	0.893	0.373

\*p>0.05

**Table 7: Relationship between Culture and Cytokines**

Cytokines	Culture results		t-test	p-value
	Positive (n=49)	Negative (n=168)		
IL_2	285.33±75.31	206.53±20.89	12.112	0.000*
IFN_G	300.76±91.63	208.52±16.94	12.404	0.000*

\*p<0.05

## **8: Relationship between GeneXpert (DS-TB and DRTB) and Cytokines**

		Cytokines (IL-10)			
Variables	N	Mean IL_2	Standard deviation	F-test	p-value
<b>GeneXpert (DRTB)</b>				<b>68.729</b>	<b>0.000*</b>
Negative (NOT DETECTED)	177	208.95	29.25		
Positive (DSTB)	36	292.87	74.99		
DRTB	4	287.87	45.12		
<b>Total</b>	<b>217</b>	<b>224.33</b>	<b>51.85</b>		
		Cytokines (IFN-y)			
Variables	N	Mean IFN_G	Standard deviation	F-test	p-value
<b>GeneXpert (DRTB)</b>				<b>74.616</b>	<b>0.000*</b>
Negative (NOT DETECTED)	177	211.16	31.63		
Positive (DSTB)	36	309.23	88.15		
DRTB	4	315.37	50.51		
<b>Total</b>	<b>217</b>	<b>229.35</b>	<b>59.84</b>		

\*p<0.05 (i.e. Significant)

**Table 9: Relationship between Phenotypic DST and Cytokines**

Phenotypic DST/Cytokines				
RIF	Sensitive	Resistant	t-test	p-value
IL_10	268.59±69.88	336.95±70.12	2.942	0.005*
IFN_Y	276.08±81.32	376.87±81.40	3.730	0.001*
<b>INH</b>				

IL_10	272.58±71.64	350.68±61.10	2.880	0.006*
IFN_Y	277.39±76.01	420.56±70.40	4.926	0.000*
<b>ETB</b>				
IL_10	275.71±70.62	393.62±24.17	3.294	0.002*
IFN_Y	286.26±79.72	463.87±49.92	4.355	0.000*
<b>LEV</b>				
IL_10	280.41±72.88	401.00±7.07	2.316	0.025*
IFN_Y	292.78±84.66	488.25±1.06	3.232	0.002*
<b>ETH</b>				
IL_10	278.90±72.93	384.00±29.86	2.462	0.018*
IFN_Y	292.03±85.43	434.66±92.81	2.791	0.008*

\*p<0.05

Key; Rifampicin, Isoniazid, ETB-Ethambutol, LEV-Levofloxacin, ETH-Ethionamide

**Table 10: Relationship between LPA and Cytokines**

<b>LPA DST/Cytokines</b>				
<b>RIF</b>	<b>Sensitive</b>	<b>Resistant</b>	<b>t-test</b>	<b>p-value</b>
IL_10	281.02±63.65	367.07±71.47	3.212	0.003*
IFN_Y	294.38±73.30	416.07±85.33	3.918	0.000*
<b>INH</b>				

IL_10	284.25±66.22	400.12±12.37	3.457	0.001*
IFN_Y	297.92±73.34	472.87±31.93	4.684	0.000*
<b>FLQ</b>				
IL_10	288.13±69.30	387.00±24.75	2.435	0.019*
IFN_Y	303.62±79.72	455.16±57.30	3.214	0.003*

\*p<0.05

Key: RIF-Rifampicin, INH, Isoniazid, FLQ-Flouroquinolone

## Discussions

HIV and TB co-infection has been described as a lethal combination, as each disease speeds up the other's progress. A realization which probably led to updating of the WHO TB/HIV treatment approach, in 2012, to place persons with HIV on TB-Preventive therapy upon confirmation of HIV.

