

Screening of different genotypes of Niger (*Guizotia abyssinica* Cass) against major insect pests

Abstract

An experiment was conducted during Pre-Rabi season in the year 2020-21, at Project Coordinating Unit Sesame and Niger, Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur (M.P.). Ecompassed of 21 promising genotypes of Niger which were screened for resistance/susceptibility against whiteflies, leafhoppers, and capsule fly. The genotypes JNS-28, JNS-9, JNC-6, NRS-1805, IGPN-15-02 and NRS-1513 were found promising against whitefly. The genotypes JNS-2017-8, IGPN-15-02, NRS-1513, NRS-1511 and JNS-2017-1 were found promising against the incidence of leafhopper. The genotypes DNS-19-24, JNS-2017-8, JNS-2016-1115, NRS-1513 and NRS-1807 were found promising against capsule fly.

Keyword: Screening, susceptibility, genotypes JNS-2017-8 and 21 promising genotypes.

1. Introduction

Niger (*Guizotia abyssinica*) is an economically important edible oilseed crop. It is crucial minor oilseed crop having therapeutic properties mainly grown in India and Ethiopia. Niger is a diploid ($2n = 2x = 30$), oil crop species belongs to Asteraceae family. It is an annual crop with a capitulum that consists of six to eight fertile female ray florets and 40–60 hermaphroditic disk florets. The corymbs cymes of heads, 5 broadly ovate-obovate outer involucre leaves, 5-nerved paleae and bigger achenes, are its main discriminating characters from other *Guizotia* species. Niger is a strictly outcrossing species with a sporophytic self-incompatibility mechanism and is mainly pollinated by insects. The earliest name given to this plant was *Verbesina oleifera*. Niger (*Guizotia abyssinica* L.f. Cass) is an important minor oilseed crop of India which developed mainly in hilly and tribal areas. It is nutritious in terms of its 38 to 42% of quality oil with 18 to 20% protein in the seed. It is the lifeline of tribal agriculture and economy in India (Panday *et al.*, 2014). An integrated pest management strategy, to increase the efficiency of fertilizer use, is being developed. Yield losses, due to various diseases and insect pests in different states, have been assessed and a

management schedule for major insect pests (niger caterpillar, niger capsule fly, cutworms, semi looper, Bihar hairy caterpillar, and aphids). One of the hurdle in the production of Niger is the massive damage caused by several insect pests under certain agro production situations. Niger crop is attacked by 24 of insect-pest species from germination to till the harvest in both Ethiopia and India. The niger capsule fly is the most serious insect pest, both in Ethiopia and India. Cultivar resistance has been identified as the most desirable and economic tactics in the management of insect pests and is the best alternative to synthetic insecticides, providing an eco-friendly, environmentally safe strategy for fruitful management of insect pests. It could be united into ecologically sound integrated pest management programmes. Before evolving insect pest management programme for a specific agro-ecosystem, it is obligatory to have basic information on the occurrence of the pests in relation to weather parameters which helps in discovering appropriate time of action and suitable method of management.

2. Materials and methods

Experiment was designed in Randomized Block Design (RBD) with three replications, with 21 genotypes. For the screening of different promising genotypes against major insect pests 21 promising genotypes were grown in three replications. weekly observations were taken started from 15 days after sowing till the harvesting of crop. From each genotype five plants were selected randomly for recording the observations. The population of sucking insect pests (leafhopper and whitefly) were observed by counting the number of nymphs and adult on three leaves/plant from top, middle and bottom of the plants. The population of capsule fly was recorded by counting the number of adult per plant.

