

Current Trends in In Vitro Maturation of Oocytes: A Review

ABSTRACT

The term “in vitro maturation” (IVM) describes the various stages of immature oocyte maturation in culture. The process of in vitro maturation (IVM) involves the collection of immature oocytes from antral follicles using an altered version of in vitro fertilization (IVF), wherein the last steps of meiosis are accomplished through in vitro culture. IVM primarily benefits high-risk patients, such as those with polycystic ovaries (PCO) and polycystic ovarian syndrome (PCOS), by lowering gonadotrophin stimulation in the patient. IVM provides further advantages for maintaining fertility, especially for individuals with oncofertility. Recent advancements in IVM research have led to markedly higher success rates and safety evidence in terms of newborn and developmental outcomes. Pre-maturation techniques and the creation of novel culture media additives have shown promise in recent times to maximize oocyte developmental stages and maturation proficiency. An essential component of later embryonic development, pregnancy, and healthy live births is the source of the immature oocytes. IVM is a successful treatment that has produced thousands of healthy births and considerable results in terms of acceptable pregnancy and implantation rates. These treatments may be beneficial for many kinds of infertile women. IVM treatment in conjunction with mild-stimulation IVF could be a good substitute for the conventional course of care. It's time to reevaluate the IVM technology and its advancement, even though IVM treatment is currently regarded as experimental. IVM and mild-stimulation IVF could end up being first-line therapy options rather than just alternatives to traditional therapies.

Keywords: Oocytes, Antral follicles, pre-maturation techniques, Implantation, In-Vitro Maturation, In-Vitro fertilization, Gonadotrophin.

INTRODUCTION

One of the most contentious parts of assisted reproductive technology is in vitro maturation, or IVM. Despite a great deal of research, it is still considered an alternative treatment and is not a part of the standard medical regimen. Nonetheless, research has demonstrated that IVM is useful for managing Ovarian Hyperstimulation Syndrome and can be applied in nearly all contexts where in vitro fertilization (IVF) is employed[1]. Before undergoing standard in-vitro fertilization or micro-injection, immature oocytes are extracted from tiny antral follicles and allowed to grow in a lab setting. This process is known as In-vitro maturation (IVM). Women with polycystic ovaries and women with normal ovaries are the two main patient categories that IVM treats. Due to their heightened sensitivity to gonadotropins, patients with polycystic ovarian

syndrome have irregular, primarily ovulatory cycles and are more likely to develop ovarian hyperstimulation syndrome. IVM has also been made available to couples with tubal, male factor, and unexplained infertility, as women with normal ovarian function may prefer to avoid the negative effects of hormone injections. Immature oocytes have been successfully developed, fertilized, and transferred into embryos in each of these patient categories. There have been reports of pregnancy rates ranging from 4% to 54%. After more than 300 births, follow-up research has revealed no significant issues with the pregnancies, delivery, or health of the children. Regarding IVM, numerous questions remain. There are currently no established markers for the ideal time to collect immature oocytes because the variables influencing follicle selection are not well understood. Moreover, the development of ideal culture conditions for maturation is challenging due to a lack of fundamental understanding of the intricate intracellular mechanisms involved in the cytoplasmic maturation of human oocytes [2].

Empirical review

Weon-Young Son, Seok-Yoon Lee, Jin-Ho Lim et al. In a study named 'Fertilization, cleavage and blastocyst development according to the maturation timing of oocytes in vitro maturation cycles' concluded that The oocyte maturation period may be a significant factor in the advancements in oocyte development technology, as it predicts the cleavage and blastocyst development of the oocytes recovered in HCG-stimulated IVM cycles[3].

In a research paper titled 'The improvement of in vitro maturation systems for bovine and porcine oocytes' Nagai T. et al found that Recently, zygotes obtained from transgenic mice overexpressing GSH synthetase were used to demonstrate the relevance of GSH for the development of zygotes in culture. In poor culture circumstances, the transgenic mice's zygotes demonstrated greater capacity Compared to the zygotes of control females who were not transgenic. Our existing maturation culture systems still seem to be subpar, and more research is needed to design a universal maturation culture system. This is because raising the GSH content in IVM oocytes increases their invitro growth following IVF. However, a major barrier to the effective development of the maturation system has been the difficulty in seeing the maturational and/or developmental capability of IVM oocytes. In a review paper, Hunter proposed that more investigation is required to identify "markers" of oocyte the ability to anticipate developmental competency. According to their birefringence, spindles in human and mouse oocytes can now be non-destructively seen using the LC polscope. Because the spindle in oocytes is very birefringent and its integrity is important for good chromosome segregation after the conclusion of meiosis, selecting structurally normal oocytes for IVF, ICSI, and nuclear transfer will be possible without harming the cells. It will enable us to forecast specific elements of the maturational and developmental competence of oocytes when applied to bovine and porcine oocytes[4].

