

IMPACT OF ZINC AND SILICA SOLUBILIZING BACTERIAL CONSORTIA ON SOIL NUTRIENT AVAILABILITY AND DIRECT SOWN PADDY

ABSTRACT

The present study highlights about the availability of nutrients in soil and uptake by rice plants by inoculating selected zinc and silica solubilizing isolates and their combinations under pot culture and field conditions. Effect of zinc and silica solubilizing bacteria and their consortia on paddy was studied under field conditions at Agricultural Research Station, Janagamaheswarapuram, Andhra Pradesh. Thirteen treatments were assessed for availability of nutrients viz., Nitrogen, Phosphorus, Potassium, Zinc and Silica in soil and concentration of Nitrogen, Phosphorus and Potassium in plant at 45, 90 and 120 days after sowing (DAS). Significantly highest nitrogen (198.9, 262.3 and 240.2 kg ha⁻¹), available phosphorus (36.7, 64.7 and 40.6 kg ha⁻¹), potassium (221.4, 349.6 and 263.5 kg ha⁻¹), zinc (0.86, 1.14 and 0.98 ppm) and silica (66.8, 98.9 and 84.8 ppm) were recorded in T₁₃ (RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3) at 45, 90 and 120DAS, respectively. In plant, nitrogen (0.89, 1.10 and 0.98 %), phosphorus (0.46, 0.67 and 0.58 %) and potassium (1.87, 2.29 and 1.98 %) were significantly highest at 45, 90 and 120DAS, respectively in T₁₃. There was increase in the available nutrient content up to 90 DAS which then decreased at 120DAS. It is inferred that consortia of two zinc solubilizing and two silica solubilizing microorganisms (T₁₃) is useful for increased availability of Nitrogen, Phosphorus, Potassium, Zinc and Silica in soil and increased uptake of NPK by rice plant which in turn reduce exogenous chemical fertilizers. It is concluded that exogenous application of bacterial consortia can be exploited to improve the nutrient status and availability in direct sown paddy.

Key words: Paddy, zinc solubilizing bacteria, silica solubilizing bacteria, Zinc and silica bacterial consortia, nutrients.

1. INTRODUCTION

India is one of the leading producers of rice crop. Rice is the basic food crop and being a tropical plant, it flourishes comfortably in a hot and humid climate. Rice is mainly grown in rain-fed areas that receive heavy annual rainfall.[21-23] That is why it is fundamentally a *kharif* crop in India. Plants need several macro and micro nutrients for their growth and development. Nitrogen, phosphorus and potassium (NPK) are the primary nutrients in commercial fertilizers. Each of these fundamental nutrients plays a key role in plant nutrition. Nitrogen is considered to be the most important nutrient, and plants absorb more nitrogen than any other element. Nitrogen is essential in making plants healthy as they develop and nutritious to eat after they're harvested. Phosphorus, is linked to a plant's ability to use and store energy, including the process of photosynthesis. It's also needed to help plants grow and develop normally. Potassium is the third key nutrient of commercial fertilizers.

40 It helps strengthen plants' abilities to resist disease and plays an important role in increasing crop
41 yields and overall quality. In rice zinc (Zn) is one of the most important micronutrients necessary for
42 the normal healthy growth and reproduction of plants. Silica is useful for proper cuticle development
43 and grain formation in rice [1]. Zinc Solubilizing Bacteria (ZnSB) and Silica Solubilizing Bacteria (SiSB)
44 and their consortia improved the bioavailable fraction of N, P, K, Zn and Si to host plant for
45 enlightening the crop growth, yield and quality [2]. It will directly effect on the crop nutrient content and
46 yield parameters. Inoculation of rice with silica solubilizing bacteria enhanced available silica in soil
47 and silica content in plant and improved rice yield. Dissolution of silicate results in rendering
48 phosphorus available for plant absorption as silica competes with phosphorus fixation sites; silica acts
49 like auxiliary for phosphorus in plants [3]. Hence, an experiment was conducted to study the
50 availability of nutrients in soil and uptake by rice plants by inoculating selected zinc and silica
51 solubilizing isolates and their combinations under pot culture and field conditions.

52 **2. MATERIALS AND METHODS**

53 Paddy variety, MTU-7029 (Swarna) was sown in the field by adopting 20cm X 10 cm spacing at ARS,
54 Jangamaheswarapuram on 8-09-2019. Thirteen treatments, replicated thrice, were imposed
55 incompletely randomized design as detailed below.

