

## Review Article

# **Recent Advances for Detecting and Addressing Plant Disease: Towards future farming**

### **Abstract**

Pests and pathogens inflict enormous financial harm to the global farming industry. Monitoring plant health and early pathogen detection are essential for facilitating successful management strategies and preventing the spread of disease. Various traditional methods and serological techniques have been found, time consuming and require handling skill, also the reliability of the result is uncertain and during asymptomatic stages it is hard to diagnose the pathogen. Hence the, innovative sensors based on host reactions assessment, phage display-based biosensors, and bio-photonics on combination with other system, remote sensing techniques integrated with spectroscopy-based approaches allow for high spatialization of data, these techniques could particularly be of immediate benefit for initial identification of infection and early control with limiting the use of Systemic Fungicides and developing a sustainable environment with high yield.

**Keywords: Bio-photonics, Bio-sensors, Pathogen, Remote Sensing and Spectroscopy**

### **Introduction**

Plant diseases contribute to huge economic losses to hectares of crop lands and post-harvest agricultural products. Plant disease epidemiology is controlled by many a large number of components, actors and factors. With the advancement of new tools for monitoring and crop modelling techniques studies of various environmental factors have become comparatively easy. Various computer-based simulation programmes have been developed via Artificial intelligence that regularly monitor the crop health. Earlier through aerial photography and imaging techniques, geography and land patterns were recognized for commercial crops as during 1920s to record the severity of cotton root rot, pictures were taken through helicopter imaging technique.

The disease management in crop is triggered by various heterogenetic factors as topographic conditions, soil condition, neighbouring fields, micro environment of crop, and sources of pathogen inoculum which often result in varied disease manifestation. Pattern of disease may vary from site to site, year to year and overtime during epidemic in a particular field. Plant disease epidemics are cyclic in nature and spread repeatedly in relation to host and environment. The inoculum development consists of fungal spores, bacterial cells, nematodes, viruses or through vectors, which gain entry and establish themselves via infection of host. The pathogen established within the host develops new inoculum, which further disseminates to susceptible sites to initiate new infection. Pathogens that produce only one cycle of infection per crop cycle are called monocyclic pathogens whereas poly cyclic pathogens produce more than one infection cycle per crop cycle. Thus, accordingly farmers have to extend the visual rating of disease incidence and severity to a nominal number of samples to decide whether there's a requirement of field spray or not.

The heterogeneity of disease spread helps to decide the quantity of fungicide sprayed during the crop growing season, which helps in minimizing the undesirable environmental

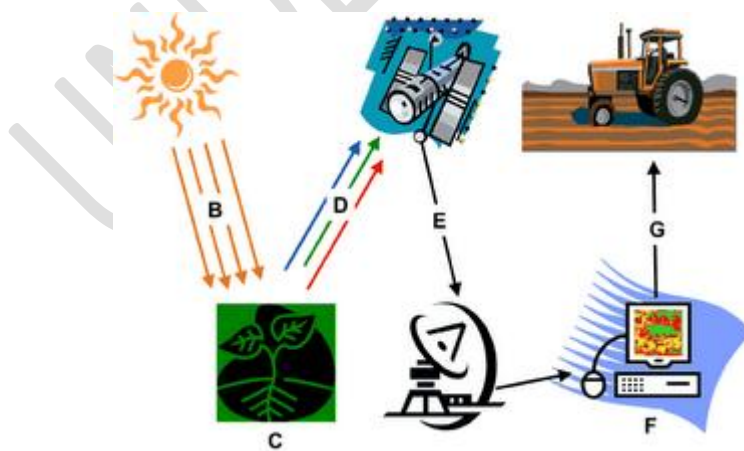
contamination by spraying only when required. This will also decrease the selection of fungicide resistance in pathogens and slow down the rate of strain variability in pathogens. Thus, essentially a site specific monitoring of plants is necessary to determine whether the plant is diseased and to what degree also whether there is a requirement of disease control. The fungicide usage was significantly reduced in France without facing any negative impact on productivity by 47%. With 59% of farm profitability.