3. Results and discussion

The aggregate of average population data of whitefly showed that all the genotypes differed significantly in respect to record the whitefly population per three leaves per plant. Among the screened genotypes the minimal population of whitefly (1.32 whitefly/three leaves) was reported from the genotype JNS-28 succeeded by (1.63 whitefly/three leaves) JNC-6 and JNS-9 (1.43 whitefly/three leaves) while the highest population (2.78 whitefly/three leaves) was recorded from the genotype JNS-2017-1 followed by (2.29 whitefly/three leaves) NRS-1511 and IGPN-2004-1 (2.28 whitefly/three leaves). The genotypes JNS-28, JNS-9, JNC-6, NRS-1805, IGPN-15-02, NRS-1513, JNS-2016-1115, DNS-19-24, IGPN-17-02 and JNS-2016-1413 were found at par to each other in respect to record the lowest population of whitefly. Present findings are in conformity with the findings of Tembhre (2005), whoscreened 78 germplasm lines of niger for resistance/susceptibility against major insect pests and reported that the entry N-KEC-7 was found highly susceptible against *B. tabaci*. Present findings are also supported by the findings of Ranganathaet al., (2016) that the genotype N-6 and N-11 were found to be tolerant against whitefly.

Table 1: Response of different genotypes of niger against whitefly

S. No.	Genotypes	White fly population/three leaves Days after sowing (DAS)								Overall mean
		15	22	29	36	43	50	57	64	
1	JNS-2017-1	3.27 (1.94)	3.16 (1.91)	3.37 (1.96)	3.00 (1.86)	3.24 (1.93)	2.90 (1.84)	2.06 (1.57)	1.29 (1.33)	2.78 (1.81)
2	NRS-1511	3.00 (1.87)	2.80 (1.82)	2.80 (1.81)	2.00 (1.58)	2.60 (1.75)	2.63 (1.76)	1.53 (1.42)	1.01 (1.22)	2.29 (1.67)
3	JNS-2017-8	3.60 (2.02)	2.60 (1.76)	1.93 (1.56)	2.47 (1.72)	1.41 (1.38)	2.10 (1.59)	1.35 (1.35)	0.97 (1.21)	2.05 (1.59)
4	NRS-1513	2.53 (1.74)	1.80 (1.52)	1.53 (1.42)	2.33 (1.68)	1.66 (1.46)	1.34 (1.35)	1.53 (1.42)	1.00 (1.22)	1.71 (1.48)
5	JNS-2017-13	2.73 (1.80)	2.67 (1.78)	2.40 (1.70)	2.60 (1.75)	1.90 (1.54)	1.09 (1.26)	1.50 (1.41)	1.35 (1.36)	2.03 (1.59)
6	IGPN-15-02	1.60 (1.45)	2.34 (1.68)	2.47 (1.72)	1.37 (1.37)	1.88 (1.54)	1.20 (1.30)	1.33 (1.35)	1.20 (1.30)	1.67 (1.47)
7	JNS-2017-17	3.53 (2.01)	3.27 (1.93)	2.47 (1.72)	1.87 (1.53)	1.93 (1.56)	1.40 (1.37)	1.55 (1.43)	1.16 (1.29)	2.14 (1.62)
8	IGPN-17-02	1.47 (1.40)	2.27 (1.66)	2.93 (1.84)	2.07 (1.60)	1.93 (1.56)	1.05 (1.23)	1.78 (1.51)	1.09 (1.26)	1.82 (1.52)
9	JNC-6	1.40 (1.38)	2.27 (1.66)	1.27 (1.33)	1.53 (1.42)	1.80 (1.52)	2.13 (1.62)	1.46 (1.40)	1.21 (1.31)	1.63 (1.46)
10	ONS-182	2.80 (1.82)	2.60 (1.76)	2.00 (1.57)	2.27 (1.65)	1.65 (1.46)	1.40 (1.37)	1.51 (1.41)	0.86 (1.16)	1.89 (1.55)

