

1 **EFFECTIVENESS OF PHOSPHORUS SOLUBILIZING BACTERIA ON ENHANCING PHOSPHORUS**
2 **AVAILABILITY FROM MINJINGU PHOSPHATE ROCK TO MAIZE IN SLIGHTLY ACID TO**
3 **NEUTRAL SOILS**

12 **Abstract**

13 The aim of this one-year field study was to evaluate the impact of phosphorus solubilizing
14 bacteria (PSB) on increasing the availability of phosphorus locked up in insoluble Minjingu
15 phosphate rock for the maize crop (*Zeamays L.*) in high soil pH. The study was carried out
16 during one wet season in Nghumbi and Mlalivillages in Kongwa District in the Dodoma Region,
17 Tanzania. Based on the pH of their soil and the limiting nutrients, two farms from Mlalivillage
18 and five from Nghumbivillage were chosen at random for field trials. To determine the
19 fertility status, composite soil samples were collected from the chosen farms and examined
20 in a lab. Target treatments included the application of PSB inoculum in combination with
21 various rates of Minjingu phosphate rock (MPR) (0, 20, 30, 40, and 60 Kg Pha⁻¹) as a basal
22 fertilizer, placed beneath maize seed and covered with a thin layer of soil. Using a fully
23 randomised block design, the treatments were set up three times. Inadequate levels of N, P,
24 K, and Zn were found in the research area's soil samples. Results indicated the main effect of
25 fields' characteristics from field No. 2 had the highest yield to other fields with significantly
26 ($P < 0.001$) highest grain yield being 4.4 t ha⁻¹, biological yield of 5.6 t ha⁻¹, and Straw P uptake
27 of 19.63 kg ha⁻¹. Using Poraninoculant (Mx) produced a negligible yield of grain and straw,
28 according to the major effect of treatments. For the studied variables, none of the treatments
29 showed any significant ($P = 0.427$) interaction effects. An intriguing revelation that the study's
30 maize response to native P is provided by interaction effects are a factor of soil and amount
31 of P released to the soil. We recommend more research on PSB for more than one season in
32 high soil pH before ascertaining the technology of farmers.

33 **Keywords:** Bio-fertilizer; cropping systems; Food crops; Soil fertility; Tanzania.

34 **1 Introduction**

35 Along with other nutritional elements, phosphorus (P) is the macronutrient that often
36 restricts plant growth and the final output. As a plant ages, phosphorus that has been
37 absorbed is moved to its fruiting sections, where the production of seeds necessitates a
38 significant energy expenditure. Soil phosphorus deficits affect normal crop maturity as well
39 as seed development. The most common nutrient deficiency in tropical African soils is
40 phosphorus, which significantly restricts Tanzania's ability to produce maize. According to
41 Nhunda et al. (2024), low-access P is a problem in Kongwa district, as evidenced by the soil's

42 acidic and alkaline conditions. Low P in the parent material and high P fixation by iron and
 43 aluminum oxides in acid are the causes of low soil P (Siles *et al.*, 2022) and precipitation by
 44 calcium in alkaline soils (Adnan *et al.*, 2017).

45 Since the 1960s, Tanzania has paid close attention to the use of phosphate rock (PR), such as
 46 Minjingu phosphate rock (MPR), as an alternative P source (Anderson, 1993; Ikerra *et al.*,
 47 1994; Szilas, 2002). Previous research findings indicated that MPR's residual effect made it
 48 superior to subsequent crops in the field after the original application (Ikerra *et al.*, 2007). The
 49 prolonged and continual gradual release of P from MPR is the cause of the high residual and
 50 long-lasting effects. Numerous studies have documented PR's capacity to release P, but little
 51 is known about how high/alkaline pH influences the material's ability to dissolve in soil.

52 Due to the lack of solubilization required to liberate P for crop use, PR is difficult to use for
 53 crop production in neutral (pH 6.6–7.3) and alkaline (pH > 7.3) soils. As soil pH climbs to 6.2,
 54 PR solubilizes swiftly at pH ranges of 4.9 to 5.5. According to Anderson *et al.* (2018), PR
 55 becomes completely insoluble at pH values higher than 6.1. Bolan and Hedley (1990)
 56 employed three forms of PR: Jordan PR (JPR), North Carolina PR (NCPR), and Nauru PR (NPR).
 57 The degree of PR dissolving at each pH was determined to follow the following order, based
 58 on the decreasing order of the chemical reactivities: As soon as the pH dropped below six,
 59 from NCPR > JPR > NPR. 5 to 3.9, the dissolution of PR increased from 29.3% to 83.5%, from
 60 18.2% to 78.9%, and from 12.5% to 60.3% for NCPR, JPR, and NPR, respectively.

61 In contrast as the pH decreased from 6.5 to 3.9, the proportion of dissolved P extracted by
 62 0.5 M NaHCO₃ decreased from 38% to 5% and the proportion of P taken up by ryegrass plant
 63 decreased from 46% to 7% (Bolan and Hedley, 1990). The decrease in plant available P
 64 corresponds to an increase in adsorption of inorganic P in low pH (Bolan and Hedley, 1990).
 65 Bolan and Hedley (1990) further noted that an increase in pH was associated with decreased
 66 degree of PR dissolution. Warren *et al.* (2009) reported that dissolution of P from PR was
 67 insignificant at pH > 6.1 while citing out the causes being pH in the media and P sorption.

68 It is necessary to investigate a unique mechanism in order to release the locked-up P in the
 69 insoluble PR when it is applied at high soil pH. It has been reported that phosphorus
 70 solubilizing bacteria convert insoluble to soluble P from PR by releasing low-weight molecular
 71 organic acids such as acetic, formic, propionic, lactic, glycolic, and fomic acids (Zaworotko *et al.*,
 72 2019). Carboxyl and hydroxyl groups from organic acids are capable of chelating with
 73 cations bound to phosphate thereby converting it into soluble forms (Zaworotko *et al.*, 2019).
 74 Additionally, the bacteria produce acidity by evolving carbon dioxide (CO₂), which causes
 75 calcium phosphate to become soluble (Yousefi *et al.*, 2011). It has been demonstrated that
 76 in soils with high pH values (> 6.2), phosphorus-solubilizing bacteria improve P availability,
 77 hence promoting crop growth (Bhattacharya and Jha, 2012).