In a study titled 'In vitro maturation and the fertilization and developmental competence of oocytes recovered from untreated polycystic ovarian patients' Alan Trounson, Carl Wood, Annette Kausche et al concluded that Because the oocytes of patients with PCO preserve their

maturational and developmental ability, immature oocyte recovery may be developed as a novel therapy option for these women [5].

Alan Penzias, M.D., et al according to a study titled 'In vitro maturation: a committee opinion' concluded that Candidates for IVM could be women who have PCOS or ovaries that resemble PCO, or who are at risk for OHSS. It is currently unclear if IVM is effective in the case of estrogen-sensitive malignancies or in women who have a short window of time to start fertility preservation before receiving possibly gonadotoxic cancer treatments. For these women, IVM offers an alternative treatment plan that has a lower patient burden because it involves fewer injections and shorter stimulation cycles, which also results in lower drug and monitoring expenses. IVM should only be performed by professionals who have received specialized training in this field. It should also always be provided with proper counseling regarding expected outcomes and informed consent. It is no longer deemed experimental to use this technology. Patients with a high AFC are the only ones who are appropriate candidates for IVM. Long-term follow-up studies of neonatal health and developmental outcomes of offspring are required, as well as large trials comparing clinical outcomes of promising newer methods of IVM versus standard IVF. At this time, however, patients should be informed that blastocyst conversion is lower and that implantation and pregnancy rates may be reduced compared with conventional IVF [6].

In a study titled 'In vitro maturation, fertilization and embryo development after ultrarapid freezing of immature human oocytes' J Wu, L Zhang, X Wang et al concluded that One potential solution to avoid iatrogenic sterility during chemotherapy or radiation therapy is cryopreservation. Furthermore, this method would enable prospective pregnancy in women with reproductive system disorders such as endometriosis, pelvic infections, cysts, and premature ovarian failure that impair ovarian function. For these patients, the use of frozen oocytes would enable the preservation of fertility [7].

Nevertheless, specific training for clinicians and embryologists is still required before IVM may be used in clinical practice. IVM is a potentially useful technique in the long future, especially. According to the study by Huixia Yang, Thomas Kolben et al titled ' Factors influencing the in vitro maturation (IVM) of human oocyte' IVM is a potentially useful technology that offers a more physiological and safe substitute for hormone stimulation, particularly when it comes to oncofertility [8].

Melanie L. Walls a b, Roger J. Hart et al according to the study titled 'In vitro maturation' found that Even though IVM success rates are still lower than those of standard IVF, there have been notable advancements in recent years, including an increase in implantation and live birth rates and a notable decline in miscarriages and early pregnancy losses as a result of the implementation of freeze-all protocols. Furthermore, with the removal of OHSS in high-risk patients, maternal outcomes seem to be better with IVM compared to normal IVF, and early positive data.[9].

Zhi-Yong Yang M.D. a, Ri-Cheng Chian Ph.D. et al according to the study titled 'Development of in vitro maturation techniques for clinical applications' concluded that The source of immature oocytes is crucial for later embryonic development and pregnancy, however, the existence of a dominant follicle does not appear to hurt the developmental competence of oocytes produced from the tiny antral follicles. Premature oocyte retrieval can be primed with FSH or HCG to increase oocyte maturation and pregnancy rates. As a result, the application of IVM technology can be expanded to treat infertile women from various backgrounds, with acceptable pregnancy and live birth rates. Several thousand healthy kids have been born thanks to IVM, although the technology is still regarded as experimental. It might be time to reevaluate IVM technology as a useful clinical intervention. As IVM treatment advances, combining natural-cycle IVF with the harvest of immature oocytes and then IVM of those immature oocytes presents an alluring option for raising the successful outcome rate. More infertile women might be able to benefit from these treatments if the procedures involved in them can be made simpler, particularly for the retrieval of immature oocytes. A good substitute for traditional treatment could be mild-stimulation in vitro fertilization (IVF) in conjunction with IVM therapy. Mild-stimulation IVF in conjunction with IVM may show to be not simply an alternative to conventional therapies but possibly a first-line option as we gather more experience and outcome data [10].

