

Revitalizing the Potential of Minor Millets: Agrarian Constraints, Possible Solutions and Future Roadmap

ABSTRACT

In contrary to the recent revival of millets like sorghum & pearl millet globally, one segment of millets is still neglected mostly in spite of their massive nutritional benefits and complementing effect on ecology; these are referred as minor millets collectively. The focal causes of its **under recognition** include lack of awareness among the consumers about its vast potential utilities and multifaceted constraints acting together to curb down its production pattern. Lack of effective research and development in the area of crop improvement via breeding and biotechnological interventions is holding back its upswing. There is very little comprehensive documentation available on these dynamics of minor millets. This paper studies those critical limiting factors while also prescribing the way out to ensure nutritional security and environmental sustainability.

Keywords: small millets, proso millet, foxtail millet, kodo millet, barnyard millet, little millet, brown top millet, sustainability

1. INTRODUCTION

Minor millets are an agronomic community of small seeded coarse-grained food crops under family Poaceae excluding Sorghum and Pearl millet. It includes Foxtail Millet (*Setaria italica*), Proso Millet/ *Cheena* (*Panicum miliaceum*), Kodo Millet (*Paspalum scrobiculatum*), Barnyard Millet / *Sanwa* (*Echinochloa crus-galli*), Little Millet/ *Kutki* (*Panicum sumatrense*), Browntop Millet (*Brachiaria ramosa*), Job's Tear (*Coix lacryma-jobi*), Fonio (*Digitaria exilis*) and Teff (*Eragrostis tef*). The term 'minor' reflects the fact that these grains are neglected in terms of any significant commercialization, acreage, production, consumption and research investment.

Minor millets have been an integral part of our culture since ages as mentioned in **main** ancient texts such as Kautilya's *Arthashastra*, *Ain-i-Akbari* etc. However, the aftermath of the green revolution pushed these minor millets into the abyss. The area declined drastically from 2.44 Mha in 1990-91 to 0.45 Mha in 2019-20 and the production shranked from 1.19 M tonne to 0.37 M tonne [1]. But lately, there is an evident paradigm shift as a result of evolved health consciousness among the consumers and its potential impact towards environmental sustainability. Minor millets are preferred over staple cereals due to their superior nutritional quality, low glycemic index, gluten free nature, competency to combat climate change, adaptability in low-input high-stressed agro ecological conditions. In the avenue of unleashing its huge potential, there are certain obstacles at both land and lab levels. This paper specifies those constraints and also broadens the horizon of crop improvement by suggesting promising prospective interventions and the way forward.

2. UTILIZATION

2.1 Minor Millets as Crop Models

Studies show that exogenous application of Selenium in Foxtail and Proso millet has several encouraging results, especially in terms of growth stimulation, increasing biomass and photosynthetic pigments which can be used as a model for studying salinity stress at different stages of crop growth [2]. Another major breakthrough comes from the recent work by Yang &

colleagues led to the development of *Xiaomi*, a Foxtail Millet mutant with fully known genome sequence ($2n=2x=18$), efficient transformation, short growth cycle and tiny plant dimensions as an ideal C4 plant model for studying various cellular, molecular, biochemical and physiological traits [3].

2.2 Minor Millets as Baby Food

Millets are well stocked with antioxidant phytochemicals, have a low glycemic index & are primarily alkaline [4]; thereby, promoting body health & providing a more diverse food basket for nutritional improvement. Proso millet is rich in lecithin which keeps the nervous system healthy. It also has the highest amount of phosphorus amongst all other millets essential for cell growth and development as well as for good immunity. Barnyard millet is rich in Iron that keeps Anaemia at bay. Little millet is rich in Phosphorus and good cholesterol, suitable for weight gain in babies. Foxtail millet has higher protein content than wheat and also has 30 times richer fiber content than rice. Healthcare experts disapprove of polishing and extensive processing of millet grains as their nutritional quality will then be put at risk.

2.3 Minor Millets for Biofuel Production

Studies have yielded encouraging results which support the use of foxtail millet as an alternative lignocellulosic feedstock for bioethanol production [5]. A comparative study of Barnyard millet, Foxtail millet & Little millet revealed that the husks of all three can be used to produce bioethanol; however, the concentration of bioethanol observed was greater in Barnyard millet husk [6]. On the other hand, Proso millet & Maize have similar starch content with fermentation efficiencies ranging from 84% to 91% whereas it was observed to be 97% in waxy type lines [7]. This proves that Proso millet has high potential to be used in fuel ethanol production.