11	JNS-28	1.01 (1.23)	1.47 (1.40)	2.07 (1.60)	1.80 (1.52)	1.37 (1.36)	1.07 (1.25)	0.94 (1.20)	0.78 (1.13)	1.32 (1.35)
12	ONS-183	2.13 (1.62)	2.73 (1.80)	2.53 (1.74)	2.93 (1.85)	1.87 (1.53)	1.33 (1.34)	1.09 (1.26)	0.98 (1.21)	1.95 (1.57)
13	JNS-2016-1115	3.00 (1.87)	2.60 (1.76)	2.07 (1.60)	1.34 (1.35)	2.03 (1.58)	1.30 (1.33)	0.97 (1.21)	0.94 (1.20)	1.78 (1.51)
14	NRS-1804	2.40 (1.70)	2.93 (1.85)	3.27 (1.94)	2.33 (1.68)	1.66 (1.47)	2.33 (1.68)	1.17 (1.29)	1.26 (1.32)	2.17 (1.63)
15	DNS-19-24	2.20 (1.64)	1.73 (1.49)	2.33 (1.68)	2.53 (1.74)	1.85 (1.53)	1.30 (1.34)	1.13 (1.27)	1.17 (1.29)	1.78 (1.51)
16	NRS-1805	1.27 (1.32)	1.87 (1.53)	2.40 (1.70)	2.26 (1.66)	1.86 (1.54)	1.39 (1.38)	1.26 (1.32)	1.08 (1.26)	1.67 (1.47)
17	JNS-2016-1413	3.13 (1.90)	2.47 (1.72)	1.80 (1.51)	1.67 (1.47)	1.63 (1.45)	1.47 (1.39)	1.40 (1.37)	1.31 (1.34)	1.86 (1.54)
18	NRS-1807	2.53 (1.74)	2.87 (1.82)	2.07 (1.60)	2.47 (1.72)	1.90 (1.54)	1.93 (1.56)	1.36 (1.36)	1.26 (1.32)	2.05 (1.59)
19	JNS-9	1.13 (1.27)	1.60 (1.45)	0.87 (1.60)	1.93 (1.54)	1.73 (1.49)	1.73 (1.49)	1.45 (1.39)	1.00 (1.22)	1.43 (1.38)
20	VNS-1802	3.07 (1.89)	2.67 (1.77)	1.93 (1.56)	1.27 (1.33)	1.62 (1.45)	2.40 (1.69)	1.80 (1.51)	1.15 (1.28)	1.98 (1.57)
21	IGPN-2004-1	2.67 (1.78)	2.67 (1.78)	2.80 (1.82)	2.81 (1.81)	2.28 (1.67)	2.53 (1.74)	1.46 (1.39)	1.03 (1.24)	2.28 (1.67)
SE(m)±		0.05	0.07	0.08	0.09	0.06	0.09	0.07	0.04	0.068
CD at 5%		0.14	0.20	0.22	0.25	0.18	0.25	0.19	0.13	0.195

* Figures in parenthesis are square root transformed value

3.1. Leafhopper (*Amrascasp.*)

Among the screened genotypes the lowest population of leafhopper (0.83 leafhopper/three leaves) was recorded from the genotype JNS-2017-8 followed by (0.86 leafhopper/three leaves) IGPN-15-02. Among the screened genotype the highest population of leafhopper (1.68 leafhopper/three leaves) was received from the genotype JNS-2016-1413 followed by (1.48 leafhopper/three leaves) IGPN-2004-1 and (1.40 leafhopper/three leaves) NRS-1804. The genotypes JNS-2017-8, IGPN-15-02, NRS-1513, NRS-1511, JNS-2017-1, JNS-2017-13, IGPN-17-02, JNC-6, ONS-182, JNS-28, NRS-1805, NRS-1807, JNS-9 and VNS-1802 were found at par to each other in respect to record the leafhopper population/three leaves. Similarly the genotypes JNS-2016-1413, DNS-19-24, NRS-1804, JNS-2017-17 and IGPN-2004-1 were also found at par to each other in respect to record the leafhopper population/three leaves.

