

# ***Original Research Article***

## **Evaluating the effect of boron & molybdenum micronutrient application methods on growth in groundnut (*Arachis hypogaea* L.)**

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### **ABSTRACT**

The present investigation was conducted during Rabi 2019 season at the central farm, Regional Research and Technology Transfer Station, Coastal Zone, OUAT, Bhubaneswar. The fresh seeds of groundnut variety Devi (ICGV - 91114) were used as sowing material. The field was laid out in randomized block design with twelve treatments and three replications. The treatments were T<sub>1</sub> (Seed priming with Borax solution @ 0.01%), T<sub>2</sub> (Seed priming with Borax solution @ 0.05%), T<sub>3</sub> (Seed priming with Borax solution @ 0.10%), T<sub>4</sub> (Foliar application of Borax @ 100 ppm), T<sub>5</sub> (Soil application of Borax @ 10 kg/ha), T<sub>6</sub> (Seed priming with Sodium Molybdate @ 0.25 g/l for 2 hours), T<sub>7</sub> (Seed priming with Sodium Molybdate @ 0.50 g/l for 2 hours), T<sub>8</sub> (Seed priming with Sodium Molybdate @ 0.75 g/l for 2 hours), T<sub>9</sub> (Foliar application of Sodium Molybdate @ 50 ppm), T<sub>10</sub> (Soil application of Sodium Molybdate @ 1.5 kg/ha), T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) and T<sub>12</sub> (Control). The results revealed that different growth phases have shown profound effect. Morphological characters of the T<sub>11</sub> showed significant increase in the plant height, number of branches, number of leaves and leaf area. Physiological analysis of treatments revealed T<sub>11</sub> significantly higher increment as compare to other treatments in the dry weight of the plant and leaf area index. It was revealed that the treatment T<sub>11</sub> has shown as superior performance over other treatments in groundnut.

*Keywords: Groundnut, Seed priming, foliar application, Soil application, Morphological characters, Boron, Molybdenum.*

### **1. INTRODUCTION**

Groundnut (*Arachis hypogaea* L.), is a leguminous crop plant which is widely cultivated in the tropics and subtropics between 40°N and 40°S latitudes. It is native to South America (Brazil). Groundnut is also known as "KING OF OILSEEDS", which is used in confectionary nut flour production, protein and peanut milk [1]. Seed oil content of groundnut is 46-52 %, while carbohydrates and protein percentage are 18 and 30% respectively. Pure form of groundnut seed oil is mainly use as cooking oil, as a salad oil and margarine while raw oil is use in soap manufacturing. Groundnut is not only an important oilseed crop of India but also an important agricultural export commodity.

There are reasons and factors that lead to low yield in groundnut, one of the major reasons for low yield is a deficiency of micronutrients. The introduction of high yielding varieties for exploiting their potential led to a widespread deficiency of micronutrients, especially for boron and molybdenum. Completely neglecting the application of micronutrients results in a reduction of yield and export quality of groundnut. Another prime reasons for the low yield of the groundnut is because of intensive cropping which leads to a deficiency of secondary and micronutrients. Seed priming has been found to involve complex molecular pathways, including changes in gene expression, enzyme activity, and metabolite synthesis[2]. Several studies have investigated the mechanisms involved in seed priming, and have identified key pathways that play a role in this process.

One of the pathways involved in seed priming is the regulation of stress-responsive genes. Seed priming has been found to induce the expression of stress-responsive genes, such as those involved in the synthesis of protective molecules like antioxidants and osmoprotectants[3]. These molecules help protect the seed against various stresses, such as dehydration, oxidative stress, and high salinity, which can occur during germination. Another pathway involved in seed priming is the modulation of

hormone signaling pathways. Seed priming has been shown to alter the levels of various plant hormones, such as abscisic acid (ABA), gibberellins (GA), and cytokinins (CK) [2]. The modulation of hormone signaling pathways can affect various aspects of seed germination, such as the regulation of dormancy, cell division, and differentiation. In addition to these pathways, seed priming has also been found to affect enzyme activity and metabolite synthesis. For example, priming has been shown to enhance the activity of enzymes involved in the breakdown of stored reserves, such as  $\alpha$ -amylase and protease [2]. Priming can also enhance the synthesis of metabolites, such as polyamines, which are involved in various processes, such as stress tolerance and cell proliferation [4]. Overall, the pathways involved in seed priming are complex and interconnected, and involve changes in gene expression, enzyme activity, and metabolite synthesis. Understanding these pathways is important for developing more effective seed priming techniques and for improving crop performance under stressful conditions.

Pre-sowing priming induces a particular physiological status in seeds and has emerged as a promising strategy to improve plant behavior in the field. Seed priming helps in various physiological, biochemical and molecular mechanisms which results in enhanced crop yield [5]. There is a strong interest for farmers and seed companies to find suitable cheap priming treatments but also to precisely identify the agronomical properties improved as a result of priming in cultivated species. Foliar spraying is beneficial because it gives fast result than application of chemical fertilizers to the soil. Foliar treatment is economical and less wasteful as compared to direct use where it absorb by soil particle and that is why less available to root system. Foliar nutrition can minimize lag time between application and uptake by the crop. Moreover, nutrition solution is absorbed by trichomes or through rupture in cuticle which can able to reach at the target site.