78 According to reports, inoculating PSB into soil contaminated with metals increased maize
 79 production (Jian *et al.*, 2008). Lin *et al.* (2009) discovered the nodulation, root, and shoot
 80 biomass of the B-inoculated maize and cowpea plants. *Compared to the control group, there*
 81 *were notably more Cepacta*. While the benefit of bacteria in raising P availability from PR
 82 are well established in acidic soils, they have not been consistently shown in Tanzanian
 83 alkaline soil studies (Zaidi *et al.*, 2009). There is a dearth of information on high pH soils, and
 84 the outcomes of the little that is available are evasive or unclear. The study aims to increase

85 maize output in Dodoma's Kongwa district by utilizing additional P from Minjingu PR, which
86 will be released into its alkaline soils. The precise goals were to assess maize performance,
87 calculate P uptake by maize, and determine how well PSB dissolved MPR in high-pH soil.

88 Maize is one of the key staple food crops and major cereal consumed in Tanzania. It is
89 estimated that the annual per capita consumption of maize in Tanzania is 112.5 kg and
90 national maize consumption is estimated to be three million tons per year (Giller *et al.* 2021a).
91 The crop is annually on an average of two million hectares or about 45% of the cultivated area
92 in Tanzania. The Dodoma region is a semi-arid area and therefore maize production is
93 hampered by drought, among other factors. Average maize production in Dodoma is about
94 0.4 tons per hectare which is far below the national average yield of just over 1 ton per hectare
95 (Giller *et al.* 2021a). This study therefore was set in this region with the aim of rising up maize
96 yield by enhancing phosphorus availability to maize crop.

97 P demand by crops needs to be taken into account especially in smallholder farmers.
98 Compared to perennial crops, crops with intense and short cycle growth, such as maize,
99 require higher P levels in soil solutions and faster absorbed P replenishment for optimal
100 production (Lino *et al.*, 2018). But because there is never enough phosphorus in agricultural
101 soils, people must rely on artificial fertilizers, which can have negative financial and
102 environmental effects. This manuscript investigates the potential synergy between Minjingu
103 rock phosphate and phosphorus solubilizing agents in high soil pH as a sustainable and
104 environmentally friendly way to increase phosphorus availability for maize cultivation (Hengli
105 *et al.*, 2017).

106 **Hypothesis:** Application of phosphorus solubilizing bacteria will significantly enhance the
107 availability of phosphorus from Minjingu phosphate rock to maize in high P soil conditions,
108 resulting to improved maize growth and yield.

109 109

110 2 Materials and Methods

111 2.1 Description of the study area

112 This study was conducted in Mlali and Nghumbi villages of Kongwa district, Dodoma region
113 located within the semi-arid zone of Central Tanzania. Kongwa district is located between
114 latitudes 5.47 to 6.26° S and longitude 36.15 to 37.08° E. Mlali village is located between
115 latitude 6° 16' 22" to 6° 17' 15" S and longitude 36° 42' 04" to 36° 47' 26" E while Nghumbi village
116 is located between latitude 6° 18' 17" to 6° 20' 36" S and longitude 36° 47' 57" to 36° 50' 58" E.
117 The location of each experimental field is indicated by using central points as shown in Table
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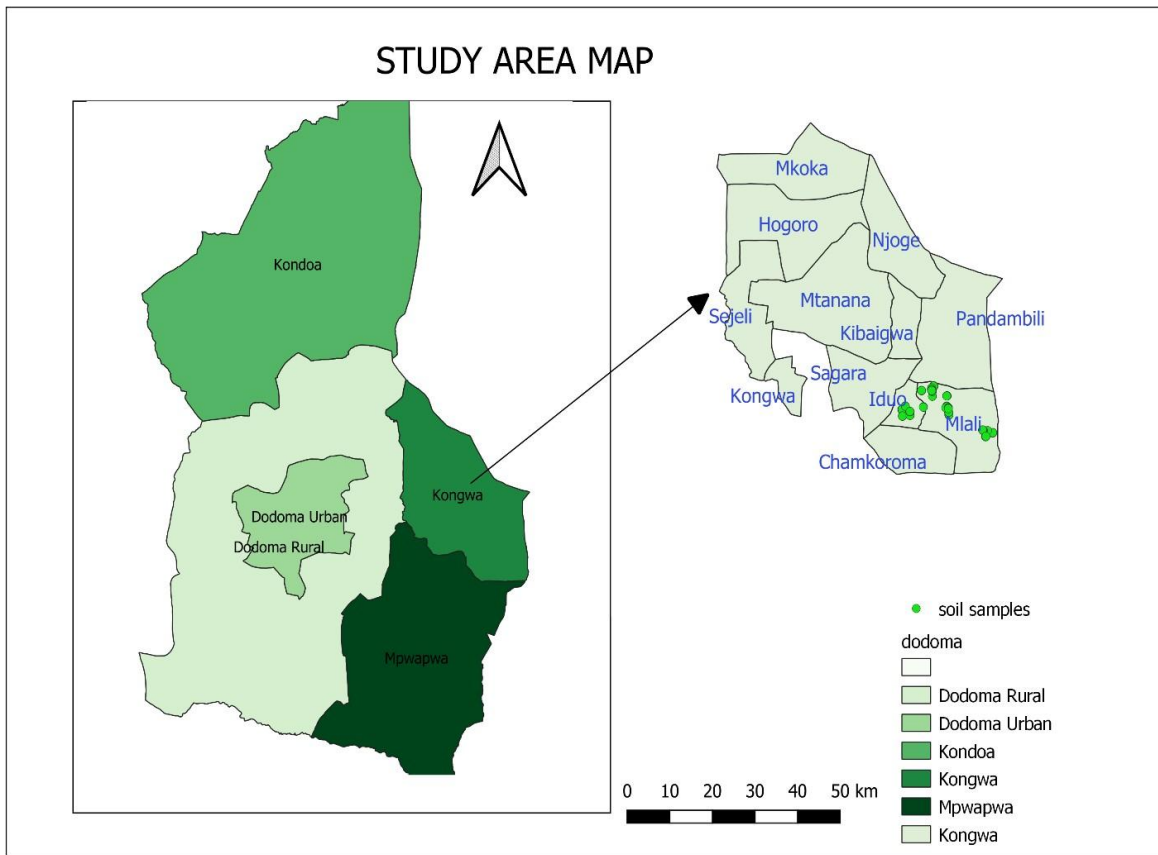
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128 **Table1:GeographicallocationsofthestudiedfieldsinKongwaDistricts,Tanzania**

| Village | FarmNo. | Coordinates | |
|---------|---------|-------------|-------------|
| Nhgumbi | 1 | S 06.31561 | E 036.82605 |
| | 2 | S 06.31456 | E 036.82698 |
| | 3 | S 06.31414 | E 036.84106 |
| | 4 | S 06.30932 | E 036.83009 |
| | 5 | S 06.30815 | E 036.81939 |
| Mlali | 6 | S 06.26317 | E 036.74073 |
| | 7 | S 06.26348 | E 036.74657 |

129 Thevillagesarecharacterisedbymediumaltitudeplainswithsomehillranges,mainly
 130 mediumtexturedsoilswithlowtomoderatefertility(Mowo*et al.*,1993).Soilsarediversebut
 131 dominatedbyhighlyweatheredandclassifiedasChromicLuvisolswithsandyloamtexture
 132 tropicalsoils(Meliyo*et al.*,2014).

