

Review Article

CULTIVATING TOMORROW: A REVIEW ON BIOSTIMULANTS AND THEIR TRANSFORMATIVE ROLE IN AGRICULTURE

Abstract

The need to feed a world population that is expanding, soil deterioration, and climate change are all serious difficulties for agriculture. Seaweed extracts, humic materials, and microbial cultures are among the natural sources of biostimulants, a family of compounds that provide a revolutionary means of augmenting plant growth and resistance. Biostimulants, as opposed to conventional fertilizers and insecticides, enhance nutrient uptake, promote stress tolerance, and stimulate natural processes to improve plant health. The various forms of biostimulants, such as amino acids, protein hydrolysates, seaweed extracts, and advantageous microbes and fungi, are examined in this review. It explores various modes of action, including enhanced interactions with the soil microbiome, hormone modulation, and antioxidant activities. Biostimulants provide a wide range of potential advantages, including higher crop yields, better resilience to environmental stresses, and a decreased need for chemical inputs, all of which support more environmentally friendly farming methods. The need for environmentally sustainable agricultural solutions is driving a sharp increase in the usage of biostimulants, according to market trends. Sales are expected to exceed 2 billion globally by 2018, thus farmers will likely embrace them more widely as they look for more affordable and ecologically friendly options. Standardizing biostimulant chemicals and comprehending their intricate interactions across many agricultural systems, however, continue to provide difficulties. This review attempts to give a thorough summary of the state of biostimulant research as it stands today, emphasizing significant discoveries, innovations in technology, and potential paths forward. This study highlights the potential of biostimulants to transform contemporary agriculture and make it more robust, sustainable, and able to meet the demands of global food security by promoting a deeper understanding of them.

Keywords: Biostimulant, seaweed extract, agriculture

1. Introduction

The probability of hunger is likely to increase by 30% by 2050 as a result of climate change and anticipated population growth (Van Dijk, Morley *et al.* 2021). Given the state of agricultural productivity today, biostimulants show promise as a tool. The most important

factor that has a major impact on agricultural yield and production, endangering both global food security and crop production systems sustainability, is the environmental condition (Swaminathan and Keshvan, 2012). A substance or combination of several naturally occurring organic compounds known as "biostimulants" can stimulate plant growth in a variety of environmental stresses (Bhupenchandra *et al.*, 2020). Over the last few decades, the application of biostimulants in agriculture has increased dramatically, and by 2018, sales of these products are expected to reach 2 billion. This is mostly because there are many different sources for these components and the final product is complicated and typically contains a large number of poorly characterized compounds. Given that biostimulants come from an extensive selection of both inorganic and biological sources (Calvo *et al.*, 2014) and unreasonable to assume that there is a single mode of action for a variety of substances, including microbial fermentations of animal or plant feedstock, living microbial cultures, macro- and micro-alga, protein hydrolysate, humic and fulvic substances, composts, manures, food, and industrial wastes prepared using widely divergent industrial manufacturing processes. Oceans and seas are often referred to as the "source of life" due to their crucial role in sustaining and supporting a wide range of ecological and biological processes. Oceans and seas are essential for sustaining life on Earth, influencing weather and climate, supporting biodiversity and providing resources and economic benefits. The current view is that newly developed synthetic fertilizers endanger both local and global ecosystems (Koli *et al.*, 2019). Among agriculture's main difficulties, today is developing ecologically friendly and sustainable solutions to meet the issue of feeding the world's rising population (Manono, 2016). The only way to meet this goal, given the steadily declining amount of arable land, is to boost crop yields while safeguarding future harvests. Plant biostimulants (PBs) are potentially a tool to mitigate climate change-induced stress and reduce the dependency on chemical fertilizers (Hunter *et al.*, 2017). Applying Plant Biostimulants is an environmentally friendly technique for farmers to maintain crop productivity while using less fertilizer (Gupta *et al.*, 2020). The biostimulants increase the yields of crops by lowering yield losses during stressful situations. This strategy can contribute to enhancing food security for a growing global population in the context of rising climate change risks (Jing Li *et al.*, 2022).

2. Bio stimulant and its importance in sustainable agriculture

Extracts obtained from organic raw materials with bioactive compounds are known as biostimulants. The term "plant biostimulant" clearly defines components, apart from fertilizers and insecticides, which, when exposed to seeds, plants, or exact formulations of growth substrates can alter the biological processes of plants in a way that provides for potential edge

to development, growth, and/or the stress reaction (Du Jardin *et al.*, 2012). The stress-response-feedback continuum in plants is an important physiological process that influences plant plasticity. Abiotic and biotic stressors are the two primary categories of stress in plants. Herbivory, pest assault, and disease-causing pathogens are examples of biotic stressors, while drought, salt, heavy metals, floods, and severe temperatures are examples of abiotic stressors. (Gull *et al.*, 2019). To increase crop production, quality, and productivity biostimulants provide biologically active chemicals that may enhance plant metabolic activity. (Qurat-UI-Ain Raza *et al.*, 2022). They seem to be a general term for anything that is good for plants but isn't a fertilizer, insecticide, or soil enhancer. They possess the ability to alter a plant's physiological processes in a way that may be advantageous to the plant's growth, development, and/or stress response (du Jardin *et al.*, 2012). By interfering with plant signalling pathways, biostimulants increase plant output by lowering the unfavorable plant reaction to stress (Patrick Brown, and Sebastian Saa 2015). In both outdoor and greenhouse crop production, the significance of biostimulants has grown significantly in recent years. While pesticides promote plant growth by lessening the negative effects of pathogens and pests on plant integrity and functionality, nutrients provide the chemical elements needed by metabolic processes for the synthesis of biomolecules and the production of biomass. Nutrients and pesticides are specifically aside in the classification of Biostimulants to create the greatest possible clarity on this distinction (du Jardin *et al.*, 2012). The morphological, physiological, and yield attributes of tomato (*Solanum lycopersicum*) plants treated with biostimulants are much better than those of control plants (Arun Kumar *et al.*, 2020). Because they function as a chemical catalyst in plants and enhance attributes like plant height, number of branches per plant, number of leaves, number of pods per plant, length of pod, number of grains per pod, yield, grain quality, plant growth regulators and biostimulants can raise yield in fenugreek (Anandhi *et al.*, 2019). The morphological parameters of Tuberose (*Polianthes tuberosa* L.) cv. Prajwal can be improved, along with the flower yield and quality, by applying biostimulants externally on the leaves and biofertilizers drenching with the soil (Arul Arasu *et al.*, 2023). Crop regulation makes use of biostimulants. The mutually beneficial relationship between nutrients and biostimulants (Anbukkarasi *et al.*, 2018).