Anne Lis Mikkelsen et al according to the study 'Strategies in human in-vitro maturation and their clinical outcome' found that The IVM protocol requires less time to complete and is comparatively straightforward. Moreover, expenses are decreased and the negative consequences of stimulation—specifically, OHSS—are removed. In contrast to traditional IVF In the future, immature oocyte retrieval in conjunction with IVM may take the place of standard stimulated inoculation in certain patients. These patients may include women with PCOS and regular cycling women referred for IVF/ICSI because of a man's reduced sperm quality. In the future, it might be feasible to combine IVM with in-vitro follicle culture. The full in vitro development of follicular oocytes is being approached. Only in mice has complete in vitro follicle growth from primordial follicle up to Graafian stage been accomplished thus far. Currently, techniques are being developed for the long-term in-vitro follicle growth of human primordial follicles; however, no viable culture method has been reported to far. Even after freeze-storage, human primordial follicles have been shown to grow into secondary follicles [11].

In the therapeutic domain of ART, IVM technology is developing with increasing success rates. While IVM may not be suitable for all types of infertile patients, for those who are gonadotropin-sensitive, it is unquestionably a safer option than IVF. According to a study by Kyung Sil Lim, Soo Jin Chae, Chang Woo Choo, Yeon Hee Ku, Hye Jun Lee, Chang Young Hur et al titled ' In vitro maturation: Clinical applications' Additionally, for a certain subset of patients, IVM in a natural cycle can be utilized as a viable option with a respectable pregnancy rate. IVF might be the best option for infertile couples as well as for getting oocytes to maintain fertility. For cancer patients, IVM is a good substitute for preserving fertility. In patients with hormone-sensitive malignancies, IVM can be used safely and without concern for high levels of estradiol when it

comes to preserving fertility. When cancer therapy should begin as soon as possible, IVM may be the best option because there is no waiting period for COS. Regarding long-term development, prenatal and obstetric outcomes, and IVM newborns, there are numerous issues. Although it is currently unable to fully evaluate the clinical safety of IVM, a number of studies have found no evidence of a higher risk of unfavourable outcomes for infants conceived following IVM in comparison to those conceived in vivo [12].

In the study titled 'In vitro maturation of human oocytes for assisted reproduction' Marcus W Jurema, Daniela Nogueira et al concluded that An Increasingly popular technique in assisted reproductive technologies is the in vitro maturation of human oocytes extracted from antral ovarian follicles. Compared to conventional controlled ovarian hyperstimulation for in vitro fertilization, this novel approach may have a number of benefits, including lower costs due to less gonadotropin and GnRH analogue use, a reduction in the risk of ovarian hyperstimulation syndrome, and a simpler protocol. While in vitro oocyte maturation for human assisted reproduction is still in its infancy, some recent studies have shown safety and efficacy results that are on par with those of regular IVF. It is possible to include in vitro maturation into an established IVF program with just a few minor alterations. A greater understanding of how to improve endometrial receptivity and immature oocyte developmental competence is critical to the technology's advancement and optimization [13].

In a study titled 'the variable success of in vitro Maturation: can we do better?' Alberto M Luciano, Federica Franciosi et al found that by using a physiological method, IVP technologies aim to increase oocyte quality and produce more oocytes with higher developmental capability. Even if they are capable of restarting meiosis on their own, oocytes derived from non-ovulatory follicles are still far from being totally competent. The goal of the majority of the research, regardless of the meiotic arrest technique employed, was to replicate in vitro the latter steps of prematuration that ordinarily take place in vivo. Prematuration has been shown to increase an oocyte's capacity for development in a number of investigations. However, regardless of follicular origin, there hasn't been much progress seen in embryonic developmental competence when oocytes are cultivated in bulk. The results of maturation systems are significantly impacted by the variety of the oocyte population at the beginning of the process, and it is important to consider the unique metabolic requirements of the oocyte during isolation. The selection of high-quality oocytes, the timing of prematuration (length of culture), and the particular environment (hormones, growth factors, molecules, etc.) for optimizing prematuration culture systems can all be aided by the identification of particular non-invasive biomarker(s) of oocyte health status and final differentiation. Simultaneously, the establishment of a customized culture system may be linked to techniques for stimulating the growth of ovarian follicles in the donor to produce oocytes that are especially appropriate for prematuration treatments that are specifically designed for them [14].