3.2. Capsule fly

The results of average population data of capsule fly revealed that all the genotypes were differed to each other in respect to record the capsule fly population

per plant. Among the screened genotypes the lowest population of capsule fly (1.11 adult /plant) was recorded from the genotype DNS-19-24 followed by (1.32 capsule fly/plant) JNS-2017-8 and JNS-2016-1115 (1.34 capsule fly/ plant). Among the screened genotype the highest population of capsule fly (2.58 capsule fly/plant) was received from the genotype JNS-2017-1 followed by (2.07 capsule fly/plant) JNS-2017-13 and NRS-1511 (1.92 capsule fly/plant), JNS-2017-17 (1.92 capsule fly/plant). The genotypes DNS-19-24, JNS-2017-8, JNS-2016-1115, NRS-1513, NRS-1807, JNS-2016-1413, ONS-182, ONS-183, IGPN-15-02, NRS-1805 and IGPN-17-02 were found at par to each other in respect to record the lowest population of capsule fly. Similarly the genotype JNS-2017-1, JNS-2017-17, NRS-1511, JNS-2017-17, JNS-9 and IGPN-2004-1, were found at par to each other in respect to record the population of capsule fly.

Table 2: Response of different genotypes of niger against whitefly

S. No.	Genotypes	White fly population/three leaves/plant Days after sowing (DAS)								Overall mean
		15	22	29	36	43	50	57	64	
1	JNS-2017-1	3.27 (1.94)	3.16 (1.91)	3.37 (1.96)	3.00 (1.86)	3.24 (1.93)	2.90 (1.84)	2.06 (1.57)	1.29 (1.33)	2.78 (1.81)
2	NRS-1511	3.00 (1.87)	2.80 (1.82)	2.80 (1.81)	2.00 (1.58)	2.60 (1.75)	2.63 (1.76)	1.53 (1.42)	1.01 (1.22)	2.29 (1.67)
3	JNS-2017-8	3.60 (2.02)	2.60 (1.76)	1.93 (1.56)	2.47 (1.72)	1.41 (1.38)	2.10 (1.59)	1.35 (1.35)	0.97 (1.21)	2.05 (1.59)
4	NRS-1513	2.53 (1.74)	1.80 (1.52)	1.53 (1.42)	2.33 (1.68)	1.66 (1.46)	1.34 (1.35)	1.53 (1.42)	1.00 (1.22)	1.71 (1.48)
5	JNS-2017-13	2.73 (1.80)	2.67 (1.78)	2.40 (1.70)	2.60 (1.75)	1.90 (1.54)	1.09 (1.26)	1.50 (1.41)	1.35 (1.36)	2.03 (1.59)
6	IGPN-15-02	1.60 (1.45)	2.34 (1.68)	2.47 (1.72)	1.37 (1.37)	1.88 (1.54)	1.20 (1.30)	1.33 (1.35)	1.20 (1.30)	1.67 (1.47)
7	JNS-2017-17	3.53 (2.01)	3.27 (1.93)	2.47 (1.72)	1.87 (1.53)	1.93 (1.56)	1.40 (1.37)	1.55 (1.43)	1.16 (1.29)	2.14 (1.62)
8	IGPN-17-02	1.47 (1.40)	2.27 (1.66)	2.93 (1.84)	2.07 (1.60)	1.93 (1.56)	1.05 (1.23)	1.78 (1.51)	1.09 (1.26)	1.82 (1.52)
9	JNC-6	1.40 (1.38)	2.27 (1.66)	1.27 (1.33)	1.53 (1.42)	1.80 (1.52)	2.13 (1.62)	1.46 (1.40)	1.21 (1.31)	1.63 (1.46)
10	ONS-182	2.80 (1.82)	2.60 (1.76)	2.00 (1.57)	2.27 (1.65)	1.65 (1.46)	1.40 (1.37)	1.51 (1.41)	0.86 (1.16)	1.89 (1.55)
11	JNS-28	1.01 (1.23)	1.47 (1.40)	2.07 (1.60)	1.80 (1.52)	1.37 (1.36)	1.07 (1.25)	0.94 (1.20)	0.78 (1.13)	1.32 (1.35)
12	ONS-183	2.13 (1.62)	2.73 (1.80)	2.53 (1.74)	2.93 (1.85)	1.87 (1.53)	1.33 (1.34)	1.09 (1.26)	0.98 (1.21)	1.95 (1.57)
13	JNS-2016-1115	3.00 (1.87)	2.60 (1.76)	2.07 (1.60)	1.34 (1.35)	2.03 (1.58)	1.30 (1.33)	0.97 (1.21)	0.94 (1.20)	1.78 (1.51)
14	NRS-1804	2.40 (1.70)	2.93 (1.85)	3.27 (1.94)	2.33 (1.68)	1.66 (1.47)	2.33 (1.68)	1.17 (1.29)	1.26 (1.32)	2.17 (1.63)
15	DNS-19-24	2.20 (1.64)	1.73 (1.49)	2.33 (1.68)	2.53 (1.74)	1.85 (1.53)	1.30 (1.34)	1.13 (1.27)	1.17 (1.29)	1.78 (1.51)
16	NRS-1805	1.27	1.87	2.40	2.26	1.86	1.39	1.26	1.08	1.67

