Original Research Article

HISTOPATHOLOGICAL STUDY IN ALLEVIATIVE EFFECTS OF GARLIC EXTRACT (ALLIUM SATIVUM) AGAINST CISPLATIN-INDUCED KIDNEY DAMAGE IN RABBITS

ABSTRACT

Cisplatin (cis-diammine-dichlorido-platinum II) is a chemotherapeutic agent used for treating different types. However, its usage is limited because it induces harmful toxicities in multiple organs, including nephrotoxicity. Garlic extract has several pharmacological activities, including antioxidant activity. The aim of the study is to alleviate theeffects of garlic extract (*Allium sativum*) against cisplatin-induced kidney damage in rabbits. Twenty-seven healthy domestic male rabbits were randomly divided into three groups, each comprising nine rabbits. The first group (Co) served as control and received a single intraperitoneal injection of normal saline solution, once per week, for six weeks, the second group (Cp) were treated with (7 mg**/** kg of body weight) therapeutic dose of cisplatin intraperitoneally once per week for six weeks and the third group (Cp+ Ga) received a dose of 250mg/kg body weightof garlic extract by oral gavages, starting from the first day of the experiment for 6 consecutive days before and 6 consecutive days after the cisplatin injection and continued daily for 6 weeks. In addition, this group was treated with (**7** mg **/** kg of body weight) therapeutic dose of cisplatin intraperitoneally once per week for six weeks. After six weeks of experiment, the animals of the experimental and control groups were anesthetized by ether inhalation, laparotomized, the renal histological sections from Cp-treated group displayed pronounced histopathological lesions, destruction of the renal tubules, slogging of almost entire epithelium due to desquamation of tubular epithelium, with pyknosis of their nuclei and swelling with multi-vacuolations of the cytoplasm with obliteration of their lumina, remarkable necrosis, and hyaline casts in renal tubules and focal area of severe hemorrhagecompared with those of control (Co) group. While sections from the kidney of animals received therapeutic dose of cisplatin and treated by garlic extract, group (Cp + Ga) revealed less to no distinct pathological

damages in renal corpuscles and renal tubules were restored nearly to the control ones. It is concluded that garlic (*Allium sativum*) extract has a powerful alleviative effect against CP-induced nephrotoxicity in rabbits.

Keywords

Cisplatin, nephrotoxicity, histopathology, garlic extract.

Introduction:

The kidney being an essential organ of metabolism and elimination of most of the toxic components, it serves as a very important site of attack by various chemical agents including cisplatin, in the form of its activated metabolite**.** Most anticancer agents cause toxicity in various organs, including the kidney, by disturbing theoxidant/antioxidant balance [(1)].

Cisplatin [Cis-diamminedichloroplatinum II (CDDP)] is a highly efficient antineoplastic drug commonly used as a first-line therapy for many solid cancers, such as stomach cancer, ovarian cancer, lung cancer, bladder cancer, and germ cell tumors[(2,3)]. The experimental cisplatin-induced nephrotoxicity was first reported in 1971[(4)]. Since then numerous studies have been published. Over the past years researchers have demonstrated that cisplatin nephrotoxicity is dose dependent and cumulative. Nephrotoxicity can be induced by single or multiple applications of cisplatin. Depending on the dosage, frequency of cisplatin injection, and cumulative dose of cisplatin, animals may develop different severity of acute (early) and chronic (advanced) kidney injury. In rodents cisplatin is usually injected intraperitoneally (ip) and less frequently intravenously (iv) or subcutaneously (sc) [(5)]. Previous studies indicated that cisplatin produced animal behavioral and morphological changes, as well as cellular and subcellular changes in kidneys [(6)].

The primary manifestations are impairment in the functioning of renal tubular epithelial cells, including inhibition of protein synthesis, oxidative stress, mitochondrial dysfunction, cytoskeleton remodeling, changes of intercellular adhesion, and apoptosis of tubular epithelial cells[(7)]. The anticancer effect of cisplatin is mediated by apoptosis and DNA-cross links with subsequent cytotoxic lesions in malignant cells[(8)].

Experimental studies on rats and mice revealed that cisplatin leads to nephrotoxicity through formation of glutathione conjugates in the bloodstream[9]. Other studies showed that cisplatin binds to DNA leading to arrest of DNA synthesis and replication resulting in cell apoptosis[10].