Tuberculosis continues to be the leading cause of mortality among PLHIV in sub-saharan Africa (Sekayi, 2023). In this study, the high isolation rate(19%) of *M.tuberculosis* among male PLHIV obtained is similar to 17.0% reported by Purushotan et al(2013) in India, but markedly lower than 37.4% rate reported by *Melkamu* (2021) in Ethiopia. The higher rate of TB/HIV co-infection cases might be explained by impairing the host's immune response to both diseases, lengthening the time it takes for TB treatments to be effective by increasing the bacilli load, which could make the bacteria resistant to anti-TB drugs, and lead to MDR-TB.

TB/HIV co-infection is a major public health problem in many parts of the world, but the prevalence of co –infection varies among countries (*Melkanu*, 2021). The risk of developing TB among the millions of people living with HIV is 18 times higher than the rest of the global population.TB remains the overall leading cause of death among people living with HIV, accounting for around one third HIV-related deaths. In this study, occurrence of *M.bovis* in PLHIV was higher among males (4.7%) when compared with (2.7%) recorded in females. However, this is in contrast to the study conducted by *Valegra* et al (2005) in Europe. The authors reported higher isolation rate of 9.5% among female subjects.

Human disease caused by *M.bovis* has been confirmed in African countries. HIV infection may be an important risk factor for *M.bovis* disease, and *M.bovis* has been associated with mortality among PLHIV (Rodwell, 2008). Non-tuberculous mycobacteria (NTM) is a common opportunistic infection in PLHIV. AS obtained in this study, the occurrence of NTM in PLHIV has been reported (Lee *et al.*, 2022). Although the prevalence is low in this study, however, NTM occurred more in males (3.8%) than in females' clients (1.8%). Similarly, NTM occurred more in the age group >40 years (3.9%). These data indicated the persistence of NTM disease even in the modern *cART* era. Lee *et al.* (2022) reported that the distribution of NTM reflects geographical diversity, wherein species vary according to region and country.

Drug-resistant TB impairs the current treatment strategies and worsens the unfavorable outcomes. The knowledge on host immune responses between drug-sensitive and drug-resistant infection is inadequate to understand the pathophysiological differences and disease severity. Secreted proteins called cytokines display versatile behavior upon infection with *M.tuberculosis* and their imbalances often tend to assist disease pathology than protection. In this study, high cytokines levels were significantly associated with anti-TB drug resistance ( $P < 0.05$ ). This is in line with the report of Pavithra *et al.* (2023) that drug-resistant tuberculosis was associated with altered cytokine levels. These TB biomarkers can be considered crucial to achieve the global TB elimination targets. The effort to understand causal factors like cytokines and their differential expression during different stages of TB is valuable in identifying unique biological signatures (Mensah *et al.*, 2021). Cytokines mount protective and pathological responses with the network of pro-inflammatory, Th1 and Th2 cytokines through interaction with complex mycobacterial antigens and RD regions (Mustafa *et al.*, (2011). Previous studies determined the augmented role of pro-inflammatory cytokines as biomarker signature for disease severity, bacterial burden, culture conversion and treatment outcomes (Mihret *et al.*, 2013; Chowdhury *et al.*, 2014; Domingo *et al.*, 2016). The susceptibility to TB disease rises on mutations in cytokines receptor which ultimately lead to defective signaling and aborts the immune surveillance and bacterial killing (Kumar *et al.*, 2019).

It was found in this study that pro and inflammatory cytokines (IL-10 and IFN- $\gamma$ ) were associated with TB ( $P < 0.05$ ). This is in-line with Marina *et al.* (2021) that the imbalance of cytokine secretion in HIV infection affects the function of the immune system and the course of

the function of the immune system and the course of the disease, increasing or suppressing viral replication (Jacobs *et al.*, 2019).

## **Conclusion**

There is high prevalence of MDR-TB among PLHIV, with Rifampicin monoresistance having the highest resistance rate among the drugs tested. Pro and inflammatory cytokines (IL-10 and IFN- $\gamma$ ) play significant role in the development of Multi Drug Resistant-Tuberculosis among people living with HIV.