2.4 Minor Millets in the Indian Food Basket

Minor millets have always been a culturally important part of our diet for centuries and are commonly consumed in the form of *roti*, *khichdi*, *upma*, *dosa*, puddings or cake. These traditional preparations possess antimicrobial, antioxidant, probiotic and prebiotic properties. Barnard & Little millet are specifically consumed during fasting [8]. In some tribal areas, grains of foxtail millet is cooked as millet rice or consumed as '*sargati*' (a stiff porridge); sprouted millet grains are used as vegetables whereas, in Uttarakhand, Barnyard millet is consumed as '*paleu*' or '*chencha*', a thick sticky savory dish [8]. Barnyard Millet is also used to make kheer & halwa. '*Juma*' is a treasured recipe of *Lahaul Spiti* wherein sheep intestine is stuffed with spiced millet flour, steam cooked and served with piping hot mutton soup [9]. Kodo millet can also be used as rice substitute.

2.5 Minor Millets as animal feed & fodder

Foxtail millet seeds have been used as bird feed and for fodder [10]. Barnyard millet has a high straw yield and fodder value even at multiple cuttings [11] (about 6.3 tons/ha) which is rich in protein (7.6%), dietary fiber (23%), ash (12%) and fat (2.0%). Proso millet combined with oats is used as a starter for calves. Its grains are pelleted with other grains for feeding multiple types of poultry. Ground Proso millet is used as lamb fattening feed and is also used in dog and hamster food.

2.6 Other Non-Conventional Uses

Millet hulls, the by-product remaining after dehulling of millet grains, is sometimes used as a filling material (as in pillows) due to their high fibrous content [12]. Barnyard Millet starch is rich in amylose and is used in combination with borage seed oil for manufacturing biodegradable films or biofilm packaging material which are resistant to microorganisms and prevents free radical formation [13]. Aqueous extract of *E. colona* can be used in biosynthesis of Silver nanoparticles as a safe and eco-friendly approach for use of nanoparticles at a large scale in varied fields such as medicine, agriculture, forensics, biotechnology, engineering etc [14]. A broad-spectrum antifungal peptide, EcAMP1 has been identified in the seeds of Barnyard millet (*E. crus-galli*) which can be used in the synthesis of novel antimicrobials through protein engineering [15].

3. MINOR MILLETS FOR SUSTAINABILITY

3.1 Climate Resilience

Climate change is already having an impact on agricultural production, food stability and nutritional security in a number of countries especially in developing countries. Up surging temperatures, unpredictable rainfall patterns and continued droughts occur during critical crop phases, resulting in shorter grain filling periods, subordinate yields and diminished biomass. On top of that, high temperatures encourage speedy evaporation, which dries out the soil and makes it more difficult for plants to get enough water, reducing their ability to absorb nutrients. Because the importance of small millets is becoming more widely recognized [16]. Millets are well known for their climatically resilient characteristics, which include adaptability to a wide range of ecological conditions, reduced irrigation requirements, improved growth and productivity under low nutrient input conditions, reduced reliance on synthetic fertilizers, and low vulnerability to environmental stresses [17].

3.1.1 Physiological benefits

Millet has a number of biochemical, morpho-physiological and molecular characteristics that make them more resistant to environmental stresses than cereals. Millets have a shorter life cycle (12-14 weeks from seed to seed) than rice and wheat, which can take 20-24 weeks. This assists people in avoiding stress. Short stature, thickened cell walls, small leaf area and the ability to form dense root systems, on the other hand, mitigate the prevalence and effects of stress conditions [18]. Millets benefit from the C4 photosynthetic trait as well. Carbon dioxide (CO₂) is concentrated in the C4 system around ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO), inhibiting oxygenation and photorespiration of ribulose 1,5-bisphosphate (RuBP) [19]. As a result, the C4 mechanism increases the concentration of CO₂ in the bundle sheath, which reduces photorespiration and increases the catalytic activity of RuBisCO in plants. Millets have increased photosynthetic rates in warm climates, resulting in instant water use efficiency (WUE) and nitrogen use efficiency (NUE) that are 1.5-4 times higher than C3 photosynthesis due to RuBisCO of C4 plants working in elevated CO₂ levels [20]. Millets benefit from C4 photosynthesis in addition to WUE and NUE, including improved ecological enactment in warm climates, more flexible biomass allocation patterns, and lower hydraulic conductivity per unit leaf area [20]. These qualities of millets make them next-generation crops with the potential to be studied in terms of the characteristics of climate-resilient plants and used to enhance major cereals.