Cisplatin is one of the most potent and widely used chemotherapeutic antitumour agents. Various agents, including antioxidants, attenuate the nephrotoxicity of this compound[11]. Natural products from plants and animals have been widely utilized all over the world either in the pure forms or crude extracts for protection from or treatment of various diseases[12]. Garlic (Allium sativum, Liliaceae) is used widely as a spice and medicinal herb not only in its native region but also all around the world. Allium sativum (Garlic) is rich in antioxidants which help destroy free radical particles that can damage cell membranes and DNA. It has been found to contain a large number of potent bioactive compounds with anticancer properties[13].

Allium sativum (Garlic) is one of the most famous plants that is widely used to combat several diseases because it contains more than 33 sulfur compounds, enzymes, minerals, amino acids, vitamins including A, B1 and C as well as fibers[13]. Garlic has a higher concentration of sulphur-containing compounds than any other member of the Allium species. This feature underlies the pungent odor of garlic and conveys the antioxidant properties of garlic preparations and many of its medicinal effects[14]. The sulphur compounds found in fresh garlic appear to be almost 1000 times more potent as antioxidants than those found in aged garlic extracts[15]. The most vital active sulfur compounds in Allium sativum are allicin and alliin which are considered the main antioxidants and scavenging free radical compounds[16].

In addition, garlic has abundant chemical compounds such as S-allyl cysteines, thiacremonone, diallyl-disulfide, diallylsulfide and others. This medicinal plant and its constituents offer a lot of benefits including anti-inflammatory, anti-cholesterolemic, anti-gastric ulcer, antimicrobial, anticancer, and antioxidant properties. Garlic also modulates the activity of several metabolizing enzymes[17].

Previous studies established the protective and antioxidant effects of garlic against cisplatin - induced acute liver and kidney injuries in male rats[18]. Aged garlic extract (20% aqueous ethanol) exerts an ameliorative effect against cisplatin induced oxidative stress and renal damage by exerting antioxidant, anti-inflammatory, and antiapoptoticeffects[19].

Treatment with an ethanolic extract of garlic might reduce cisplatin-induced nephrotoxicity in rats by decreasing serum levels of kidney biomarkers such as urea, uric acid and creatinine by increasing the activities of antioxidant enzymes[20].

Razo-Rodriguez et al., 2008 were found that the garlic powder feeding was able to prevent by 40–59% the alterations in the markers of renal injury studied, by 33% the histological damage, and by 38–75% the increase in markers of oxidative and nitrosative stress. It concluded that the ability of garlic powder to ameliorate cisplatin-induced renal injury is associated with its antioxidant properties[21]. Consequently, the current investigation was designed to evaluate the role of garlic extract on the nephrotoxicity induced by therapeutic dose of cisplatin in rabbits.

Materials and Methods:

The present experiment was conducted on twenty-seven healthy domestic male rabbits (Oryctolaguscuniculusdomesticus), weighing 1.5– 2kg, and were housed in the animal house of the Histology Department, Faculty of Medicine, University of Benghazi. They were acclimatized for 2 weeks prior to experimentation. This was done to enable adaptation to the surroundings and enforce a daily routine with 12 to 13 h of light to maintain the colony's circadian biorhythms. The animals were allowed unrestricted access to food and water ad libitum. The rabbits were examined daily for possible behavioral and gross morphological or physical changes.The rabbits were randomly divided into three groups, each comprising nine rabbits.

The first group (Co) served as control and received a single intraperitoneal injection of normal saline solution (once per week, for six weeks) for the duration of the experiment to simulate the effect of injection.

The second group (Cp) were treated with a single (**7** mg/ kg of body weight) therapeutic dose of cisplatin intraperitoneally once per week for six weeks.

The third group (Cp+Ga) animals of this group received a dose of 250 mg/kg body weightof garlic extract by oral gavages, starting from the first day of the experiment for 6 consecutive days before and 6 consecutive days after the cisplatin injection and continued daily for 6 weeks.

In addition, this group was treated with a single (7 mg / kg of body weight) therapeutic dose of cisplatin intraperitoneally once per week for six weeks.

The concentration of cisplatin dose and garlic extract was selected based on previous studies [19,22,23,24].

For histological analysis, after six weeks of experiment, the animals of the experimental and control groups were anesthetized by ether inhalation, laparotomized, the kidney samples were removed and divided into segments. Segments from all groups were placed in 10% formal saline for 48 hours for histopathological assessment. After a period of 48 hours of fixation, tissue processing was done by using paraffin technique, the slides were stained with Hematoxylin and Eosin (H&E). Then sections were examined under a light microscope [25].