## **Compliance with ethical standard**

The study proposal was examined, approved, and permission for work was granted by the ethical committee of University College Hospital/University of Ibadan. All study participants were educated on the purpose of conducting the research, after which verbal/ written consent was obtained from each participants.

## **References**

Chowdhury, I. H. (2014). Alteration of serum inflammatory cytokines in active pulmonary tuberculosis following anti-tuberculosis drug therapy. *Molecular Immunology*, 62, 159–168.

Dayyab M.F., Iliyasu G. Garba A.B., Aliyu U.I., Shuaib M.N. Bajehson M.M., Daiyab I., Akpala O., Remilekun O., Habib G.A., & For Kano TB Concilium Expert (2022). Emerging threat of drug-resistant tuberculosis and trends in the era of COVID-19: A descriptive study from northwestern Nigeria. *Journal of Clinical tuberculosis and other mycobacterial diseases*, 28, 100319. <https://doi.org/10.1016/j.jctube.2022.100319>.

Domingo-Gonzalez, R., Prince, O., Cooper, A., & Khader, S. A. (2016). Cytokines and Chemokines in Mycobacterium tuberculosis Infection. *Microbiology spectrum*, 4(5), 10.1128/microbiolspec.TBTB2-0018-2016. <https://doi.org/10.1128/microbiolspec.TBTB2-0018-2016>

Elewski BE, Baddley JW, Deodhar AA, et al. Association of Secukinumab Treatment With Tuberculosis Reactivation in Patients With Psoriasis, Psoriatic Arthritis, or Ankylosing Spondylitis [published correction appears in JAMA Dermatol. 2021 Jan 1;157(1):124. doi: 10.1001/jamadermatol.2020.5204]. JAMA Dermatol. 2021;157(1):43-51. <https://doi.org/10.1001/jamadermatol.2020.3257>

Giri, P. A., Deshpande, J. D., & Phalke, D. B. (2013). Prevalence of Pulmonary Tuberculosis Among HIV Positive Patients Attending Antiretroviral Therapy Clinic. North American journal of medical sciences, 5(6), 367–370. <https://doi.org/10.4103/1947-2714.114169>

Kumar, N. P., Fukutani, K. F., Shruthi, B. S., Alves, T., Silveira-Mattos, P. S., Rocha, M. S., West, K., Natarajan, M., Viswanathan, V., Babu, S., Andrade, B. B., & Kornfeld, H. (2019). Persistent inflammation during anti-tuberculosis treatment with diabetes comorbidity. eLife, 8, e46477. <https://doi.org/10.7554/eLife.46477>.

Kumar, S., Sharma, C., Kaushik, S. R., Kulshreshtha, A., Chaturvedi, S., Nanda, R. K., Bhaskar, A., Chattopadhyay, D., Das, G., & Dwivedi, V. P. (2019). The phytochemical bergenin as an adjunct immunotherapy for tuberculosis in mice. The Journal of biological chemistry, 294(21), 8555–8563. <https://doi.org/10.1074/jbc.RA119.008005>

Lee EH, Chin B, Kim YK, Yoo JS, Choi YH, et al. (2022) Clinical characteristics of nontuberculous mycobacterial disease in people living with HIV/AIDS in South Korea: A multi-center, retrospective study. PLOS ONE 17(11): e0276484. <https://doi.org/10.1371/journal.pone.0276484>

Melkamu R., Berhane N., Jacobs B.K.M., Mohammed R., Kassa M., Yeshanew A., Fikre, H., Atnafu, S., van Henten, S., van Griensven, J. & Pareyn, M. (2023). PCR for detection of Leishmania donovani from microscopically negative tissue smears of suspected patients in Gondar, Ethiopia. PLoS Negl Trop Dis 17(2): e0011128. <https://doi.org/10.1371/journal.pntd.0011128>.

Mensah GI, Boakye AN, Basingnaa A, Owusu E, Antwi-Baffour S, Ofori MF, Addo KK, Jackson-Sillah D, Adekambi T. Identification of Serum Cytokine Biomarkers Associated with

Multidrug Resistant Tuberculosis (MDR-TB). *Immuno*. 2021; 1(4):400-409. <https://doi.org/10.3390/immuno1040028>.

