

Phylogenetic Analysis to Detect COVID Superspreaders

ABSTRACT

Aims: Detection of superspreading events by phylogenetic analysis of nucleotide sequences from a population of individuals collected from a narrow time interval.

Study design: Retrieve nucleic acid sequences, construct multiple sequence alignments, and build phylogenetic networks to determine sources of infection.

Place and Duration of Study: This study was performed at the Delaware Biotechnology Institute of the University of Delaware over the period: June-August, 2022. The data used were from the GIS AID database.

Methodology: Sequences for analysis were sampled from the GISAID initiative's open access SARS-CoV-2 genome database. We selected for high quality nucleotide sequences submitted by Delaware labs between March 18 and April 14, 2021, an important time span of 4 weeks which saw the Alpha variant spread rapidly in the Delaware population.

Results: Four sources accounted for 215 of the 401 sequences. In other words, 54% of all cases were rooted in just five sources.

Conclusion: Thus, superspreading seems to have a major impact upon the proportion of individuals in a population affected with COVID.

Keywords: COVID, superspreaders, phylogenetic networks,

1. INTRODUCTION

"It is now generally thought that superspreading is very common in epidemics, with a rough rule of thumb being that 20% of a population causes 80% of disease cases." (Brauer, 2019 abc) .

Contact tracing by interviewing patients infected with a virus has long been a critical aspect of public health approaches to epidemiology. Since the development of sequencing of pathogen genomes and of powerful mathematical and computational procedures for inferring the evolutionary history of the spread of infections, we have a more direct method of inferring who was infected by whom. Popa *et al.* (2020) noted that "Superspreading events shaped the coronavirus disease 2019 (COVID-19) pandemic." They reported that: "Our results integrating epidemiological and sequencing data emphasize that phylogenetic analyses of SARS-CoV-2 sequences empower robust tracing from interindividual to local and international spreading events. ... This study underscores the value of combining epidemiological approaches with virus genome sequencing to provide critical information to help public health experts track pathogen spread." While a follow-up study by Martin and Koelle (2021) was critical of some of Popa *et al.*'s interpretations, they concluded that: "Small bottleneck sizes also mean that infections generally start off with very little, if any, viral genetic diversity, such that acute infections will likely be characterized by low levels of viral diversity except in instances of superinfection consistent with other recent studies." We

believe that the results of these two studies and others (James, Pitchford, and Plank, 2007; Edholm *et al.*, 2018; Hasan *et al.*, 2020; Jha, Kumar, and Rai, 2020; Goyal, Reeves, and Schiffer, 2021; Popa *et al.*, 2020; Attwood *et al.*, 2022; Li *et al.*, 2022; Müller and Hösel, 2023) make it both easier and highly beneficial to examine other local populations to determine the impact of superspreading more generally. Therefore, we examined a sample from our state of Delaware because we felt that three important criteria are met: (a) a sufficient set of sequence data had been collected to have a reasonable size; (b) the sequences were available for over a period of rapid spread of the disease; and (c) a new variant occurred which was rapidly spreading during a short time frame.

Unfortunately few epidemiological studies account for the significant role of superspreading. In particular, phylogenetic detection of superspreading is understudied particularly when insufficient sequencing is monitoring the course of infection in populations. Only by collecting and evolutionarily analyzing the sequences from the viruses can we infer the fine-scale dynamics of viral spread.

METHODOLOGY:

Phylogenetic analysis is able to contribute to epidemiological studies in six major different ways (Jungck *et al.*, 2006).

- (1) Determining the origin of a pandemic;
- (2) Identifying new variants as containing sufficiently different mutations, they have different levels of infectivity, morbidity, and mortality.
- (3) Determining when such variants evolved (Wang *et al.*, 2020).
- (4) Determining the rate of mutation (Chakraborty *et al.*, 2021 (Figure 1) and Robeva and Jungck, 2023).
- (5) The intensity of selection.
- (6) Determining where such variants evolved.

These investigations fall into three categories identified in different literatures with different taxonomic names that focus on:

- (1) time: molecular clocks or chronocladistics.
- (2) space: phylogeography or topocladistics.
- (3) genealogy: common ancestry or patrocladistics.

Associated with each inquiry are a variety of different phylogenetic methods. For example, in order to build a molecular clock, an important assumption about the distance matrix of differences between sequences inferred from a multiple sequence alignment should satisfy an ultra-metric condition (see Table I).