133 Theslectedvillageshaveundulatingtorollingplainsandplateauxwithanaltituderanging
 134 from700to900metresabovesealevel(masl).Rainsareusuallyerraticwithvariabilityintheir
 135 onset,distribution,andintensity(Mongiet *al.*,2010).Therestudyareahasaverageannual
 136 rainfallrangingfrom500to800mm(Meliyo*et al.*,2014).Seasonaldistributionsofraincanbe
 137 verysporadicwith48%oftherainfallingtowardsthe endofthegrowing seasongiving little
 138 advantagetocropgrowthandyield(Kimaro*et al.*,2009).



140 Figure1:MapofDodomaregionshowingstudydistrictandvillagesinwhichexperiment
141 wasconducted

142 2.2 Siteselectionofthestudysite

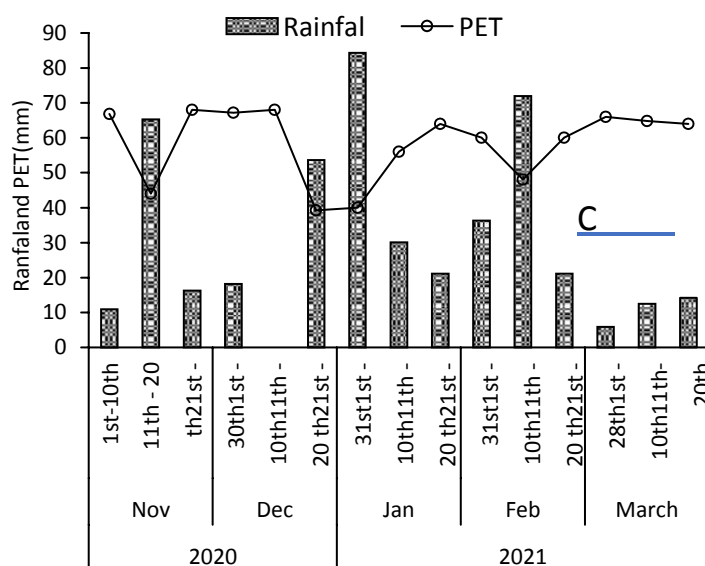
143 In ordertolearnaboutthecroppinghistoryofthefieldsandtogainanunderstandingofthe
 144 techniquesinvolvedinproducingmaize,suchasthe kindsutilized,plantingdates,and
 145 establishmentofagrowingseason,areconnaissancemissionwascarriedoutinseven
 146 significantregionsintheKongwadistrict.Additionally,focusgroups(FGDs)andconsultative
 147 sessionswithvillageleadersandindividualfarmerswereheld.Thissurveyrevealedthat
 148 farmersemploylocalkindsofmaize,theresearchareahastwotothreemonthsof rainfall,
 149 andDecember istheidealtimetoputuptrials.

150 Compositesoilsamplesweretakenat0–20cmdepthsin24surveyedfields,which
 151 correspondedto theKongwadistrict'smaize-growingzones.Afterthesesoilswereanalyzed
 152 inalab,afielddystudyincludingmaizeasthetestcropandPRinjectedwithP-solubilizing
 153 bacteriawaseventuallylimitedtosevenfieldsbasedonthepHofthesoil.As showninFigure
 154 2,dailydataontemperatureandrainfallweregatheredfromNovember2020untilApril2021,
 155 whentheexperimentcametoaconclusion.Whenmaizeplantswereintheexperimental
 156 fields,therewasnonlinear(polynomial)dropintemperatureorrainfall.

157 AsshowninFigure2,dailyrainfalldatawasgatheredfromtherainfallstationthatUSAID
 158 erectedintheexperimentalregionaspartoftheIIITA-AfricaRISINGESAProjectin2019 from
 159 November2020toApril2021.Ten(10)dayintervalsareusedtoillustratetherainfalldata.

160 When maize plants were in the experimental fields, there was no linear (polynomial) drop in
 161 temperature or rainfall. The closest experimental allocation used by Casper (2002) to collect
 162 evapotranspiration data provided the evapotranspiration data.

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165 **Figure 2:** Trends of rainfall (mm) and evapotranspiration in the study area during period of
 166 experimentation with maize crop – Kongwa district. L= shows the planting date, B= is the
 167 drought period during vegetative growth, C= drought period during flowering and grain
 168 filling

169 **Table 2:** Identity of isolated PR-solubilizing species based on nucleotide database on
 170 American National Institute of Health (NIH) NCBI genetic database (GenBank) (source: SUA
 171 soil laboratory published by Kwasiema, 2021).

| Isolate | Species | Accession number | Nucleotide identity | Country | Source rhizosphere |
|---------------|------------------------------|------------------|---------------------|----------|--------------------|
| Fg1 | <i>Fusarium proliferatum</i> | MZ497514 | 100 | Tanzania | Maize |
| Mk10 | <i>Burkholderia</i> sp | MZ502221 | 99.9 | Tanzania | Maize |
| NA19a | <i>Klebsiella</i> sp | MZ502673 | 99.9 | Tanzania | Maize |
| Klm3 | <i>Burkholderia</i> sp | MZ502220 | 99.9 | Tanzania | Maize |
| MbMz1 | <i>Klebsiella</i> sp | MZ502668 | 99.8 | Tanzania | Maize |
| SI-Sp1 | <i>Klebsiella</i> sp | MZ502674 | 99.8 | Tanzania | Sweet potato |
| NA4a | Unidentified | | | Tanzania | Irish potato |
| NA4b | <i>Klebsiella</i> sp | MZ502671 | 99.8 | Tanzania | Irish potato |
| SUApp3 | <i>Klebsiella</i> sp | MZ502675 | 99.7 | Tanzania | Sweet pepper |
| MdG1 | <i>Klebsiella varriicola</i> | MZ502670 | 99.8 | Tanzania | Banana |

| | | | | | |
|-------------|------------------------------|----------|------|----------|--------------|
| NA5 | <i>Klebsiella varriicola</i> | MZ502672 | 99.9 | Tanzania | Sweet potato |
| MdE4 | <i>Klebsiella varriicola</i> | MZ502669 | 100 | Tanzania | Common bean |

172 2.3 Experimental design, treatment, and field experimentation

173 Seven fields (two at Mlali and five at Nghumbi) with the desired soil pH in the study area were
 174 selected. Trials using maize as a test crop in seven selected fields were established. The
 175 treatment combinations were PSB inoculum co-applied with MPR at different rates, as shown
 176 in table 2.