Mihret, A., Bekele, Y., Bobosha, K., Kidd, M., Aseffa, A., Howe, R. & Walzl, G. (2013). Plasma cytokines and chemokines differentiate between active disease and non-active tuberculosis infection. *Journal of Infection*, 66(4), 357-365. <https://doi.org/10.1016/j.jinf.2012.11.005>.

Mustafa, S., Javed, H., Hashmi, J. et al . Emergence of mixed infection of Beijing/Non-Beijing strains among multi-drug resistant Mycobacterium tuberculosis in Pakistan. *3 Biotech* 6, 108 (2016). <https://doi.org/10.1007/s13205-016-0423-9>

Naidoo C.C., Nyawo G.R., Wu B.G., Gerhard W., Robbin M.W., Segal L.N. and Theron G. (2019): The Microbiome and tuberculosis: state of the art, potential applications, and defining the clinical research agenda. *The Lancet Respiratory Medicine* 2019, 7(10): 892-906.

Pavithra S., Rajamanickam A., Thiruvengadam K., Natarajan A.P., Hissar S., Dhanapal M., Thangavelu B., Jayabal L., Ramesh P.M., Ranganathan U.D., Babu S. and Bethunaickam R. (2023). Cytokine upsurge among drug-resistant tuberculosis endorse the signatures of hyper inflammation and disease severity. *ScientificReports*, 13(1): 785. <https://doi.org/10.1038/s41598-023-27895-8>

Peraman, R., Sure, S.K., Dusthacker, V.N.A. et al. Insights on recent approaches in drug discovery strategies and untapped drug targets against drug resistance. *Futur J Pharm Sci* 7, 56 (2021). <https://doi.org/10.1186/s43094-021-00196-5>

Rodwell, T. C., Moore, M., Moser, K. S., Brodine, S. K., & Strathdee, S. A. (2008). Tuberculosis from *Mycobacterium bovis* in binational communities, United States. *Emerging infectious diseases*, 14(6), 909–916. <https://doi.org/10.3201/eid1406.071485>

Sampath, P., Rajamanickam, A., Thiruvengadam, K., Natarajan, A. P., Hissar, S., Dhanapal, M., Thangavelu, B., Jayabal, L., Ramesh, P. M., Ranganathan, U. D., Babu, S., & Bethunaickan, R. (2023). Cytokine upsurge among drug-resistant tuberculosis endorse the signatures of hyper inflammation and disease severity. *Scientific reports*, 13(1), 785. <https://doi.org/10.1038/s41598-023-27895-8>.

Sekayi, W., Namyalo, E., Namayanja, J., & Kungu, J. M. (2023). Prevalence and predictors of tuberculosis among HIV patients who completed isoniazid preventive therapy (IPT) at Reach out Mbuya community health initiative. *Scientific reports*, 13(1), 17602. <https://doi.org/10.1038/s41598-023-44649-8>.

Valerga, M., Viola, C., Thwaites, A., Bases, O., Ambroggi, M., Poggi, S., & Marino, R. (2005). Tuberculosis por *Mycobacterium bovis* en una mujer con SIDA [*Mycobacterium bovis* tuberculosis in a female patient with AIDS]. *Revista Argentina de microbiologia*, 37(2), 96–98.

Vilchèze, C., & Jacobs, W. R., Jr (2019). The Isoniazid Paradigm of Killing, Resistance, and Persistence in *Mycobacterium tuberculosis*. *Journal of molecular biology*, 431(18), 3450–3461. <https://doi.org/10.1016/j.jmb.2019.02.016>.

World Health Organisation (2023a). Global Tuberculosis Report 2023. <https://www.who.int/teams/global-tuberculosis-programme/tb-reports/global-tuberculosis-report-2023>.

World Health Organization (2022). global tuberculosis report 2022. <https://www.who.int/teams/global-tuberculosis-programme/tb-reports/global-tuberculosis-report-2022>