Boron is one of the key micronutrients required by the plant for its growth and development. It has known that peanut need for boron is a bit higher than other legumes crops. Boron has the ability to increase photosynthetic and enzymatic activity in the peanut plant. It also involves in protein and nucleic acid metabolism. Boron maintains the structural integrity of the plant and protects the plasma membrane from external damage [6]. It also helps in sugar transport, division and elongation of a cell, involve in the transport of auxin and metabolism in roots and improve ATPase activity [7]. Boron deficiency causes pollen grain germination. Pollen tube growth and viability of pollen grains are also affected due to boron deficiency [8]. Boron is the only element which is available in soil solution and plant can easily take up from the soil as a non-ionized molecule at a suitable pH range [9].

Molybdenum involves in nitrogenase - an enzyme that is responsible for the nitrogen fixation process by bacteria symbiotically with legumes crops. It also plays a key role in nitrogen metabolism, protein synthesis, and Sulphur metabolism. Molybdenum is required in pollen formation so Mo deficient plant will cause effect in their fruits and pollen grains formation. It is also important for the absorption and translocation of iron in the plants [10]. In India, Micronutrient application to crop plants is done mainly as a basal application in soil or through foliar sprays. Here in this research, application of B and Mo through seed priming, foliar spray, soil application as well as their combined effect is assed and their result on growth parameters are shown here.

## **2. METHODOLOGY**

### **2.1 Experimental Site and Layout**

The present investigation was conducted during Rabi 2019 season at the central farm, Regional Research and Technology Transfer Station, Coastal Zone, OUAT, Bhubaneswar. The farm is situated at N 20°15.885' and E 85°48.024' with an accuracy of  $\pm 9$  feet as recorded in GARMIN GPSmap 76CSx device. The fresh seeds of groundnut variety Devi (ICGV - 91114) were used as sowing material. The details of the experiment site is as shown in table 1.

**Table 1.** Details of the experiment site

Location	Central Research Station, OUAT
Elevation	25.9 meter
Land form	Medium land
Slope	0.1 N-S
Parent material	Laterite
Ground water table	1.5 m
Erosion	el
Surface drainage	Well-drained

The field was laid out in randomized block design with twelve treatments and three replications with groundnut. The treatments were T<sub>1</sub> (Seed priming with Borax solution @ 0.01%), T<sub>2</sub> (Seed priming with Borax solution @ 0.05%), T<sub>3</sub> (Seed priming with Borax solution @ 0.10%), T<sub>4</sub> (Foliar application of Borax @ 100 ppm), T<sub>5</sub> (Soil application of Borax @ 10 kg/ha), T<sub>6</sub> (Seed priming with Sodium Molybdate @ 0.25 g/l for 2 hours), T<sub>7</sub> (Seed priming with Sodium Molybdate @ 0.50 g/l for 2 hours), T<sub>8</sub> (Seed priming with Sodium Molybdate @ 0.75 g/l for 2 hours), T<sub>9</sub> (Foliar application of Sodium Molybdate @ 50 ppm), T<sub>10</sub> (Soil application of Sodium Molybdate @ 1.5 kg/ha), T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) and T<sub>12</sub> (Control). Seed priming treatments were given for the period a two hours, followed by the drying of the seeds. The treated seeds were sown in the field to evaluate the efficacy of the treatment.

## 2.2 Climatic Conditions

Climate is humid throughout the season. Values of relative humidity are about 80% to 87% in the morning and 3% to 85% in the afternoon during the southwest monsoon season. Mean maximum temperature of coldest month (Dec.) was 28.9 °C. Mean minimum temperature of coldest month (Dec.) was 15.4 °C. In the summer and post monsoon seasons, there were moderate clouds. Sky was clear or lightly clouded in the winter. Thunderstorms were observed throughout the year except December. Fog occurred during winter season.

- Max. precipitation – 21 mm (26 April, 2020)
- Max. relative humidity – 90.39 (3 Jan, 2020)
- Max. Temperature – >38.1 °C (End of March and April)
- Min. Temperature – 9.6 (29 December, 2019)
- Max. wind speed – 6.0 m/s

## 2.3 Measurement and Analysis

Various morpho-physiological parameters at various growth stages were recorded. Five plants were selected randomly from each plot and their height was measured from ground level to the tip of the plant. Observation was recorded at 30, 60 and 90 days after sowing (DAS) of the seeds. The average height of the 5 plants were considered for the final record. The number of branches per plant was counted from five plants from each plot at 30, 60 and 90 DAS, respectively. While, Number of leaves was calculated only at 30 DAS. Five trifoliate leaves from each treatment were collected from sample plants and leaf area was measured by leaf area meter and the average was calculated in square centimeter. Leaf area was recorded at 60 and 90 DAS, respectively. This leaf area values were worked out to calculate the leaf area index (LAI) using the formula written in the table 2. LAI is defined as the one-sided green leaf area per unit ground surface area. LAI was calculated by dividing leaf area per plant by the land area occupied by a single plant. Dry matter contents were recorded at 30, 60 and 90

