

Effect of Bio-agents on Hatching and Mortality of Root –Knot Nematode (*Meloidogyne javanica*)

Comment [A1]: Revise as the reference also has a same kind of title....

ABSTRACT:

Investigations were carried out in *in vitro* condition to evaluate the antagonistic effect of fungal and bacterial bio-agents. For effective management of root-knot nematode experiment was conducted in laboratory condition using the culture filtrate (5, 10 and 20 per cent concentrate) of bio-agents (*viz.*, *Trichoderma viride*, *Trichoderma harzianum*, *Purpureocillium lilacinum*, *Pochonia chlamydosporia* and *Pseudomonas fluorescens*). Experimental results showed that all the tested bio-agents significantly reduced the per cent hatched juveniles and increased the per cent mortality of juveniles. Among the tested bio-agents *T. viride* was found most effective treatment with minimum per cent hatched juveniles and maximum per cent mortality on eggs and juveniles of *Meloidogyne javanica*.

Key Words: Root-knot nematode, *Meloidogyne* spp., Bio-agent, Hatching, Mortality.

INTRODUCTION

Nematodes are hidden enemy of plant. Among the nematodes, root-knot nematode, *Meloidogyne* spp. is an important pest of vegetables all around the world and causes severe damage to brinjal crop [Kamran *et al* 2010] in India, the root-knot nematode was first reported by [Barber 1901] on tea (*Thea sinensis*) roots from Devala territory of Tamil Nadu, South India. [In Rajasthan, [Arya 1957] reported it for the first time from Jodhpur, while [Yadav and Naik 1966] found *Meloidogyne* spp. and its wide distribution in the soils of Rajasthan. The nematode infected plant shows reduced root system with less feeder roots [Anwar and Mckenry 2010]. Overall, plant-parasitic nematodes cause 21.3% crop losses amounting to Rs. 102040 million (1.58 billion USD) annually; the losses in 19 horticultural crops were assessed at Rs. 50,225 million, while for 11 field crops it was estimated at Rs. 51,815 million. Among the vegetable crops comparatively more losses recorded in tomato (Rs. 6035.2 million), brinjal (Rs. 3499.12 million) and okra (2480.86 million) [Kumar *et al* 2020]. Management of nematode is much difficult as compare to other pathogen because nematodes mainly attack underground parts of

Comment [A2]: Check sentence

plants [Sikora and Fernandez 2005]. The nematodes control mainly depended on synthetic nematicides [Akthar and Malik 2000]. Although, nematicides are efficient and fast acting, yet they are currently being reappraised as there are relatively unaffordable to many small-scale farmers. The potential negative effect on environment and ineffectiveness after prolonged use have led to a total ban or restricted use of most chemical nematicides and an urgent need for safe and more effective alternatives [Zukerman and Esnard 1994]. Biological control appears the alternative strategies for management of plant parasitic nematodes in the soil [Agyarko and Asante 2005] [Collange *et al* 2011]. Application of microorganisms antagonistic to root-knot nematodes or compounds produced by these microbes could provide an additional option for managing the damage caused by root-knot nematodes. Fungi and bacteria are among the most dominant soil-borne groups in natural soil ecosystem and some of them have shown great potential as biological control agents for root-knot nematodes [Kerry 2000]. So, the present investigation was carried out on test the effect of different bio- agents' cultural filtrate on root-knot nematode.

Comment [A3]: Control of nematodes

MATERIALS AND METHODS

The efficacy of bio-agents tested against hatching and mortality of juveniles of root-knot nematode in laboratory condition.

Preparation and maintenance of pure culture of *M. javanica*: -

M. javanica infected brinjal plant were uprooted from the pure culture plots of Division of Nematology, RARI, Durgapura, Jaipur and brought to the laboratory. The roots were first rinsed carefully in water to remove adhering soil particles. Egg masses were carefully detached from roots, and then egg masses were kept in distilled water in watch glasses at room temperature for hatching.

Preparation of culture filtrates of fungal and bacterial bio-agents: -

The potato dextrose agar (PDA) for fungal agents and nutrient agar (NA) for bacterial agents were prepared, inoculated with respective bio-agents in 100 ml conical flasks followed by incubation at 30 °C in a shaker for 48 hrs. The cultures were centrifuged at 6000 rpm for 20-30 minutes. The supernatant was kept as a stock solution of cent per cent concentration. Next grade of 5, 10 and 20 per cent concentration were made by dilution with distilled water.