Table I. Phylogenetic Analysis Software: Assumptions and Applications

Approach	Software	Assumptions	Output
UPGMA <u>(Unweighted Pair Group Method with Arithmetic Mean) algorithm</u>	DendroUPGMA http://genomes.urv.cat/UPGMA/	an important assumption about the distance matrix of differences between sequences inferred from a multiple sequence alignment should satisfy an ultrametric condition; namely, that the rate of mutation is constant over time.	Molecular Clock
Neighbor Joining	MEGA 11: Molecular Evolutionary Genetics Analysis https://www.megasoftware.net/ (Tamura, Stecher, and Kumar, 2021)	pairwise distance estimation plus Bootstrap re-sampling strategy	Distance based phylogenetic tree; i.e., the lengths of each interior edge of the tree is labeled with a distance
Maximum Likelihood	MEGA 11:	Bootstrapping of multiple runs	Confidence of each bifurcation in the phylogenetic tree given as a percentage

Phylogeography	<p>EvoLaps</p> <p>http://www.evolaps.org/</p> <p>Chevenet, F., Fargette, D., Guindon, S. <i>et al.</i> EvoLaps: a web interface to visualize continuous phylogeographic reconstructions. <i>BMC Bioinformatics</i> 22, 463 (2021).</p>	<p>Uses longitude and latitude coordinates for where each</p>	<p>Produces a phylogenetic tree whose terminal vertices are locations</p>
Character-based cladistic approach	<p>Mesquite Version 3.81</p> <p>https://mesquiteproject.org/</p> <p>Maddison, W. P. and D.R. Maddison. (2023). Mesquite: a modular system for evolutionary analysis.</p>	<p>Maximum parsimony;</p> <p>Coding substitutions as transitions, transversions; Noting deletions, insertions, etc.</p>	<p>Good for identifying what happened on each interior edge of the phylogenetic tree</p>
Phylogenetic Networks	<p>SplitsTree</p> <p>https://software-ab.cs.uni-tuebingen.de/download/splitstree4/welcome.html</p> <p>Huson, Klopper, and Bryant. (2008).</p>	<p>When many <u>assumptions about the distance matrix</u> are not met, it is often important to ask how tree-ed is your data.</p> <p>Split decomposition test of quartets</p>	<p>Phylogenetic Network rather than a Phylogenetic Tree</p>

Phylogenetic networks are better than phylogenetic trees when particularly important biological features thought to underlie viral evolution such as recombination and horizontal gene transfer occur. Because several assumptions were not met in our distance matrices from our multiple sequence alignments, we built a phylogenetic network using the Splitstree software (Huson and Bryant, 2006).

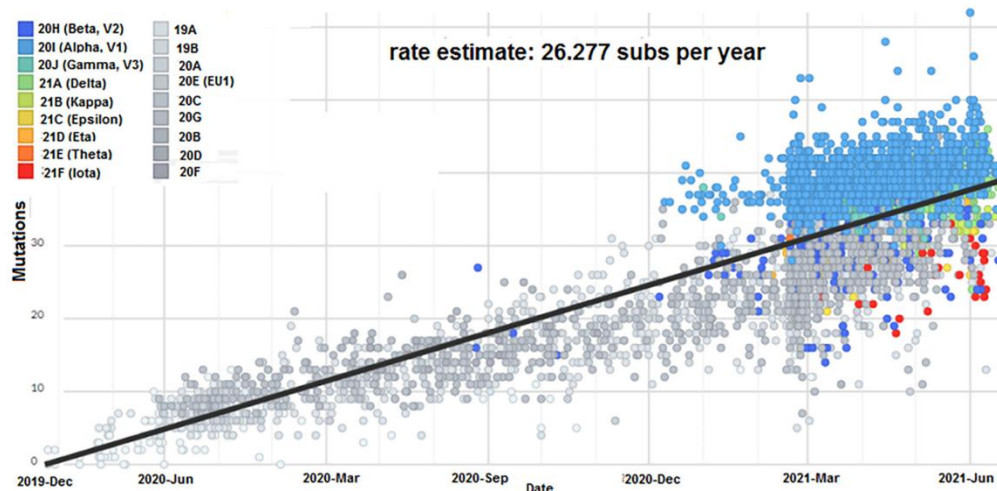


Figure 1. The mutation rate of the COVID 19 virus was determined by Chakraborty *et al.* (2021) to be over 26 substitutions per genome per year. Their legend: “Scatterplot showing the genome diversity cluster of all circulating lineages between December 2019 and June 2021 through the Nextstrain server, using GISAID data.” Creative Commons license level 4.