		(1.32)	(1.53)	(1.70)	(1.66)	(1.54)	(1.38)	(1.32)	(1.26)	(1.47)
17	JNS-2016-1413	3.13 (1.90)	2.47 (1.72)	1.80 (1.51)	1.67 (1.47)	1.63 (1.45)	1.47 (1.39)	1.40 (1.37)	1.31 (1.34)	1.86 (1.54)
18	NRS-1807	2.53 (1.74)	2.87 (1.82)	2.07 (1.60)	2.47 (1.72)	1.90 (1.54)	1.93 (1.56)	1.36 (1.36)	1.26 (1.32)	2.05 (1.59)
19	JNS-9	1.13 (1.27)	1.60 (1.45)	0.87 (1.60)	1.93 (1.54)	1.73 (1.49)	1.73 (1.49)	1.45 (1.39)	1.00 (1.22)	1.43 (1.38)
20	VNS-1802	3.07 (1.89)	2.67 (1.77)	1.93 (1.56)	1.27 (1.33)	1.62 (1.45)	2.40 (1.69)	1.80 (1.51)	1.15 (1.28)	1.98 (1.57)
21	IGPN-2004-1	2.67 (1.78)	2.67 (1.78)	2.80 (1.82)	2.81 (1.81)	2.28 (1.67)	2.53 (1.74)	1.46 (1.39)	1.03 (1.24)	2.28 (1.67)
	SE(m)±	0.05	0.07	0.08	0.09	0.06	0.09	0.07	0.04	0.068
	CD at 5%	0.14	0.20	0.22	0.25	0.18	0.25	0.19	0.13	0.195

* Figures in parenthesis are square root transformed value.