One ml of sterilized double distilled water was added on fully grown fresh mother culture of bio-agents *Trichoderma viride*, *Trichoderma harzianum*, *Purpureocillium lilacinum*, *Pochonia chlamydosporia* and *Pseudomonas fluorescens* than scraped with a spade to produce slurry and then transferred to 99 ml of distilled water to prepare a suspension that was referred as stock solution. From this stock solution 10 ml suspension was transferred into 90 ml distilled water that was referred as 2nd dilution suspension

Effect of bio-agents on root-knot nematode in laboratory condition:

Effect of bio-agent on hatching of *M. javanica*

To test the effect of bio-agents on nematode hatching the experiment was conducted in completely randomized design (CRD) with four replications. One healthy average sized egg mass of *M. javanica* were collected from infected brinjal roots and kept in glass cavity block (1 egg mass/ cavity block) containing 3 ml of bio-agents formulation 5, 10 and 20 per cent (*Trichoderma viride*, *Trichoderma harzianum*, *Purpureocillium lilacinum*, *Pochonia chlamydosporia* and *Pseudomonas fluorescens*) concentrations respectively. A distilled water control was also maintained simultaneously. Number of juveniles hatched were recorded after 24, 48 and 72hrs under binocular microscope.

The per cent of egg hatching was calculated by using formula:

$$\text{Hatching per cent} = (C/T) \times 100 \%$$

Where,

C = number of parasitized nematodes after 24, 48 and 72hrs exposure.

T = total number of nematodes in a cavity block.

Effect of bio-agent on mortality of *M. javanica*

The experiment on test the effect of bio-agents on nematode mortality was laid out in completely randomized design (CRD) with four replications. Freshly hatched second stage juveniles of *M. javanica* were transferred to different cavity blocks (10 juveniles/ cavity block) containing 3 ml of bio-agents formulation *i.e.*, 5, 10 and 20 per cent (*Trichoderma viride*, *Trichoderma harzianum*, *Purpureocillium lilacinum*, *Pochonia chlamydosporia* and *Pseudomonas fluorescens*) concentrations respectively. A distilled water control was maintained

simultaneously. Per cent juvenile mortality rate was counted at intervals of 24, 48 and 72 hrs. The dead juveniles attained the shape of straight line and the mortality was ensured by touching the juvenile with a fine needle.

The per cent mortality was calculated by using formula:

$$\text{Per cent mortality} = (C/T) \times 100$$

Where,

C = number of parasitized nematodes after 24, 48 and 72hrs exposure.

T = total number of nematodes in a cavity block.

STATICAL ANALYSIS.

After completion of experiment, data were statically analyzed for interpretation of finding. The critical deference was calculated for comparison of treatment for significant at 5 % level of significance. Summary table along with SEM± and CD were worked out and presented in chapter “Experimental Results”.

Comment [A4]: ?

RESULT AND DISCUSSION

Effect of bio-agents were tested on the hatching and mortality of *M. javanica* under laboratory conditions. 5, 10 and 20 per cent concentration of bio-agents culture filtrate were used for hatching and mortality of *M. javanica*. Number of hatched larvae out from egg masses and dead larvae were observed after 24, 48 and 72 hrs. Data presented in table-1 and 2 showed that all the bio-agents significantly decreased number of hatched juveniles and increased mortality compared to untreated check. The minimum number of hatched juveniles and maximum number of dead juveniles were observed with the *T. viride* at 20 per cent concentration followed *P. lilacinum* and *P. fluorescens* respectively at 20 per cent concentration. While, maximum number of hatched juveniles and no mortality were observed in untreated check

Effect of bio-agents on per cent hatching:-

Hatching at 5 per cent: - Data presented in table-1 showed that all the bio-agents significantly decreased number of hatched juveniles compared to untreated check. The minimum number of hatched juveniles (24.38per cent) were observed with the *T. viride* after 72 hrs followed by *P. lilacinum* (25.39per cent) and *P. fluorescens* respectively (26.88per cent) after 72 hrs. While, maximum number of hatched juveniles (75.78per cent) were observed in untreated check.