177 **Table 3.** Treatment combinations used in a study

| S/N | Treatment | Treatment symbol | Description |
|-----|-----------|--------------------------------|--|
| 1 | T1 | P ₀ M ₀ | treatment control |
| 2 | T2 | P ₃₀ M ₀ | MPR at 30 kg ha ⁻¹ |
| 3 | T3 | P ₀ M _x | Only PSB |
| 4 | T4 | P ₂₀ M _x | MPR at 20 kg ha ⁻¹ with PSB |
| 5 | T5 | P ₄₀ M _x | MPR at 40 kg ha ⁻¹ with PSB |
| 6 | T6 | P ₆₀ M _x | MPR at 60 kg ha ⁻¹ with PSB |

178 There were three replications of the treatments, for a total of eighteen plots. Using maize as
 179 the test crop and three replications or blocks of treatments in each of these seven fields, the
 180 study used a randomized complete block design (RCBD). Every test plot measured 4 m in
 181 length and 3 m in breadth, or 12 m². Between December 25th and 30th, two maize seeds
 182 (variety Situka) were planted in each hole. As a result, there were four rows and thirteen holes
 183 in a row, with a 90 cm gap between rows and a 30 cm gap within rows. Applying the PSB
 184 inoculum under maize seed, various rates of P from MPR (0, 20, 30, 40, and 60 kg ha⁻¹) were
 185 combined with a basal fertilizer and adding a little layer of soil on top. Every planting hole
 186 received 5 mL of solution containing the inoculum. Limiting nutrients like N and S were fixed
 187 by using Yara Amidas, which was divided into two dressings: a basal dressing and a top
 188 dressing (N 40% and S 5.5%).

189 189

190 2.4 Data collection

191 2.4.1 Laboratory soil analysis

192 Before field selection and the commencement of field trials, the soils were assessed for total
 193 nitrogen using the micro-Kjeldahl method (Bremner and Mulvaney, 1982). Available P was
 194 extracted using the Bray-1 method (1982), and its color was determined using a
 195 spectrophotometer using the molybdenum blue method (Murphy and Riley, 1962).
 196 Exchangeable K was found in ammonium acetate filtrates using a flame photometer. The
 197 same filtrate included exchangeable Ca and Mg, as determined by atomic adsorption
 198 spectrophotometry.

199 199

200 2.4.2 Laboratory analysis of plants samples

201 Following the completion of the plant growth cycle, fully grown maize plants were taken out
 202 of each plot, their individual biomass was measured, the cobs were threshed, and the dry
 203 grain weight was computed. In order to calculate P absorption and P utilization efficiency, the
 204 cobs and straws were chopped, cleaned, and processed in the lab using Moberg's (2002)
 205 guidelines for P concentration analysis. The P absorption by maize straw was then calculated
 206 using Equation 1's methodology.

$$207 \quad P_{\text{uptake}} (\text{kg ha}^{-1}) = P_{\text{conc. in straw}} (1/100) \times \text{straw yield} (\text{kg ha}^{-1}) \times 1000 \quad (1)$$

208 Equation 2's biomass yield per unit of nutrient uptake was used to calculate P uptake by
 209 straw.

$$210 \quad P_{\text{uptake in straw}} (\text{kg kg}^{-1} \text{P}) = \frac{BY_f}{N_f} \quad (2)$$

211
212

211 Where BY_f is the biological/straw yield (kg ha^{-1}) and N_f is the (P) uptake by the straw.
 212 Biologically yield is defined as the total dry matter accumulation of a plant material.

213 Furthermore, P use efficiency (PUE) was calculated by using amount of P in straws/uptake
 214 and maize biomass yield as shown in Equation 3.

$$215 \quad \text{Straw P use efficiency} (\text{kg kg}^{-1}) = \frac{\text{Straw yield in kg}}{\text{Straw P uptake in kg}} \quad (3)$$

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218 2.4.3 Limiting nutrients of the study soils in Kongwa district

219 In order to comprehend the characteristics of each field under study and the particular
 220 nutrients that are likely to restrict crop growth and development, limiting nutrients for each
 221 field were selected and arranged based on farm number. The acquired soil data were used to
 222 assess each field's limiting nutrients and field features.

223 2.5 Statistical analysis

224 In assessing phosphorus concentration (%P), total P uptake, P use efficiency, biological yield,
 225 and grain yield the fixed main effects were the farmer's field characteristics and treatments,
 226 whereas blocks were treated as random effect. A TWO-WAY analysis of variance (ANOVA)
 was performed and the model in Equation 4 was used.

$$227 \quad Y_{ij} = \mu + \alpha_i + \beta_j + (\alpha\beta)_{ij} + \varepsilon_{ij} \quad (4)$$

228 Where Y_{ij} is the observed response variable in the ij th factors; μ is the overall (grand) mean;
 229 α_i and β_j are the main effects of the factors farmer's field characteristics and treatments,
 230 respectively; $(\alpha\beta)_{ij}$ is the two-way (first order) interactions between the factors; ε_{ij} is the
 231 random error associated with the observation of response variable in the ij th factors.

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236 3 Results and Discussion

237 3.1 Fertility status of the selected fields in study area

238 3.1.1 Soil pH

239 Table 3 shows the pH range of the soils within the research area, which was 6.48 for slightly
 240 acidic soils and 7.7 for slightly alkaline soils. The native P content and pH of the soil have an
 241 impact on how well phosphate-solubilizing microorganisms work (Wakelin, 2007).

242 The solubilization of phosphates in soil is facilitated by microbial secretion of low molecular
 243 weight organic acids, which alters pH and causes phosphate to become soluble (Chenet al.,
 244 2006). Due to anion exchange, these organic acids have the ability to dissolve phosphates or
 245 chelate the Ca, Fe, or Al ions that are connected to the phosphates (Gyaneshwari et al., 2002).
 246 The pH of the soils in the fields under investigation does not support Minjingu PR
 247 solubilization for P release for plant uptake. In this regard, the use of solubilizing bacteria is
 248 an important alternative to enhance plant P nutrition (Gyaneshwari et al., 2002; van der
 249 Heijden et al., 2008).