UNDER PEER REVIEW

Table 3: Response of different genotypes of niger against capsule fly

S. No.	Genotypes	Capsule fly population/plant Days after sowing (DAS)				Overall mean
		45	52	59	66	
1	JNS-2017-1	2.80 (1.80)	3.01 (1.87)	2.69 (1.78)	1.82 (1.52)	2.58 (1.75)
2	NRS-1511	2.67 (1.78)	1.70 (1.48)	2.01 (1.58)	1.29 (1.33)	1.92 (1.56)
3	JNS-2017-8	1.40 (1.37)	1.20 (1.33)	1.28 (1.37)	1.21 (1.28)	1.32 (1.34)
4	NRS-1513	1.74 (1.49)	1.20 (1.30)	1.28 (1.27)	1.21 (1.30)	1.36 (1.36)
5	JNS-2017-13	2.60 (1.76)	2.53 (1.74)	1.90 (1.53)	1.25 (1.31)	2.07 (1.60)
6	IGPN-15-02	1.74 (1.49)	1.40 (1.38)	1.48 (1.36)	1.37 (1.37)	1.50 (1.41)
7	JNS-2017-17	3.20 (1.92)	1.33 (1.34)	1.82 (1.50)	1.32 (1.34)	1.92 (1.55)
8	IGPN-17-02	1.74 (1.49)	1.27 (1.32)	1.49 (1.40)	1.37 (1.36)	1.46 (1.40)
9	JNC-6	1.74 (1.49)	2.33 (1.68)	1.79 (1.50)	1.36 (1.36)	1.80 (1.52)
10	ONS-182	1.26 (1.32)	1.45 (1.39)	1.98 (1.57)	1.34 (1.35)	1.50 (1.41)
11	JNS-28	2.40 (1.70)	1.20 (1.30)	2.07 (1.59)	1.11 (1.26)	1.69 (1.48)
12	ONS-183	1.54 (1.40)	1.42 (1.38)	1.75 (1.47)	1.22 (1.30)	1.48 (1.41)
13	JNS-2016-1115	1.00 (1.22)	1.26 (1.32)	1.94 (1.55)	1.17 (1.29)	1.34 (1.35)
14	NRS-1804	1.73 (1.49)	2.13 (1.62)	1.81 (1.50)	1.34 (1.35)	1.75 (1.50)
15	DNS-19-24	1.13 (1.27)	1.01 (1.23)	1.28 (1.33)	1.02 (1.23)	1.11 (1.27)
16	NRS-1805	2.40 (1.70)	1.34 (1.34)	1.26 (1.32)	1.04 (1.24)	1.51 (1.42)
17	JNS-2016-1413	1.46 (1.39)	1.65 (1.47)	1.55 (1.43)	1.28 (1.33)	1.49 (1.41)
18	NRS-1807	1.73 (1.49)	1.25 (1.32)	1.42 (1.39)	1.16 (1.18)	1.39 (1.37)
19	JNS-9	2.80 (1.81)	2.40 (1.70)	1.34 (1.35)	1.06 (1.17)	1.90 (1.55)
20	VNS-1802	1.34 (1.35)	1.81 (1.52)	1.63 (1.45)	1.97 (1.57)	1.69 (1.48)
21	IGPN-2004-1	2.46 (1.72)	1.98 (1.58)	1.46 (1.41)	1.54 (1.42)	1.86 (1.54)
	SE(m)±	0.08	0.05	0.10	0.06	0.07
	CD at 5%	0.22	0.15	0.29	0.18	0.21

* Figures in parenthesis are square root transformed value.

4. Conclusions

The 21 promising genotypes of Niger were screened for resistance/susceptibility against whiteflies, leafhoppers, and capsule fly. It is assessed that the genotypes JNS-28, JNS-9, JNC-6, NRS-1805, IGPN-15-02 and NRS-1513 were found promising against whitefly. The genotypes JNS-2017-8, IGPN-15-02, NRS-1513, NRS-1511 and JNS-2017-1 were found promising against the incidence of leafhopper. The genotypes DNS-19-24, JNS-2017-8, NS-2016-1115, NRS-1513 and NRS-1807 were found promising against capsule fly. It has been concluded that JNS-2017-1 genotype is highly resistant to whitefly and capsule fly and JNS-2016-1413 was resistant to Leafhopper. JNS-28, JNS-2017-8, DNS-19-20 were susceptible genotype with respect to whitefly, leafhopper and capsule fly.