3. Formulations of Biostimulants

The utilisation of cold cellular-burst technology is responsible for the elevated concentrations of these two biomolecules that promote plant development in the Kelpak® samples. To find the ideal concentrations of eckol and phloroglucinol for plant growth in various liquid seaweed fertilisers, more research is needed (Rengasamy *et al.*, 2016). The

impact of foliar and soil drench applications of liquid seaweed extracts (LSEs) derived from *Ulva lactuca*, *Caulerpa sertularioides*, *Padina gymnospora*, and *Sargassum liebmannii* as biostimulants on tomato (*Solanum lycopersicum*) germination and growth under laboratory and greenhouse conditions. **Liquid seaweed extracts (LSE)** increase fruit development and maintain plant growth (Hernández-Herrera et al. 2014).

4. Classification and various types of BS

There are various kinds of biostimulants, including inorganic chemicals, microbial biostimulants, plant and algae extracts, and acid-based biostimulants. Based on the source of raw material it is classified as including microbes, protein hydrolysates, seaweed extracts, humic and fulvic acid compounds (Miguel Baltazar *et al.*, 2021). These several kinds of biostimulants each have advantages in terms of improved fertility, enhanced vigour, benefits to plant health, and improved crop quality (Papantzikose *et al.*, 2024). Biostimulants include a wide range of compounds, including extracts from seaweed, humic substances, complex organic materials, beneficial chemical components, inorganic salts, chitin, and chitosan derivatives, Antitranspirants, free amino acids, and other compounds containing nitrogen (du Jardin *et al.*, 2012). A foliar spray containing different biostimulant components, including as phytohormones, humic acid, and seaweed extract, was used. It promotes natural processes that improve crop quality, tolerance to abiotic stress, and nutrient uptake. Biostimulants are an efficient way of influencing physiological and biochemical features that lead to increased yield and growth (Kavipriya and Boominathan., 2018).

5. Role of seaweed extract-based biostimulant

As a result of their advantageous effects on soil, both macro- and microalgae have long been employed to increase plant productivity and food production in different parts of the world. Algae and the soil ecosystem surely interact in complex ways, and the benefits that result vary depending on the crop and the local environmental factors. This has led to a great deal of conjecture on the mechanisms at play and the veracity of the results that have been published. It has been 60 years since the first seaweed extract was produced commercially for use in agriculture. For the first time, these aqueous extracts made it possible to directly apply soluble seaweed components to certain plant organs like leaves and roots. Seaweed extracts have opened up new and interesting possibilities for use in both plant and animal applications. (Craigie *et al.*, 2011).

The seaweeds are photoautotrophic marine algae with a variety of traits. These multicellular organisms are largely derived from water-based environments and provide a wealth of possible applications as well as being economically viable, renewable, and harmless

to the environment resources. (Stirk *et al.*, 2020). The proportionate effectiveness of SWE use in the agricultural sector demonstrates the beneficial effects on plant development, yield, nutritional quality, and bioactive content. Application of **Seaweed Extract (SWE)** is therefore linked to a greater capacity of plants to withstand both abiotic and biotic stresses. (Guinan *et al.*, 2012). Many macroalgae are employed in the synthesis of biostimulants; however, in the past few years, *Ascophyllum nodosum*, *Ecklonia maxima*, and *Kappaphycus alvarezii*, together with the genera *Gracilaria* spp., got particular interest. (Shukla *et al.*, 2021 & 2024). Red seaweeds include *Kappaphycus alvarezii*, *Gracilaria edulis*, *Acanthophora spicifera*, *Gelidium robustum*, and *Gracilaria parvispora*, and brown seaweeds include *Ascophyllum nodosum*, *Fucus* spp., *Laminaria setchellii*, *Sargassum hildebrandtii*, *Turbinaria* spp., *Macrocystis pyrifera*, *Sargassum horridum*, *Ecklonia arborea*, *Durvillaea antarctica*, and *Sargassum horridum* (Ali *et al.*, 2021). One of the most popular biostimulants made from different seaweed species is seaweed extract (SWE). The main component of commercially marketed SWEs is polysaccharides, which make up between 30 and 40 percent of their fractions (Battacharyya *et al.*, 2015).

The *Ascophyllum nodosum* extract (ANE), application of ANE (Sealicit™) to soybean in Brazil and Canada indicates the favorable benefits of anti-shattering, pod dehiscence, firmness, seed weight, and eventually seed yield in soybean (*Glycine max*) (Lukasz Langowski *et al.* 2021). Spraying seaweed extract at seedling, stem elongation, and early mature stage improves enzyme activity to increase sucrose % on sugarcane stems. (Diwen Chen *et al.* 2021). **Ascophyllum nodosum extract (ANE)** influences the mechanism of nitrogen (N) uptake in plants, assimilated in plant parts. In a better way, it allowed to reduce the N fertilization and simultaneously increase the yield of barley. It maintains **Nutrient Use Efficiency (NUE)**, and reduces **Nitrogen (N)** fertilizer application by up to 27%. In another way it leads to sustainable, profitable, and eco-friendly agriculture (Oscar Goni *et al.*, 2021)

6. Role of Humic acid as biostimulant

Humic acid primarily contributes to increased root growth, as well as morphological and physiological alterations in roots and shoots associated with nutrient intake, distribution, and assimilation. It may additionally trigger modifications in primary and secondary plant metabolism associated with resistance to abiotic stress. It was formed by non-renewable resources such as peat and coal (Canellas *et al.*, 2015). These proteins were analyzed further using bioinformatic techniques, and their biological functions protein synthesis, folding and elongation, and energy and metabolism were organized into three primary categories. Additionally, these scientists were able to connect the found proteins to several biological

processes, including respiration, the metabolism of energy and cell walls, protein synthesis, folding, and degradation, as well as responses to heat, inorganic chemicals, and cell transport and division. These findings might clear the way for a deeper comprehension of the molecular pathways that **Humic Acid (HA)** favorably affects. The quantity of carbohydrates and the majority of free amino acids in the roots were significantly reduced in a metabolomic investigation that also used *A. thaliana* and treatments with **Humic Acid (HA)** as a biostimulant (Conselvan *et al.*, 2018). A rise in protein content in the roots and leaves, most likely as a result of increased protein synthesis and metabolic activity, may be contributing to the plants treated with **Humic Acid** growing at a faster rate (Byun *et al.*, 2021). Humic acid, when used as a biostimulant in cucumber crops, boosts crop growth parameters such as plant height, stem diameter, and biomass content, as well as ammonia and potassium storage capacity and availability of P status. He concludes that 1% **Humic Acid** applied to substrates (cocopeat) improves plant development. (Jingcheng Xu *et al.*, 2021). Advantages of **Humic Acid** on growth indices, carotenoid content, antioxidant activity, and nutritional value of yarrow in field and greenhouse conditions. In alkaline soils with low pH, yarrow can grow and become more antioxidant-active by applying HA (Hassan Bayat *et al.*, 2021). In maize roots, humic compounds have been shown to encourage cell wall elongation and loosening (Jindo *et al.*, 2011).