In 'In vitro maturation rates in young premenarche patients' Gilad Karavani, Natali Schachter-Safrai et al concluded that the usefulness of the current IVM approach in this age group is called

into doubt because IVM done following ovarian tissue cryopreservation in premenarche girls, and notably in very young girls (4 years and younger), provides dramatically lower maturation rates compared with postmenarche patients. To evaluate the adaptation of the IVM approach for young girls, more research is needed [15].

Oocyte in vitro maturation (IVM) lessens the requirement for ovarian hyperstimulation brought on by gonadotrophin and the health hazards that go along with it, but its clinical adoption has been hampered by the unacceptable low rates of conception and pregnancy. We present the creation of a brand-new in vitro system that simulates physiological oocyte maturation (SPOM). According to the study by F.K. Albus, M. Sasseville, et al titled 'Simulated physiological oocyte maturation (SPOM): a novel in vitro maturation system that substantially improves embryo yield and pregnancy outcomes' With its novel approach to IVM, SPOM significantly enhances oocyte developmental outcomes while simulating certain aspects of oocyte maturation in vivo. When SPOM is modified for therapeutic use, it ought to have a big impact on managing infertility and benefit patients greatly [16].

The term "In vitro maturation" (IVM) describes the process of immature oocytes maturing in culture, whether or not they have been subjected to brief gonadotropin doses. According to the study by Sauerbrun-Cutler, May-Tal et al titled 'In Vitro Maturation and Its Role in Clinical Assisted Reproductive Technology' Since the 1970s, around 5000 live births have been attributed to IVM. IVM is now only used for individuals who have been carefully chosen and are at risk for ovarian hyperstimulation syndrome or who are contraindicated for hormone delivery. IVM is a safer and less complicated option to traditional IVF, while having a lower pregnancy rate. Prior to widespread adoption, further research should concentrate on increasing implantation and live birth rates [17].

In a research paper titled 'Gene expression profiling of human oocytes following in vivo or in vitro maturation' Gayle M Jones, David S Cram, Bi Song et al found that Developmental competence discrepancies between in vivo and in vitro developed oocytes have a biological basis, which has been elucidated by global gene expression profiling utilizing microarrays and bioinformatics analysis. The overabundance of transcripts found in oocytes retrieved from gonadotrophin-stimulated cycles and maturing in vitro at the immature germinal vesicle stage is most likely the result of dysregulation in either post-transcriptional modification or gene transcription. For any embryos generated from these oocytes, either approach would lead to an erroneous temporal usage of genes, potentially resulting in developmental incompetence [18].

Johan EJ Smitz, Jeremy G Thompson, Robert B Gilchrist et al in a study titled 'The promise of in vitro maturation in assisted reproduction and fertility preservation' concluded that To make this patient-friendly procedure a regular part of infertility treatment and fertility maintenance, a novel technique to in vitro maturation (IVM) is needed. Compared to traditional in vitro fertilization/intracytoplasmic sperm injection, current methods of IVM never record an implantation rate of more than 10 to 15% per embryo transplanted, which is two to three times

lower and results in greater rates of early pregnancy losses. The utilization of pharmacological substances that enable the synchronization of nuclear and cytoplasmic maturation processes within the oocyte is the fundamental component of this cutting-edge culture approach. To encourage a longer contact between the immature oocyte and appropriately conditioned cumulus cells, a longer oocyte maturation period is justified. If a new method of IVM is successfully introduced, it will require less fertility medications and require less intrusion into the patient's everyday life in terms of intravaginal ultrasonography and blood hormone level monitoring. The new IVM conditions will, in the end, lower the overall cost of treatment while also reducing a wide range of minor and significant issues in assisted reproductive technology. Cancer patients who wish to preserve gonadal tissue before receiving medication that severely impairs eventual germ-cell competence can benefit from this procedure's limited invasiveness [19].