Hatching at 10 per cent: - Data revealed that the minimum number of hatched juveniles (20.70per cent) were observed with the *T. viride* after 72 hrs followed *P. lilacinum* (21.17per cent) and *P. fluorescens* respectively (23.44per cent) after 72 hrs. Whereas, maximum number of hatched juveniles (75.78per cent) were observed in untreated check. All the bio-agents significantly decreased number of hatched juveniles over untreated check.

Hatching at 20 per cent: - Data showed that *T. viride* after 72 hrs was observed most effective treatment with minimum number of hatched juveniles (14.53) followed *P. lilacinum* (16.41per cent) and *P. fluorescens* respectively (20.00 per cent) after 72 hrs. However, untreated check was observed least effective treatment with maximum number of hatched juveniles (75.78 per cent).

Table 1. Effect of bio-agent on per cent hatching of root-knot nematode juveniles

Concentration	5%			10%			20%			
	Treatment	24 h	48 h	72 h	24 h	48 h	72 h	24 h	48 h	72 h
<i>T. viride</i>		19.45	21.48	24.38	16.72	18.28	20.70	12.03	13.59	14.53
<i>T. harzianum</i>		25.00	26.41	28.44	22.97	24.84	26.25	20.31	22.03	23.36
<i>P. lilacinum</i>		21.56	23.59	25.39	18.20	19.77	21.17	13.75	15.00	16.41
<i>P. chlamydosporia</i>		23.91	25.39	27.50	21.72	23.05	24.30	18.52	20.08	21.56
<i>P. fluorescens</i>		21.95	24.06	26.88	20.16	21.72	23.44	16.72	18.44	20.00
Control		31.95	55.16	75.78	31.95	55.16	75.78	31.95	55.16	75.78
SEM±		1.16	1.479	1.965	1.34	1.630	2.048	1.324	1.680	2.044
CD 5%		3.45	4.39	5.837	3.97	4.844	6.084	3.93	4.993	6.072
CV %		9.69	10.08	11.31	12.18	12.02	12.82	14.02	13.97	14.29

* Average of four replications

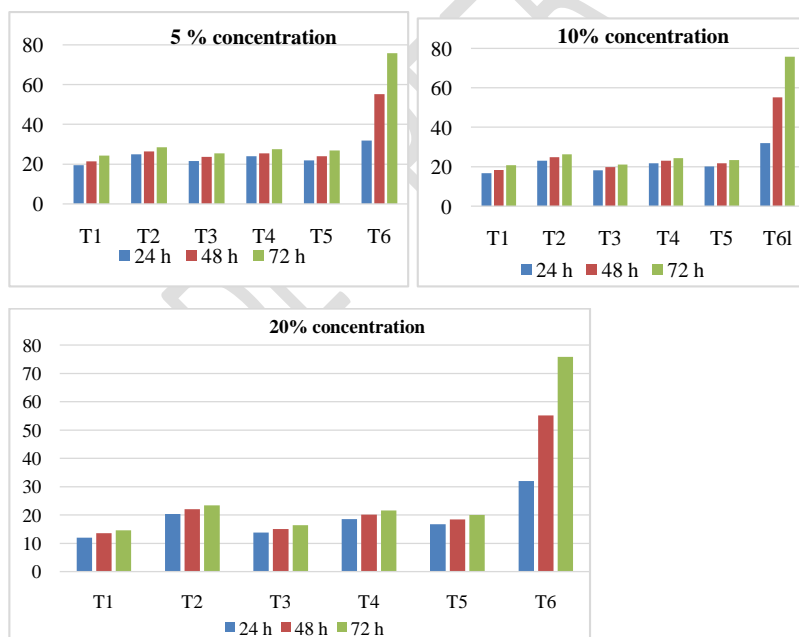


Fig.1 Effect of bio-agents on per cent hatching of root-knot nematode juveniles

Effect of bio-agents on per cent mortality: -

Mortality at 5 per cent: - All the bio-agents significantly increased per cent mortality of juveniles over untreated check. Data presented in table-2 showed that the maximum juvenile's mortality per cent (52.50 per cent) were observed with the *T. viride* after 72 hrs followed by *P. lilacinum* (40.00 per cent) and *P. fluorescens* (35.00 per cent). While, juvenile's mortality was not observed in untreated check.