56 **Treatment details:**

57 T₁: RDF (Control)

58 T₂: RDF + ZnSO₄

59 T₃: RDF + Calcium silicate

60 T₄: RDF + ZnSO₄ + Calcium silicate

61 T₅: RDF + ZnKJJ-4

62 T₆: RDF + ZnPGG-1

63 T₇: RDF + SiKPP-1

64 T₈: RDF + SiPYY-3

65 T₉: RDF + ZnKJJ-4 & ZnPGG-1

66 T₁₀: RDF + SiKPP-1 & SiPYY-3

67 T₁₁: RDF + ZnKJJ-4+ SiKPP-1

68 T₁₂: RDF + ZnPGG-1+ SiPYY-3

69 T₁₃: RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3

70 Where,

71 RDF = Recommended dose of fertilizer

72 ZnKJJ-4, ZnPGG-1, SiKPP-1 and SiPYY-3: Efficient zinc and silica solubilizing isolates.

73 **2.1 Estimation of nutrient (N, P, K, Zn and Si) content in soil samples**

74 **2.1.1 Soil sample collection and processing**

75 Soil samples collected at 45, 90 and 120 DAS, from all the 13 treatments were dried under shade,
76 gently ground with wooden hammer, sieved through 2 mm sieve and stored in labelled new polythene
77 lined cloth bags for analysis. Processed soil samples were used for analysing available nutrients by
78 adopting standard procedures.

79 **2.1.2 Available nutrients (nitrogen, phosphorus, potassium, zinc and silica) in soil**

80 Nitrogen was estimated by alkaline permanganate method as outlined by [4] and expressed in kg ha⁻¹.
81 Available phosphorus content of soil was determined by Olsen's method as described by [5] and
82 phosphorus in the extract was determined using spectrophotometer at 680 nm wavelength. It was
83 expressed in kg ha⁻¹. Available potassium in the soil was determined by ELICO flame photometer after
84 extracting the soil with neutral normal ammonium acetate as described by [6]. It was expressed in kg
85 ha⁻¹. The available zinc in the digested soil samples was determined by atomic absorption
86 spectrophotometer after making proper standards as described by [7]. It was expressed in ppm. The
87 available silica in the soil samples was determined by 0.5M acetic acid extractant method after making
88 proper standards as described by [8]. It was expressed in ppm.

89 **2.2 Estimation of nutrient (N, P and K) content in plant samples**

90 **2.2.1 Collection, preparation and analysis of plant samples**

91 The plant samples were collected at 45 DAS, 90 DAS and 120 DAS, washed thoroughly with distilled
92 water and dried under shade. Then, they were dried in hot air oven at 65°C till a constant weight was
93 obtained. Dried plant samples were ground in a wooden pestle and mortar and stored in polythene
94 bags for further chemical analysis. N, P, K, Zn and Si contents were estimated by following standard
95 methods.

96 **2.2.2 Digestion of plant sample**

97 Powdered whole plant samples were separately treated with concentrated HNO₃ overnight for pre
98 digestion. Then, the pre-digested samples were treated with diacid mixture [HNO₃:HClO₄ (9:4 ratio)]
99 and digested on sand bath at low temperature till colourless white precipitate was obtained. The
100 residue was dissolved in 6N HCl, filtered, made to known volume by using 6N HCl. This was used for
101 further nutrient analysis. The following analyses were carried out from the diacid digested samples of
102 whole plant.

103 **Nitrogen content in plant**

104 Nitrogen concentration in plant samples were estimated by Kelplus method [9] after digesting the
105 organic matter by H₂SO₄ and H₂O₂.

106 The nitrogen content in plant sample is calculated as follows:

107 Weight of sample = 0.1g

108 Normality of H₂SO₄ = 0.02

109 Titration value (TV) = Sample titration value – Blank titration value

110 % N in plant sample = $\frac{T.V \times 0.00028 \times 100}{0.1} = 0.28 \times T.V$

111 Concentration of macronutrients was expressed as % and micro nutrients in ppm

112 **Phosphorus content in plant**

113 The phosphorus content in the digested plant samples was determined by Vanadomolybdo-
114 phosphoric acid yellow colour method using spectrophotometer at 470 nm wave length [10].