DAS by uprooting five plants randomly from each treatment. These samples were oven dried at 60 °C till a constant weight was recorded on per plant basis. Crop growth rate (CGR) is defined as the rate of dry matter production per unit land area per unit time. The observations were recorded at different intervals of 60 and 90 days after sowing. Relative growth rate (RGR) is growth rate relative to size. It is also called as the exponential growth rate or the continuous growth rate. The observations were recorded at 60 and 90 days after sowing. Net assimilation rate (NAR) is a value that relates plant productivity to plant size. It is obtained by dividing the rate of increase in dry weight by leaf size (usually leaf area).

**Table 2.** Formulas used to assess the morpho-physiological growth parameters in the experiment

Sr. No.	Growth parameters	Formula used	References
1	Leaf area index	$LAI = \frac{\text{Leaf area}}{\text{Ground area}}$	Sestak et al.[11]
2	Crop Growth Rate (g dm <sup>2</sup> day <sup>-1</sup> )	$CGR = \frac{W_2 - W_1}{t_2 - t_1} \times \frac{1}{A}$	Watson[12]
3	Relative Growth Rate (mg g <sup>-1</sup> day <sup>-1</sup> )	$RGR = \frac{\ln W_2 - \ln W_1}{t_2 - t_1}$	Blackman[13]
4	Net Assimilation Rate (g m <sup>-2</sup> day <sup>-1</sup> )	$NAR = \frac{W_2 - W_1}{A_2 - A_1} \times \frac{\log A_2 - \log A_1}{t_2 - t_1}$	Radford[14]

Where,

W<sub>1</sub>= Dry matter production plant<sup>-1</sup>(g) at time t<sub>1</sub>  
W<sub>2</sub>= Dry matter production plant<sup>-1</sup>(g) at time t<sub>2</sub>  
A = Ground area covered by the plant (dm<sup>-2</sup>)  
A<sub>1</sub>= Ground area covered by the plant (dm<sup>-2</sup>) at time t<sub>1</sub>  
A<sub>2</sub>= Ground area covered by the plant (dm<sup>-2</sup>) at time t<sub>2</sub>  
ln = natural logarithm  
log A<sub>2</sub> – log A<sub>1</sub>= natural log difference leaf area

### 3. RESULTS AND DISCUSSION

#### 3.1 Plant Height

The investigation on the impact of seed priming, basal and foliar application of micronutrients was studied for the plant. The plant height of five randomly selected plants in each plot were measured at 30 DAS, 60 DAS, and 90 DAS and the mean values have been presented in table 3. Plant height reported non-significant at 30 DAS. It was observed that treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) has shown the significant higher plant height of 33.0 cm, and 42.60 cm at 60 DAS and 90 DAS, respectively. Among all the treatments, T<sub>11</sub> was observed 21.32 %, and 16.85 % higher than the control at 60 DAS and 90 DAS, respectively. The next significantly higher was observed by treatment T<sub>5</sub> having a value of 32.50 cm and 42.60 cm at 60 and 90 DAS, respectively. It was observed that treatment T<sub>5</sub> shown the second-highest percent increase which was 19.49 % and 17.68 % as compared to control at all three intervals of 60 and 90 DAS, respectively. The lowest plant height was recorded in the untreated control.

Plant height is one of the important characters of the plant which indicates the vegetative growth and the changes in this character may brought by different seed priming, basal and foliar application practices which definitely influence this growth parameter. The height of plant was more in T<sub>11</sub> treatment due to the application of additional Boron and Molybdenum through seed priming treatments.

Similar results of increase in plant height with combined application of boron and molybdenum were reported by Dugger[8], Duyingjonget al.[15] and Nayak et al.[16].

**Table 3. : Effect of micronutrients on plant height at 30, 60 and 90 DAS**

Treatments	30 DAS (cm)	60 DAS (cm)	90 DAS (cm)
T <sub>1</sub> - Seed priming with B @ 0.01 %	7.87	28.90	38.10
T <sub>2</sub> - Seed priming with B @ 0.05%	7.90	29.40	39.43
T <sub>3</sub> - Seed priming with B @ 0.10%	8.10	30.00	40.70
T <sub>4</sub> - Foliar app. of B @ 100 ppm (20 DAS and 30 DAS)	8.20	31.60	41.70
T <sub>5</sub> - Soil application of Borax @ 10 kg/ha	8.37	32.50	42.60
T <sub>6</sub> - Seed priming with Mo @ 0.25 g/l for 2 hrs	7.70	28.30	36.70
T <sub>7</sub> - Seed priming with Mo @ 0.50 g/l for 2 hrs	7.83	29.20	36.80
T <sub>8</sub> - Seed priming with Mo @ 0.75 g/l for 2 hrs	7.80	29.80	38.30
T <sub>9</sub> - Foliar app. of Mo 50 ppm (20 DAS and 30 DAS)	7.70	30.00	39.17
T <sub>10</sub> - Soil app. of Mo @ 1.5 kg/ha	7.80	30.50	39.60
T <sub>11</sub> - T <sub>2</sub> + T <sub>7</sub>	8.63	33.00	42.30
T <sub>12</sub> - CONTROL	7.60	27.20	36.20
SE.m (±)	0.43	1.11	1.45
C.D. (0.05)	NS	3.26	4.27
C.V.	9.42	6.41	6.41