6. References

- Acharya VS and Singh AP. Screening of cotton genotypes against whitefly, *Bemisia tabaci* Genn. Journal of Cotton Research and Development. 2007; 21(1): 111-114.
- Anonymous. Niger technology for maximizing production. All India coordinated research project on sesame and niger (ICAR), JNKVV Campus, Jabalpur (M.P). 2014; 1-4p.
- Bulcha W. *Guizotia abyssinica* (L.f.) Cass. Record from PROTA (Plant Resources of Tropical Africa), Wageningen, Netherlands. 2007; 24p.
- Cemal K. Screening of Turkish sesame (*Sesamum indicum* L.) landraces for whitefly and lodging resistance. International gap agriculture & livestock congress. 25-27 April 2018, Turkey. 45.
- Chakravati BN. Niger seed (*Guizotia abyssinica* Cass) as they rainy season host plant of *Perigeacapis Gn*, Indian Journal Entomology. 1947; 10: 129-130.
- Chatterjee PM, Shanowly and Das, Ayan. Screening of Different Genotypes of okra (*Abelmoschus esculentus* L.) against Leafhopper (*Amrasca biguttulabiguttula* L.) and Whitefly (*Bemisia tabaci* G.) under New Gangetic Alluvial Zone of West Bengal, International Journal of Current and Applied Sciences. 2019; 8(3): 1087-1095.

- Chauhan RS, Sharma AK, Kamal R and Ali A. Screening of mungbean (*Vignaradiata*L.)germplasm against major sucking pest. Journal of Pharmacognosy and Phytochemistry.2018;7(5): 1784-1787
- Das P and KhandweN. 2005. Screening of niger varieties against major pest in Satpura plateau of Madhya Pradesh. National seminar on strategies for enhancing production and expert of Sesame and Niger, April 7th and 8th, Agri. Res. Station, RAU Mandor, Jodhpur (Rajasthan).2005; 49.
- Ganguly J, Ganguly RN,Vaishampayan SM and Verma R. Identity of the migratory phase of cabbage semilooper on the basis of reproductive development and Fat body content.Journal Advancement Zoology.1989;10 (2): 88- 94.
- Hooda VS.Dhankhar, B. S. and Singh, Ram.2011. Evaluation of okra cultivars for field resistance to the leafhopper *Amrascabiguttulabiguttula* (Ishida), International Journal of Tropical Insect Science, 17(4):323 – 327.
- Jahangir AH, Muhammad W and Jamil M. Screening of okra genotypes (*Abelmoschusesculentus* L.) against jassid (*Amrascabiguttulabiguttula* Ishida) under agro-climatic conditions of Bahawalpur, Pakistan.ARPN Journal of Agricultural and Biological Science. 2017; 12(12): 352-359.
- Mandloi I. To study the effect of different dates of sowing on the incidence of major pests of niger.M.sc.Thesis, JNKVV,Jabalpur.2020; 1-75.
- ManivannanA, Philip RS, Karthikeyan S and Pillai AK.Screening of cotton genotypes against leafhopper, *Amrascabiguttulabiguttula* (Ishida) (Homoptera: Cicadellidae), Journal of Entomology and Zoology Studies.2017; 5(6): 1305-1310.
- Moorie AO, Sequeria RV and Woodger TA. Susceptibility of crop plants to *Bemisiatabaci* (Gennadins) B-biotype (Hemiptera, Aleyrodidae) in control. Queensland australian Entomologist. 2004; 31(2):69-74.
- Nagaraj G and Patil HS.Quality and oil composition of Niger (*Guizotiaabyssinica*L.f. Cass). A Review.J.oilseed Res.2004;21 (2):224-229.
- Narayanan, ES. Insect pest of niger and methods of their control in niger and safflower in Karnataka.1961;6(7):119-120.