115
$$\% P \text{ in plant sample} = \text{sample conc. in ppm} \times \frac{\text{Final volume (50 ml)} \times 100 \times 100}{\text{Wt of sample (1g)} \times \text{aliquot (5ml)} \times 10^6}$$

118 **Potassium content in plant**

119 The plant samples for K estimation were digested by diacid through wet digestion. The
120 samples are then read in flame photometer using filter for K [6].

$$\begin{aligned} 121 & \qquad \qquad \qquad 100 \times 100 \\ 122 & \quad \% \text{ K in plant sample} = R \times \frac{\qquad \qquad \qquad}{\qquad \qquad \qquad} \\ 123 & \qquad \qquad \qquad \text{Wt. of sample (1g)} \times 10^6 \\ 124 & \qquad \qquad \qquad = R \times 0.01 \end{aligned}$$

125 Where R = concentration of K in ppm obtained from standard curve

126 3. RESULTS AND DISCUSSION

127 3.1 Influence of zinc and silica solubilizing bacterial consortia on available nutrients in 128 the soil

129 3.1.1 Available nitrogen in soil

130 Available nitrogen content was 142.03 kg ha⁻¹ (Table 1) in the initial soil sample. At 45 DAS all
131 the treatments showed increased available nitrogen content compared to initial stage. Highest
132 available nitrogen was recorded in T₁₃, RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3 (198.9 kg
133 ha⁻¹) followed by T₁₂ (RDF + ZnPGG-1 + SiPYY-3) *i.e.*, 196.7 kgha⁻¹, which were on par. Available
134 nitrogen content further increased with highest recorded in T₁₃ (262.7 kg ha⁻¹) at 90DAS, followed by
135 T₁₂ (258.7 kg ha⁻¹) and both were on par. At 120 DAS available nitrogen content decreased to 240.2
136 kg ha⁻¹ in T₁₃ followed by T₁₂ (238.5 kg ha⁻¹) and T₁₁ (237.2 kg ha⁻¹) and were superior to other
137 treatments, (Table 2).

138 Similar findings were obtained by [11] where biofertilizers and the recommended dose of
139 fertilizers expanded the soil available nitrogen (63 %). The most effective treatment was ZSB + PSB +
140 KRB + RDF [12] reported increased available nitrogen with the applied fertilizers and biofertilizers.

141 3.1.2 Available phosphorus in soil

142 Phosphorus helps a plant convert other nutrients into usable building blocks to grow. Available
143 phosphorus content was initially 26.28 kg ha⁻¹ (Table 1). At 45 DAS all the treatments exhibited
144 increased available content of phosphorus than initial. The highest phosphorus availability was
145 registered in T₁₃, RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3 (36.7 kg ha⁻¹) followed by T₁₂
146 (RDF + ZnPGG-1 + SiPYY-3) *i.e.*, 35.4 kg ha⁻¹ and were on par. At 90 DAS also increased available
147 phosphorus was observed in T₁₃ (64.7 kg ha⁻¹), on par with T₁₁ (RDF + ZnKJJ-4 + SiKPP-1) *i.e.*, 62.4
148 kg ha⁻¹ and T₁₂ (RDF + ZnPGG-1 + SiPYY-3) (59.6 kg ha⁻¹). At 120 DAS available phosphorus content
149 decreased to 40.6 kg ha⁻¹ in T₁₃ and found superior to T₁₂ (38.5 kg ha⁻¹) (Table 2). An increasing trend
150 until 90 DAS followed by decreasing trend by 120 DAS though more than 45 DAS was observed in
151 general for all the available major nutrients.

152 Present results indicates that available phosphorus content increased slightly and depleted
153 gradually in all the treatments with insufficient dose of phosphatic fertilizers. Inoculated zinc and silica
154 microbial consortium stimulated root length development under reduced phosphorus supply with
155 stabilized ammonium by 52 %. This was accompanied by the increased auxin production capacity of
156 rhizosphere bacteria [13].

157 3.1.3 Available potassium in soil

158 Potassium has significant role in the regulation of water in plants (osmoregulation). Potassium
159 influences both uptake of water through plant roots and its loss through the stomata. Available
160 potassium content was 202.14 kg ha⁻¹ (Table 1) in the initial soil sample. At 45 DAS all the treatments
161 showed increased potassium compared to initial stage. The highest available potassium was recorded
162 in T₁₃, RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3 (221.4 kg ha⁻¹) followed by T₁₂ (RDF +
163 ZnPGG-1 + SiPYY-3) *i.e.*, 220.9 kg ha⁻¹ and T₁₁ (220.6 kg ha⁻¹) and were on par. At 90 DAS, all the
164 treatments showed increased potassium availability than 45 DAS; significantly highest available
165 potassium was recorded in T₁₃ (349.6 kg ha⁻¹). At 120 DAS amount of available potassium decreased
166 to 263.5 kg ha⁻¹ in T₁₃ but significantly higher than other treatments (Table 2).