The present detection and diagnosis of plant disease currently rely on visual rating of plants, and in case of doubt specialized techniques as nucleic acid assay, ELISA, qPCR or lab based serological technologies are used. But these techniques cover only a representative sample and are often time taking. The indirect remote sensing methods as thermography, fluorescence imaging and spectral techniques allows repeated monitoring of potentially crop of interest. The Gas chromatography and electronic nose (e-nose) techniques require the volatile organic samples or VOCs released by infected plant or the infecting pathogen which detect the presence of disease in individual field.

Diseases (caused by fungus, oomycetes, bacteria, and viruses) have been estimated to account for 16 percent and 11 percent of the feasible crop yield. Plants must be protected from diseases and other pests, it is widely agreed. Integrated pest management (IPM) systems combine mechanical, biological, and chemical instruments with other supporting technologies to achieve effective, efficient, and sustainable pest management.

In agricultural crops, disease control often assumes a uniform pattern of disease spread, thus crops are sprayed at uniform application rates. However, crop heterogeneity resulting from changes in soil conditions, topography, adjoining fields, microclimatic conditions, and pathogen inoculum sources frequently leads in a heterogeneous distribution of illnesses displayed as patches, gradients, or random patterns. Remote sensing can be employed as a first step in disease management at a given location, as well as to profile plant genotype responds to pathogen assault.

Integrated pest management (IPM) systems combine mechanical, biological, and chemical instruments with other complementary technologies to achieve effective, efficient, and long-term pest management.



(Source: NDSU, 2004)

**Fig:1** Plants (C) receive electromagnetic energy (B) from the sun (A). The leaves allow some of the electromagnetic radiation to pass through. The reflected energy (D) is

**picked up by the satellite's sensor. After then, the information is sent to the ground station (E). The data is examined (F), and field maps are displayed (G).**

### Scope

For a prolonged period, crop diseases detection and diagnosis were based on human assessment and the operator's specialised knowledge. Various serological techniques Such as DNA based technologies are often time consuming and required significant improved facilities and skill for plant pathogendiagnosis and management.

Whereas Sensors should be unbiased, exact, fast, and available 24 hours a day, seven days a week.Plant disease sensors can be used once for quality control (for example, by the food industry or quarantine authorities) or they can be linked into autonomous systems for constant monitoring of products for plant pathogens, i.e., examining and maintaining a complete record.A systemic assessment of a crop using technical sensors can enable the operator to take action when illnesses are detectable or exceed action threshold levels. Sensors ought to be able to

- A) recognising a pathogen-caused change in the health state of the crop
- B) determining the disease
- C) determining the disease's severity

The ability to distinguish between different potential diagnosis based on disease-specific signs is required for disease diagnosis.For imaging systems, quantifying typical illness symptoms (disease severity) and assessing leaves infected by many pathogens is simple, but non imaging sensors and sensors with insufficient spatial resolution face a problem.Disease detection is the impression of a variation from a healthy crop/fruit in some situations. There is no need to identify or quantify sickness in these situations.For the detection of plant diseases, the Global Plant Protection Convention's protocols combine phenotype, immunological, and genomic techniques; these methods give complimentary information(Martinelli *al.et.*,2015).Physical sensors allow for the employment of autonomous systems for quarantine inspections, which can be used as a first step in identifying suspect material that can then be tested and validated using molecular techniques for pathogen detection.

Table-1 Disease identification: possible methods, parameters, and users

scope	Application	Environmental circumstances	User
Monitoring of quarantine	Plant quality and safety inspections	Semi-controlled	Exporters, importers, and regulatory agencies are all involved.
Plant product quality assurance, particularly for postharvest illnesses		controlled	Food enterprises
Forstry	Crop health is being closely monitored.	Woods,forest plantations	Forest management on both private and public lands
Field Crops		Field	Contractors in the agricultural industry

Speciality crops grown in greenhouse		Semi-controlled	Gardeners
Plant genotype responses phenotyping	Disease resistance selection in crops	Semi-controlled	Plant breeders

Source-Oerke.*et. at.*,2020

Post - harvest sensing of the safety and quality of (processed) plant products in food business comprises assessing ripeness, colour, and storage compatibility; Detecting flaws, bruising, and diseases in fruits and vegetables, as well as determining mycotoxin contamination from fungus, such as *Aspergillus flavus* in maize kernels(Yao *et. at.*,2013).Sensors and high-throughput platforms thrive in highly regulated environments. The sorting and grading procedures necessitate real-time decision-making based on sensor data.Under controlled conditions and in field stations, crop germplasm can be phenotyped for disease susceptibility.