7. Role of fulvic acid as biostimulant

The FA are characterized by high concentrations of carboxylic groups (COOH), low concentrations of aromatic structures, and high concentrations of phenolic substances (Canellas *et al.*, 2015). In spring wheat (*Triticum aestivum*), barley (*Hordeum vulgare*), and sugar beetroot (*Beta vulgaris*), **Fulvic Acid (FA)** biostimulants were found to enhance germination while also increasing shoots and increasing the dry weight of shoots and roots. Through faster cellular water potential restoration, osmotic adjustment, increased stomatal conductance, photosynthetic activity, and photoassimilate production, higher efficiency in energy dissipating mechanisms, which reduced **Reactive Oxygen Species (ROS)** generation, and higher activity of antioxidant enzymes, fulvic acids induced soybean plants to recover from water deficits more effectively (Rosa *et al.*, 2020). When given at various phenological stages, fulvic acid may mitigate the harmful effects salt stress causes to the photosynthetic system in soybean plants. The benefits of **Fulvic Acid (FA)** on yarrow's growth indices, carotenoid content, antioxidant activity, and nutritional content in both greenhouse and field settings. Applying **Fulvic Acid (FA)** to yarrow can help it grow and become more antioxidant-active, especially in alkaline soils with low (Hassan Bayat *et al.*, 2021).

8. Role of microorganisms-based biostimulant

It's possible that biostimulants based on bacteria and fungi can help ameliorate the negative environmental effects of agriculture (Nuti *et al.*, 2015). Additionally, microbes are essential to the phyllosphere, rhizosphere, and endosphere of plants because they increase the availability of certain nutrients and make them easier for plants to absorb. The symbiotic relationship between the two has been vital for the evolution of both (Vandenkoornhuysen *et al.*, 2015). Many microorganisms, including *Arthrobacter* spp, *Pseudomonas* spp, *Rhodococcus* spp, *Enterobacter* spp, *Ochrobactrum* spp, *Acinetobacter* spp, *Bacillus* spp, *Rhizobium* spp, and *Streptomyces* spp, have been thoroughly investigated to explore their possible function as biostimulants; some of these species have even been commercialized (Rodriguez *et al.*, 2020).

Several soils were used to extract *Bacillus* spp and *Pseudomonas* spp, and this not only serves as bio fungicides to support the health of plants and soil (Caulier *et al.*, 2018). The use of *Enterobacter* strain 15S may have solubilized calcium phosphate ($\text{Ca}_3(\text{PO}_4)_2$) to assist plants in overcoming P shortage, whereas the association of maize activated plant metabolism. Based on nutritional status, this mechanism enhances plant growth and P nutrition. The amount of chlorophyll was assessed using the chlorophyll meter (SPAD). (Mónica Yorlady Alzate Zuluaga *et al.*, 2021).

9. Role of Peptone and Protein biostimulant on crops

Fruit qualities such as sugar deposits, overall acidity, carotenoid levels, and vitamins C and E were preserved when peptone was utilized as a biostimulant. Peptone promotes vegetative growth and fruit yield by increasing cytokinin accumulation and photosynthetic pigments in plants. Peptone treatment effectively increases tomato yield in greenhouse conditions when N priming is used. (Tania Mesa *et al.*, 2022). Protein-based biostimulants derived from enzymatic hydrolysis may be a suitable strategy for enhancing productivity and improve the utilization of specific nutrients, including nitrogen. (Colla *et al.*, 2017). Through the application of Protein hydrolysates, it improves in photosynthetic rate, root growth, and upregulation of genes linked to antioxidant activity, photosynthesis, nutrient uptake, and primary metabolisms; increased biomass, concentration of soluble sugars, and levels of chlorophyll and phenolic content in plants. Elevated levels of terpenes, alkaloids, phenylpropanoids, and chemicals containing nitrogen (Andrea ertani *et al.*, 2017). Enhanced stomatal conductance and photosynthetic rate during drought stress; lessening the adverse impacts of stress on transpiration rates and gas exchange. Enhance the drought tolerance and increase the chlorophyll content in broccoli (kaluzewicz *et al.*, 2017). Increased the amount of

plant biomass and leaf chlorophyll by stimulating the proliferation of microorganisms that promote plant growth (Luziatelli *et al.*, 2019).

Table 1. Cumulative potential benefits of biostimulants in Agricultural sector

Biostimulant	Plant Species	Plant Response	References
	<i>Arabidopsis thaliana</i>	Increased heat stress tolerance through activation of heat-shock protein genes; elevated amounts of proteins involved in energy metabolism and cell walls; respiration, folding, protein synthesis, degradation, reaction to heat and inorganic chemicals, cell trafficking and division, and a drop in the concentration of amino acids and carbohydrates.	Cha,J.-Y.; Kang <i>et al.</i> , 2020
	<i>Brassica napus</i>	Reduction in soluble carbohydrates, linolenic and erucic acid; increase in yield and chlorophyll percentage; enhanced oil quality, plant net the processes of photosynthesis gas exchange rate, and electron transport flux.	Kuan Qin <i>et al.</i> , 2018
Humic acids	<i>Capsicum annuum L.</i>	Enhanced chlorophyll content resulted in enhanced net photosynthesis; enhanced root development and increased plant biomass under drought stress while quickly decreasing leaf stomatal conductance and transpiration rates.	Nasiri <i>et al.</i> , 2021
	<i>Phaseolus vulgaris L.</i>	It alters the stress-related gene expression	Taspinar <i>et al.</i> , 2017
	<i>Solanum tuberosum</i>	Improved plant growth, nutrient transport, and photosynthetic indices under drought stress; increased tuber production and plant biomass.	Yang Man-hong <i>et al.</i> , 2020
	<i>Zea mays</i>	Enhanced efficiency in using water and nitrogen; increased expression of genes involved in water transport, nutrient absorption, and nitrate transporters	de Azevedo <i>et al.</i> , 2019
Fulvic acid	<i>Arabidopsis thaliana</i>	Reduced production of reactive oxygen species (ROS), oxidative as well as induce drought stress tolerance,	Rasul <i>et al.</i> , 2021