Choice of period of study: Early on we had access to the work of Hockstein *et al.* (2021) who examined the spread of infection in Delaware. They were able to show that students in a historically black university were safer on campus than in the general population before vaccines became available by doing masking, social distancing, contact tracing, and frequent testing. In their comparison with campus and state data, a second peak of the pandemic occurred in Delaware and on their campus between March 18 and April 14, 2021 (Figure 2).

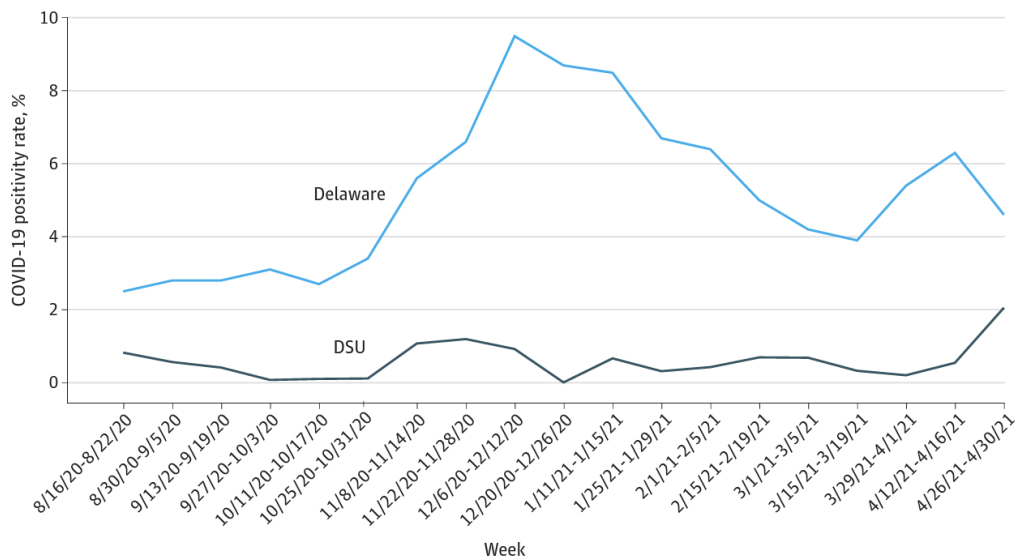


Figure 2. COVID-19 positivity rates of Delaware State University population versus that in the general Delaware population (Hockstein *et al.*, 2021; personal permission).

Source of sequences: GISAID provides open access to sequence data on the coronavirus causing COVID-19 from around the world (<<https://gisaid.org/>>). We selected a genetically diverse dataset of 401 nucleotide sequences. The full genome is roughly 30 kilobases. We decided to focus on the spike protein as it is the primary target of vaccines (Zhu *et al.*, 2021). Thus we chose a region of 3.8k bases which covered the spike protein gene. The sequences were aligned with the MUSCLE (<bi.ac.uk/Tools/msa/muscle/>) multiple sequence alignment tool and the resulting data matrix was entered into the SPLITSTREE software to generate a phylogenetic network.

RESULTS:

Our phylogenetic network of 401 sequences from Delaware COVID patients over the period between March 18 and April 14, 2021 collected from the GISAID database of the region of the COVID virus genome covering the spike protein is shown in Figure 3.