Result:

Clinical observations:

All the following observations were seen, 60 to 120 minutes after injection of the treated rabbits with therapeutic dose of cisplatin, group (Cp): The animals exhibited excessive sleeping, diarrhea, abdominal swelling, loss of appetite, fearless behavior, decrease movement, rapid breathing, and finally we were noticed some animals death after second and \ or third doses of drug.

The dead animals were replaced by additional rabbits, injecting the same dose of cisplatin for the desired duration. While treatment with garlic extract (Cp+Ga) group ameliorated all above mentioned clinical signs but they did not return to normalcy.No clinical signs were seen in the control (Co) group. There was no mortality among the animals of the control group (Co) andprotective group (Cp+Ga).

Histological and histopathological findings:

Control animals group (Co group):

Histological examination of the kidney sections of rabbit in the control group (Co) revealed normal morphological characteristics in terms of preserved architecture of the renal tissue. The kidney was covered by a thin superficial fibrous connective tissue capsule composed of fine collagen and reticular fibers, outer cortex and inner medulla. The outer cortex was highly vascular; the inner medulla was slightly thick and less vascular.

The nephron was the functional unit of the kidney; each nephron was consisting of renal corpuscles, proximal convoluted tubules, loop of Henle, distal convoluted tubules and collecting tubules.

The sections was extended from the capsule to the medulla showing variations in distribution of renal corpuscle from the superficial which had little renal corpuscles to the mid-cortical region which had more renal corpuscle and the juxtamedullary region had less renal corpuscles than the mid cortical and cortical region **(Fig. 1a).**

The renal corpuscle was consisting of a tuft of capillaries, the glomerulus, surrounded by a double layered cup - shaped Bowman's capsule. The proximal convoluted tubules (PCT) had narrow lumen, were lined with darkly acidophilic high cuboidal epithelium with rounded nuclei and a brush border. The distal convoluted tubules (DCT) were lined with faint acidophilic cubical epithelial cells with rounded nuclei, and their lumen apparently wider than proximal convoluted tubules (PCT).

The Henle loop (nephron loop), was consisting of the thin and thick segments that are more evident in medulla. The thin segment of Henle loop is lined by simple squamous epithelium tissue, while the thick segment is lined by cuboidal epithelium.

The collecting tubules were continuing from the terminal part of distal tubule and converged in the renal cortex to form bundles of tubules called medullary rays. The collecting tubules lined by cuboidal epithelial tissue with relatively large nucleus occupy the entire cell and reset on the visible basement membrane. The cytoplasm of cuboidal cells was pale and had a dark oval nucleus **(Fig. 1b, c).**

Treated animals group (Cp group):

Sections from kidney tissue of rabbits treated with therapeutic dose of cisplatin group (Cp) were showing extensive disruption of tissue architecture, which was leading to complete destruction of the normal pattern of the renal tissue. Kidney tissue sections revealed marked morphological disturbances at the cortico-medullary zone, wereshown glomerular congestion and atrophy, desquamation with shedding of the tubular epithelial lining cells. There were extensive multifocal-to-diffuse cortical tubular degeneration and acute necrosis**(Fig.2).**

These damages were encountered by the presence of atrophy of glomerular tuft, wide capsular space, thickening of parietal layer of Bowman's capsule as well as focal interstitial nephritis. There was excessive accumulation of homogenous exudates casts in both proximal convoluted tubules (PCT) and distal convoluted tubules (DCT). Intertubular hemorrhage was also observed **(Fig.3).**

The degenerative changes in the proximal tubules that consisted of hydropic degeneration, pyknotic nuclei, cytoplasmic vacuolization, evident loss of the brush border, necrosis and apoptosis of tubular cells, and desquamation of necrotic epithelial cells filling the tubular lumens and forming hyaline casts. These morphological changes were more pronounced in the proximal tubules but were also induced in distal and collecting tubules as well **(Fig.4).**

High magnification cross sections in the structure of rabbit kidney treated with therapeutic dose of Cisplatin of (Cp) group showed a severe atrophy of glomerulus, which was apparent due to the reduction in its size withdistension of Bowman's space. Shrunken renal corpuscle and degeneration of the parietal epithelial cells of Bowman's capsule and some glomeruli exhibiting thickening of the basement membrane was also observed. Cellular debris in the tubular lumen and increased tissue in the interstitium was also an indication of Cisplatin-induced renal necrosis.