and downregulation of genes associated with growth impairment under stress

	<i>Beta vulgaris</i>	Enhanced germination parameters, including soluble sugar content, root size, and yield	Braziene <i>et al.</i> , 2021
	<i>Glycine max</i> L.	Enhanced stomatal conductance, photosynthetic efficiency and activity, chlorophyll content, and antioxidant activity under drought stress and root growth	Do Rosário Rosa <i>et al.</i> , 2021
	<i>Allium cepa</i>	Yield growth and a decrease in downy mildew severity, Improving the onion's chlorophyll and carotenoid concentrations, as well as its plant height, leaf count, bulb diameter, protein content, and sulfur content.	Hidangmayum <i>et al.</i> , 2017
	<i>Lycopersicon esculentum</i>	Improve photosynthetic indices, pollen viability, and thermo tolerance at both normal and high temperatures; higher fruit yield and chlorophyll content. Increase fruit yield components in both salinity-stressed and normal circumstances	Di Stasio <i>et al.</i> , 2020
	<i>Malus domestica</i>	It minimises the alternate bearing habit	Spinelli <i>et al.</i> , 2009
Seaweed extracts	<i>Solanum melongena</i>	Greater productivity and growth of vegetation	Abd El-Gawad and Osman (2014)
(<i>Ascophyllum nodosum</i>)	<i>Triticum aestivum</i> L.	Enhanced germination circumstances and produced better grain quality and yield.	Braziene <i>et al.</i> , 2021
	<i>Vitis vinifera</i> L.	Increased berry number, anthocyanin, phenolic content, plant biomass, yield, Nitrogen (N), and soluble sugar concentration all without adversely affecting the quality of the berries.	Frioni <i>et al.</i> , 2018
	<i>Oryza sativa</i>	Improved germination, seedling vigour, and root growth; increased yield parameters, grain number, protein, and nutritional content in the grain, plant biomass, and chlorophyll content.	Layek <i>et al.</i> , 2018

Seaweed extracts (Ecklonia maxima)	<i>Phaseolus vulgaris</i> L.	Improve better nutritional quality, higher yield, and antioxidant activity in the seeds	Kocira <i>et al.</i> , 2020
	<i>Saccharum officinarum</i>	Improve brix content in the juice	Karthikeyan. K <i>et al.</i> , 2017
	<i>Zea mays</i>	Increased yield, photosynthetic pigments, antioxidants, grain quality, and protein content under drought stress Improve plant growth, antioxidant activity, and nutrient uptake, under ideal conditions	Khanjan Trivedi <i>et al.</i> , 2018
Kappaphycus alvarezii	<i>Solanum esculentum</i>	Fruit production, length, diameter, and nutritional quality also increased	Zodape <i>et al.</i> (2008)
	<i>Solanum tuberosum</i> L	Enhanced plant growth indices; higher yield, higher quality yield; higher concentration of nutrients; higher ascorbic acid and soluble sugar content.	Patel <i>et al.</i> , (2018)
	<i>Saccharum officinarum</i>	Enhanced plant growth and juice brix content; increased plant production.	Karthikeyan <i>et al.</i> , 2017
		Improved plant stress tolerance; increased root development, photosynthetic pigment concentration, RWC, amino acid content, proline, and soluble sugar content	Khantika Patel <i>et al.</i> , 2018
	<i>Zea mays</i>	Reduced photosystem damage and lipid peroxidation; improved yield parameters, photosynthetic pigments, antioxidants, grain quality, and protein content; under drought stress. Unfavorable circumstances, increased yield characteristics and quality, enhanced nutrient uptake, enhanced plant growth, reduced lipid peroxidation, and ROS production.	Trivedi <i>et al.</i> , 2023

10. Physiology and metabolism

Seaweed extracts and their chemical constituents have a special effect on plants' metabolic regulatory pathways, which are being illuminated by recent developments in seaweed extract research that involve gene expression analysis. In spinach treated with a commercial brown algal extract, there was an increase in total soluble protein, antioxidant properties, content of phenolics and flavonoids, and transcript abundance of regulatory enzymes involved in nitrogen metabolism (cytosolic glutamine synthetase, glutathione reductase, and betaine aldehyde dehydrogenase and choline monooxygenase) (Fan *et al.*, 2013). The synthesis of chloroplasts increased, chlorophyll degradation decreased, and senescence was delayed, all of which contributed to the rise in chlorophyll concentration. It enhances maize productivity despite its low phosphorus uptake (Jannin *et al.*, 2013). The problem of alternating bearing under nutrient-deprived conditions was resolved by applying a commercial seaweed extract to apple trees (variety Fuji), but not under normal nutrient management conditions. Furthermore, the biostimulant minimized fruit production oscillations in alternate-bearing trees by improving plant growth, fruit production, and fruit quality (Spinelli *et al.*, 2009). It seems that the presence of a hormone or signaling component in a commercial seaweed extract derived from *A. nodosum* may contribute to its effect on alternate bearing under nutrient-deprived situations.

11. Uptake of nutrients

It is also known that different seaweed extracts have an impact on the regulation of genes that are crucial for nutrient uptake. For instance, *A. nodosum* extract enhanced auxin transport and nitrogen sensing by upregulating the expression of the nitrate transporter gene NRT1 (Krouk *et al.*, 2010). Furthermore, it is known that certain chemical components of brown seaweed extract stimulate the growth and root colonization of helpful soil fungi. suggest that in addition to having **Arbuscular Mycorrhiza (AM)** stimulatory chemicals, red and green algae aid in the growth of mycorrhizal fungi in higher plants (Kuwada *et al.*, 2006). It has been found that applying brown seaweed extract topically increases grapevine copper uptake. Water stress tolerance, a rapid recovery in rehydrated plants, and the maintenance of a greater leaf water potential and stomatal conductance during the stress period were all induced by the extract with remarkable efficacy (Turan and Köse, 2004). Seaweed extracts include nutrients that are easily absorbed by leaves via their hydrophilic cuticle pores and stomata. Environmental factors that impact stomata opening, cuticle and cell wall permeability, and temperature, humidity, or light intensity also impact the absorption of these mineral nutrients from the leaf surface. It was demonstrated that foliar application of seaweed extract products,

including those of a commercial *A. nodosum* extract, enhanced the Copper (Cu) uptake in grapevine under conditions of nutrient deficiency. This was likely due to increased cell membrane permeability.

12. Abiotic stress tolerance

Additionally, the effects of intensive agricultural methods are a major factor in the frequent occurrence of unfavorable conditions for crop plant growth and development. Globally, abiotic stressors are responsible for significant losses in crop productivity. Salinity alone has the potential to severely reduce the productivity of important food-producing crops. (Zhu, 2000). Applying an *A. nodosum* extract formulation, for instance, reduced the leaves' osmotic potential, a crucial marker of osmotic tolerance, and enhanced grapes' ability to withstand freezing temperatures. In greenhouse experiments, a commercial extract of *A. nodosum*-treated vegetables, bedding plants, and turf crops significantly reduced water use (better water use efficiency), increased leaf water content, and improved drought-wilted plant recovery when compared to controls. Two weeks apart, root treatments of a commercial *Ascophyllum* extract to almonds increased the treated plots' negative mid-day stem-water potential (Little and Neily, 2010).