**250 Table 4: Level of some chemical properties and their ratings for the studied soils in
 251 selected fields in Kongwa district**

| VILLAGE | Farm No. | Soil pH _{H2O} | TN | OC | Ext. P (Olsen) | S | Zn | Ca | Mg | K |
|---------|----------|------------------------|--------|--------|------------------------|--------|-------|-----------------------------|-------|-------|
| | | | (%) | | (mg kg ⁻¹) | | | (cmol(+) kg ⁻¹) | | |
| NGHUMBI | 1 | 7.05 | 0.15l | 1.11l | 38.61h | 10.33m | 1.98m | 9.14h | 3.41h | 0.25m |
| | 2 | 7.06 | 0.09vl | 1.11l | 53.51h | 16.72m | 3.74h | 8.76h | 3.40h | 0.29m |
| | 3 | 7.1 | 0.07vl | 0.74l | 29.86h | 17.41m | 1.91m | 5.21h | 2.38m | 0.33m |
| | 4 | 6.69 | 0.11l | 1.26m | 19.79l | 13.07m | 1.78m | 10.04h | 4.06h | 0.32m |
| | 5 | 6.99 | 0.05vl | 0.59vl | 22.09m | 21.73h | 3.86h | 6.92h | 3.05h | 0.21l |
| MLALI | 6 | 7.22 | 0.02vl | 0.30vl | 11.11l | 21.31h | 0.94l | 2.47m | 1.16m | 0.24l |
| | 7 | 7.63 | 0.04vl | 0.63l | 9.19l | 14.43m | 0.82l | 5.29h | 1.23m | 0.16l |

252 Key: M=moderate, h=high, vl=very low, vh= very high, l=low, exch. Ca=exchangeable Ca,
 253 ext. P=extractable phosphorus, TN =total nitrogen (Nhunda et al 2024).

254 3.1.2 Available phosphorus

255 According to Landon (1991), the results demonstrated that 54% of the chosen fields had
 256 sufficient available P, while 46% of the fields had insufficient available P (Table 4). Low P in
 257 some fields may result from precipitation by high Ca measured in the examined soils, or it may
 258 be related to intrinsic low P in the parent material. The primary determinant of phosphate
 259 concentration in the soil's liquid phase is the amount of calcium present in the soil solution
 260 (Gyaneshwari et al., 2002).

261 Microorganisms are essential to the soil P cycle and are involved in the mediation of P
 262 availability (Kannapiran and Ravindran, 2012; Turan et al., 2006). According to Adesemoye
 263 and Kloepper (2009), phosphate-solubilizing microorganisms can increase crop uptake and
 264 production by solubilizing inaccessible soil P. Numerous studies have shown that adding P-
 265 solubilizing bacteria or fungus to soil can increase its availability (Chenet al., 2006; Adesemoye

266 and Kloepper, 2009). Numerous autotrophic and heterotrophic soil microorganisms have
 267 been found to solubilize mineral phosphorus and to contribute to the mobilization of soil P in
 268 forms that are soluble for plants (Chen et al., 2006).

269 Sumner and Farina (1986) reported that maize responded favorably to the combination of
 270 plant-available soil P and N that was applied to the crop. While Ca and Mg were sufficient in
 271 all of the soils, the exchangeable K in the soils of the fields under study ranged from low (0.16
 272 cmol(+)kg⁻¹) to medium (0.32 cmol(+)kg⁻¹) (Table 4).

273 273

274 3.1.3 Limiting nutrients in the soils of experimental fields

275 Table 5 presents the findings of the categorization of limiting nutrients in the soils of the
 276 tested areas. The nutrients were categorized and rated based on their distinct chemical
 277 characteristics, as listed in Table 4. The limiting nutrients in the soils of the experimental fields
 278 were found to be N, P, K, and Zn. All soils were lacking in nitrogen (N), which was fixed by
 279 adding fertilizer that contained N (Yara Amidas). The most frequent food crop grown by
 280 smallholder farmers in the study area is maize, but in adequate soil fertility—which results
 281 from little to no external nutrient inputs—has prevented maize from producing at its
 282 maximum yield.

283 **Table 5: Limiting nutrients in each of the selected fields in the studied area in Kongwa**
 284 **district**

| Village | Owner | Soil pH _{H2O} | Limiting nutrients |
|---------|-------|---------------------------|-----------------------|
| Nghumbi | 1 | 7.05 | N |
| | 2 | 7.06 | N |
| | 3 | 7.1 | N |
| | 4 | 6.69 | N |
| | 5 | 6.99 | N and K |
| Mlali | 6 | 7.22 | N, P, Zn and K |
| | 7 | 7.63 | N, P, Zn and K |

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286 4 Maize Performance in the Study fields

287 4.1 Effect of field characteristics

288 In this section, the performance of maize was evaluated according to the specific field
 289 selection parameters, like the pH of the soil and the limiting nutrients that were found in each
 290 field. ANOVA was used to assess the P concentration, P absorption, P usage efficiency,
 291 biological productivity, and grain yield across fields (Table 6 and 7). Table 3.4 and 3.5 show
 292 that the farmer's field features significantly affected grain, biomass production, and P
 293 concentrations at a p-value of less than 0.001, while the primary treatment effect is
 294 insignificant at a p-value of less than 0.001, according to the ANOVA table.

Table6:Analysisofvariance(ANOVA)of Pconcentration,totalPuptake,andPuseefficiencyinmaizeasaffectedbyfarmer’sfieldcharacteristics, treatments,andtheirinteractions inKongwadistrict

| Sourceofd.fvariation | | Pconcentration | | | | TotalPuptake | | | | Puseefficiency | | | |
|----------------------|-----|----------------|--------|------|-------|--------------|--------|-------|--------|----------------|-------|------|-------|
| | | s.s. | m.s. | v.r. | Fpr. | s.s. | m.s. | v.r. | Fpr. | s.s. | m.s. | v.r. | Fpr. |
| Replication | 2 | 0.0516 | 0.0258 | 4.31 | | 68.79 | 34.39 | 1.92 | | 95550 | 47775 | 6.31 | |
| Farmer | 6 | 0.1016 | 0.0169 | 2.83 | 0.059 | 1800.46 | 300.08 | 16.77 | <0.001 | 300204 | 50034 | 6.61 | 0.003 |
| Residual | 12 | 0.0717 | 0.0060 | 0.85 | | 214.68 | 17.89 | 1.11 | | 90849 | 7571 | 0.78 | |
| Treatment | 5 | 0.0140 | 0.0028 | 0.4 | 0.848 | 88.22 | 17.64 | 1.09 | 0.373 | 21965 | 4393 | 0.45 | 0.811 |
| Farmer×Treatment | 30 | 0.0927 | 0.0031 | 0.44 | 0.993 | 302.17 | 10.07 | 0.62 | 0.924 | 201465 | 6715 | 0.69 | 0.869 |
| Residual | 70 | 0.49 | 0.007 | | | 1131.86 | 16.17 | | | 681168 | 9731 | | |
| Total | 125 | 0.82158 | | | | 3606.18 | | | | 1391202 | | | |