Where, B=Boron, Mo=Molybdenum, ppm= parts per million

### 3.2 Number of Branches

The study examined the effects of seed priming, basal, and foliar application of micronutrients on the plant. The number of branches of 10 randomly selected plants in each plot was counted at 30, 60, and 90 DAS and the mean values have been presented in table 4. It was observed that treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) has shown significantly higher number of branches viz. 6.20 and 9.17 at 30 DAS and 90 DAS, respectively. The number of branches recorded significantly higher under treatment T<sub>5</sub> at 60 DAS. The lowest number of branches was counted in untreated control (T<sub>12</sub>) at 30 DAS and 90 DAS. At 60 DAS, treatment T<sub>1</sub> recorded the lowest number of branches. It was observed from table 4 that treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) has reported significantly higher number of branches. Sahu[17] also observed similar results when treated with combined application of Zn, B and Mo.

**Table 4. Effect of micronutrients on number of branches at 30, 60 and 90 DAS**

Treatments	30 DAS	60 DAS	90 DAS
T <sub>1</sub> - Seed priming with B @ 0.01 %	5.30	7.40	7.68
T <sub>2</sub> - Seed priming with B @ 0.05%	5.40	7.60	7.80
T <sub>3</sub> - Seed priming with B @ 0.10%	5.50	7.90	8.20
T <sub>4</sub> - Foliar app. of B @ 100 ppm (20 DAS and 30 DAS)	5.50	8.10	8.30
T <sub>5</sub> - Soil application of Borax @ 10 kg/ha	5.80	8.30	8.40
T <sub>6</sub> - Seed priming with Mo @ 0.25 g/l for 2 hrs	5.20	7.60	7.80
T <sub>7</sub> - Seed priming with Mo @ 0.50 g/l for 2 hrs	5.23	7.70	7.90
T <sub>8</sub> - Seed priming with Mo @ 0.75 g/l for 2 hrs	5.40	7.80	8.00
T <sub>9</sub> - Foliar app. of Mo 50 ppm (20 DAS and 30 DAS)	5.60	7.80	8.10
T <sub>10</sub> - Soil app. of Mo @ 1.5 kg/ha	5.70	7.90	8.10
T <sub>11</sub> - T <sub>2</sub> + T <sub>7</sub>	6.20	8.20	9.17
T <sub>12</sub> - CONTROL	5.20	7.70	7.57
SE.m (±)	0.17	0.17	0.26
C.D. (0.05)	0.49	0.51	0.76
C.V.	5.27	3.87	5.59

### 3.3 Leaf Area and Number of Leaves

Plant leaf area was calculated at different growth stages (60 DAS and 90 DAS) presented in table 5. It was observed that significantly higher mean value of leaf area shown by treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) of 1350.0 cm<sup>2</sup> plant<sup>-1</sup> and 1500.0 cm<sup>2</sup> plant<sup>-1</sup> at 60 DAS and 90 DAS, respectively. The lowest leaf area was observed in untreated control (T<sub>12</sub>) at 60 DAS and 90 DAS. The investigation on the impact of seed priming, basal and foliar application of micronutrients was made for number of leaves per plant at 30 DAS and the observations have been depicted in table 5. The result showed that the significantly higher mean value regarding number of leaves were recorded at 30 DAS in the treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) of 10.48 leaves per plant. The lowest number of leaves was recorded in untreated control (T<sub>12</sub>) at 30 DAS.

The data on leaf area of groundnut observed at interval of 60 DAS and 90 DAS that presented in table 5 as influenced by different levels and method of boron and molybdenum application. This increment in leaf area is also supported by the finding of Geethanjali et al. [18], Sharma et al. [19] and Sushmitha et al. [20]. The more the number of the leaves, more photosynthesis will occur and more productivity of the plant. The data on number of leaves are presented in table 5. The result showed that the significantly higher number of leaves were recorded at 30 DAS in the treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) of 10.48 numbers per plant. This result of the combined seed priming of boron and molybdenum are agreed with Sharma et al. [19].