Abiotic stress has become more common in recent years, mostly as a result of climate change, which has led to an extraordinary rise in extreme weather events and patterns. Additionally, the effects of intensive agricultural methods are causing unfavorable conditions for crop plant growth and development to spread widely. Abiotic stress is responsible for significant global crop output losses. Kobayashi et al., 2008 reported that abiotic stresses such as salinity, drought, waterlogging, and high temperatures are some of the primary factors that influence the quantity and quality of horticultural crops. Zhang et al., 2007 proposed that an increase in **Potassium (K⁺)** uptake and the presence of "cytokinin-like" molecules in the extract were responsible for of this uptake.

Applying seaweed extracts to creeping bentgrass increased its heat tolerance. These abiotic stresses are predicted to have a greater adverse impact by the real climate change scenario, raising major concerns about crop productivity and, consequently, global food security (Rouphael *et al.*, 2018). The biostimulant based Glycine betaine (GB)- enhances the crop abiotic stress tolerance (drought), and treated plants get higher photosynthetic activity, through the accumulation of lipids and thickening of leaf resulting in ass increase **Water Use Efficiency (WUE)**. **Generalized Additive Mixed Modeling (GAMM)** was used to evaluate the photosynthetic efficiency. (Giulia Antonucci *et al.*, 2021). Among these, GAMMs have been implemented successfully in a wide range of applied science areas. (Murase *et al.*, 2009;

Ohana-Levi et al., 2020). Similarly, the impacts of GB on water-use efficiency (WUE) have received less attention. (Ahmed *et al.*, 2019).

13. Conclusion

In conclusion, by improving plant growth, stress tolerance, and nutrient uptake, biostimulants outperform conventional fertilizers and pesticides in sustainable agriculture. They are essential for combating rising food prices, degraded soil, and climate change. The types, processes, and revolutionary potential of biostimulants in agriculture are reviewed in this paper. Growing worldwide sales of biostimulants are expected, reflecting a shift in farming practices toward more environmentally friendly options. For widespread application, nevertheless, issues like product standardization and comprehension of biostimulant interactions in diverse agricultural systems need to be resolved. Developing application guidelines and elucidating biostimulant mechanisms should be the main goals of future studies. Effective integration requires cooperation between industry, politicians, and researchers. To sum up, biostimulants have the potential to completely transform agriculture by improving plant health and yield while lowering the need for chemical inputs. Using biostimulants will result in robust, environmentally friendly farming systems that will improve both environmental sustainability and global food security.

Disclaimer (Artificial intelligence)

Option 1:

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts. – I hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts.

Option 2:

Author(s) hereby declare that generative AI technologies such as Large Language Models, etc have been used during writing or editing of manuscripts. This explanation will include the name, version, model, and source of the generative AI technology and as well as all input prompts provided to the generative AI technology

Details of the AI usage are given below:

- 1.
- 2.
- 3.

References

1. Ahmed, R., Liu, G., Yousaf, B., Abbas, Q., Ullah, H., & Ali, M. U. (2020). Recent advances in carbon-based renewable adsorbent for selective carbon dioxide capture and separation-A review. *Journal of Cleaner Production*, 242, 118409.
2. Ali, O., Ramsubhag, A., & Jayaraman, J. (2021). Biostimulant properties of seaweed extracts in plants: Implications towards sustainable crop production. *Plants*, 10(3), 531.
3. Alzate Zuluaga, M. Y., Martinez de Oliveira, A. L., Valentinuzzi, F., Tiziani, R., Pii, Y., Mimmo, T., & Cesco, S. (2021). Can inoculation with the bacterial biostimulant *Enterobacter* sp. strain 15S be an approach for the smarter P fertilization of maize and cucumber plants?. *Frontiers in Plant Science*, 12, 719873.
4. Anandhi, M., Ramar, A., Jegadeeswari, V., & Srinivasan, S. (2019). Effect of biostimulants and growth regulators on plant growth and herbage yield of fenugreek (*Trigonella foenum-graecum* L.). *Journal of Pharmacognosy and Phytochemistry*, 8(3), 3364-3367.
5. Anbukkarasi, V., Prabu, M., Paramaguru, P., Pugalendhi, L., & Jeyakumar, P. (2018) Effect of Nutrients and Biostimulants on Growth, Yield and Quality of Tomato (*Solanum lycopersicon*). *International Journal of Current Microbiology and Applied Sciences*
6. Baltazar, M., Correia, S., Guinan, K. J., Sujeeth, N., Bragança, R., & Gonçalves, B. (2021). Recent advances in the molecular effects of biostimulants in plants: An overview. *Biomolecules*, 11(8), 1096.
7. Battacharyya, D., Babgohari, M. Z., Rathor, P., & Prithiviraj, B. (2015). Seaweed extracts as biostimulants in horticulture. *Scientia horticultrae*, 196, 39-48.
8. Bayat, H., Shafie, F., Aminifard, M. H., & Daghighi, S. (2021). Comparative effects of humic and fulvic acids as biostimulants on growth, antioxidant activity and nutrient content of yarrow (*Achillea millefolium* L.). *Scientia Horticulturae*, 279, 109912.
9. Bayat, H., Shafie, F., Aminifard, M. H., & Daghighi, S. (2021). Comparative effects of humic and fulvic acids as biostimulants on growth, antioxidant activity and nutrient content of yarrow (*Achillea millefolium* L.). *Scientia Horticulturae*, 279, 109912.
10. Bhupenchandra, I., Devi, S. H., Basumatary, A., Dutta, S., Singh, L. K., Kalita, P. & Borah, K. (2020). Biostimulants: Potential and prospects in agriculture. *Int. Res. J. Pure Appl. Chem*, 21, 20-35.