Phenotyping devices need advanced hardware, yet they deliver high throughput and frequently monitor the plants numerous times during the growing cycle without needing immediate data analysis. While administering the target pathogen vaccine, disorder identification is not necessary, but disease quantification is essential.

### Theoretical foundation and definition

By detecting the electromagnetic radiation reflected/backscattered or emitted by the Earth's surface, Remote sensing is a technique for collecting information on an object without physical touch(Jong and Meer, 2006) In this application, RS is an indirect assessment technique that can monitor vegetation conditions from a distance and determine the spatial breadth and patterns of vegetation attributes and plant health.sensors classified as active or passive sensors create artificial radiation and detect reflected or backscattered energy, whereas passive sensors monitor reflected solar radiation or emitted thermal radiation (passive sensors)example of active remote sensing instrument include Radar and lidar.Various passive instruments used to measure solar radiation in the visible(VIR range400-700 ),near infrared (NIR range 700- 1100 nm) and short infrared (SWIR-1100-2500 nm) and the thermal infrared (TIR-3 to 15 µm) region are used in remote sensing for plant disease detection.

Variables defining canopy structure, such as leaf area and orientation, spatial arrangement, and roughness, as well as the optical, dielectric, or thermal properties of the plant element, all have an impact on the spectral signature of vegetation (Baret *et. at.*,2007).

Also Checking ripeness, colour, and storage appropriateness of fruits and vegetables; detecting flaws, bruising, and diseases of fruits and vegetables; and assessing mycotoxin contamination from fungi, such as *Aspergillus flavus* in maize and peanut kernels are all examples of quality control in the food sector.Sensors and high-throughput platforms thrive in highly regulated environments. The sorting and grading procedures necessitate real-time decision-making based on sensor data.Thus, for caring out such operation smoothly sensor-based data appreciately increase the shelf life of marketable products.

### Disease sensing and data processing

Options for recording the technical sensor data on diseases manifestation is gaining popularity. The availability of structural and physiological features of tree tops damaged by insect pests and fungal infections is likely to be enhanced by spectral information and 3D data from remote sensing (Stone *et. at.*, 2017). Digital imaging, chlorophyll fluorescence, spectrum imaging, thermographic imaging, and volatile chemical detection are all common sensor systems. Less commonly used techniques include magnetic resonance imaging, soft X-ray imaging, and ultrasound. Non-invasive approaches provide a lot of promising and inspection analysis for post-harvest underutilized crops (Ruiz *et. at.*, 2010). These techniques are categorized 1. disease identification and severity evaluation using correlation and regression analysis; 2. generation of disease-specific indices using general spectral vegetation indices (SVIs); 3. data-mining methods used to data processing and feature extraction for data dimensionality reduction 4. automated/Machine learning (ML). Plant-pathogen interactions have a wide range of uses, from disease monitoring to the prediction of molecular pathogen effectors *spp.* (Sperschneider *et. at.*, 2020). Robust ML performance necessitates a thorough understanding of both methodology and biology.