In a study titled 'Clinical definition paper on in vitro maturation of human oocytes' Michael H Dahan, Seang Lin Tan et al found that Human oocyte in vitro maturation (IVM) is a 25-year-old reproductive procedure that is becoming more and more well-liked. However, due in part to some of these variations in protocols, the IVM techniques utilized in different clinics vary greatly, leading to highly varying pregnancy rates. These variations include the use of gonadotrophin therapy for a few days to maintain modest follicle growth and the use of HCG triggering in certain cycles before oocyte retrieval. The number of embryos transferred, the cleavage-stage embryo or blastocyst transfer, and the patient selection (including those with polycystic ovaries or low ovarian reserve) are additional significant determinants. Regarding IVM and its implications, clinicians also disagree significantly in their perspectives. It was decided to create this study in an effort to bring uniformity when comparing treatments and IVM outcomes because of the wide diversity in regimens. The harvest of oocytes from small and intermediate-sized follicles in an ovary before the largest follicle has exceeded 13 mm in mean diameter is the clinical definition of in vitro fertilization (IVM) that was defined. It is important to highlight the usage of brief gonadotrophin stimulation. However, it should be stated that metaphase II oocytes also have the potential to be collected at that time in the cycles associated with either HCG or GnRH agonist priming. Many feel this is not IVM because some mature oocytes are retrieved, therefore, we recommend renaming this procedure either natural cycle IVF or modified natural cycle IVF (if gonadotrophin stimulation is given) with early triggering, combined with IVM. The percentage as well as the absolute number of mature oocytes at retrieval should be indicated. The use of these titles will allow transparency when comparing results of IVM cycles [20].

In 'Prospects for Oocyte Banking and In Vitro Maturation' Roger G Gosden et al concluded that In the middle of the menstrual cycle, before the dominant follicle appears, oocytes are collected for in vitro maturation (IVM). The follicles typically have a diameter of 8–10 mm; smaller follicles, measuring 3–7 mm, still contain oocytes that can mature in vitro, but their chances of success are significantly lower, most likely due to cytoplasmic immaturity. Success with IVM—in vitro fertilization—is predicted by the quantity of oocytes recovered (IVF or ICSI). Ovarian

polycystic patients typically harvest over 10 oocytes from unstimulated ovaries; the quantity of oocytes harvested is a predictor of clinical pregnancy rates following in vitro fertilization. Aspirating immature oocytes at a lower pressure than usual and without flushing prevents the follicles from collapsing too quickly, which speeds up the recovery of the eggs. Nevertheless, because the cumulus cell mass around the oocyte is unmucified absent gonadotropin stimulation, the efficiency is lower than that of periovulatory follicles. In order to maintain the physiological connections between the oocyte and its somatic cell envelope, the cumulus–oocyte complex is often cultivated intact (28). The best times for equilibration for the oocyte and its relatively small cumulus cells are probably going to differ, which makes cryopreservation of the complete cumulus–oocyte complex challenging. Additionally, immersion in hypertonic CPAs results in cell shrinking, which may impair granulosa cell functions and the oocyte’s gap junction communication—which is thought to be essential for the transmission of regulatory and nutritional molecules... It is unlikely that results can be improved by vitrification, which calls for the cumulus-oocyte complex to equilibrate at even greater CPA concentrations. Currently, the success rate of IVM in young women with polycystic ovaries is reported to be between 25% and 30% per cycle; the rate of miscarriages is high (29). These rates, which are likely to be further impaired if the oocytes are cryopreserved prior to IVM, are only half of those attained at the finest clinical centers with IVF using fresh oocytes [21].