Mortality at 10 per cent: - The maximum per cent mortality of juveniles (60.00 per cent) were observed with the *T. viride* after 72 hrs followed by *P. lilacinum* (47.00 per cent) and *P. fluorescens* (42.00 per cent). Whereas, juvenile's mortality was not observed in untreated check. All the bio-agents significantly increased per cent mortality of juveniles over untreated check.

Mortality at 20 per cent: - Data presented in table-2 showed that all the bio-agents significantly increased per cent mortality of juveniles over untreated check. Among all the tested bio-agents *T. viride* was observed best treatment with maximum per cent mortality of juveniles (80.00 per cent) after 72hrs followed by *P. lilacinum* (65.00 per cent) and *P. fluorescens* (60.00 per cent). However, untreated check was observed least effective with no mortality of juveniles.

Table 2 Effect of bio-agent on root-knot nematode juveniles per cent mortality

Concentration	5.00%			10.00%			20.00%		
	24 h	48 h	72 h	24 h	48 h	72 h	24 h	48 h	72 h
<i>T. viride</i>	35.00	45.00	52.50	45.00	50.00	60.00	55.00	65.00	80.00
<i>T. harzianum</i>	12.50	17.50	22.50	15.00	25.00	27.50	25.00	32.50	40.00
<i>P. lilacinum</i>	25.00	32.50	40.00	35.00	40.00	47.50	45.00	50.00	65.00
<i>P. chlamydozporia</i>	17.50	22.50	27.50	25.00	32.50	37.50	32.50	42.50	55.00
<i>P. fluorescens</i>	22.50	30.00	35.00	32.50	40.00	42.50	42.50	45.00	60.00

Control	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
SEm±	2.43	2.700	3.173	2.57	3.281	3.118	2.500	2.764	4.082
CD 5%	7.22	8.02	9.428	7.63	9.748	9.264	7.43	8.212	12.130
CV %	25.92	21.97	21.45	20.21	21.00	17.40	15.00	14.11	16.33
* Average of four replications									

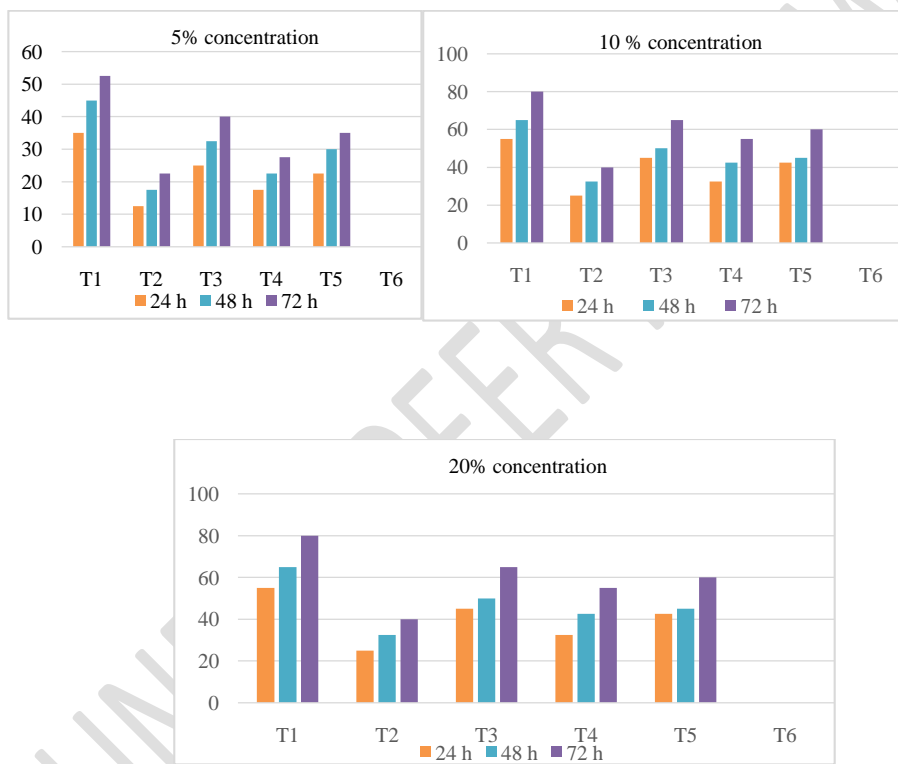


Fig 2. Effect of bio-agents on per cent mortality of root-knot nematode juveniles.