167 Similar observations was made by earlier works of [14] reported increased chlorophyll content
168 by inoculated bacteria, soluble and rock potassium. Many microorganisms like zinc and silica
169 solubilizers in the soil, apart from zinc and silica, they can solubilize 'unavailable' forms of K bearing
170 minerals, such as micas, illite and orthoclases by excreting organic acids which either directly dissolve
171 rock K or chelate silicon ions to bring the K into solution [15].

172 **3.1.4 Available zinc in soil**

173 Available zinc content was 0.48 ppm (Table 1) in the initial soil sample. At 45 DAS all the treatments
174 showed increased available zinc content compared to initial. Highest available zinc was recorded in
175 T₁₃, RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3 (0.86 ppm), followed by T₁₂ (RDF + ZnPGG-1
176 + SiPYY-3) *i.e.*, 0.83 ppm. Available zinc content further increased in all the treatments at 90DAS.
177 Higher available zinc content was recorded in T₁₃ (1.14 ppm) at 90 DAS. At 120 DAS available zinc
178 content was decreased among the treatments over 90 DAS and significantly highest zinc availability
179 was recorded in T₁₃ (0.98 ppm), followed by T₁₂ (0.96 ppm) (Table 3).

180 The above results were in agreement with [16] where growth and yield parameters of paddy
181 showed a significant increase in the treatment that received combination of MZSB 6, MZSB 8 and
182 75% recommended dose of fertilizer (RDF) as compared to control and other treatments.

183 **3.1.5 Available silica in soil**

184 Available silica content was 46.0 ppm (Table 1) in the initial soil sample. At 45 DAS all the
185 treatments showed increased available silica content compared to initial stage. Highest available silica
186 was recorded in T₁₃, RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3 (66.8 ppm) followed by T₁₂
187 (RDF + ZnPGG-1 + SiPYY-3) and T₁₁ (RDF + ZnKJJ-4 + SiKPP-1) *i.e.*, 65.9 and 65.7 ppm and found
188 on par. All the treatments at 90 DAS showed increased available silica content than initial and 45
189 DAS, T₁₃ recorded highest (98.90 ppm), followed by T₁₂ and T₁₁ (97.7 and 97.3 ppm) and were on par.
190 At 120 DAS available silica content decreased among the treatments over 90 DAS and significantly
191 highest silica availability was recorded in T₁₃ (84.8 ppm), T₁₂ and T₁₁ recorded 82.6 and 82.4 ppm,
192 respectively (Table 3).

193 Available silica was observed highest in T₁₃. Similar results were observed with [17] where
194 application of silica solubilizing bacteria increased availability of silica in soil by 12.45 - 60.15 % more
195 over the control. It might be due to the silica solubilizing microorganisms present in the soil influenced
196 the available silica content in soil by additional application externally.

197 **3.2 Influence of zinc and silica solubilizing bacterial consortia on nutrient**
198 **concentration in plants**

199 **3.2.1 Percent nitrogen in plant**

200 Percent nitrogen in plant was influenced by the zinc and silica solubilizing bacterial isolates
201 and their consortia by easy availability of the nutrients. Percent N significantly differed among the
202 treatments. Highest nitrogen concentration of 0.87 %, 1.10 % and 0.98 % was recorded in T₁₃ (RDF +
203 ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3) followed by T₁₂ (0.87 %, 1.08% and 0.95%) at 45, 90 and
204 120DAS, respectively, and 0.95% in T₁₁ at 120DAS, which were on par. Control recorded the least
205 nitrogen of 0.81% (Table 4).

206 Growth enhancement of inoculated plants could be due to the higher N accumulation by
207 bacterial N₂ fixation and better root growth, which might have promoted the greater uptake of water
208 and nutrients. Similar results were found by [18] where addition of SSB-enriched biofertilizer to clay
209 substrate significantly increased the content of total nitrogen, phosphorus and potassium in the leaves
210 of *Brassica juncea*. They concluded that SSB-enriched biofertilizer improved the photosynthetic
211 function of *B. juncea*.