In a research paper titled ‘Oocyte in vitro maturation: physiological basis and application to clinical practice’ Robert B Gilchrist, Johan Smitz et al found that For a significant number of infertile couples globally who currently lack access to therapy due to socioeconomic constraints, the simplicity of infertility treatment provided by IVM holds promise. The shorter treatment duration of IVM, its reduced hormone intake requirement, its independence from the OPU cycle stage, its lack of complications and side effects, and its lower cost compared to traditional IVF suggest that the barrier to ART treatment should decrease significantly, allowing infertile couples to choose to start a family earlier. In contrast to IVF, IVM requires an extra one to two days of culture time in the lab and a separate oocyte search technique, but the latter requires significantly less technical knowledge to master than, say, ICSI or embryo biopsy. The knowledge that IVM requires little or no hormone injections may also significantly lower the barrier to gamete banking, or social freezing, among young women. Despite being triggered by a GnRH agonist, ovarian superovulation usually has some negative effects on most patients. This is because the goal is to optimize oocyte recovery per retrieval, which frequently results in significant ovarian swelling and discomfort in the days leading up to and following the procedure. The IVM cycles mitigate these negative impacts. According to a recent study, half of all women who are at risk of hyper-response base their choice of therapy (IVM vs. IVF) on the procedure that carries the least amount of side effects. This highlights the fact that treatment success, or a live birth, is not the only crucial factor in these women’s reproductive care [22].

Robert B. Gilchrist et al According to the study titled ‘Recent insights into oocyte-follicle cell interactions Provide opportunities for the development of new Approaches to in vitro maturation’

Recent years have seen significant advancements in understanding long-standing disputes on the specific cellular and molecular mechanisms governing Maturation of eggs in vivo. Novel signaling pathways that were mainly unidentified a decade ago, specifically the oocyte signaling pathway utilizing GDF9/BMP15–SMAD2/3 and the EGF-like ERK1/2 cascade peptide, have now been thoroughly characterized. This new information must now be used by oocyte IVM experts and doctors in novel IVM systems for genetic improvement in domestic animals and clinical Treatments for infertility that aim to increase oocyte developmental competence and generate a large number of healthy embryos and progeny [23].

In a study titled ‘ In vitro maturation of human immature oocytes for fertility preservation’ Ri Cheng Chian, Peter S Uzelac, Geeta Nargund et al concluded that The primary means of preserving female fertility is by the cryopreservation of ovarian tissues, oocytes, or embryos. The significant rise in the frequency of babies delivered from vitrified oocytes suggests that oocyte cryopreservation is becoming one of the most significant intervention choices. For some cancer patients, however, oocyte cryopreservation combined with conventionally regulated ovarian hyperstimulation may not be an option due to grave concerns over the potential impact of hormone-induced ovarian stimulation on the likelihood of cancer recurrence. Furthermore, a patient receiving immediate gonadotoxic cancer treatment might not have enough time to undergo hormonal ovarian stimulation. Therefore, for women who are unable to undergo ovarian stimulation or who are unable to postpone their gonadotoxic cancer therapy, the extraction of immature oocytes from ovaries without ovarian stimulation, followed by in vitro maturation and vitrification, represents a promising option for preserving fertility. It is possible to harvest immature oocytes from the ovaries both in the luteal and follicular phases, which increases the likelihood of preserving fertility. Another promising method of preserving fertility in young women with cancer is the combination of cryopreserved ovarian tissue and the collection of immature oocytes from the tissue, followed by oocyte vitrification through in vitro maturation [24].

In ‘ The Place of In Vitro Maturation in Assisted Reproductive Technology’ Lan N Vuong, Tuong M Ho, Robert B Gilchrist, Johan Smits et al found that By collecting and maturing immature cumulus-oocyte complexes in vitro, without the requirement for regulated ovarian stimulation and ovulation triggering, in vitro maturation (IVM) is an assisted reproductive technology (ART). IVM has several advantages over in vitro fertilization (IVF), such as minimal or no stimulation, cheaper prescription costs, and less strain on the patient. Nevertheless, IVM’s early clinical results weren’t the best. Clinical trials from more recent times have shown that roughly 40% of live births occur after IVM. The effectiveness of IVM is being increased by utilizing new IVM culture systems. These have been used extensively for many years on animals, and they are now demonstrating potential in the clinical context. Patients who are at risk of OHSS (such as women with polycystic ovary syndrome), have limited time for ovarian stimulation, or when prolonged increases of estradiol are contraindicated (such as oncofertility causes) are more likely to benefit from IVM versus IVF. IVM’s relative effectiveness in comparison to IVF has been the

primary deterrent to its use to date, and worries about the health of the babies born after IVM have also been raised. Nevertheless, no distinctions have been found between congenital defects caused by IVM and other ARTs. Furthermore, both standard operating procedures and experience are deficient... Improved clinician training, increased and better-funded research in the field, and enhanced IVM recognition by reproductive specialists are some strategies to surmount obstacles to its utilization. All things considered, IVM provides a worthwhile substitute for ART in some patient groups. Pregnancy outcomes that are comparable to those following IVF seem to be achievable with new methods of IVM. The ultimate goal of improving fertility outcomes is to raise financing for IVM research and to improve the training and education of fertility specialists to increase the usage of IVM in the future [25].