3.1.2 Low Carbon Footprint

Wheat emits the most greenhouse gases of any major cereal crop, at approximately 4 tonnes CO₂ eq/ha, followed by rice and maize (at approximately 3.4 tonnes CO₂ eq/ha). Millet has a low carbon footprint per hectare of 3,218 kilograms, which contributes to mitigating climate change. Millets have the lowest carbon equivalent emission (CEE) of any crop (878 kg C ha⁻¹) [21].

3.1.3 Drought adaptability

Certain strategies are devised by minor millets to withstand drought conditions which makes it a perfect crop for dryland agriculture. In case of little millet, shoot length is decreased but root length is increased under moisture stress conditions along with significant accumulation of antioxidants, Reactive Oxygen Species (ROS) scavenging enzymes, superoxide, catalase, glycine betaine (GB) & increased concentration of total free amino acid [22]

3.2 Effect on Soil

Several researchers have studied and demonstrated that millet cultivation influences soil health and fertility positively. A study revealed that after 5 years of monocropping Proso millet, the soil had higher levels of total nitrogen, available nitrogen, phosphorus and potassium than the soil in which common buckwheat and common beans were cultivated [23]. Millet based farming system has proved to be a sustainable approach to improve soil health and soil organic matter content [24]. Such farming practices improve soil physical properties and enhance soil water holding capacity which is beneficial in rainfed areas. Millets with deep rooted system improve physical characteristics of soil such as soil compactness, soil porosity and soil aggregation and also enable them to assess moisture and nutrients from the deeper soil layers, making them drought tolerant crops [25]. Intercropping millets with legumes can improve soil organic carbon, soil aggregate stability and soil microbial activity which are key indicators of soil health. Minor millets with nitrogen fixing ability can contribute to soil fertility by improving nutrient availability and physicochemical properties [26]. Millet cultivation can also check soil erosion as the dense root system holds soil compactly [27]. Overall, the Minor millet inclusive farming system has a significant positive impact on soil health, making it an essential crop for sustainable agriculture practice in different regions.

4.0 CONSTRAINTS

4.1 Constraints in the Pathway of Genetical Improvement

The narrow genetic base is one of the massive barriers in the way of minor millet genetic improvement. If we talk about small millet germplasm conserved in gene banks, Foxtail Millet has the highest germplasm conserved (44761) while Guinea Millet has least (3). Conservation of germplasm is essential before losing them forever [28]. Availability of Genome sequencing is crucial for the possible genetic improvement in any crop. Among small millets, only Foxtail Millet has complete annotated genome sequencing available whereas Proso Millet and Barnyard Millet has draft genome sequencing; but no genome sequencing is there for Little Millet and Kodo Millet [29].

4.2 Constraints in the Pathway of Biotechnological Improvement

Due to regional or economic factors, biotechnology-based minor millets improvement has been limited. Production is increased by the conventional breeding methods for selection and controlled hybrid and there are already cultivars available which are resistant to biotic and abiotic stresses. That is the reason for neglected improvement in the novel traits of minor millet [30]. Productivity of the minor millet is very low than major millets and cereals and that could be related to cultivating environment and practices [31]. Due to challenges with plant regeneration and subpar transformation efficiency, biotechnology of minor millets has trailed behind that of the major cereal crops. The responsiveness of millets to

transformation methods is still quite low. For any of the millet species, there are not any model cultivars that can transform at an efficient rate [30].