Congestion of blood vessel, necrosis of renal tubular epithelium with pyknosis of their nuclei and swelling with multi vacuolations of the cytoplasm with obliteration of their lumina and focal area of severe haemorrhage was observed **(Fig.5).**

There was destruction of the renal tubules, slogging of almost the entire epithelium due to desquamation of tubular epithelium. The lumina of distal and proximal convoluted tubules contain hyaline casts of dead cells and congested blood vessels can be also detected. The renal medulla showed dilated collecting tubules stuffed with R.B.Cs???.Severeinterstitial inflammation and edema in renal cortex and outer medulla **(Fig.6).**

Protective animals group (Cp + Ga):

Sections from the kidneys of animals received therapeutic dose of cisplatin and treated by garlic extract, (Cp + Ga) group, revealed less histological damages in renal corpuscles and renal tubules compared with that of rabbits in the (Cp) group. There was reduced the nephrotoxic effect of Cisplatin and no distinct pathological changes except mild glomerular congestion and turbidity of the epithelial cells of the proximal convoluted tubules. Mild tubular degeneration with luminal dilatation and inflammation were seen within the renal cortex **(Fig.7).**

There were slight atrophy and vacuolation of glomerular tuft with slight distension of Bowman's capsule, also the lumen of the proximal convoluted tubules appeared filled with debris **(Fig.8).**

In some areas within the renal cortex, there were slight tubular necrosis and moderate tubular degeneration was observed in proximal and distal convoluted tubules of the renal parenchymal structure. Extensive congestion with in loop of Henle and collecting tubules was also noticed **(Fig.9).**

In some tubules in rabbit kidneys received a therapeutic dose of cisplatin and treated by garlic extract; (Cp + Ga) group were showing apparent normal renal parenchyma, but still some glomeruli appeared with slight atrophy. Transparent hyaline casts can be also noticed in the lumen of renal tubules in decreased accumulation with occasional degenerative changes when compared to cisplatin treated rabbits of (Cp) group **(Fig.10).**

Discussion:

Cisplatin (cis-dichlorodiammine-platinum (II), plays a highly effective role on a diverse spectrum of malignancies and yet is one of the most potent agents in resisting tumors. It is widely used to treat malignancies of various solid tumors such as head and neck, esophageal, bladder, testicular, ovarian, and small cell lung cancer[26]. Despite the positive effects of platinum compounds, they are poisons. Patients receiving these agents experience severe side effects that limit the dose which can be administered.However, it has been reported that the use of cisplatin is often limited because of a number of adverse effects, including nausea, vomiting, sensitivity reactions, ototoxicity, neurotoxicity, and bone marrow suppression. Among these adverse effects, nephrotoxicity is the most important limiting factor in cancer treatment using cisplatin[27].

The kidney plays a major role in the maintenance of constant volume and composition of the extra-cellular fluid hence does the basic functions of glomerular filtration, tubular reabsorption and tubular secretion. The kidney being an essential organ of metabolism and elimination of most of the toxic components, it serves as very important site of attack by various chemical agents including cisplatin, in the form of its activated metabolite and thus drug-induced nephrotoxicity is a frequent entity in clinical medicine[28].

The nephrotoxicity of cisplatin has been recognized since its introduction over 25 years ago.

Once cisplatin enters the cell it exerts its cytotoxic effect by losing one chloride ligand, binding to DNA to form intra-strand DNA adducts, and inhibiting DNA synthesis and cell growth. The DNA

lesions formed from cisplatin-induced DNA damage activate DNA repair response via NER (nuclear excision repair system) by halting cisplatin-induced cell death by activation of ATM (ataxia telangiectasia mutated) pathway. However, because cisplatin-induced DNA damage activate several signal transduction pathways that can facilitate or prevent apoptosis[29].

Cisplatin produces reactive oxygen species such as hydroxyl radicals, hydrogen peroxide and singlet oxygen and superoxide ions, compromises natural anti-oxidant defense by inhibiting anti-oxidant enzymes and increasing reactive oxygen species, and also leads to lipid peroxidation in membranes and a decrease in protective enzyme activities against peroxidation. This increased oxidative stress, which results in DNA injury, is responsible for the resulting nephrotoxicity[30].