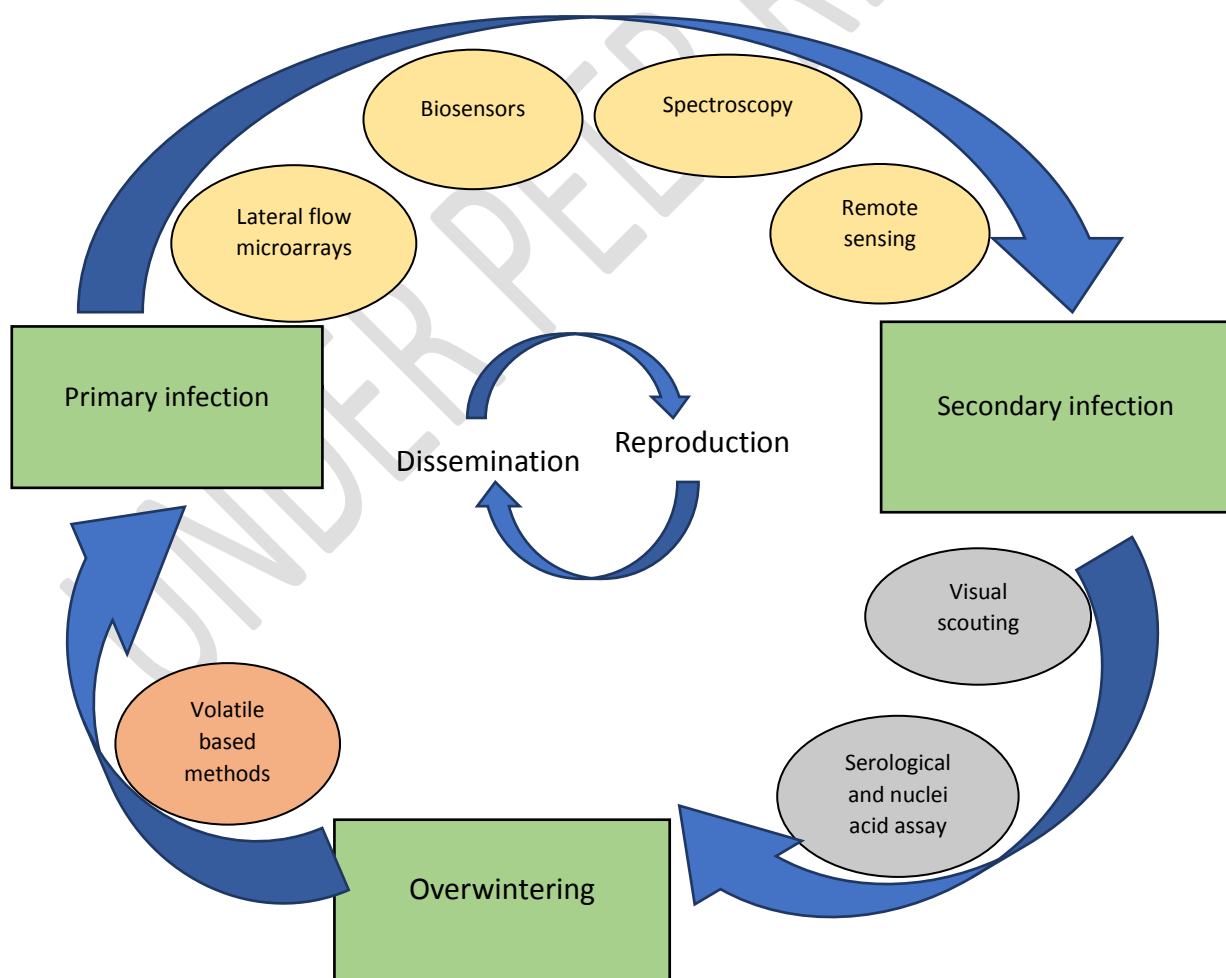


Fig.2: Methods indicated for time of detection and diagnosis of Polycyclic diseases

### Sensor used for plant disease

Sensors are classified by (1) the electromagnetic spectrum range they cover, such as visible (VIS), near-infrared (NIR), short-wave infrared, thermal infrared, and radar; (2) the scale/platform they operate on, such as remote sensing strictly, airborne and spaceborne, UAV, ground-based/proximal, and microscopic; and (3) the recording principle they use, such as passive sensors that record radiation emitted by an object (thermography) or solar radiation reflectance (RGB, spectrum cameras)—active sensors [LIDAR (light detecting and ranging), SAR (specific absorption rate), fluorescence] emit a unique detecting radiation and recording its changes as a result of interactions with the target object—(4) the type of data recording (imaging vs. non-imaging). (Oerke *et. al.*, 2018)