Key:d.f.=degreesoffreedom; s.s.=sumofsquares; m.s.= meansumofsquares; v.r.= variance;Fpr.=test-Fprobability

Table 7: Analysis of variance (ANOVA) of biological yield and grain yield in maize as affected by farmer’s field characteristics, treatments,and theirinteractionsinKongwadistrict

| Sourceofvariation | d.f. | Measuredvariablesinmaize andstatisticalparameters | | | | | | | | | | | |
|-------------------|------|---|-------|-------|--------|-----------------|-------|-------|--------|------|------|------|------|
| | | Biologicalyield | | | | Maizegrainyield | | | | | | | |
| | | s.s. | m.s. | v.r. | Fpr. | s.s. | m.s. | v.r. | Fpr. | s.s. | m.s. | v.r. | Fpr. |
| Replication | 2 | 4.513 | 2.26 | 1.1 | | 0.15 | 0.07 | 0.19 | | | | | |
| Farmer | 6 | 218.26 | 36.38 | 17.78 | <0.001 | 88.74 | 14.79 | 37.18 | <0.001 | | | | |
| Residual | 12 | 24.56 | 2.05 | 1.36 | | 4.77 | 0.40 | 0.9 | | | | | |
| Treatment | 5 | 3.66 | 0.73 | 0.49 | 0.785 | 2.40 | 0.48 | 1.09 | 0.375 | | | | |
| Farmer×Treatment | 30 | 49.66 | 1.66 | 1.1 | 0.364 | 13.86 | 0.46 | 1.05 | 0.427 | | | | |
| Residual | 70 | 105.39 | 1.51 | | | 30.94 | 0.44 | | | | | | |
| Total | 125 | 406.03 | | | | 140.87 | | | | | | | |

Key:d.f.=degreesoffreedom; s.s.=sumofsquares; m.s.= meansumofsquares; v.r.= variance;Fpr.=test-Fprobability

The chosen fields' maize yields were as follows: farm No. 2 > No. 5 > No. 4 > No. 6 > No. 1 = No. 6 > No. 3. According to the above sequence, field No. 2 had the highest yield of maize of all the fields, with a significant difference ($P < 0.001$) from the other fields. This was indicated by the major effect of field features. Field No. 2 produced 4.0 t ha⁻¹ of grain (Table 7). Given that the soils in field No. 2 only contained one limiting component, N (Table 4), which was also well-corrected, the soil's favorable qualities were most likely the reason of the significantly high yield. The natural phosphorus content of the field was high, at 53.4 mg kg⁻¹.

1. Field No. 5 had the second-highest maize grain production (3.3 t ha⁻¹), with two limiting nutrients present there. N and K are among these nutrients; N was rectified. As shown in Table 3 (Przygocka et al., 2020), a little limitation of K (0.21), which was near to a moderate level, could be the likely cause of the notable lower yield at this field next to No. 2. Grain yields in fields No. 4 (2.6 t ha⁻¹) and No. 6 (2.2 t ha⁻¹) were the next highest yields, after field No. 5. Field No. 6 had four limiting nutrients, namely N, P, K, and Zn, whereas Field No. 4 had just one limiting nutrient, N (Table 4). It is suggested that the primary reason for field No. 6's low grain yield is these limiting minerals. Furthermore, the No. 1 and No. 7 fields had low grain yields (1.8 t ha⁻¹).

1). Field No. 1 contained just one limiting nutrient (N), but field No. 7 had four (N, P, K, and Zn).

The limiting nutrients in No. 7 field are thought to be the reasons for low grain yield while in field No. 1 would have been affected by bad weather (drought). Field No. 3 (1.5 t ha⁻¹) had the lowest grain yield and had a contrasting trend in that yield did not seem to relate to limiting nutrients since N was the only inadequate nutrient and was corrected. In this field the reason could presumably be lack of soil moisture caused by drought experienced in the experimental site during vegetative stage in the early January to mid-February and during flowering to grain filling in March (Fig. 2). According to Figure 2, soil moisture stress occurred during all times when evapotranspiration exceeded rainfall. The order of biological/straw yields was as follows: No. 2 > No. 4 > No. 5 > No. 6 > No. 3 > No. 7 > No. 1. Similar to the patterning in grain yield, this trend is also reflected in the fields' limiting nutrient sequence and number, with the exception of field No. 3, which is affected by low soil moisture. These results, with the exception of fields Nos. 3 and 5, are in line with the field characteristics classified according to the limiting nutrients. The trend in P uptake results was No. 2 ≈ No. 4 ≈ No. 5 ≈ No. 6 ≈ No. 3 ≈ No. 7 ≈ No. 1. The majority of the phosphorus in maize crops is found in the grains, as has been demonstrated and documented. This has an impact on the nutritional content of the grain, protein, and micronutrients (Wafula et al., 2018). According to Wu et al. (2015), the critical concentration of P in maize straw is 0.26% by mass, of which 60 to 80% is in the form of phytate. In maize straw, a P absorption of 15 to 30 kg ha⁻¹ is ideal (Assefa et al., 2021).

Field No. 2's modest increase in P concentration (i.e., 0.27%) is most likely the result of the soil's high native P content (53.5 mg kg⁻¹), which the plants may have absorbed early in the maize plant's growth. This condition was also noted in field No. 5, where the native P content of the soil was adequate (22.1 mg kg⁻¹) for the growth of maize. It was discovered that fields with lower native P concentrations, such as Nos. 6 and No. 7 (9.2 mg kg⁻¹), had low P concentrations of 0.07 to 0.17% and 0.07 to 0.17%, respectively. These results support the findings of Gomez-Munoz et al. (2018), who found that high native P increases grain-based crop yields. Other researchers have also noted comparable outcomes of increased crop yield on soil with high native P (e.g., Mehrvarz, 2008; Krey et al., 2013; Saleque et al., 2014; Leggett et al., 2015; Santana, 2016; Selvi, 2017; Raymond et al., 2019).