11. Braziene, Z., Paltanavicius, V., & Avizienytė, D. (2021). The influence of fulvic acid on spring cereals and sugar beets seed germination and plant productivity. *Environmental research*, 195, 110824.
12. Braziene, Z., Paltanavicius, V., & Avizienytė, D. (2021). The influence of fulvic acid on spring cereals and sugar beets seed germination and plant productivity. *Environmental Research*, 195, article number 110824. doi: 10.1016/j.envres.2021.110824
13. Brown, P., & Saa, S. (2015). Biostimulants in agriculture. *Frontiers in plant science*, 6, 671.
14. Byun, M. Y., Kim, D., Youn, U. J., Lee, S., & Lee, H. (2021). Improvement of moss photosynthesis by humic acids from Antarctic tundra soil. *Plant Physiology and Biochemistry*, 159, 37-42.
15. Calvo, P., Nelson, L., & Kloepper, J. W. (2014). Agricultural uses of plant biostimulants. *Plant and soil*, 383, 3-41.
16. Canellas, L. P., Olivares, F. L., Aguiar, N. O., Jones, D. L., Nebbioso, A., Mazzei, P., & Piccolo, A. (2015). Humic and fulvic acids as biostimulants in horticulture. *Scientia horticulturae*, 196, 15-27.
17. Canellas, L. P., Olivares, F. L., Aguiar, N. O., Jones, D. L., Nebbioso, A., Mazzei, P., & Piccolo, A. (2015). Humic and fulvic acids as biostimulants in horticulture. *Scientia horticulturae*, 196, 15-27.
18. Caulier, S., Gillis, A., Colau, G., Licciardi, F., Liépin, M., Desoignies, N., ... & Bragard, C. (2018). Versatile antagonistic activities of soil-borne *Bacillus* spp. and *Pseudomonas* spp. against *Phytophthora infestans* and other potato pathogens. *Frontiers in microbiology*, 9, 143.
19. Cha, J. Y., Kang, S. H., Ali, I., Lee, S. C., Ji, M. G., Jeong, S. Y., ... & Kim, W. Y. (2020). Humic acid enhances heat stress tolerance via transcriptional activation of Heat-Shock Proteins in Arabidopsis. *Scientific reports*, 10(1), 15042.
20. Chen, D., Zhou, W., Yang, J., Ao, J., Huang, Y., Shen, D., & Shen, H. (2021). Effects of seaweed extracts on the growth, physiological activity, cane yield and sucrose content of sugarcane in China. *Frontiers in Plant Science*, 12, 659130.
21. Colla, G., Hoagland, L., Ruzzi, M., Cardarelli, M., Bonini, P., Canaguier, R., & Rouphael, Y. (2017). Biostimulant action of protein hydrolysates: Unraveling their effects on plant physiology and microbiome. *Frontiers in plant science*, 8, 2202.

22. Conselvan, G. B., Fuentes, D., Merchant, A., Peggion, C., Francioso, O., & Carletti, P. (2018). Effects of humic substances and indole-3-acetic acid on Arabidopsis sugar and amino acid metabolic profile. *Plant and Soil*, 426, 17-32.
23. Craigie, J. S. (2011). Seaweed extract stimuli in plant science and agriculture. *Journal of applied phycology*, 23, 371-393.
24. de Azevedo, I. G., Olivares, F. L., Ramos, A. C., Bertolazi, A. A., & Canellas, L. P. (2019). Humic acids and *Herbaspirillum seropedicae* change the extracellular H⁺ flux and gene expression in maize roots seedlings. *Chemical and biological technologies in agriculture*, 6, 1-10.
25. Di Stasio, E., Cirillo, V., Raimondi, G., Giordano, M., Esposito, M., & Maggio, A. (2020). Osmo-priming with seaweed extracts enhances yield of salt-stressed tomato plants. *Agronomy*, 10(10), 1559.
26. Dijk, M. V., Morley, T., Rau, M. L., & Saghai, Y. (2021). Van Dijk et al.(2021), A meta-analysis of projected global food demand and population at risk of hunger for the period 2010–2050, data and scripts.
27. do Rosário Rosa, V., Dos Santos, A. L. F., da Silva, A. A., Sab, M. P. V., Germino, G. H., Cardoso, F. B., & de Almeida Silva, M. (2021). Increased soybean tolerance to water deficiency through biostimulant based on fulvic acids and *Ascophyllum nodosum* (L.) seaweed extract. *Plant Physiology and Biochemistry*, 158, 228-243.
28. du Jardin, P. (2012). The Science of Plant Biostimulants—A bibliographic analysis, Ad hoc study report.
29. El-Gawad, A., El-Shazly, M. M., & Ahmed, A. E. importance of biofertilization and marine algal extract in improving growth and productivity of faba bean cultivated under new valley conditions.
30. Ertani, A., Schiavon, M., & Nardi, S. (2017). Transcriptome-wide identification of differentially expressed genes in *Solanum lycopersicon* L. in response to an alfalfa-protein hydrolysate using microarrays. *Frontiers in Plant Science*, 8, 1159.
31. Fan, D., Hodges, D. M., Critchley, A. T., & Prithiviraj, B. (2013). A commercial extract of brown macroalga (*Ascophyllum nodosum*) affects yield and the nutritional quality of spinach in vitro. *Communications in soil science and plant analysis*, 44(12), 1873-1884.
32. Frioni, T., Sabbatini, P., Tombesi, S., Norrie, J., Poni, S., Gatti, M., & Palliotti, A. (2018). Effects of a biostimulant derived from the brown seaweed *Ascophyllum*

nodosum on ripening dynamics and fruit quality of grapevines. *Scientia Horticulturae*, 232, 97-106.

33. Goñi, O., Łangowski, Ł., Feeney, E., Quille, P., & O'Connell, S. (2021). Reducing nitrogen input in barley crops while maintaining yields using an engineered biostimulant derived from *Ascophyllum nodosum* to enhance nitrogen use efficiency. *Frontiers in plant science*, 12, 664682.
34. González-Pérez, B. K., Rivas-Castillo, A. M., Valdez-Calderón, A., & Gayosso-Morales, M. A. (2022). Microalgae as biostimulants: A new approach in agriculture. *World Journal of Microbiology and Biotechnology*, 38(1), 4.
35. Guinan, K. J., Sujeeth, N., Copeland, R. B., Jones, P. W., O'brien, N. M., Sharma, H. S. S., ... & O'sullivan, J. T. (2012, November). Discrete roles for extracts of *Ascophyllum nodosum* in enhancing plant growth and tolerance to abiotic and biotic stresses. In *I World Congress on the Use of Biostimulants in Agriculture 1009* (pp. 127-135).
36. Gull, A., Lone, A. A., & Wani, N. U. I. (2019). Biotic and abiotic stresses in plants. *Abiotic and biotic stress in plants*, 7, 1-9.
37. Gupta, G., Dhar, S., Dass, A., Sharma, V. K., Shukla, L., Singh, R., & Verma, G. (2020). Assessment of bio-inoculants-mediated nutrient management in terms of productivity, profitability and nutrient harvest index of pigeon pea–wheat cropping system in India. *Journal of Plant Nutrition*, 43(19), 2911-2928.
38. Hendrawan, Y., Rohmatulloh, B., Prakoso, I., Liana, V., Fauzy, M. R., Damayanti, R., ... & Al Riza, D. F. (2021, November). Classification of large green chilli maturity using deep learning. In *IOP Conference Series: Earth and Environmental Science* (Vol. 924, No. 1, p. 012009). IOP Publishing.
39. Hernández-Herrera, R. M., Santacruz-Ruvalcaba, F., Ruiz-López, M. A., Norrie, J., & Hernández-Carmona, G. (2014). Effect of liquid seaweed extracts on growth of tomato seedlings (*Solanum lycopersicum* L.). *Journal of applied phycology*, 26, 619-628.
40. Hidangmayum, A., & Sharma, R. (2017). Effect of different concentrations of commercial seaweed liquid extract of *Ascophyllum nodosum* as a plant bio stimulant on growth, yield and biochemical constituents of onion (*Allium cepa* L.). *Journal of Pharmacognosy and Phytochemistry*, 6(4), 658-663.
41. Jannin, L., Arkoun, M., Etienne, P., Lainé, P., Goux, D., Garnica, M., ... & Ourry, A. (2013). Brassica napus growth is promoted by *Ascophyllum nodosum* (L.) Le Jol. seaweed extract: microarray analysis and physiological characterization of N, C, and S metabolisms. *Journal of plant growth regulation*, 32, 31-52.