- Navneet and Tayde, AR.2018. Screening of okra genotypes against white fly [*Bemisiatabaci*(Gennadius)] and YVM Virus. Journal of Entomology and Zoology Studies, 6(1): 565-568.
- Panday AK, Bisen R, Jain S and Sahu R. 2019. Effect of climatic factors on population intensity of insect pests damaging niger crop. XIX International Plant Protection Congress IPPC2019 Hyderabad, Telangana.278.
- Panday AK, Sharma S, Bisen R, Jain S, Malviya M and Ranganatha ARG.2014. Niger improvement : Current status and future strategies .J. oilseed res.,31(2): 95-113.
- Patil HS, Ranganatha ARG, Paroha S and Tripathi A. Niger [*Guizotiaabyssinica* (L.f.) Cass.]. In: Breeding of Field Crop. Agrobios (India), Jodhpur. 2012; 567-587.
- Rai BK. Pests of oilseed crops in India and their control. Publ. Indian Council Agricultural Research, New Delhi.1976; 121.
- Rai HS and Tiwari RC. Screening of Niger germplasm lines against niger capsule fly (*Dioxyasororcula*Weid). Geobios, 1995; 22(4):174-176.
- Rai HS. Pest complex of niger (*Guizotiaabyssinica* Cass) and screening of its available germplasm against major pests. M.Sc. (Agri.) Thesis submitted to JNKVV, Jabalpur. India.1984;120.
- Ranganatha ARG. Strategies for Sesame and Niger Developmental Programmes in India. Paper presented in DAC-GOI review, New Delhi. The Indian society of oilseeds research. 2009; 32(2):95-193.
- Ranganatha ARG, Panday AK Bisen R. Jain S and Sharma Shikha. Breeding Oilseed Crops for Sustainable Production.2016; 169-199 .DOI:10.1016/B978-0-12-801309-0.00008-2.
- Saleem M, Hassan WM and Jamil M. Screening of sunflower (*Helianthus annus* L.) varieties against insect pests, predators' populations, and their yield comparison under semi-arid climatic conditions. Journal of Entomology and Zoology Studies 2017; 5(3): 1219-1225.
- Saxena RN. Screening of promising germplasm lines of niger (*Guizotiaabyssinica*Cass.) against the major insect pests. M.Sc. (Agri.) Thesis submitted to JNKVV, Jabalpur.200;1-6p.

- Shaikh AA, Patel DR and Patel JJ. Screening of different genotypes/cultivars against sucking pests infesting brinjal, AGRES – An International e-Journal, 2013. 2(1) :51-57.
- Shrinivasan S and Yuvaraja B. In Incidence of *Prospaltacapensis* Gn. and its natural enemies on safflower Insect Environment. 2004; 10(3):115-116.
- Shrivastava N. Insect pest of niger and their integrated management. Proceeding of workshop cum Seminar on niger production technology held at ZARS, Chhindwara (M.P.) during 28th Sept.- 1st Oct. 1995; 44-50.
- Shah SJ, Khan SB, Zulfiqar Ali, Shah Sajid Ali, Ullah A, Bakhsh K, Ahmed ND, Naseer A and Mastoi AT. Screening of mustard varieties against sucking insect pests of mustard. Pure and Applied Biology. 2020; 9(2):1522-1533.
- Tembhre, H. Screening of germplasm of niger (*Guizotia abyssinica* Cass.) against the major insect pests. M.Sc. (Agri.) Thesis submitted to JNKVV, Jabalpur. 2005; 1-85p.
- Vles RO and Gottenbos JJ. Nutritional characteristics and food uses of vegetable oils. Oil Crops of the World, McGraw-Hill Publishing Company, New York. 1989; 63-86.
- Yadav, Amrita. To study the succession, population dynamics and estimate avoidable yield losses caused by insect pests in Niger crop. M.Sc. (Agri) Thesis., JNKVV, Jabalpur. 2017; 1-9.
- Misal D, Vaibhav, Davane SS and Mane BS. Improvement of yield in *Guizotia abyssinica* (L.F.) Cass. by using backcross method. World Journal of Advanced Research and Reviews, 2021, 12(02), 261–266. Article DOI: <https://doi.org/10.30574/wjarr.2021.12.2.0579.2021>.