4.3 Constraints Related to Insect Pests

Insect pests can cause significant damage to these crops, resulting in reduced yields and financial losses for farmers. There are over 150 species of insects worldwide that feed on millets, with 116 of them being found in India [32, 33]. Insects attack and damage millet crops at different growth stages of the plant, resulting in lower production, declining productivity, and poor-quality grain [34, 35]. In India, insect attacks account for 10–20% losses in yield in millets [36]. Minor millets are more often attacked by shoot flies, stem borer, armyworm, termites, and the grain weevil. One of the main pests that affect millets is the shoot fly. There are many species of shoot flies (Diptera: Muscidae) that attack different types of minor millets in India [37]. It is a very destructive pest that results in considerable losses in yield. During the seedling stage, larvae cut the growing point, which results in a dead heart, and at the reproductive stage, they consume ear heads and destroy panicles [38].

The stem borer is a moth with a filthy, brownish appearance that is mostly active at night. Caterpillars consume leaves, bore through stems to cause dead hearts, and dig into ear heads. It damages the crop starting in the second week after germination up to crop maturity. The early instar larvae eating in the whorl are what generate the irregularly formed holes on the leaves. A central shoot's drying, which results in "dead hearts", is seen and there is also significant stem tunneling. Tunneling of the peduncle causes it to shatter, which causes whole or partial chaffy panicles [38]. The armyworm (*Spodoptera spp.*) is a polyphagous pest that attacks many crops, including millets. The larvae feed on leaves, stems, and ears, causing defoliation and yield losses. Termites are a social insect that lives in underground colonies and attacks both young seedlings and mature plants. Plants that are infected droop and eventually die. The grain weevil is a common pest of stored millet grains. The adults lay eggs on the grains, and the larvae feed inside, causing damage and reducing grain quality.

5. POTENTIAL SOLUTIONS AND FUTURE ROADMAP

5.1 Breeding Objectives

Glutinous (waxy/sticky) varieties are comparatively more preferred by consumers for the Foxtail Millet and Proso Millet. To detect the waxy genotype, specially designed molecular markers are required [39]. The wild species should be studied beyond only the weed science point of view. Cyclopiazonic acid, produced in Kodo seed infested by *Aspergillus flavus* and *A. tamarii* is responsible for the 'Kodua poisoning' in man and animals [40]. The breeding target should be to develop variety resistant to these two fungi. For orchards and agroforestry, shade tolerant varieties should be the target [28]. Grain loss due to lodging is a major problem in Kodo Millet which can be overcome by improving culm strength [41]. Breeding for larger seeds is helpful to improve milling recovery in small millets [42]. Growers don't prefer additional intervention to control diseases for obvious economic reasons; thus, particularly for orphanage crops like minor millets, use of disease resistant varieties is advantageous. A list of already developed resistant varieties against major diseases of different minor millets is provided in Table 1.

Table 1. Resistant Varieties/ Entries in Minor Millets

Crop	Disease	Causal Organism	Resistant Varieties/Entries	Reference
BARNYARD MILLET	Head Smut	<i>Ustilago crus-galli</i>	ABM 4-1, VL 207, VL 208	[43]
			PRJ 1	[45]
	Grain Smut	<i>Ustilago panici-frumentacei</i>	PRB 402, S 841, TNAU 92, VL 216, VL 219	[43]
			PRB 901, PRB 903, TNAU 141 and TNAU 155	[45]
FOXTAIL MILLET	Rust	<i>Uromyces-setaria italicae</i>	SiA 3164, SiA 3205	[46]

			DHFtMV 2-5, SiA 3221, TNSI 266, SiA 3156, SiA 3164	[47]
	Blast	<i>Pyricularia setaria</i>	SiA 3164, SiA 3205	[46]
			DHFtMV 109-3, DHFtMV 2-5, DHFtMV 55-3, SiA 3223	[47]
			SR 118, SR 102, ISc 709, 701, 703, 710, 201, JNSc 33, 56, RS 179	[48]
			Foxtail- 49, 96, 132, 160, 200, 237, 267, 295, 362, 364, 663, 717, 774, 784, 838, 936, 1013, 1037, 1137, 1162, 1665, 1725, Check SIA - 3156.	[49]
	Banded Blight	<i>Rhizoctonia solani</i> kuhn.	SiA 2863, ISC 74	[50]
			VFMC-391	[51]
KODO MILLET	Head Smut	<i>Sorosporium paspali</i>	KMV 8, KMV 20, JK 41, JK 62, JK 65, JK 106, JK 13	[52]
	Udbatta	<i>Ephelis sp.</i>	IPS 45, 196, 342, 365, 368, 381, 387, 140, Niwas 1	[53]
PROSO MILLET	Leaf Spot	<i>Bipolarispanici-miliacei</i>	RAUM-7	[54]
LITTLE MILLET	Grain Smut	<i>Macalpinomyces sharmae</i>	DPI 2386, DPI 2394, PLM 202, OLM 203, CO 2	[55]