Xueqi Gong, Hemei Li, Yiqing Zhao et al in a study titled 'The improvement and clinical application of human oocyte in vitro maturation (IVM)' IVF was not invented; oocyte in vitro maturation (IVM) is a long-standing method. IVM has been examined extensively, but for nearly thirty years, its efficiency has been low. Regarding the advantages of IVM, numerous noteworthy advancements in recent years have increased IVM's efficiency and usage. The most significant development in recent years is the establishment of biphasic IVM. Pre-IVM culturing phase and IVM phase are included in biphasic IVM. It has been demonstrated that the CNP-mediated pre-IVM culture technique is effective for non- or minimally stimulated immature oocytes. This is the biggest advancement in this field in recent decades. Pre-IVM culturing phase and IVM phase are included in biphasic IVM. It has been demonstrated that the CNP-mediated pre-IVM culture technique is effective for non- or minimally stimulated immature oocytes. This is the biggest advancement in this field in recent decades. IVM can be utilized in the clinic to prevent ovarian hyperstimulation syndrome (OHSS) in PCOS patients. Furthermore, this technique can help some people with unique disorders (resistant ovarian syndrome) whose reproductive issues cannot be resolved by IVF. Furthermore, this technique can help some people with unique disorders (resistant ovarian syndrome) whose reproductive issues cannot be resolved by IVF. Oocytes in tiny antral follicles are lost during the majority of fertility preservation operations. IVM can, however, seize this type of oocyte and preserve its reproductive potential. IVM can be seamlessly integrated with fertility preservation techniques used in clinics to increase the effectiveness of fertility preservation. Shortly, IVM, a practical and appealing technology, might find widespread application around the globe [26].

Some fertility institutions provide the safe and efficient treatment of in vitro maturation, which involves harvesting immature oocytes from unstimulated ovaries for assisted reproduction. According to the study by G Durga Rao, Seang Lin Tan et al titled 'In vitro maturation of oocytes' concluded that As a result, the process avoids hazards including ovarian hyperstimulation syndrome, adverse drug reactions, and ovarian stimulation using pricey gonadotropins. Additional benefits include a shorter treatment plan and fewer monitoring scans than with in vitro fertilization. Women with polycystic ovaries and multiple antral follicles were the initial candidates under consideration; however, the indications are expanding to encompass

women who typically have low-quality embryos in repeated cycles and who do not respond well to stimulation. At McGill Reproductive Center, we are currently effectively adopting two novel uses for in vitro maturation: oocyte donors and fertility preservation, particularly in cancer patients receiving gonadotoxic medication. When young women without partners require this procedure to preserve their fertility, it is coupled with oocyte vitrification. Up until the age of 35, women undergoing IVM for infertility therapy had a 38% clinical pregnancy rate each cycle, while recipients of IVM egg donation had a 50% clinical pregnancy rate per cycle [27].

In the researcher paper titled 'In vitro maturation of oocytes: uncommon indications' Michael Grynberg, Hady El Hachem et al found that Retrieval of immature oocytes from unstimulated ovaries, followed by in vitro maturation (IVM) was initially proposed to avoid the risks and side effects of exogenous gonadotropin administration. Therefore, during the past decades, IVM was mainly offered to patients with polycystic ovary syndrome (PCOS) at high risk of ovarian hyperstimulation syndrome (OHSS). However, the development of fertility preservation has recently opened new perspectives in the field of IVM. The present review summarizes uncommon indications of IVM, which is a viable option to treat infertility in patients with ovarian resistance to FSH, but may also be considered to preserve fertility in leukemia as well as before ovarian transposition and endometrioma excision [28].