42. Jindo, K., Martim, S. A., Navarro, E. C., Pérez-Alfocea, F., Hernandez, T., Garcia, C., ... & Canellas, L. P. (2012). Root growth promotion by humic acids from composted and non-composted urban organic wastes. *Plant and Soil*, 353, 209-220.
43. KAŁUŻEWICZ, A., KRZESIŃSKI, W., SPIŻEWSKI, T., & Zaworska, A. (2017). Effect of biostimulants on several physiological characteristics and chlorophyll content in broccoli under drought stress and re-watering. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 45(1), 197-202
44. Karthikeyan, K., & Shanmugam, M. (2017). The effect of potassium-rich biostimulant from seaweed *Kappaphycus alvarezii* on yield and quality of cane and cane juice of sugarcane var. Co 86032 under plantation and ratoon crops. *Journal of Applied Phycology*, 29(6), 3245-3252.
45. Kavipriya, R., & Boominathan, P. (2018). Influence of biostimulants and plant growth regulators on physiological and biochemical traits in tomato (*Lycopersicon esculentum* Mill.). *Madras Agricultural Journal*, 105(march (1-3)), 1.
46. Kobayashi, F., Maeta, E., Terashima, A., Kawaura, K., Ogihara, Y., & Takumi, S. (2008). Development of abiotic stress tolerance via bZIP-type transcription factor LIP19 in common wheat. *Journal of Experimental Botany*, 59(4), 891-905.
47. Kocira, A., Lamorska, J., Kornas, R., Nowosad, N., Tomaszewska, M., Leszczyńska, D., ... & Tabor, S. (2020). Changes in biochemistry and yield in response to biostimulants applied in bean (*Phaseolus vulgaris* L.). *Agronomy*, 10(2), 189.
48. Koli, P., Bhardwaj, N. R., & Mahawer, S. K. (2019). Agrochemicals: harmful and beneficial effects of climate changing scenarios. In *Climate change and agricultural ecosystems* (pp. 65-94). Woodhead Publishing.
49. Krouk, G., Lacombe, B., Bielach, A., Perrine-Walker, F., Malinska, K., Mounier, E., ... & Gojon, A. (2010). Nitrate-regulated auxin transport by NRT1.1 defines a mechanism for nutrient sensing in plants. *Developmental cell*, 18(6), 927-937.
50. Kumar, K. A., Jeyakumar, P., Ravichandran, V., Swarnapriya, R., & Kalaiselvi, T. (2020). Effect of bio-stimulants on the morpho-physiological and yield traits of tomato (*Solanum lycopersicum* L.). *International Journal of Ecology and Environmental Sciences*. 4,682-686
51. Kuwada, K., Kuramoto, M., Utamura, M., Matsushita, I., & Ishii, T. (2006). Isolation and structural elucidation of a growth stimulant for arbuscular mycorrhizal fungus from *Laminaria japonica* Areschoug. *Journal of applied phycology*, 18, 795-800.

52. Łangowski, Ł., Goñi, O., Marques, F. S., Hamawaki, O. T., Da Silva, C. O., Nogueira, A. P. O., ... & O'Connell, S. (2021). *Ascophyllum nodosum* extract (Sealicit™) boosts soybean yield through reduction of pod shattering-related seed loss and enhanced seed production. *Frontiers in plant science*, *12*, 631768.
53. Layek, J., Das, A., Idapuganti, R. G., Sarkar, D., Ghosh, A., Zodape, S. T., ... & Meena, R. S. (2018). Seaweed extract as organic bio-stimulant improves productivity and quality of rice in eastern Himalayas. *Journal of Applied Phycology*, *30*, 547-558.
54. Li, J., Van Gerrewey, T., & Geelen, D. (2022). A meta-analysis of biostimulant yield effectiveness in field trials. *Frontiers in Plant Science*, *13*, 836702.
55. Little, H., & Neily, W. (2010). Effect of commercial extracts of *Ascophyllum nodosum* on plant water relations.
56. Luziatelli, F., Ficca, A. G., Colla, G., Baldassarre Švecová, E., & Ruzzi, M. (2019). Foliar application of vegetal-derived bioactive compounds stimulates the growth of beneficial bacteria and enhances microbiome biodiversity in lettuce. *Frontiers in Plant Science*, *10*, 60.
57. Manono, B. (2016). Agro-ecological role of earthworms (Oligochaetes) in sustainable agriculture and nutrient use efficiency: a review. *Journal of Agriculture and Ecology Research International*, *8*(1), 1-18.
58. Man-Hong, Y., Lei, Z., Sheng-Tao, X., McLaughlin, N. B., & Jing-Hui, L. (2020). Effect of water soluble humic acid applied to potato foliage on plant growth, photosynthesis characteristics and fresh tuber yield under different water deficits. *Scientific Reports*, *10*(1), 7854.
59. Mesa, T., Polo, J., Casadesús, A., Gómez, Í., & Munné-Bosch, S. (2022). Application of a biostimulant (Pepton) based in enzymatic hydrolyzed animal protein combined with low nitrogen priming boosts fruit production without negatively affecting quality in greenhouse-grown tomatoes. *Frontiers in Plant Science*, *13*, 828267.
60. Murase, K., Mészáros, P., & Zhang, B. (2009). Probing the birth of fast rotating magnetars through high-energy neutrinos. *Physical Review D—Particles, Fields, Gravitation, and Cosmology*, *79*(10), 103001.
61. Nuti, M., & Giovannetti, G. (2015). Borderline products between bio-fertilizers/bio-effectors and plant protectants: the role of microbial consortia. *J. Agric. Sci. Technol. A*, *5*, 305-315.