5.2 Genetic Intervention

For improvement under adverse condition, priority traits for breeding should be incorporated [39]. Comparative genomic approach can be applied efficiently for gene mining and identifying molecular markers. As several small millets lack complete genome sequencing, the genome of conventional crops can be used for study [29]. 16 genotypes (TNPSc 86, TNPSc 155, TNPSc 217, Podivaragu, Adari, Pacheri etc.) are reported which can be utilized to improve culm strength in Kodo Millet [41]. Interspecific hybridization can be a promising pathway towards genetic enhancement. The F1 as a result of Interspecific hybridization between Japanese and Indian barnyard millet shows improved plant height, high culm branching and increased number of tillers in comparison to its parents [56]. MutMap+ can be used to identify genomic regions associated with various biotic and abiotic resistance [42]. The availability of Online Genomic Database for minor millets is important for further research related to crop improvement. Some of the available genomic databases are - Millet Genome Database, *Setaria italica* Genome Database (SiGDB), Foxtail Millet Marker Database (FmMdb), Foxtail Millet MicroRNA Database (FmMiRNADb), Foxtail Millet Transcription Factor Database (FmTFDb), Foxtail Millet Transposable Elements-Based Marker Database (FmTEMDb) [57].

5.3 Biotechnological Interventions

The goals of improvement of minor millets using biotechnology include improved use of natural resources, biotic and abiotic stress resistance development, and quality enhancement for increased consumer acceptance.

5.3.1 Improving Productivity

One of the key physiological elements contributing to plant productivity is a characteristic relating to photosynthesis [58]. The primary characteristics that affect photosynthetic capability at the level of the entire plant are the canopy architecture, leaf morphology, and vascular architecture. Although little research has been done on this topic, it may be possible to increase agricultural photosynthesis and, eventually, production by understanding the genetic diversity in traits associated to photosynthesis in both crop and wild species [59]. Organelles involved in light perception, gene expression, the manufacture of lipids, pigments, and proteins, and the expression of numerous transporters must work closely together at the cellular level. A full understanding of the physiological underpinnings of the developmental features is necessary to engineer them in order to increase photosynthetic efficiency and, consequently, yield. Understanding the above-mentioned mechanism in model species and important cereal crops has advanced quite a bit. There are, however, few instances of this comprehension in small millet [60].

5.3.2 Transgenic plant improvement

Minor millet has less regeneration and transformation efficiencies [61]. Though transgenic plants were produced by particle bombardment of foxtail millet pollen and inflorescence, but the transformation efficiency was low [62]. For gene transfer use of *Agrobacterium* is more efficient than the biolistic gun. *Agrobacterium* mediated transformation can produce a high number of stable and low copy number transgenic events with fewer DNA rearrangements, transfer larger DNA segments into recipient cells and transgene expression is more stable over generations than with the direct gene transfer method [63]. Using this improved transformation technique, the SBgLR gene, which encodes a lysine-rich protein from potato (*Solanum tuberosum* L. cv. Desiree), was effectively introduced into Foxtail millet cv. Jigu 11.

5.3.3 Future Prospects

Investigation has been going on changing in protein quality and quantity in minor millets. Studies required on the effect of processing methods on the biological functions of millet protein. There is little information about the changes in physiological, biochemical, and structural characteristics associated with photosynthesis in small millets, thus more research is required for increasing the productivity of small millets [60].

5.4 Insect Pest Management

The late-sown crop is more commonly infested. One of the simplest and most cost-effective ways to control it is to seed early when the monsoon begins. It can be controlled by adopting cultural practices such as timely sowing, maintaining proper plant spacing, and intercropping with legumes or non-host crops, which can help reduce the impact of shoot fly in little millet [64]. Chemical control measures such as the use of insecticides can also be effective in managing shoot flies in minor millets. Use of imidacloprid (Confidor 200 SL) at 0.5 mL/L or thiamethoxam (Actara 25 WG) at 0.3 g/L has been found effective against shoot flies in minor millets [65].