Cisplatin causes damage to nuclear and mitochondrial DNA and production of reactive oxygen species (ROS) which lead to activation of both mitochondrial and non-mitochondrial pathways of apoptosis and necrosis. Mitochondrial energetic are also disrupted by cisplatin and may contribute to nephrotoxicity[31]. Previous reports also demonstrate that cisplatin interferes with mitochondrial function and causes injury because it plays an important role in maintaining calcium homeostasis, Na+ - K+ - ATPase activity and ATP synthesis[32].The cellular pathways of cisplatin injury to kidney cells have been examined primarily *in vitro* using freshly isolated or cultured renal tubular epithelial cells. *In vitro*, low concentrations of cisplatin preferentially result in apoptotic cell death while at higher concentrations necrosis ensues[33].*In vivo* administration of nephrotoxic doses of cisplatin produces a large increase in both necrosis and apoptosis in the kidney[34].

The primary manifestations are impairment in the functioning of renal tubular epithelial cells, including inhibition of protein synthesis, oxidative stress, mitochondrial dysfunction, cytoskeleton remodeling, changes of intercellular adhesion, and apoptosis of tubular epithelial cells. It has been suggested that oxidative damage related to oxidative stress may also be one of the causes of cisplatin -induced nephrotoxicity. Therefore, it is important to prevent nephrotoxicity of cisplatin[35].

The result of current study were shown several clinical signs appear in group of animals treated intrapretonially by therapeutic dose of cisplatin (Cp group) as excessive sleeping, diarrhea, loss in ability to taste food and decrease movement were noticed, compared to the control group, might

be attributed to the direct damage of cisplatin on renal tubular cells resulting in decreased water and sodium reabsorption with subsequent polyuria, dehydration[36, 37]. Another study postulated that the decreased body weight in cisplatin injected rats might be attributed to gastrointestinal toxicity resulting in lost appetite, ingestion and assimilation of food[9].

Glinsukon et al., 1986 were suggested that it could in turn be ascribed to drug induced toxicity, psychological pressures and the necrotizing effects of the drug on the digestive system. Furthermore, the drug also affects the mucous lining of the gastro-intestinal tractand increased metabolic rate, which were considered side effects of the chemotherapy[38]. The clinical observations seen in our study were corroborate the findings of previous researchers; King and Berry 2001[39];Leite et al., 2009[40];Rickenbacher et al. 2011[41];Hesham and Ghobara, 2013[42]; Aboraya et al., 2022[43] andElbeltagy et al., 2022[23].

The results of the present investigation showed that cisplatin toxicity produced significant structural changes in the kidney of group Cp, cisplatin treated group, in the form ofextensive disruption of tissue architecture that was leading to complete destruction of the normal pattern of the renal tissue. There was glomerular congestion and atrophy with wide capsular space, thickening of parietal layer of Bowman's capsule, desquamation with shedding of the tubular epithelial lining cells, extensive multifocal-to-diffuse cortical tubular degeneration and acute necrosis, excessive accumulation of homogenous exudates casts in both proximal and distal convoluted tubules with hydropic degenerative changes in their cells, pyknotic nuclei and cytoplasmic vacuolization, evident loss of the brush border, necrosis and apoptosis of tubular cells. Intertubular hemorrhage and severe interstitial inflammation and edema in renal cortex and outer medulla have also been noticed.

The changes obtained in the present study run parallel with the report documented by many authors who reported the toxicity of cisplatin on the kidney;Shirwaikar et al., 2003[44]; Ravindra et al.,

2010[45]; Perše and VeIeriT-Haler, 2018[46]; Aboraya et al., 2022[43]; Elbeltagy et al., 2022[23] .

Cornelison and Reed, 1993[47]; Meyer and Madias, 1994[48]and Vickers et al., 2004[49]were mentioned that, the site of injury in kidneys involves either (the distal tubules and collecting ducts or the proximal and distal tubules but the glomerulus has no obvious morphologic changes.

However in current study, the glomerular damage, including atrophy, shrinkage, and degeneration of the parietal epithelial cells of Bowman's capsule and some glomeruli exhibiting thickening of the basement membrane was clearly appear in rabbit kidneys of cisplatin treated group. The sites affected probably depend on dose and time duration of drug administration.

In the present work, sections of rabbit kidney treated with therapeutic dose of cisplatin, were observed destruction of the renal tubules, slogging of almost the entire epithelium due to desquamation of tubular epithelium. These changes in the present study are in agreement with the prior observations made by Kim et al.,1995[50] using electron microscopy, wherein it was reported that cisplatin treatment in rabbits caused nephrotoxicity, which was evidenced by significant loss of brush-border microvilli of the renal tubules which is supposed to be responsible for reducing the area for active glucose reabsorption. Furthermore, Vickers et al., 2004 showed acute nephrosis of the tubular epithelium induced by cisplatin in vivo and were reproduced in both human and rat kidney slices[49].