### Spectral information

Imaging systems with spatial resolutions ranging from a few hundred to millions of pixels per image can record (a) one waveband or the sum of all wavebands in the 400–700 nm spectrum (panchromatic); (b) the three basic colour components red, green, and blue (typical bandwidth 60–80 nm, e.g., smartphone RGB cameras); and (c) additional (NIR) bands (multispectral, discrete, and somewhat ambiguous). (d) narrow spectral bands over a continuous spectral range (hyperspectral, narrow wavebands 1 nm spectral resolution). As opposed to RGB data, hyperspectral data has additional spectral information. Each pixel is a vector with a dimensionality equal to the number of wavebands recorded. The implications of scale and environment on the collection and analysis of hyperspectral data on plant diseases, as well as the translation of this technology from controlled circumstances to the field, were explored by Thomas *et. al.*, (2018). Under normal growing conditions, powdery mildew, for example, is virtually undetectable. Thermography may be ideal for pre-symptomatic detection of pathogen activities within plant tissue because this passive technology is particularly sensitive to early changes in transpiration (Mahlein *et. al.*, 2016; Oerke *et. al.*, 2011).

### Thermography

Infrared thermography is a technique for determining the surface temperature of leaves, plants, or crop canopies based on their water status, particularly stomatal and cuticular transpiration (Costa *et. al.*, 2013). Root infections, reduced water movement within stems, changing stomatal aperture, and changes in cuticular conductance are examples of diseases impacting the plant's water status that can be detected thermally (Oerke *et. al.*, 2018).

### Fluorescence of Plants

Pathogen attack affects the plant's photosynthetic apparatus, such as pigments, the electron transport chain, and Calvin cycle enzymes, either directly (necrosis) or indirectly (feedback regulation of the electron transport chain) by reducing photosynthetic leaf area and chlorophyll degradation (chlorosis) chain of transportation. For high-throughput phenotyping, a combination of pulse–amplitude–modulation chlorophyll fluorescence systems and image analysis utilising dark-adapted plants was suitable for assessing the impacted leaf area (Rousseau *et. al.*, 2015). As early as four days after inoculation, fluorescence spectra were beneficial in distinguishing brown rust–infected tissue from healthy wheat tissue (Tischler *et.*

*et. al.*, 2018). Similarly, the quantum yield of photosystem II and nonphotochemical quenching could be used to identify the reactions of barley genotypes with different levels of resistance to *Blumeriagraminis* f. sp. *hordei* (Brugger *et. al.*, 2018).

### **Electronic Nose**

Various Volatile Organic Compounds (VOC) are emitted by Plants on Disease and insect manifestation. E-noses can detect (some of) these VOCs; for specific applications, gas chromatographic headspace analytics or tailored commercial e-noses (e-sensing) are available. Disease-specific VOCs of fire blight of apple, grey mould of tomato, and powdery mildew of tomato have been found as phenylethyl alcohol, -copaene, and fluoro-aliphatic hydrocarbons, respectively (Wilson *et. al.*, 2018). Also, E-noses are used to monitor food quality and production operations, as well as to detect fruit and vegetable postharvest infections.

### **Gathering other informations by sensors**

Plant illnesses and plant products can be detected using nuclear magnetic resonance (NMR) and X-ray imaging techniques (Sankaran *et. al.*, 2015). Internal bruising and Spraying disease signs in potato tubers were investigated using NMR imaging, as well as the difference between belowground damage caused by *Heterodera schachtii* and *Rhizoctonia solani* in sugar beets (Thybo *et. al.*, 2004; Hullnhutter *et. al.*, 2012). An additional underlying layer of melon seeds infected with Cucumber green mottle mosaic virus was discovered using optical coherence tomography (Lee *et. al.*, 2011). Similarly, on persimmon leaves, biophotonic examination with a 1,310-nm swept-source optical-coherence tomograph was effective in detecting morphological variations between healthy leaves and those with round leaf spots (Wijesinghe *et. al.*, 2016).