Stemborer can be managed by removing and destroying the stubble of the previous crop and by slicing the stems to stop it from further spreading. Biological control measures such as the use of natural enemies can be effective in managing stem borer in millets. Research has shown that the use of egg parasitoids such as *Trichogramma chilonis* and *Bracon brevicornis* can effectively control the stem borer population in millet [66]. Chemical control measures such as the use of insecticides can also be effective in managing the stem borer in millets. Research has shown that the use of insecticides such as carbaryl and quinalphos can effectively control the stem borer population in millets [38].

Cultural practices such as crop rotation, early planting, and timely harvesting can help reduce the armyworm (*Spodoptera spp.*) population. Insecticides such as Chlorpyrifos, Spinosad, and Emamectin Benzoate are also effective in controlling armyworms [67]. Cultural control measures such as deep ploughing and crop rotation can help reduce termite populations. Deep ploughing can expose termites to predators and reduce their numbers. Crop rotation with non-host crops can also help to reduce the incidence of termite infestation. Irrigate the crop when there is low moisture in the field. In areas where termites are prevalent, using chlorpyrifos at 1–1.5 mL per liter of water is recommended for termite control [68]. Proper storage practices such as cleaning, drying, and fumigation can help storage infestation. Grain weevils thrive in warm temperatures, so it's important to keep the stored millets in a cool, dry place. The ideal storage temperature for millets is around 10°C to 15°C [69].

6. GOVERNMENT SUPPORT

The Department of Agriculture and Farmers Welfare (DA&FW) is implementing a Sub-Mission on Nutri-Cereals (Millets) under National Food Security Mission (NFSM) to enhance area report, production & productivity of millets [70]. There has been survey conducted to know the inclusiveness of minor millets and its accessibility to millets-based offerings, various states are conducting campaigns. In Odisha, Odisha Millet Mission (OMM) launched in 2018 aims to revive millets in 15 districts; in Karnataka, they have launched initiatives for Millets as “The Food of the Future” with incentive to farmers Rs. 10000/ha for cultivation of millets. They are also encouraging organic farming & millet promotion through “Savayava Bhagya

Yojana". To endorse and create awareness, they organized National and International trade fairs [71]. In order to promote millets, Chhattisgarh state is the only state where Kodo, Kutki and Ragi are being procured at Minimum Support Price (MSP); Kodo-Kutki is being procured at Rs 3,000 per quintal to help the tribal and other farmers of the state [72].

The Government of India has taken certain initiatives to promote Minor Millets throughout the nation such as Initiative for Nutritional Security through Intensive Millet Promotion (INSIMP) under Rashtriya Krishi Vikas Yojana (RKVY), Rainfed Area Development Program (RADP), National Food Security Mission (NFSM), National Nutrition Mission (NNM) etc. The Integrated Child Development Services (ICDS) and the Mid-Day Meal Scheme (MDMS) have included minor millets in their menu to promote their consumption among children to combat malnutrition and hidden hunger. The government also plans to set up Centers of Excellence for bringing these ancient crops to the centerstage.

7. CONCLUSION

The revitalization of minor millets holds **great** potential for addressing the urgent issues of global food security, environmental sustainability and climate change. In order to unleash its complete potential, Minor Millets should be utilized in a multidimensional way for satisfying the holistic needs. To disseminate its awareness extensively, genuine efforts need to be made by all the actors of the ecosystem. Conferences, symposiums and seminars should be organized frequently to integrate the minds of policy makers, stakeholders & researchers. The government should encourage the creation of entrepreneurial ventures in the field of minor millets. For addressing the neoteric agrarian challenges, minor millets can play a pivotal role by intensifying the cropping system as a cover crop, catch crop or inter-crop. Both conventional breeding methods and biotechnology should be employed **simultaneously** for achieving maximum crop improvement. The emphasis should be on developing male sterility line, mutation breeding, genomic assisted breeding, proteomics, metabolomics, Agrobacterium-assisted transformation etc. The roadmap for the future must be built on the foundation of research, innovation and policy support to ensure a sustainable future for all.