Our study found that the histology of renal sections exhibited remarkable vacuolation, necrosis, desquamation of epithelial cells, and hyaline casts in renal tubules after intraperitoneal treatment with therapeutic doses of cisplatin The toxic effects of cisplatin in the present study were similar to those discovered by Badary et al.,2005 who found that intravenous cisplatin administration caused abnormal kidney function in male wistar albino rats, as evidenced by markedly increased levels of serum blood urea nitrogen (BUN) and creatinine, which are associated with pathologies of the kidney, compared with a control group[26].

The results of the present investigation showed necrosis of renal tubular epithelium with pyknosis of their nuclei and swelling with multi-vacuolations of the cytoplasm with obliteration of their lumina and focal area of severe haemorrhage was observed. These findings are in agreement with Erkurt et al., 2009 who reported acute tubular necrosis has been seen in 30% of patients after even the first course of cisplatin therapy[51]. It had been reported that the nephrotoxicity induced by cisplatin in childhood cancer therapy is accompanied with renal tubular damage, excess elimination of low molecular weight peptides and decreased excretion of some glycoproteins[52]. Another study on cisplatin revealed that about 25% of patients have a significant increase in the

levels of blood nitrogen following 1-2 weeks of treatment which is considered as a main cause for glomerular and tubular damage[181]. Pinta and Lippard suggested that the accumulation of platinum metabolites of cisplatin inside the renal tubule is implicated in induction of nephrotoxicity[53]. Moreover, frequent doses of cisplatin can directly induce oxidative stress renal tubular and glomerular cells resulting in inflammation and apoptosis[54]. However, studies suggest that inflammation, oxidative stress injury and apoptosis probably explai
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part of cisplatin injury. Oxidative stress has been recognised as a major pathway of cisplatin nephrotoxicity. Antioxidant enzymes are inhibited by cisplatin and renal activities are reduced significantly. The reactive oxygen species (ROS) produced directly act on cell components including lipids, proteins and DNA and destroy their structure. The consequence of this is oxidative stress and cell death. The involvement of oxidative stress in cisplatin-induced toxicity may be further supported by the fact that many antioxidants have been reported to prevent cisplatin-induced nephrotoxicity [55].

The disturbance in renal functions by cisplatin is mainly due to its ability to suppress protein synthesis by the renal tubular cells [56] or to promote lipid peroxidation and liberation free radicals in renal tubular cells [57].

Lipid peroxidation (LPO) is a key process in many pathological events and it is induced by Oxidative stress. It is regarded as one of the fundamental mechanisms of cellular damage caused by free radicals having reacted with lipids causing peroxidation that eventually results in the release of products such as malondialdehyde (MDA), hydrogen peroxide (H2O2) and hydroxyl radicals. All cellular components are susceptible to attack by ROS, particularly by hydroxyl radicals[58].

The free radical theory is based on the evidence that living organisms (aerobes) produce oxygen-centered free radicals, capable of inducing irreversible damage to biological structures. These are formed inside body cells when oxygen is used in metabolic processes in order to produce energy. Mitochondrial respiration produces reactive oxygen species (ROS) by leakage of intermediates from the electron transport chain. These molecules are highly unstable because they have an

unpaired electron; they therefore seek to achieve a stable state by appropriating electrons from nearby molecules, these in turn become unstable and so on, thus creating an instability chain reaction[59]. Usually, the harmful activity of a small percentage of these free radicals is inhibited by the natural antioxidants occurring in the cell , such as enzymatic group (superoxide dismutase, catalase, glutathione peroxidase) or by non-enzymatic groups, such as (vitamins E, C, A and natural herbal plants). When, however, the amount of free radical increases (due to chemical toxic, chemotherapeutic drugs, radiation exposure, persistent chronic inflammation, etc.) the pool of antioxidants is saturated and the excess of free radicals damages biological structures. At first the damage is evident in mitochondria, which have the potential to affect the DNA and the RNA causing cell mutations and destruction, but also on proteins and lipids. Endothelial cells and tissue cells are the most affected by oxidative stress. Fortunately, organisms utilize both enzymatic and nonenzymatic antioxidant defenses to minimize cell injury[13]. Further studies explained that cisplatin can induce apoptosis with progressive accumulation of DNA and inhibition of DNA repair pathways, through generation of reactive oxygen species[56].