Apart from the maize grain yield data, there was a significant difference ($P=0.003$) in P usage efficiency between the fields, although no discernible trend was seen. These results indicate the heterogeneity in nutrient levels. In this study, PUE ranged from 195.1 to 351 kg kg⁻¹. According to a study by Baligaret al. (2001), PUE ranged extremely high, from 400 to 500 kg kg⁻¹. The favorable conditions in the study by Baligaret al. (2001) included sufficient soil moisture, which facilitated PSB function and ultimately led to strong plant absorption. This resulted in a high PUE. It is evident that the PUE data are quite low when comparing this range to those found in this investigation. Due to the drought that the experimental site experienced, poor dry matter yield is most likely what causes the usually low PUE levels. The dimensionless ratio of harvested P agricultural products (P yield) to the mass of all P inputs into the system during the specified period is known as phosphorus usage efficiency, or PUE (Chowdhury and Zhang, 2021). According to Chen and Graedel (2016), it is also known as the ratio of P input conversion into valuable plant exports, such as harvested crops. As per Baligaret al. (2001), dividing dry matter yield (kg) by nutrient accumulation/uptake (kg) is one way to represent the efficiency of phosphorus consumption in plants. In this study, PUE has been defined as follows. PUE is significant to the crop production system because it serves as a gauge for the agricultural production system's P management status and its effects on environmental preservation and food security.

4.2 Treatments influence on the performance of maize

Data on grain yield, biological yield, P concentration, straw uptake, and use efficiency are shown in Table 8.

Table 8: P concentration, Straw P uptake, and P use efficiency in maize as affected by farmer's field characteristics and treatments in the study sites in Kongwa district.

| Farmer's- No. | Treatments | Grain yield (tha ⁻¹) | Biological yield (tha ⁻¹) | P concentration (%) | Straw P uptake (kg ha ⁻¹) | P use efficiency (kg kg ⁻¹) |
|------------------|--------------------------------|-------------------------------------|---|---------------------------|--|---|
| 2 | | 4.0a | 5.6a | 0.12a | 10.81a | 246.8bcd |
| 4 | | 2.6c | 4.7ab | 0.08ab | 7.96ab | 219.4cd |
| 1 | | 1.8de | 1.5e | 0.08ab | 2.38d | 351.8a |
| 3 | | 1.5e | 3.0cd | 0.10ab | 5.18bcd | 195.1d |
| 5 | | 3.3b | 4.0bc | 0.09ab | 7.21abc | 272.1bc |
| 7 | | 1.8de | 2.2de | 0.11a | 4.03cd | 294.8ab |
| 6 | | 2.2cd | 3.4c | 0.07b | 5.48bcd | 296.1ab |
| S.E.D. | | 0.22 | 0.477 | 0.011 | 1.351 | 29 |
| P-value | | <0.001 | <0.001 | 0.059 | <0.001 | 0.003 |
| | P ₀ M ₀ | 2.3a | 3.5a | 0.10a | 5.82a | 256.4a |
| | P ₂₀ M ₀ | 2.4a | 3.4a | 0.07a | 9.85a | 280.6a |
| | P ₀ M _x | 2.7a | 3.6a | 0.09a | 6.59a | 267.5a |
| | P ₂₀ M _x | 2.4a | 3.3a | 0.07a | 5.48a | 271.5a |
| | P ₄₀ M _x | 2.5a | 3.8a | 0.08a | 7.05a | 247a |
| | P ₆₀ M _x | 2.4a | 3.4a | 0.09a | 5.78a | 285.3a |
| | S.E.D. | 0.2037 | 0.379 | 0.011 | 1.251 | 30.44 |
| | P-value | 0.441 | 0.785 | 0.848 | 0.373 | 0.811 |

Key: S.E.D. = Standard errors of differences of means. Means in each column bearing different letter(s) differ significantly at 5% error rate; otherwise, are not statistically different.

In this investigation, the experimental treatment combinations involved applying MPR at varying rates in conjunction with the PSB inoculum. The primary impact of the treatments was found to be negligible across all sites. The first possible explanation is that the study site experienced a drought that started three weeks after emergence and lasted for approximately 35 days in January and February, which resulted in low soil moisture levels, which prevented the P from MPR from dissolving properly (Fig. 2). Second, because of the low soil moisture content, using PSB as an inoculum did not significantly aid in the solubilization of P from MPR. Low soil moisture affects grain yield and total grain content because it decreases PSB activity and P diffusion in the soil and from the soil to plant roots (Frossard et al., 2000). In a field experiencing such a severe drought during the vegetative stage, inoculums sown into the soil would not be able to survive. According to Schutz (2018), in sufficient soil moisture, phosphate solubilizing bacteria have a strong effect when administered P fertilizer at rates ranging between 40 and 60 P kg Pha⁻¹.

Table 9: Data for grain yield, biological yield, P concentrations, P uptake and P use efficiency in maize as affected by the interactions between farmer's field characteristics and treatments

| Farmer's- No. | Treatments | Grain yield (tha ⁻¹) | Biological yield (tha ⁻¹) | P concentration (%) | Straw P uptake (kg ha ⁻¹) | P use efficiency (kg kg ⁻¹) |
|------------------|--------------------------------|--|---|---------------------------|---|---|
| 3 | P ₀ M ₀ | 1.11 | 2.9f-m | 0.13ab | 6.23f-j | 179.2bcd |
| 3 | P ₂₀ M ₀ | 1.4jkl | 2.6g-m | 0.13ab | 6.7f-j | 211.2bcd |
| 3 | P ₀ M _x | 1.8g-l | 3.4c-m | 0.17ab | 8.1e-j | 218bcd |
| 3 | P ₂₀ M _x | 1.4jkl | 3.2d-m | 0.13ab | 9.33c-j | 149.3d |
| 3 | P ₄₀ M _x | 1.9f-l | 3.0f-m | 0.13ab | 7.93e-j | 247.5bcd |
| 3 | P ₆₀ M _x | 1.3kl | 3.1e-m | 0.10ab | 8e-j | 165.7cd |
| 1 | P ₀ M ₀ | 1.4jkl | 1.1m | 0.17ab | 4.17j | 326.2a-d |
| 1 | P ₂₀ M ₀ | 1.9f-l | 1.6lm | 0.13ab | 5.33hij | 460.8a |
| 1 | P ₀ M _x | 1.7g-l | 1.5lm | 0.13ab | 4.97ij | 335.5a-d |
| 1 | P ₂₀ M _x | 1.6h-l | 1.4m | 0.17ab | 5.3hij | 293.4a-d |
| 1 | P ₄₀ M _x | 2.6c-k | 2.2j-m | 0.17ab | 7.57e-j | 358.2abc |
| 1 | P ₆₀ M _x | 1.4jkl | 1.2m | 0.13ab | 4.17j | 337a-d |
| 4 | P ₀ M ₀ | 2.9c-h | 4.6a-j | 0.13ab | 12.93a-i | 235.1bcd |
| 4 | P ₂₀ M ₀ | 2.1e-l | 4.2b-k | 0.17ab | 10.8b-j | 195.9bcd |
| 4 | P ₀ M _x | 2.8c-i | 5.3a-f | 0.17ab | 14.07a-g | 202.3bcd |
| 4 | P ₂₀ M _x | 2.9c-h | 4.7a-i | 0.13ab | 11.7a-j | 263.4bcd |
| 4 | P ₄₀ M _x | 2.4d-l | 5.0a-h | 0.20ab | 11.73a-j | 226.4bcd |
| 4 | P ₆₀ M _x | 2.7c-j | 4.6a-j | 0.17ab | 13.73a-g | 193.2bcd |
| 5 | P ₀ M ₀ | 3.0b-g | 4.2b-k | 0.20ab | 14.4a-f | 215.1bcd |
| 5 | P ₂₀ M ₀ | 3.2b-f | 3.6b-m | 0.17ab | 12a-j | 293.2a-d |
| 5 | P ₀ M _x | 3.9abc | 4.8a-i | 0.20ab | 14.93a-e | 261.3bcd |
| 5 | P ₂₀ M _x | 3.4a-e | 3.3c-m | 0.13ab | 10.27b-j | 329.4a-d |
| 5 | P ₄₀ M _x | 2.4d-l | 4.2b-k | 0.17ab | 13.63a-g | 201.1bcd |
| 5 | P ₆₀ M _x | 4.2ab | 3.9b-l | 0.17ab | 13.27a-h | 332.7a-d |
| 2 | P ₀ M ₀ | 4.2ab | 6.0ab | 0.20ab | 17.37abc | 248.6bcd |
| 2 | P ₂₀ M ₀ | 3.6a-d | 4.5a-j | 0.20ab | 13.2a-h | 307.7a-d |
| 2 | P ₀ M _x | 4.4a | 5.1a-g | 0.27a | 18.13ab | 257bcd |
| 2 | P ₂₀ M _x | 3.7abc | 5.7abc | 0.20ab | 16.93a-d | 225.1bcd |
| 2 | P ₄₀ M _x | 3.9abc | 5.6a-e | 0.23ab | 19.63a | 213bcd |
| 2 | P ₆₀ M _x | 3.8abc | 6.8a | 0.20ab | 17.23abc | 229.3bcd |
| | S.E.D. | 0.5389 | 1.031 | 0.06748 | 3.309 | 79.04 |
| | P-value | 0.427 | 0.364 | 0.993 | 0.924 | 0.869 |