### **Ground based systems**

Despite the development of effective sensing techniques and technologies, most of them still require a controlled environment for data gathering in order to prevent false positives. An autonomous ground-vehicle robot designed for high-throughput in-field agricultural row-crop phenotyping was used to evaluate plant height and canopy closure data using the normalised difference vegetation index (NDVI) assessment using HIS (Underwood *et. al.*, 2017).

### **Conclusion**

The ability of remote sensing systems to detect plant diseases varies. Although highly sensitive to pathogen-induced changes in plant metabolism, chlorophyll fluorescence and thermography lack the ability to identify illnesses and distinguish them from abiotic symptoms and effects from arthropod activities (Oerke *et. al.*, 2014). Spectral data, when combined with spatial data from photos and data on VOCs generated by diseased plants, appears to be useful for disease detection and categorization. However, thermography and fluorescence can be employed in crop monitoring to detect anomalies, followed by a visual check of problematic plants or locations.

Sensing is a tool for determining the severity of a condition, not a requirement or an active control mechanism for disease treatment. Sensors cannot replace the use of fungicides and mechanical disease control devices, but they can help direct (and focus) the actuator(s) to the plants or areas that need attention, hence assisting in the reduction of the amount of chemical

used. Only when effective curative treatment alternatives, like as systemic fungicides, are available it is feasible to detect polycyclic diseases. Sensors can be used to estimate disease-related production losses, determine which plants should be removed from the cultivated area, and evaluate which sections of the field should be kept uncultivated in the growing season due to the presence of soilborne diseases that are resistant to other methods of management, such as crop rotation. However, employing these techniques in individual farm is costly but cumulative efforts for large cultivable areas both with Governmental policies and individual deed successful results with low yield loss and high eco- friendly environment can be achieved.

## References

- Baret F, Houlès V, Guèrif M 2007 Quantification of plant stress using remote sensing observations and crop models: the case of nitrogen management. *J Exp Bot* 58:869–880. doi:10. 1093/jxb/erl231
- Bock CH, Poole GH, Parker PE, Gottwald TR. 2010. Plant disease severity estimated visually, by digital photography and image analysis, and by hyperspectral imaging. *Crit. Rev. Plant Sci.* 29:59–107
- Bravo C, Moshou D, West J, McCartney A, Ramon H. 2003. Early disease detection in wheat fields using spectral reflectance. *Biosyst. Eng.* 84:137–45
- Brugger A, Kuska MT, Mahlein AK. 2018. Impact of compatible and incompatible barley: *Blumeriagraminisf.sp. hordei* interactions on chlorophyll fluorescence parameters. *J. Plant Dis. Prot.* 125:177– 86
- Costa JM, Grant OM, Chaves MM. 2013. Thermography to explore plant–environment interactions. *J. Exp. Bot.* 64:3937–49
- De Jong S, van der Meer FD 2006 Remote sensing image analysis: including the spatial domain. Springer editions, Dordrecht
- Hillnhuetter C, Sikora RA, Oerke EC, van Dusschoten D. 2012. Nuclear magnetic resonance: a tool for imaging belowground damage caused by *Heterodera schachtii* and *Rhizoctonia solani* on sugar beet. *J. Exp. Bot.* 63:319–27
- Lee C, Lee SY, Kim JY, Jung HY, Kim J. 2011. Optical sensing method for screening disease in melon seeds by using optical coherence tomography. *Sensors* 11:9467–77
- Mahlein AK, Alisaac E, Al Masri A, Behmann J, Dehne HW, Oerke EC. 2019. Comparison and combination of thermal, fluorescence, and hyperspectral imaging for monitoring *Fusarium* head blight of wheat on spikelet scale. *Sensors* 19:2281
- Martinelli F, Scalenghe R, Davino S, Panno S, Scuderi G, 2015. Advanced methods of plant disease detection. A review. *Agron. Sustain. Dev.* 35:1–25
- Oerke EC, Froehling P, Steiner U. 2011. Thermographic assessment of scab disease on apple leaves. *Precis. Agric.* 12:699–715.
- Oerke EC, Mahlein AK, Steiner U. 2014. Proximal sensing of plant diseases. In *Detection and Diagnostics of Plant Pathogens. Plant Pathology in the 21st Century*, Vol. 5, ed.