Aydogan et al., 2008 added that the significant elevation in the renal biomarkers is mainly attributed to the direct cytotoxic effect of cisplatin on the glomerular and tubular structures through the excessive liberation of ROS[60]. Serum creatinine and urea levels are considered the major biomarkers that efficiently mark the glomerular filtration rate. A significant elevation of renal biomarkers was recorded in cisplatin-treated animals[37].

Elbeltagy et al., 2022 reported that the renal damage of cisplatin was apparent from the increased levels of serum urea and creatinine[23]. The elevation in the serum levels of renal biomarkers might be in part due to impaired renal function, tubular obstruction with back-leakage of the renal tubules, inhibition of protein synthesis in the tubular cells or the direct toxic effect of cisplatin on the glomerular and tubular structures through the generation of ROS and enhancement of lipid peroxidation in renal tubules[57].

An early study by Daugaard et al., 1988 confirmed that cisplatin metabolites can inhibit the mechanism involving electrolytes reabsorption especially in the distal segment of the nephron[61]. Furthermore, cisplatin can induce disturbance in renal tubular reabsorption of potassium[62].

Another study clarified that cisplatin can inhibit the activity of antidiurtic hormone leading to hypernatremia[63].

Antioxidants, or free radical scavengers, have the ability to protect cells and tissues exposed to cisplatin from toxicity. Several natural from plants have been widely utilized all over the world either in the pure forms or crude extracts antioxidants were able to improve cisplatin-induced nephrotoxicity via significant enhancement of enzymatic antioxidant activities[64].

Garlic (Allium sativum L.) is one of the World's oldest medicines and has been employed as an antioxidant agent. It is the most famous plant that is widely used to combat several diseases because it contains more than 33 sulfur compounds, enzymes, minerals, amino acids, vitamins including A, B1 and C as well as fibers. The most vital active sulfur compounds in Allium sativum are allicin and alliin which are considered the main antioxidants and scavenging free radical compounds[65].Biochemical studies have demonstrated that garlic extract acts as antioxidants to protect cells against reactive oxygen species (ROS). Garlic extract exerts antioxidant action by scavenging reactive oxygen species, enhancing the cellular antioxidant enzymes, and protects DNA against free radical-mediated cell damage and mutations[66].

The results of the current study, sections from the kidney of animals received therapeutic dose of cisplatin and treated by garlic extract, group (Cp+Ga) revealed less histological damages in renal corpuscles and renal tubules compared with that of rabbits in the (Cp) group. There was reduced the nephrotoxic effect of cisplatin and no distinct pathological changes, except mild glomerular congestion and turbidity of the epithelial cells of the proximal convoluted tubules. Mild tubular degeneration with luminal dilatation and inflammation were seen within the renal cortex. There were slight atrophy and vacuolations of glomerular tuft with slight distension of Bowman's capsule. These findings are following other previous investigators; Nasr and Saleh, 2014[19]; Essam et al., 2014[22]; Nasr and Ibrahim, 2015[24] Abdel-Daim et al., 2020[18] and El-Beltagy et al., 2022 [23].Previous reports have demonstrated that garlic extract can prevent oxidative stress and exerts ameliorative effect against various toxic agents through its vital antioxidant and free radical scavenger's constituents.Other studies revealed that garlic has powerfulcytoprotective effects on the cells of vital body organs[67].

Razo-Rodriguez et al., 2008 were found that the garlic powder feeding was able to prevent by 40–59% the alterations in the markers of renal injury studied, by 33% the histological damage, and by

38–75% the increase in markers of oxidative and nitrosative stress. They concluded that the ability of garlic powder to ameliorate cisplatin-induced renal injury is associated with its antioxidant properties[21].

While Dorrigiv et al., 2020 stated that the garlic and its major components can ameliorate the toxicity of different agents in brain, kidney, blood, liver, embryo, spleen, pancreas, heart, reproductive system in part through radical scavenging, antioxidant effect, reducing lipid peroxidation, anti-inflammatory, cytoprotective activities, increase protein synthesis in damaged tissues and suppressing apoptosis. It is rich in antioxidants which help destroy free radicals that can damage cell membranes and DNA[17].