212 **3.2.2 Percent phosphorus in plant**

213 Zinc and silica solubilizing bacterial isolates have the ability to solubilize P to some extent,
214 these microorganisms help for the growth and development of the crop and also elevated crop
215 tolerance under water deficit condition. At 45 DAS significant concentration of plant phosphorus was
216 found in T₁₃: RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3 (0.46 %), followed by T₁₂ (RDF +
217 ZnPGG-1 + SiPYY-3) *i.e.*, 0.44 %. At 90 DAS significantly higher plant phosphorus concentration was
218 recorded in T₁₃ (0.67 %). At 120 DAS highest phosphorus concentration was obtained in T₁₃ (0.58 %),
219 followed by T₁₂ (0.56 %) and T₁₁ (0.55 %) and statistically they were on par (Table 4).

220 Phosphorus (%) was highest in T₁₃ treatment at all the intervals studied. Similar findings were
221 observed by [19] where zinc solubilizing bacterial isolates *i.e.*, *Pseudomonas striata* along with
222 *Pseudomonas fluorescence* strains showed phosphate solubilizing ability apart from zinc, resulted in
223 significant increase in percent of phosphorus in plant compared to individual inoculations in paddy.

224 **3.2.3 Percent potassium in plant**

225 Osmoregulation is maintained by the potassium concentration in the plant. More the
226 concentration of potassium, higher the osmoregulation, and helps the plant during transpiration. At 45
227 DAS and 90 DAS higher plant potassium concentration was obtained in T₁₃: RDF + ZnKJJ-4 &
228 ZnPGG-1 + SiKPP-1 & SiPYY-3 (1.87 % and 2.29 %), followed by T₁₂: RDF + ZnPGG-1 + SiPYY-3
229 (1.85 % and 2.28 %), respectively. At 120 DAS significant maximum potassium concentration was
230 recorded in T₁₃ (1.98 %), followed by T₁₀ (RDF + SiKPP-1 & SiPYY-3) and T₁₂ (RDF + ZnPGG-1 +
231 SiPYY-3) *i.e.*, 1.96 % (Table 4).

232 Present results revealed that a higher value of potassium concentration was noticed in
233 treatments those received potassium along with N or P or combinations at all the growth stages. The
234 concentration of potassium decreased slowly from 90 to 120 DAS. Besides silicon, silicate minerals
235 contain potassium, calcium, magnesium, iron and zinc and therefore inoculation of Silica solubilizing
236 bacteria (SiSB) to soil benefit the crop by releasing several of these nutrients [20]. By the action of

237 SSB potassium availability was more in soil that showed direct impact on the percent potassium in the
238 plant.

239 4. CONCLUSION

240 Nitrogen, phosphorus and potassium are essential for crop growth and development in
241 paddy, where as zinc and silica nutrients improve the grain quality and quantity. Zinc and
242 silica solubilizing bacteria and their consortia showed significant effect on available nitrogen,
243 phosphorus, potassium, zinc and silica in soil as well as nitrogen, phosphorus and potassium
244 concentration in plant compared to individual zinc and silica solubilizing microorganisms. It
245 is concluded that exogenous application of bacterial consortia can be exploited to improve the
246 nutrient status and availability in direct sown paddy.

247

248 **Table 1. Initial Physico-chemical and microbiological properties of experimental field soil**

Soil property	Field
Available N (kg ha^{-1})	142.03
Available P (kg ha^{-1})	26.28
Available K (kg ha^{-1})	202.14
Available Zn (ppm)	0.48
Available Si (ppm)	46.0

Table 2. Influence of zinc and silica solubilizing bacterial consortia on available soil nutrients, (N, P and K, kg ha⁻¹) in direct sown paddy