62. Ohana-Levi, N., Munitz, S., Ben-Gal, A., & Netzer, Y. (2020). Evaluation of within-season grapevine evapotranspiration patterns and drivers using generalized additive models. *Agricultural Water Management*, 228, 105808.
63. Papantzikos, V., Papanikou, A., Stournaras, V., Mpeza, P., Mantzoukas, S., & Patakioutas, G. (2024). Biostimulant Effect of Commercial Rhizobacteria Formulation on the Growth of *Vitis vinifera* L.: Case of Optimal and Water Deficit Conditions. *Applied Biosciences*, 3(1), 151-164.
64. Patel, K., Agarwal, P., & Agarwal, P. K. (2018). *Kappaphycus alvarezii* sap mitigates abiotic-induced stress in *Triticum durum* by modulating metabolic coordination and improves growth and yield. *Journal of Applied Phycology*, 30, 2659-2673.
65. Qin, K., & Leskovar, D. I. (2018). Lignite-derived humic substances modulate pepper and soil-biota growth under water deficit stress. *Journal of Plant Nutrition and Soil Science*, 181(5), 655-663.
66. Rasul, F., Gupta, S., Olas, J. J., Gechev, T., Sujeeth, N., & Mueller-Roeber, B. (2021). Priming with a seaweed extract strongly improves drought tolerance in *Arabidopsis*. *International journal of molecular sciences*, 22(3), 1469.
67. Raza, Q. U. A., Bashir, M. A., Rehim, A., Ejaz, R., Raza, H. M. A., Shahzad, U., ... & Geng, Y. (2022). Biostimulants induce positive changes in the radish morphology and yield. *Frontiers in Plant Science*, 13, 950393.
68. Rengasamy, K. R., Kulkarni, M. G., Papenfus, H. B., & Van Staden, J. (2016). Quantification of plant growth biostimulants, phloroglucinol and eckol, in four commercial seaweed liquid fertilizers and some by-products. *Algal research*, 20, 57-60.
69. Rosa, S. D., Silva, C. A., & Maluf, H. J. G. M. (2020). Phosphorus availability and soybean growth in contrasting Oxisols in response to humic acid concentrations combined with phosphate sources. *Archives of Agronomy and Soil Science*, 66(2), 220-235.
70. Roupshael, Y., Kyriacou, M. C., & Colla, G. (2018). Vegetable grafting: A toolbox for securing yield stability under multiple stress conditions. *Frontiers in Plant Science*, 8, 2255.
71. Shukla, P. S., & Prithiviraj, B. (2021). *Ascophyllum nodosum* biostimulant improves the growth of *Zea mays* grown under phosphorus impoverished conditions. *Frontiers in plant science*, 11, 601843.

72. Shukla, P. S., Nivetha, N., Nori, S. S., Kumar, S., Critchley, A. T., & Suryanarayan, S. (2024). A biostimulant prepared from red seaweed *Kappaphycus alvarezii* induces flowering and improves the growth of *Pisum sativum* grown under optimum and nitrogen-limited conditions. *Frontiers in Plant Science*, *14*, 1265432.
73. Spinelli, F., Fiori, G., Noferini, M., Sprocatti, M., & Costa, G. (2009). Perspectives on the use of a seaweed extract to moderate the negative effects of alternate bearing in apple trees. *The Journal of Horticultural Science and Biotechnology*, *84*(6), 131-137.
74. Spinelli, F., Fiori, G., Noferini, M., Sprocatti, M., & Costa, G. (2009). Perspectives on the use of a seaweed extract to moderate the negative effects of alternate bearing in apple trees. *The Journal of Horticultural Science and Biotechnology*, *84*(6), 131-137.
75. Stirk, W. A., & van Staden, J. (2020). Potential of phytohormones as a strategy to improve microalgae productivity for biotechnological applications. *Biotechnology Advances*, *44*, 107612.
76. Swaminathan, M. S., & Kesavan, P. C. (2012). Agricultural research in an era of climate change. *Agricultural Research*, *1*, 3-11.
77. Taspinar, M. S., Aydin, M., Sigmaz, B., Yildirim, N., & Agar, G. (2017). Protective role of humic acids against picloram-induced genomic instability and DNA methylation in *Phaseolus vulgaris*. *Environmental Science and Pollution Research*, *24*, 22948-22953.
78. Trivedi, K., Anand, K. V., Vaghela, P., & Ghosh, A. (2018). Differential growth, yield and biochemical responses of maize to the exogenous application of *Kappaphycus alvarezii* seaweed extract, at grain-filling stage under normal and drought conditions. *Algal research*, *35*, 236-244.
79. Trivedi, K., Anand, K. V., Vaghela, P., Critchley, A. T., Shukla, P. S., & Ghosh, A. (2023). A review of the current status of *Kappaphycus alvarezii*-based biostimulants in sustainable agriculture. *Journal of Applied Phycology*, *35*(6), 3087-3111.
80. Turan, M., & Köse, C. (2004). Seaweed extracts improve copper uptake of grapevine. *Acta Agriculturae Scandinavica, Section B-Soil & Plant Science*, *54*(4), 213-220.
81. Vandenkoornhuyse, P., Quaiser, A., Duhamel, M., Le Van, A., & Dufresne, A. (2015). The importance of the microbiome of the plant holobiont. *New Phytologist*, *206*(4), 1196-1206.
82. Xu, J., Mohamed, E., Li, Q., Lu, T., Yu, H., & Jiang, W. (2021). Effect of humic acid addition on buffering capacity and nutrient storage capacity of soilless substrates. *Frontiers in Plant Science*, *12*, 644229.

83. Zheng, X., Yu, T., Chen, R., Huang, B., & Wu, V. C. H. (2007). Inhibiting *Penicillium expansum* infection on pear fruit by *Cryptococcus laurentii* and cytokinin. *Postharvest biology and technology*, 45(2), 221-227.
84. Zhu, J. K. (2000). Genetic analysis of plant salt tolerance using *Arabidopsis*. *Plant physiology*, 124(3), 941-948.
85. Zodape, S. T., Mukhopadhyay, S., Eswaran, K., Reddy, M. P., & Chikara, J. (2010). Enhanced yield and nutritional quality in green gram (*Phaseolus radiata* L) treated with seaweed (*Kappaphycus alvarezii*) extract.

UNDER PEER REVIEW

UNDER PEER REVIEW