**Table 5.** Effect of micronutrients on leaf area at 30, and 60 DAS and number of leaves at 30 DAS

Treatments	Leaf area (cm <sup>2</sup> plant <sup>-1</sup> )		Number of leaves
	30 DAS	60 DAS	30 DAS
T <sub>1</sub> - Seed priming with B @ 0.01 %	1134.00	1421.00	9.41
T <sub>2</sub> - Seed priming with B @ 0.05%	1182.67	1456.00	9.62
T <sub>3</sub> - Seed priming with B @ 0.10%	1236.00	1461.00	9.84
T <sub>4</sub> - Foliar app. of B @ 100 ppm (20 DAS and 30 DAS)	1261.00	1484.00	10.26
T <sub>5</sub> - Soil application of Borax @ 10 kg/ha	1311.00	1498.00	10.18
T <sub>6</sub> - Seed priming with Mo @ 0.25 g/l for 2 hrs	1211.67	1346.00	9.43
T <sub>7</sub> - Seed priming with Mo @ 0.50 g/l for 2 hrs	1261.00	1351.00	9.82
T <sub>8</sub> - Seed priming with Mo @ 0.75 g/l for 2 hrs	1273.00	1394.00	10.11
T <sub>9</sub> - Foliar app. of Mo 50 ppm (20 DAS and 30 DAS)	1281.00	1477.00	10.37
T <sub>10</sub> - Soil app. of Mo @ 1.5 kg/ha	1290.00	1468.00	10.09
T <sub>11</sub> - T <sub>2</sub> + T <sub>7</sub>	1350.00	1500.00	10.48
T <sub>12</sub> - CONTROL	1090.00	1320.00	9.41
SE.m (±)	49.54	42.21	0.25
C.D. (0.05)	145.31	123.81	0.75
C.V.	6.92	5.11	4.46

### 3.4 Leaf Area Index

The calculation of Leaf Area Index (LAI) is presented in Table 6, specifically for the growth stages occurring at 60 and 90 days after sowing (DAS). The significantly higher LAI was obtained under treatment T<sub>11</sub> which was 4.5 (60 DAS) and 5.0 (90 DAS). The lowest mean value of LAI was observed in untreated control (T<sub>12</sub>) that is 3.63 (60 DAS) and 4.40 (90 DAS).

Leaf area index is a measure of leafiness per unit ground area and relates to the photosynthetic efficiency level. It is a significant variable for growth and yield as it is an important determinant of light interception and transpiration. In table 6, the LAI data determined at various growth phases were provided. Results revealed that T<sub>11</sub> shows significantly increased value compared to control in both

intervals. This corroborates with the results of Geethanjali et al.[18], Sharma et al.[19] and Sushmitha et al.[20].

**Table 6.**Effect of micronutrients on leaf area index at 60 and 90 DAS

Treatments	Leaf area index	
	60 DAS	90 DAS
T <sub>1</sub> - Seed priming with B @ 0.01 %	3.78	4.74
T <sub>2</sub> - Seed priming with B @ 0.05%	3.94	4.85
T <sub>3</sub> - Seed priming with B @ 0.10%	4.12	4.87
T <sub>4</sub> - Foliar app. of B @ 100 ppm (20 DAS and 30 DAS)	4.20	4.95
T <sub>5</sub> - Soil application of Borax @ 10 kg/ha	4.37	4.99
T <sub>6</sub> - Seed priming with Mo @ 0.25 g/l for 2 hrs	4.04	4.49
T <sub>7</sub> - Seed priming with Mo @ 0.50 g/l for 2 hrs	4.20	4.50
T <sub>8</sub> - Seed priming with Mo @ 0.75 g/l for 2 hrs	4.24	4.65
T <sub>9</sub> - Foliar app. of Mo 50 ppm (20 DAS and 30 DAS)	4.27	4.92
T <sub>10</sub> - Soil app. of Mo @ 1.5 kg/ha	4.30	4.89
T <sub>11</sub> - T <sub>2</sub> + T <sub>7</sub>	4.50	5.00
T <sub>12</sub> - CONTROL	3.63	4.40
SE.m (±)	0.16	0.14
C.D. (0.05)	0.48	0.41
C.V.	6.92	5.11

### 3.5 Dry Weight

The dry weight of the plant of each treatment was estimated and their mean values were presented in table 7. From the results, it was found that the treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) shows the significantly higher values of the dry weight of the plant at all intervals (30, 60, and 90 DAS) which was 2.51, 18.67, and 26.41 g plant<sup>-1</sup>, respectively. The treatment T<sub>11</sub> reported 31.59 %, 15.84 %, and 22.04 % increment as compared to untreated control. The lowest mean dry weight of the plant was reported by untreated control (T<sub>12</sub>) that is 1.91, 16.12, and 21.64 g plant<sup>-1</sup> at 30, 60, and 90 DAS, respectively.

Since plants have a high composition of water and the level of water in a plant will depend on the amount of water in its environment, using dry weight as a measure of plant growth tends to be more reliable. Data related to dry weight at different growth interval are presented in table 7. Boron is known as helpful in sugar transport, division and elongation of cell, involve in transport of auxin and metabolism in roots and improve ATPase activity [7]. Overall boron application helps in enhancing growth of the plant. Application of molybdenum also leads to increase in dry weight and growth of the plant. This statement is in tune with reports of Singh et al.[21].