Treatment s	Available nitrogen (kg ha ⁻¹)			Available phosphorus (kg ha ⁻¹)			Available potassium (kg ha ⁻¹)		
	45 DAS	90 DAS	120 DAS	45 DAS	90 DAS	120 DAS	45 DAS	90 DAS	120 DAS
T ₁	179.1	218.1	208.2	26.9	48.8	28.7	212.4	330.1	251.4
T ₂	181.4	224.9	212.6	28.4	50.1	30.2	213.6	331.8	252.9
T ₃	180.7	220.5	210.8	30.5	51.0	31.4	213.4	332.2	253.3
T ₄	184.1	230.3	218.6	29.5	52.1	30.6	215.4	333.4	254.2
T ₅	182.8	228.5	214.4	30.6	54.0	31.5	215.7	333.5	254.5
T ₆	186.6	232.2	221.1	28.8	52.4	30.7	215.8	333.7	255.5
T ₇	190.2	248.5	233.2	30.9	56.6	31.8	216.6	334.6	256.1
T ₈	190.8	247.1	230.2	31.7	58.5	32.7	217.4	335.7	256.7
T ₉	191.5	248.3	232.4	31.0	55.7	32.2	218.1	336.2	257.1
T ₁₀	192.3	250.5	235.6	32.9	58.5	35.3	218.8	338.6	258.4
T ₁₁	194.2	254.8	237.2	33.3	62.4	36.4	220.6	340.8	260.0
T ₁₂	196.7	258.7	238.5	35.4	59.6	38.5	220.9	341.4	261.8
T ₁₃	198.9	262.3	240.2	36.7	64.7	40.6	221.4	349.6	263.5
SE(m)	2.32	2.50	2.14	1.23	2.04	2.02	1.15	2.12	1.01
CD(p=0.05)	6.97	7.35	6.43	3.70	6.12	6.06	3.44	6.37	3.04
CV	3.14	1.80	2.43	4.14	6.34	5.23	3.48	4.13	2.15

249 T₁: RDF (Control), T₂: RDF + ZnSO₄, T₃: RDF + Calcium silicate, T₄: RDF + ZnSO₄ + Calcium silicate, T₅: RDF + ZnKJJ-4, T₆: RDF + ZnPGG-1, T₇: RDF + SiKPP-1,
250 T₈: RDF + SiPYY-3, T₉: RDF + ZnKJJ-4 & ZnPGG-1, T₁₀: RDF + SiKPP-1 & SiPYY-3, T₁₁: RDF + ZnKJJ-4 + SiKPP-1, T₁₂: RDF + ZnPGG-1 + SiPYY-3, T₁₃: RDF +
251 ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3

Table 3. Influence of zinc and silica solubilizing bacterial consortia on available soil micro nutrients, Zn and Si (ppm) in direct sown paddy

Treatments	Available Zinc (ppm)			Available Silica (ppm)		
	45 DAS	90 DAS	120 DAS	45 DAS	90 DAS	120 DAS
T ₁	0.60	0.91	0.69	57.2	90.1	72.1
T ₂	0.61	0.94	0.74	58.4	91.2	74.2
T ₃	0.69	0.98	0.79	59.6	92.2	75.2
T ₄	0.68	0.97	0.76	60.8	93.3	76.4
T ₅	0.67	0.96	0.78	62.2	94.5	78.3
T ₆	0.69	0.99	0.79	60.4	93.1	77.2
T ₇	0.74	1.02	0.81	62.3	94.7	79.7
T ₈	0.76	1.05	0.84	63.4	95.2	80.4
T ₉	0.77	1.08	0.86	63.7	95.6	80.6
T ₁₀	0.78	1.09	0.88	64.6	96.4	81.2
T ₁₁	0.81	1.10	0.93	65.7	97.3	82.4
T ₁₂	0.83	1.11	0.96	65.9	97.7	82.6
T ₁₃	0.86	1.14	0.98	66.8	98.9	84.8
SE(m)	0.00	0.01	0.01	0.63	0.43	0.54
CD (p=0.05)	0.01	0.03	0.02	1.84	1.28	1.62
CV	1.43	1.67	1.35	1.74	1.37	1.42

252 *T₁: RDF (Control), T₂: RDF + ZnSO₄, T₃: RDF + Calcium silicate, T₄: RDF + ZnSO₄ + Calcium silicate,*
 253 *T₅: RDF + ZnKJJ-4, T₆: RDF + ZnPGG-1, T₇: RDF + SiKPP-1, T₈: RDF + SiPYY-3, T₉: RDF + ZnKJJ-4 &*
 254 *ZnPGG-1, T₁₀: RDF + SiKPP-1 & SiPYY-3, T₁₁: RDF + ZnKJJ-4 + SiKPP-1, T₁₂: RDF + ZnPGG-1 + SiPYY-3,*
 255 *T₁₃: RDF + ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3*
 256

Table 4. Influence of zinc and silica solubilizing bacterial consortia on N, P and K concentration (%) in direct sown paddy plants