- ML Gullino, PJM Bonants, pp. 55–68. Dordrecht, The Netherlands: Springer Netherlands
- Oerke EC, Steiner U. 2010. Potential of digital thermography for disease control. In *Precision Crop Protection: The Challenge and Use of Heterogeneity*, ed. EC Oerke, R Gerhards, G Menz, RA Sikora, pp. 167–82. Dordrecht, Neth.: Springer
- Oerke EC. 2018. Precision crop protection systems. In *Precision Agriculture for Sustainability*, ed. J Stafford, pp. 347–97. Cambridge, UK: Burleigh Dodds Science
- Oerke, E.C., 2020. Remote sensing of diseases. *Annual review of phytopathology*, 58, pp.225-252.
- Rousseau C, Belin E, Bove E, Rousseau D, Fabre F, 2013. High throughput quantitative phenotyping of plant resistance using chlorophyll fluorescence image analysis. *Plant Methods* 9:17
- Rousseau C, Hunault G, Gaillard S, Bourbeillon J, Montiel G, 2015. Phenoplant: a web resource for the exploration of large chlorophyll fluorescence image datasets. *Plant Methods* 11:24
- Ruiz-Altisent M, Ruiz-Garcia L, Moreda GP, Lu RF, Hernandez-Sanchez N, 2010. Sensors for product characterization and quality of specialty crops: a review. *Comput. Electron. Agric.* 74:176–94
- Sankaran S, Khot LR, Zuniga Espinoza C, Jarolmasjed S, Sathuvalli VR, 2015. Low-altitude, high-resolution aerial imaging systems for row and field crop phenotyping: a review. *Eur. J. A*
- Sperschneider J. 2020. Machine learning in plant-pathogen interactions: empowering biological predictions from field scale to genome scale. *New Phytol.* <https://doi.org/10.1111/nph.15771>
- Stone C, Mohammed C. 2017. Application of remote sensing technologies for assessing planted forests damaged by insect pests and fungal pathogens: a review. *Curr. For. Rep.* 3:75–92
- Thomas S, Kuska MT, Bohnenkamp D, Brugger A, Alisaac E, 2018. Benefits of hyperspectral imaging for plant disease detection and plant protection: a technical perspective. *J. Plant Dis. Prot.* 125:5–20
- Thybo AK, Jespersen SN, Laerke PE, Stodkilde-Jorgensen HJ. 2004. Nondestructive detection of internal bruise and spraing disease symptoms in potatoes using magnetic resonance imaging. *Magn. Reson. Imaging* 22:1311–17
- Tischler YK, Thiessen E, Hartung E. 2018. Early optical detection of infection with brown rust in winter wheat by chlorophyll fluorescence excitation spectra. *Comput. Electron. Agric.* 146:77–85
- Underwood J, Wendel A, Schofield B, McMurray L, Kimber R. 2017. Efficient in-field plant phenomics for row-crops with an autonomous ground vehicle. *J. Field Robot.* 34:1061–83

Wijesinghe RE, Lee SY, Kim P, Jung HY, Jeon M, Kim J. 2016. Optical inspection and morphological analysis of Diospyros kaki plant leaves for the detection of circular leaf spot disease. *Sensors* 16:1282

Wilson AD. 2018. Applications of electronic-nose technologies for noninvasive early detection of plant, animal and human diseases. *Chemosensors* 6:45

Yao H, Hruska Z, Kincaid R, Brown RL, Bhatnagar D, Cleveland TE. 2013. Detecting maize inoculated with toxigenic and atoxigenic fungal strains with fluorescence hyperspectral imagery. *Biosyst. Eng.* 115:125–35

<https://www.ag.ndsu.edu/>

UNDER PEER REVIEW