Anusuya et al., 2013, reported that the protective effect of garlic extract could be attributed to its anti-inflammatory effect that decreased the renal edema induced by cisplatin. These denote the mild tubular degeneration with luminal dilatation and inflammation that were seen in current study.

In the animals pretreated with garlic extract, a remarkable improvement in the serum level of the renal biomarkers was recorded. These results confirm the ability of garlic extract to improve the renal functions through its antioxidant and free radical scavenger activities[20].

Nasr and Ibrahim, 2015 stated that the significant increase in lipid peroxidation accompanied with marked reduction in level and activity of the antioxidant enzymes in the cisplatin -treated animals, whereas pretreatment with garlic extract produced a remarkable improvement in level and activity of renal antioxidant enzymes, with a reduction in lipid peroxidation. These findings provide evidence about the antioxidant effect of garlic extract against cisplatin -induced oxidative stress and lipid peroxidation[24]. Concomitant with the results of our work, the ultrastructure study of Nasr and Ibrahim, 2015 of renal tissues in garlic extract -pretreated rats revealed normal glomeruli, preserved apical microvilli,and restoration of nuclear structure of the proximaltubular epithelial cells. Also, no evidence of inflammatory cell infiltration was observed in garlic extract -pretreated animals. This could be considered a proof of the anti-inflammatory effect of garlic extract. Thus, the inflammation could be considered as a mechanism in cisplatin-induced nephrotoxicity[24].

The authors added that different nuclear apoptotic changes, in the form of membrane budding, cell shrinkage, and marginal condensation of chromatin were observed in cisplatin -treated renal tissue. Disappearance of such morphological changes in the renal tissues of garlic extract -pretreated rats gives a clue to theantiapoptotic property of garlic extract[24].

A number of health benefits of garlic depend on its antioxidant activity. Garlic extracts and components obtained from garlic bulbs were shown to prevent oxidative modification of DNA, protein and lipids by scavenging reactive oxygen species (ROS), increasing the cellular antioxidant enzymes and enhancing glutathione levels inside the cells[68].

The anticancer mechanisms of action of these garlic-derived nontoxic organosulfur -containing compounds include altering mitochondrial permeability, inhibiting angiogenesis, enhancing antioxidative and proapoptotic properties, and regulating cell proliferation. All these effects of garlic's sulfur-compounds have been demonstrated in various human cancers[69]. It was found that the protective mechanism of garlic extract on cisplatin nephrotoxicity may be associated with its scavenging activity, antioxidant potential, anti-inflammatory, andantiapoptotic properties thereby enhancing the activity of antioxidant enzymes and reduce mitochondrial oxidative stress, restored mitochondrial respiratory enzyme activities and attenuated expressions of apoptosis and inflammation related proteins, thus forming the molecular basis for protective mechanism of garlic extract against cisplatin-induced nephrotoxicity.

Conclusion:

According to current study, we conclude that the cisplatin which is a chemotherapeutic drug used in the treatment of various cancer types was determined to cause renal toxicity in domestic rabbits received intrapritoneal dose equivalentto human therapeutic dose. This toxicity manifested by extensive disruption of tissue architecture that was leading to complete destruction of the normal pattern of the renal tissue. There was glomerular congestion and atrophy with wide capsular space, thickening of parietal layer of Bowman's capsule, desquamation with shedding of the tubular epithelial lining cells, extensive multifocal-to-diffuse cortical tubular degeneration and acute necrosis, excessive accumulation of homogenous exudates casts in both proximal and distal convoluted tubules with hydropic degenerative changes in their cells, pyknotic nuclei and cytoplasmic vacuolization, evident loss of the brush border, necrosis and apoptosis of tubular cells. Intertubular hemorrhage and sever interstitial inflammation and edema in renal cortex and outer

medulla. It seems that cisplatin can induce weaken the antioxidant defense systems in tissue of the kidney and to increase oxidative stress together with inflammation induces apoptosis and, on the other hand, the possibility of recovery. It was also determined that garlic extract co-treatment was markedly efficient in reducing reactive oxygen spaces accumulation and strengthening endogenous antioxidant systems leading to a considerable attenuation of the renal toxicity as evidenced by histopathological examinations.

Recommendation:

It was concluded that garlic extract application can be considered as a supportive adjuvant therapy able to reduce or prevent the nephrotoxic effects ofcisplatin by inhibition of oxidative stress. Therefore, we recommend the patients to using the garlic as antiapoptotic and antioxidant supporter against cancer drugs.

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