Treatments	Nitrogen (%)			Phosphorus (%)			Potassium (%)		
	45 DAS	90 DAS	120 DAS	45 DAS	90 DAS	120 DAS	45 DAS	90 DAS	120 DAS
T ₁	0.70	0.93	0.81	0.32	0.53	0.38	1.71	2.10	1.84
T ₂	0.71	0.95	0.83	0.35	0.54	0.41	1.73	2.14	1.86
T ₃	0.73	0.97	0.84	0.36	0.55	0.44	1.74	2.16	1.85
T ₄	0.76	0.98	0.86	0.37	0.56	0.46	1.76	2.17	1.88
T ₅	0.77	0.99	0.87	0.38	0.57	0.45	1.78	2.19	1.89
T ₆	0.81	1.02	0.93	0.37	0.58	0.47	1.80	2.20	1.90
T ₇	0.82	1.03	0.92	0.38	0.60	0.51	1.82	2.23	1.93
T ₈	0.83	1.05	0.90	0.40	0.61	0.52	1.81	2.22	1.92
T ₉	0.85	1.06	0.93	0.41	0.62	0.54	1.83	2.24	1.94
T ₁₀	0.86	1.07	0.94	0.42	0.62	0.53	1.85	2.26	1.96
T ₁₁	0.85	1.06	0.95	0.43	0.63	0.55	1.84	2.25	1.95
T ₁₂	0.87	1.08	0.96	0.44	0.64	0.56	1.85	2.28	1.96
T ₁₃	0.89	1.10	0.98	0.46	0.67	0.58	1.87	2.29	1.98
SE(m)	0.01	0.01	0.01	0.00	0.00	0.01	0.02	0.01	0.01
CD(p=0.05)	0.02	0.02	0.02	0.06	0.01	0.02	0.05	0.02	0.02
CV	1.15	2.18	1.47	1.85	2.18	1.28	2.55	1.25	1.47

257 *T₁: RDF (Control), T₂: RDF + ZnSO₄, T₃: RDF + Calcium silicate, T₄: RDF + ZnSO₄ + Calcium silicate,*
 258 *T₅: RDF + ZnKJJ-4, T₆: RDF + ZnPGG-1, T₇: RDF + SiKPP-1, T₈: RDF + SiPYY-3, T₉: RDF + ZnKJJ-4 & ZnPGG-*
 259 *1, T₁₀: RDF + SiKPP-1 & SiPYY-3, T₁₁: RDF + ZnKJJ-4 + SiKPP-1, T₁₂: RDF + ZnPGG-1 + SiPYY-3, T₁₃: RDF +*
 260 *ZnKJJ-4 & ZnPGG-1 + SiKPP-1 & SiPYY-3*

261 **Disclaimer (Artificial intelligence)**

262 Author(s) hereby declare that NO generative AI technologies such as Large Language Models
263 (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of
264 manuscripts.