**Table 7.**Effect of micronutrients on dry weight at 30, 60 and 90 DAS

Treatments	Dry weight (g plant <sup>-1</sup> )		
	30 DAS	60 DAS	90 DAS
T <sub>1</sub> - Seed priming with B @ 0.01 %	1.98	16.28	22.02
T <sub>2</sub> - Seed priming with B @ 0.05%	2.15	16.49	22.86
T <sub>3</sub> - Seed priming with B @ 0.10%	2.23	17.07	23.67
T <sub>4</sub> - Foliar app. of B @ 100 ppm (20 DAS and 30 DAS)	2.12	16.57	24.46
T <sub>5</sub> - Soil application of Borax @ 10 kg/ha	2.45	17.92	25.64
T <sub>6</sub> - Seed priming with Mo @ 0.25 g/l for 2 hrs	1.96	16.98	22.50
T <sub>7</sub> - Seed priming with Mo @ 0.50 g/l for 2 hrs	2.05	16.44	22.01
T <sub>8</sub> - Seed priming with Mo @ 0.75 g/l for 2 hrs	2.15	16.52	22.81
T <sub>9</sub> - Foliar app. of Mo 50 ppm (20 DAS and 30 DAS)	2.17	16.78	23.98
T <sub>10</sub> - Soil app. of Mo @ 1.5 kg/ha	2.07	17.84	25.57
T <sub>11</sub> - T <sub>2</sub> + T <sub>7</sub>	2.51	18.67	26.41
T <sub>12</sub> - CONTROL	1.91	16.12	21.64

SE.m (±)	0.12	0.51	1.06
C.D. (0.05)	0.36	1.51	3.11
C.V.	9.81	5.25	7.76

### 3.6 Crop Growth Rate

The crop growth rate (CGR) was calculated for 60 DAS to 90 DAS growth period and shown in the form of a bar chart in figure 1 for the easy comparison of the data. Treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) was observed as a maximum crop growth rate among all the treatments. Treatment T<sub>11</sub> calculated 0.3023 g dm<sup>-2</sup> day<sup>-1</sup> as compared to 0.1841 g dm<sup>-2</sup> day<sup>-1</sup> in control T<sub>12</sub>. The lowest value of crop growth rate was recorded by untreated control T<sub>12</sub>.

The crop growth rate and relative growth rate (RGR) was calculated for 60 DAS to 90 DAS growth period and shown in the form of a bar chart in figure 1 and 2 that reveals that Treatment T<sub>11</sub> as a maximum growth rate. It may be due to that priming with micronutrients enable seeds to uptake water at a faster rate and encourages it for different metabolic processes and Boron is already reported that it activate key enzymes such as phosphorylase, α-amylase etc. which might stimulate cell elongation, meristematic growth and increase chlorophyll content leading to overall growth. Geethanjali et al. (2015)[18] found similar results on the relative growth rate of groundnut.

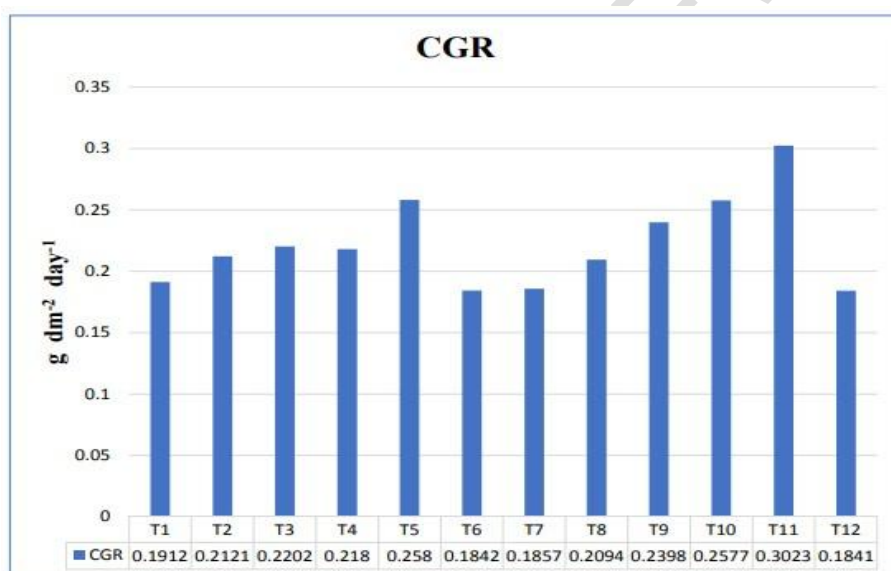


Fig. 1. Crop growth rate as influenced by the different levels and methods of borax and sodium molybdate applications

### 3.7 Relative Growth Rate

Treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) was observed as a maximum relative growth rate among all the treatments. Treatment T<sub>11</sub> calculated 14.52mg g<sup>-1</sup> day<sup>-1</sup> as compared to 9.823mg g<sup>-1</sup> day<sup>-1</sup> in control T<sub>12</sub>. The lowest value of relative growth rate was recorded by untreated control T<sub>12</sub>. Balla et al.[22] reported improved CGR, RGR, and yield with the combination treatment of boron and molybdenum in groundnut.