DISCUSSION

The results obtained were similar with the findings of [Singh and Mathur *et al* 2010] found culture filtrates of *A. strictum* was very effective against the nematode in regards to egg parasitism, egg hatching inhibition and mortality compared to controls. *A. strictum* was caused greater mortality of the second stage juveniles (J₂). *A. terreus* did not show egg parasitism but was found to be highly toxic against second stage juveniles (J₂) causing high mortality. [Sharma *et al* 2014] showed that *Paecilomyces lilacinus* culture filtrate from Karanj cake medium killed 100% *Meloidogyne incognita* larvae while only 78.28% mortality was recorded by Czapeck-Dox filtrate within 12 h of exposure. [Annapurna *et al* 2018] showed *T. harzianum* was caused highest egg hatch inhibition and juvenile mortality of *M. incognita*. The culture filtrate of *T. harzianum* showed the highest activity with a LC50 value at 96 hrs of exposure. [Popal *et al* 2020] also studied the efficacy of different bio-agents on egg-hatching and larval mortality of *M. incognita* after 24, 48 and 72 hours of inoculation. *T. viride* was superior over *P. penetrans* as against untreated check in regards to egg-hatching. While, regarding larval mortality of the nematode *P. lilacinus* and *P. penetrans* were at par with each other and were superior over *T. viride*. [Kumari *et al* 2021] tested bio-control agents on hatching and juvenile mortality of *M. incognita* after 24, 48, 72 and 96 hrs exposure period and found that *T. viride* and *T. asperellum* were at par and significantly effective on hatching inhibition and larval mortality of *M. incognita*. Maximum inhibition of egg hatching and larval mortality of root-knot nematode recorded with *T. harzianum* after 72 hours of incubation. *T. harzianum* and *T. viride* were able to colonize *M. incognita* eggs and second stage juveniles and female. *In vitro* studies demonstrated that both tested isolates were effective in causing nematode mortality compared with the control [Jegathambai *et al* 2011] [Naserinasab *et al* 2011] [Devi and Bora *et al* 2018] [Prasad and Ravichandra *et al* 2018]. All bio-agent showed distortion of juveniles was observed in most of the eggs in the present study. The well-known observations suggested that the inhibitory effect of the bio-agents on hatching of the nematode larvae may be due to the nematotoxic metabolites like chitinase and other lytic enzymes like proteases and lipases that cause break down of egg shell and facilitate egg penetration for successful establishment [Li *et al* 2005] [Kalele *et al* 2010]

J. Study reported that mortality of root-knot nematode increased with increase in exposure time as well as the concentration of culture filtrate. Mortality of second stage juveniles by these bio-agents might be due to release of lytic enzymes like chitinases, lipases and acetic acid in the filtrates that cause breakdown of nematode cuticle proteins [Annapurna *et al* 2018].

CONCLUSION

Number of hatched larvae out from egg masses and dead larvae were observed after 24, 48 and 72 hrs. All the bio-agents significantly decreased number of hatched juveniles and increased mortality. The minimum number of hatched juveniles and maximum number of dead juveniles were observed with the *T. viride* at 20 per cent concentration followed *P. lilacinum* and *P. fluorescens* respectively at 20 per cent concentration after 72 hours.

Comment [A5]: Needs revision
This looks like results.....

References :-

- Agyarko, K. and Asante, J.S. (2005). Nematode dynamics in soil amended with neem leaves and poultry manure. *Asian Journal of Plant Sciences* **4**: 426-428.
- Akhtar, M. and Malik, A. (2000). Roles of organic soil amendments and soil organisms in the biological control of plant-parasitic nematodes. A review. *Bioresource Technology* **74**: 35-47.
- Annapurna, M., Bhagawati, B. and Kurulkar, U. (2018). *In-vitro* efficacy of native fungal bio-agents against *Meloidogyne incognita*. *International Journal of Current Microbiology and Applied Sciences* **7(11)**: 396-410.
- Anwar, S. and Mckenry, MV. (2010). Incidence and reproduction of *Meloidogyne incognita* on vegetable crop Genotype. *Pakistan Journal of Zoology* **42**: 135-141.
- Arya, H.C. (1957). Root-knot diseases of tomatoes in Jodhpur. *Science and Culture* **22**:391-393.
- Barber, C.A. (1901). A tea *ell* worm disease in South India. *Dept. Land Record, Madras Agricultural Branch* 2. Bull. No. **45**.
- Collange, B., Navarrete, M., Peyre, G., Mateille, T. and Tchamitchian, M. (2011). Root-knot nematode (*Meloidogyne*) management in vegetable crop production. The challenge of an agronomic system analysis. *Crop Protection* **30**: 1251–1262.