In a study titled 'Ultrastructure of human oocytes after in vitro maturation' Rubens Fadini, Mario Mignini Renzini et al concluded that every oocyte exhibited homogeneous organelle distribution in its typical ooplasm. The appearance of mitochondrial morphology was consistent throughout maturation circumstances. In all oocytes, cortical granules were observed to be normally arranged in a single, largely continuous row directly beneath the ooplasm. After IVM, microvilli were in good condition. Not all oocytes included vacuoles, and when they were, they were usually connected to lysosomes. In vivo matured oocytes were frequently reported to include mitochondria-vesicles (MV) complexes and aggregates of mitochondria-smooth endoplasmic reticulum (M-SER). But in IVM oocytes, big MV complexes somewhat took the place of M-SER aggregates [29].

In 'Signaling mechanisms and their regulation during in vivo or in vitro maturation of mammalian oocytes' Patrycja Strączyńska, Krzysztof Papis et al found that Currently, one of the most successful ways to treat infertility is in vitro fertilization (IVF). Extracorporeal maturation of oocytes, also known as in vitro maturation, or IVM, can be an alternative to the widely utilized ovarian hyperstimulation. The oocyte's cytoplasmic nuclear, and genomic maturity are necessary for fertilization and healthy embryonic development. The granulosa, cumulus, and oocyte cells communicate bi-directionally through maternal signals and the ovarian follicle microenvironment, which affects the development, maturation, and acquisition of oocyte development capability. The high quantity of cAMP during oogenesis in mammals inhibits meiosis in the oocyte at prophase I of the meiotic division. The action of granulosa cell-produced C-type natriuretic peptide (CNP, NPPC) keeps this level at this level. In cumulus cells, the CNP binds to the NPR2 receptor and initiates the synthesis of cyclic guanosine monophosphate

(cGMP). By inhibiting phosphodiesterase 3A (PDE3A), which is responsible for low MPF activity, cGMP that enters the oocyte through gap junctions raises the level of MPF activity. The CNP/NPR2 complex is less active during the LH surge of the reproductive cycle, which lowers the amounts of cGMP in cumulus cells and, as a result, in the oocyte. Lower levels of cGMP unblock PDE3A's hydrolytic activity, which lowers the amount of cAMP within the oocyte. As a result, meiosis resumes and MPF is activated. Prematuration and the actual maturation process are the two processes of the newest IVM techniques, known as SPOM, NFSOM, or CAPA IVM. The percentage of mature oocytes in vitro and the percentage of correctly grown embryos in both animals and humans have significantly increased as a result of their consideration of the function of cAMP in first suppressing and then unblocking oocyte development [30].

CONCLUSION

In vitro maturation, or IVM, has become a hot issue in assisted reproduction because of its many clinical applications. Often used with disappointing outcomes, in vitro fertilization (IVF) superovulation cycles use oocytes from the germinal vesicle stage and germinal vesicle breakdown/metaphase I. There are significant differences between the biological aspects of the actual IVM technique and this so-called rescue in vitro oocyte maturation. Immature oocytes are extracted from tiny antral follicles of unprimed or minimally stimulated cycles in the latter case. This is done either as a simple substitute for traditional IVF in patients who are normo-ovulatory or to prevent ovarian hyperstimulation syndrome in high-risk patients. Mature oocytes in culture at various stages, with or without brief exposure to gonadotropins, are referred to as being in vitro matured (IVM). For healthy live births, pregnancy, and continued embryonic development, the source of the immature oocytes is crucial. IVM is a successful treatment that has already shown notable results, including acceptable rates of pregnancy and implantation and has given birth to several thousand healthy children. Combining a natural-cycle in vitro fertilization (IVF) therapy with immature-oocyte retrieval and then IVM of those immature oocytes is an appealing prospect for enhancing the already good outcome as IVM treatment development proceeds. Different categories of infertile women can benefit from these treatments if the procedures for immature-oocyte retrieval can be made simpler. IVM treatment in conjunction with mild-stimulation IVF could be a good substitute for the conventional course of care. It's time to reevaluate the IVM technology and its advancement, even though IVM treatment is currently regarded as experimental. IVM and mild-stimulation IVF could end up being first-line therapy options rather than just alternatives to traditional therapies.

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