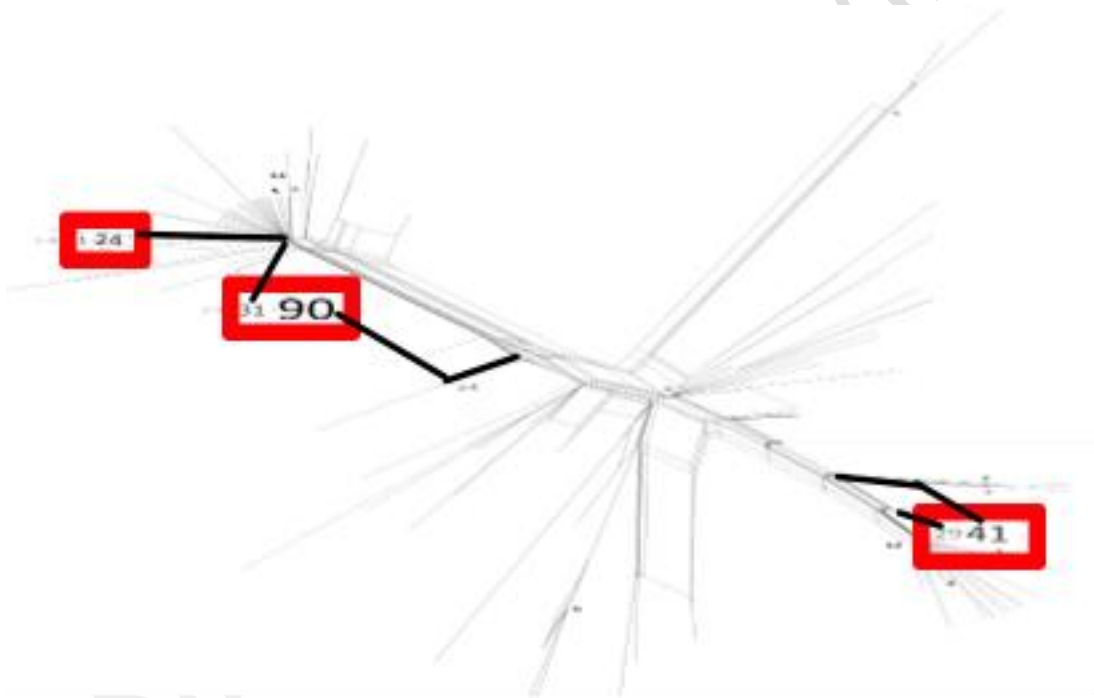


Figure 3. Splits tree generated phylogenetic network of 401 sequences from Delaware COVID patients over the period March 18 to April 14, 2021 collected from the GISAID database.

If we take the five major branches of duplicate sequences ($90 + 41 + 31 + 24 + 29$), these account for 215 out of the 401 cases, or 54% of the cases. This is far above Brauer's (2019 abc) estimate of 20%. We believe that this is consistent with the hypothesis of superspreaders being responsible for a significant fraction of infections in a restricted population in a short interval of time.

Conclusion:

Within a genetically diverse dataset of 401 nucleotide sequences, large amounts of duplicate Spike sequences were present. Since we were investigating fairly long lengths — 3.8 kilobases — of these nucleotide sequences, we believe that the presence of so many identical sequences is consistent with the hypothesis that they originated from the same or highly similar genetic sources and hence are evidence of superspreading.

Super-spreading provides a plausible explanation of the large number of identical sequences observed in the analysis, positing, for example, that many of the 90 infectious cases highlighted in week 4 of Figure 3 could be traced back to an earlier case which may have been one recorded in a previous week. If this data were corroborated by patient data and contact tracing of identical sequences, it would confirm the likely hypothesis that super-spreading is visible at the nucleotide level of a dataset and can be identified using phylogenetic analysis. Franke *et al.* (2022) subsequent to our work give a larger framework for the transmission of COVID-19 in Delaware but they do not explicitly address the issue of superspreaders.

These findings provide grounds for investment in future studies assessing whether phylogenetic analysis could be used in the estimation of contact tracing based off sequence data rather than patient data.

Subsequent to our work, Taube, Miller, and Drake (2022) published “An open-access database of infectious disease transmission trees to explore superspreader epidemiology.” Their work will make it much easier for individuals to build upon our and others’ work in phylogenetic analysis of COVID transmission. Such studies are crucial for setting public health policy.

In addition, we are aware that there is some controversy about the use of phylogenetic network analysis. Chookajorn raised an alarm about a previous phylogenetic network analysis by Foster *et al.* (2020a): “As an evolutionary biologist working in a developing country, I have experienced firsthand how sensational findings can influence decision-making processes by diverting time and resources to control virus strains deemed to be “more aggressive.” In the fog of war, scarce resources are allocated in haste, and the developing world does not have well-informed science advisers sitting in every key meeting to help provide balanced scientific viewpoints. The scientific community, as a whole, needs to be extra cautious in interpreting new findings related to corona virus disease 2019 (COVID19), and any potential misinformation must be promptly addressed.” While Foster *et al.* (2020b) submitted a rejoinder, we believe that the sensitivities such important policy ramifications need heterogeneous stakeholders with multiple perspectives to be at the decision table.

Finally, we are particularly worried that since public concern about the ongoing COVID pandemic has decreased and concomitantly there has been less funding for sequencing current strains in the broad international community that we are facing a situation where even doing phylogenetic analysis of available sequences may be insufficient to identify many newly evolving variants and their associated levels of infectivity, morbidity, and mortality.

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