- Devi Gitanjali and Bora, L.C. (2018). Effect of some bio-control agents against root-knot Nematode (*Meloidogyne incognita* race2). *International Journal of Environment, Agriculture and Biotechnology* **3(5)**: 52-58.
- Jegathambai, V., Wilson Wijeratnam, R.S. and Wijesudera, R.L.C. (2011). Effect of *Trichoderma viride* strain NRRL6418 and *Trichoderma harzianum* (Hypocera lixil TWC1) on *Livistonia rotundifolia* root-knot nematode *Meloidogyne incognita*. *Journal of Entomology* **8**:229-239.
- Kalele, D.N., Affokpon, A., Coosemans, J. and Kimenju, J. W. (2010). Suppression of root-knot nematodes in tomato and cucumber using biological control agents. *African Journal of Horticulture Sciences* **3**:72-80.
- Kamran, M., Anwar, S.A., Javed, M., Khan, S.A. and Sahi, GH. (2010). Incidence of root-knot nematodes on tomato in Sargodha, Punjab, Pakistan. *Pakistan Journal of Nematology* **28**: 253-262.
- Kerry, B.R. (2000). Rhizosphere interactions and the exploitation of microbial agents for the biological control of plant-parasitic nematodes. *Annual Review of Phytopathology* **38**: 423-441.
- Kumar, Vinod, Khan M.R., Walia, R.K. (2020) Crop loss estimations due to plant-parasitic nematodes in major crops in India. *The National Academy of Sciences, India*.
- Kumari, Manju; Sharma, M.K.; Baheti, B.L.; Nama, C.P. and Chandrawat, B.S. (2021) Reaction of locally isolated bio-agents on hatching and mortality of root-knot nematode, *Meloidogyne incognita*. *Journal of Entomology and Zoology Studies*. **9(2)**: 1276-1281
- Li B, Xie GL, Soad A, Coosemans J. (2005). Suppression of *Meloidogyne javanica* by antagonistic and plant growth promoting rhizobacteria. *Journal of Zhejiang University Science*. **B**, 496-501.
- Naserinasab, F., Sahebani, N. and Etebarian, H.R. (2011). Biological control of *Meloidogyne javanica* by *Trichoderma harzianum* BI and salicylic acid on tomato. *African Journal of Food Science* **5(3)**:276-280.
- Popal, N. A. (2020). Evaluation of bio-agents on egg hatching and larval mortality of root-knot nematode (*Meloidogyne incognita*). *International Journal of Research in Agricultural Sciences* **7 (2)**: 2348 – 3997.

- Prasad, G.R. and Ravichandra, N.G. (2018). Evaluation of indigenous antagonists on inhibition of egg hatching and larval mortality of *Meloidogyne incognita* infecting carrot under in vitro. *International Journal of Current Microbiology and Applied Sciences* **7(5)** 917-924.
- Sharma, Abhishek, Sharma, Satyawati and Dalela, Manu (2014). Nematicidal activity of *Paecilomyces lilacinus* 6029 cultured on karanja cake medium. *Microbial Pathogenesis* **75**: 16-20.
- Sikora, R.A. and Fernández, E. (2005). Nematode's parasites of vegetables. In: Liuc, M., Sikora, R.A., Bridge, J. (Eds.), *Plant Parasitic Nematodes in Subtropical and Tropical Agriculture*. CAB International, Wallingford (GBR) pp. 319–392.
- Singh, Satyandra and Mathur, Nita (2010). In vitro studies of antagonistic fungi against the root-knot nematode, *Meloidogyne incognita*. *Biocontrol Science and Technology* **20 (3)**: 275-282.
- Yadav, B.S. and Naik, S.M.P. (1966). Nematodes associated with economic plants of South East Plateau of Rajasthan. *Labdev Journal of Science and Technology* **4**: 184-186.
- Zukerman, B.M. and Esnard, J. (1994). Biological control of plant nematodes current status and hypothesis. *Japanese Journal of Nematology* **24**: 1–13.