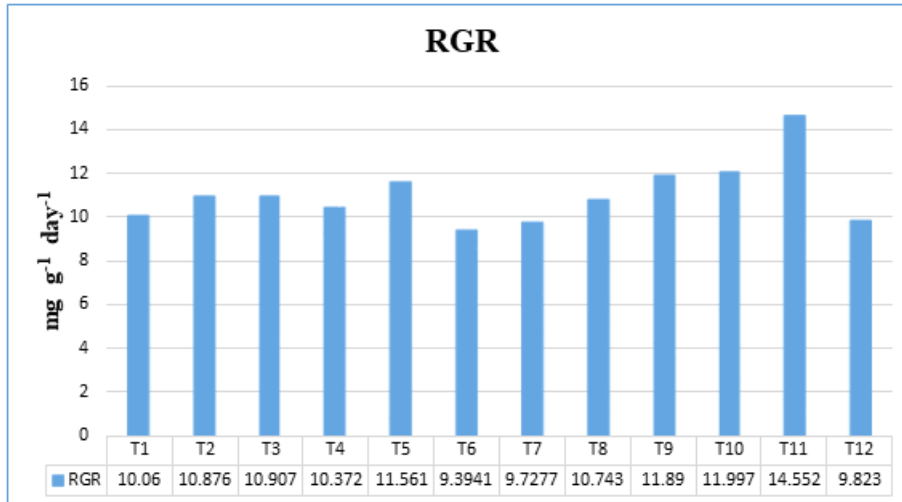


Fig. 2. Relative growth rate as influenced by the different levels and methods of borax and sodium molybdate applications

### 3.8 Net Assimilation Rate

Treatment T<sub>11</sub> (T<sub>2</sub> + T<sub>7</sub>) found as maximum net assimilation rate among all the treatments. Treatment T<sub>11</sub> calculated 2.2351 g m<sup>-2</sup> day<sup>-1</sup> as compared to 1.9876 g m<sup>-2</sup> day<sup>-1</sup> in control T<sub>12</sub>. The lowest value of net assimilation rate of 1.9207 g m<sup>-2</sup> day<sup>-1</sup> was recorded by treatment T<sub>1</sub>.

Borate ion forms complex with numerous polyhydroxy and related substances. It assists in transport of growth regulators within the plant and helps in polymerization of phenolic compounds. Molybdenum is vital for N<sub>2</sub>-fixation, which has assured greater N supply to the crop and activation of enzyme nitrate reductase, a soluble molybdoflavoprotein in plant [23], leading to increased crop growth and production.

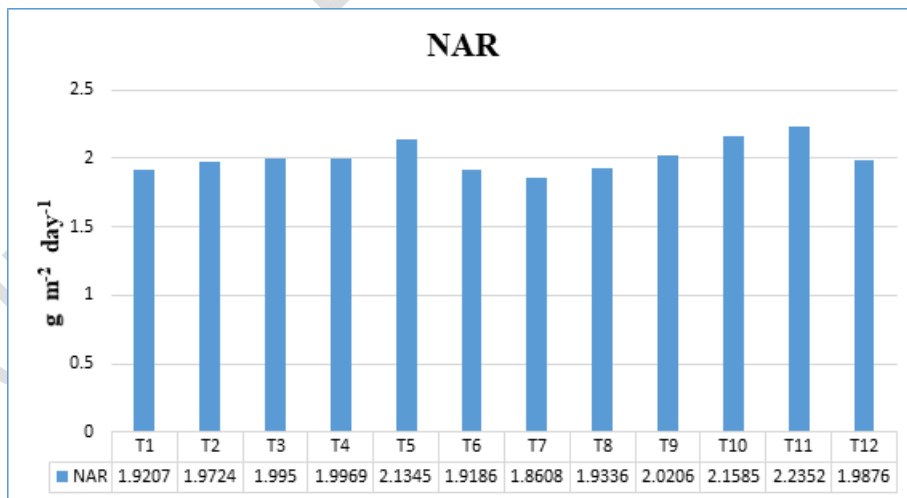


Fig. 3. Net assimilation rate as influenced by the different levels and methods of borax and sodium molybdate applications

#### 4. CONCLUSION

The application of micronutrients is of paramount importance in groundnut crop cultivation. Notably, Boron and Molybdenum play significant roles in the growth and physiological processes of this crop. Based on the findings of this study, it can be deduced that seed priming with micronutrients, specifically Borax (0.05%) + Sodium Molybdate (0.50 g/l) for a duration of 2 hours, followed by subsequent drying to reach the original moisture content before sowing, proved to be an effective method for delivering micronutrients to the crops, ultimately resulting in enhanced crop growth.