265 REFERENCES:

- 266 1. Rodriguiz H, Gonzalez T, Goire I and Bashan Y. Gluconic acid production and phosphate
267 solubilization by the plant growth promoting bacterium *Azospirillum* spp. *Nature*. 2004; 91:
268 552-555.
- 269 2. Thompson JP. Correction of dual phosphorous and zinc deficiencies of linseed with cultivars
270 of vesicular-arbuscularmycorrhizal fungi. *Soil Bio and Bioche*. 1996; 28: 941-951.
- 271 3. Sang MK, Muhammad W, Raheem S, Young HY and Sajjad A. Isolation and characterization
272 of a novel silicate-solubilizing bacterial strain *Burkholderiae burnea* CS4-2 that promotes
273 growth of japonica rice (*Oryza sativa* L. cv. Dongjin). *Soil Sci and Plant Nutri*. 2017; 63(3).
274 233-241.
- 275 4. Subbiah BV and Asija CL. A rapid procedure for the estimation of available nitrogen in soils.
276 *Curre Sci*. 1956; 25: 259-260.
- 277 5. Olsen SR, Code CL, Watanable FS and Dean LA. Estimation of available phosphorus in soils
278 by extraction with sodium bicarbonate. *United States Development Agency, Washington DC,*
279 *Circular Number*. 1954; 939. 1-19.
- 280 6. Jackson ML. *Soil Che Analy*. Prentice- Hall of India private limited. New Delhi.1973.
- 281 7. Lindsay WL and Norvell WA. Development of DTPA soil test for zinc, iron, manganese and
282 copper. *Soil Sci Socie of American Jour*. 1978; 41:421-428.
- 283 8. Snyder GH. Development of a silicon soil test for Histol-grown rice. Belle Grade EREC
284 Research report EV-1991-2, University of Florida, Belle grade, FL. 1991.
- 285 9. Piper CS. *Soil and Plant Analysis*. Hons publishers, Bombay. 1966.
- 286 10. Tandon HLS. Analysis of plant material for macro and micronutrients. In method of analysis of
287 soils, plant, water and fertilizer. 1993. pp. 49-82.
- 288 11. Anusha A, Kadali SK, Udayasaukar A and Thakurs KD. Influence of biofertilizers on uptake of
289 NPK in soils and eggplant. *Inter Jour of Analy Micro and Appl Sci*. 2017; 6 (12): 1259-1263.
- 290 12. Yuvaraj K. Effect of biofertilizers and inorganic fertilizers on soil health, growth and yield of
291 Rice (*Oryza sativa* L.) crop. M. Sc. (Agri.) Thesis, Punjab Agricultural University, Ludhiana,
292 India. 2016.
- 293 13. Bradacova K, Kandler E, Berger N, Ludewig U and Neumann G. Microbial consortia
294 inoculants stimulate early growth of maize depending on nitrogen and phosphorus
295 supply. *Plant, Soil and Enviro*. 2020. 66 (3): 105-112.
- 296 14. Prajapati MC and Modi HA. Isolation of two potassium solubilizing fungi from ceramic industry
297 soils. *Life Sci Leaflets*. 2012. 5: 71-75.
- 298 15. Bennett PC, Choi WJ and Rogers JR. Microbial destruction of feldspars. *Mineralogical*
299 *Magazine*. 1998; 8 (62A): 149-150.
- 300 16. Manasa S, Mahadevaswamy G, Yadav R, Nagaraj MN and Gundappagol RC. *In vivo* efficacy
301 of zinc solubilizing bacteria on available zinc content, growth and yield attributes of paddy
302 (*Oryza sativa* L.). *Jour of Agril and Eco Res Intern*. 2019; 1-9.
- 303 17. Chandrakala C, Voleti SR and Bandeppa S. Silicate solubilization and plant growth promoting
304 potential of *Rhizobium* sp. Isolated from Rice Rhizosphere. *Silicon*. 2019; 11, 2895-2906.
- 305 18. Maleva WM, Hussein MM, Mehanna HM and El-Moneim DA. Bacteria polysaccharides elicit
306 resistance of wheat against some biotic and abiotic stress. *Inter Jour of Pharma Sci Rev and*
307 *Res*. 2017; 29 (2): 50: 292-298.
- 308 19. Alagawadi AR and Gaur AC. Inoculation of zinc solubilizing pseudomonas bacterial isolates
309 their effect on yield of paddy. *Tropi Agril*. 2012; 5:16-18.
- 310 20. Muralikannan N. Biodissolution of silicate, phosphate and potassium by silicate solubilizing
311 bacteria in rice ecosystem. M. Sc (Ag) thesis submitted to Tamil Nadu Agricultural University.
312 Coimbatore. 1996; pp: 125.

- 313 21. Babu, S. Vinod, A. Vijaya Gopal, N. Trimurtulu, G. Kishore
314 Babu, and S. L. Bhattiprolu. 2024. "Efficacy of Silica Solubilizing
315 Bacteria As Plant Growth Promoting Rhizobacteria and Their
316 Biochemical Characteristics". *International Journal of Environment
317 and Climate Change* 14 (6):199-210.
318 <https://doi.org/10.9734/ijecc/2024/v14i64221>
319
- 320 22. Chandrakala, C., Voleti, S. R., Bandeppa, S., Sunil Kumar, N., & Latha, P. C. (2019). Silicate
321 solubilization and plant growth promoting potential of Rhizobium sp. isolated from rice
322 rhizosphere. *Silicon*, 11(6), 2895-2906.
323
- 324 23. Raturi, G., Sharma, Y., Rana, V., Thakral, V., Myaka, B., Salvi, P., ... & Deshmukh, R. (2021).
325 Exploration of silicate solubilizing bacteria for sustainable agriculture and silicon
326 biogeochemical cycle. *Plant Physiology and Biochemistry*, 166, 827-838.
327

UNDER PEER REVIEW