Key: S.E.D. = Standard errors of differences of means. Means in each column bearing different letter(s) differ significantly at 5% error rate; otherwise, are not statistically different.

Table 10: Data for grain and biological yield, P concentration, straw P uptake, P use efficiency in maize as affected by the interactions between farmer's field characteristics and treatments

| Farmer's- No | Treatments | Grain yield | Biomass yield | P concentration | Straw P uptake | P use efficiency |
|-----------------|--------------------------------|----------------------|----------------------|--------------------|------------------------|------------------------|
| | | (tha ⁻¹) | (tha ⁻¹) | (%) | (kg ha ⁻¹) | (kg kg ⁻¹) |
| 6 | P ₀ M ₀ | 1.9f-l | 2.3i-m | 0.13ab | 6.1g-j | 317.1a-d |
| 6 | P ₂₀ M ₀ | 2.4d-l | 4.1b-k | 0.13ab | 11.87a-j | 220.2bcd |
| 6 | P ₀ M _x | 2.3e-l | 2.5h-m | 0.10ab | 7.37e-j | 318.7a-d |
| 6 | P ₂₀ M _x | 2.1e-l | 3.3c-m | 0.07b | 7.17e-j | 317.3a-d |
| 6 | P ₄₀ M _x | 2.7c-j | 5.6a-d | 0.13ab | 13.77a-g | 233.7bcd |
| 6 | P ₆₀ M _x | 2.0f-l | 2.6g-m | 0.17ab | 8.27e-j | 369.9ab |
| 7 | P ₀ M ₀ | 1.6h-l | 3.4c-m | 0.10ab | 7.27e-j | 273.5a-d |
| 7 | P ₂₀ M ₀ | 2.1e-l | 3.0f-m | 0.13ab | 9.07d-j | 275a-d |
| 7 | P ₀ M _x | 2.2e-l | 2.4i-m | 0.17ab | 10.67b-j | 279.9a-d |
| 7 | P ₂₀ M _x | 1.7h-l | 1.6lm | 0.17ab | 5.3hij | 322.4a-d |
| 7 | P ₄₀ M _x | 1.5i-l | 1.2m | 0.17ab | 7.9e-j | 249bcd |
| 7 | P ₆₀ M _x | 1.7g-l | 1.8klm | 0.07b | 4.83ij | 369.2ab |
| | S.E.D. | 0.5389 | 1.031 | 0.067 | 3.309 | 79.04 |
| | P-value | 0.427 | 0.364 | 0.993 | 0.924 | 0.869 |

Key: S.E.D. = Standard errors of differences of means. Means in each column bearing different letter(s) differ significantly at 5% error rate; otherwise, are not statistically different.

Grain yields were marginally greater in field No. 2 with P₀M_x (4.4 t ha⁻¹) treatments than in P₄₀M_x (3.9 t ha⁻¹) treatments, but there was no significant ($P = 0.427$) interaction effect between field features and treatments on the measured variables (Table 9 and 10). Based on this discovery and in accordance with earlier findings documented in the main effect of treatment, natural P, rather than the given MPR and inoculant combinations, was the primary source of P in the crops. These results simply that the nutritional status of the soil, particularly P, N, and K, affects maize productivity in the study area. An intriguing discovery revealed by interaction effects is that soil and the quantity of P released or supplied to the soil determine how maize responds to native P inputs in the study area. These results are consistent with the findings of Frossard et al. (2000), who observed that a variety of factors, including soil conditions, influence crop performance.

5 Conclusions and Recommendations

In every field where maize crop production systems were evaluated, nitrogen was the limiting nutrient element. In addition to N, other limiting nutrients were noted in fields No. 5 and No. 6, which had limited potassium (K) and P, respectively, and field No. 7, which had limited P, K, and Zn. With the highest grain production, biological yield, and total P uptake among all other fields, field No. 2's maize performance was statistically the best, according to the major effect of the field's features. In addition, there were notable variations in P usage efficiency among the fields, which could be attributed to variations in nutrient levels. The primary outcome of the treatments showed that using P or inoculants independently did not increase grain yield in a way that was encouraging. Grain yields in field No. 2 were significantly greater

than in other fields because of its high native P in soils, even though there were no significant interactions on effects of field features and treatment combination on the measured variables in maize. These results imply that native P and soil nutrient status, particularly N and K, affect maize productivity in the studied area.

Before the technique is suggested for farmers to use, we advise doing additional field trials spanning multiple seasons to determine the significance of solubilizing bacteria in boosting P availability from MPR for increase of maize yield.

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