#### REFERENCES

1. Woodroof JG. Nuts as a source of edible oil. *Technological Advances in Improved and Alternative Sources of Lipids*. Springer US, Boston, MA. 1994;150-176.
2. Kaur S, Gupta AK, Kaur N. Seed priming: a comprehensive review on its techniques, applications, and benefits. *Plant Growth Regul.* 2020;90:401-436.
3. El-Maarouf-Bouteau H, Sajjad Y, Bazin J, Langlade N, Cristescu SM, Balzergue S, Baudouin E, Bailly C. Reactive oxygen species, abscisic acid and ethylene interact to regulate sunflower seed germination. *Plant Cell Environ.* 2015;38:364-374.
4. Roychoudhury A, Basu S, Sarkar SN, Sengupta DN. Comparative physiological and molecular responses of a common aromatic indica rice cultivar to high salinity with non-aromatic indica rice cultivars. *Plant Cell Rep.* 2008;27:1395-1410. <https://doi.org/10.1007/s00299-008-0556-3>
5. Diya A, Beena R, Jayalekshmy VG. Physiological, Biochemical and Molecular Mechanisms of Seed Priming: A Review. *Legume Research - An International Journal.* 2021;1-8. <https://doi.org/10.18805/LR-4638>
6. Ismail C, Volkar R. Boron nutrition of crops in relation to yield and quality- a review. *Plant Soil.* 1997;193:71-73.
7. Gupta UC. *Handbook of Plant Nutrition*. Barker A V., Pilbeam DJ (eds). Taylor and Francis Groups, New York, 2007;241-268.
8. Dugger WM. *Functional Aspects of Boron in Plants. Trace Elements in the Environment*. American Chemical Society. 1973;112-129.
9. Oertli JJ, Grgurevic E. Effect of pH on the Absorption of Boron by Excised Barley Roots. *Agron J.* 1975;67:278-280. <https://doi.org/https://doi.org/10.2134/agronj1975.00021962006700020028x>
10. Subba R, Adinarayana V. *Molybdenum Research in Agricultural production*. FDCO, New Delhi. 1995;115-117.
11. Sestak Z, Catský J, Jarvis PG. *Plant photosynthetic production. Manual of methods*. Plant photosynthetic production Manual of methods, 1971.
12. Watson DJ. The physiological basis of variation in yield. *Advances in agronomy.* 1952;4:101-145.
13. Blackman VH. The compound interest law and plant growth. *Ann Bot.* 1919; 33:353-360.
14. Radford PJ. Growth analysis formulae-their use and abuse 1. *Crop Sci.* 1967;7:171-175
15. Duyingqiong Q, Xinrong L, Jianghua H, Zhoyao H, Xiaohong Z. Effect of B and Mo on the growth, development and yield of peanut. *Plant NutrFert Sci.* 2002;8:233-235.

16. Nayak SC, Sarangi D, Mishra GC, Rout DP. Response of groundnut to secondary and micronutrients. 2009.
17. Sahu SK. Available boron and molybdenum status of some lateritic and alluvial soil of Orissa growing groundnut and its response to molybdenum on lateritic soil. *Environment and Ecology*. 1998;16:772-775.
18. Geethanjali K, Ashoka RY, Narasimha RKL, Madhuvani P. Effect of foliar application of ethrel and boron on morphological parameters, growth characteristics and yield in groundnut (*Arachis hypogaea* L.). *International Journal of Food, Agriculture and Veterinary Sciences*. 2015;5:120-125.
19. Sharma MK, Jat RA, Sree Ganesh S. Effect of Micronutrients and Biofertilisers on Morpho-physiological Parameters and Productivity of Summer Groundnut *Arachis hypogaea* L. *Indian Journal of Fertilisers*. 2017;13:56-59.
20. Sushmitha BP, Hanumanthappa DC, Mudalagiriappa, Kalyanamurthy KN, Shree Harsha Kumar SS. 2018. Response of groundnut (*Arachis hypogaea* L.) to nano boron. *Green Farming*. 2018;9(5):925-927.
21. Singh AL, Chaudhari V, Basu MS. Boron Nutrition and Boron Application in Crops: Boron Deficiency and its Nutrition of Groundnut in India. *Advances in Plant and Animal Boron Nutrition: Proceedings of the 3rd International Symposium on all Aspects of Plant and Animal Boron Nutrition*. Springer. 2007;149-162.
22. Balla H, Singh V, Tiwari D, Shaik MA and Jonnagorla L. Effect of boron and molybdenum on growth rate and yield of groundnut (*Arachis hypogaea* L.). *J PharmacognPhytochem*. 2020;9(6):1416-1419.
23. Tandon HLS. Sulphur fertilizer for Indian Agriculture-A guide Book Fertilizer Development and Consultation organization, New Delhi. 1995: 1-23.