REFERENCES

1. Sathish Kumar M, Lad YA, Mahera AB. Trend Analysis of Area, Production and Productivity of Minor Millets in India. *Biological Forum – An International Journal*. 14(2): 14-18(2022)
2. Rasool A, Shah WH, Tahir I, Alharby HF, Hakeem KR, Rehman R. Exogenous application of selenium (Se) mitigates NaCl stress in proso and foxtail millets by improving their growth, physiology and biochemical parameters. *Acta physiologiae plantarum*. 2020 Jul; 42:1-3.
3. Yang Z, Zhang H, Li X, Shen H, Gao J, Hou S, Zhang B, Mayes S, Bennett M, Ma J, Wu C. A mini foxtail millet with an Arabidopsis-like life cycle as a C4 model system. *Nature plants*. 2020 Sep;6(9):1167-78.
4. Malik S, Krishnaswamy K, Mustapha A. Physical properties of complementary food powder obtained from upcycling of Greek yogurt acid whey with kodo and proso millets. *Journal of Food Process Engineering*. 2021 Nov;44(11): e13878.
5. Tekaligne TM, Woldu AR, Tsigie YA. Bioethanol production from finger millet (*Eleusine coracana*) straw. *Ethiopian Journal of Science and Technology*. 2015 Nov 20;8(1):1-3.
6. Ashwini RN. Bioethanol Production from Husks of Different Small Millets (Doctoral dissertation, University of Agricultural Sciences, GKVK).
7. Rose DJ, Santra DK. Proso millet (*Panicum miliaceum* L.) fermentation for fuel ethanol production. *Industrial Crops and Products*. 2013 May 1;43:602-5.
8. Bhat BV, Arunachalam A, Kumar D, Tonapi VA, Mohapatra T. Millets in the Indian Himalaya. *Indian Council of Agricultural Research, New Delhi*. 84p. 2019.
9. Bhalla TC. Traditional foods and beverages of Himachal Pradesh. *International Journal of Traditional Knowledge*. 2007 January;6(1):17-24
10. Chandrasekara A, Naczka M, Shahidi F. Effect of processing on the antioxidant activity of millet grains. *Food Chemistry*. 2012 Jul 1;133(1):1-9.
11. Bandyopadhyay BB. Evaluation of barnyard millet cultivars for fodder yield under single and double cut treatments at higher elevation of hills. *Agricultural Science Digest*. 2009;29(1):66-8.
12. Tran G. Millet hulls. *Feedipedia*, a programme by INRAE, CIRAD, AFZ and FAO. 2015. <https://www.feedipedia.org/node/15695>
13. Cao TL, Yang SY, Song KB. Characterization of barnyard millet starch films containing borage seed oil. *Coatings*. 2017 Nov 1;7(11):183.
14. Lakshman Kumar D, Siva Sankar S, Venkatesh P, Hepcy Kalarani D. Green synthesis of silver nanoparticles using aerial parts extract of *Echinochloa colona* and their characterization. *European Journal of Pharmaceutical Medical Research*. 2016;3(4):325-8.

15. Nolde SB, Vassilevski AA, Rogozhin EA, Barinov NA, Balashova TA, Samsonova OV, Baranov YV, Feofanov AV, Egorov TA, Arseniev AS, Grishin EV. Disulfide-stabilized helical hairpin structure and activity of a novel antifungal peptide EcAMP1 from seeds of barnyard grass (*Echinochloa crus-galli*). *Journal of Biological Chemistry*. 2011 Jul 15;286(28):25145-53.
16. Singh RK, Muthamilarasan M, Prasad M. Biotechnological approaches to dissect climate-resilient traits in millets and their application in crop improvement. *Journal of Biotechnology*. 2021 Feb 10; 327:64-73.
17. Kole C, Muthamilarasan M, Henry R, Edwards D, Sharma R, Abberton M, Batley J, Bentley A, Blakeney M, Bryant J, Cai H. Application of genomics-assisted breeding for generation of climate resilient crops: progress and prospects. *Frontiers in plant science*. 2015 Aug 11;6:563.
18. Li P, Brutnell TP. *Setaria viridis* and *Setaria italica*, model genetic systems for the Panicoid grasses. *Journal of experimental botany*. 2011 May 1;62(9):3031-7.
19. Aubry S, Brown NJ, Hibberd JM. The role of proteins in C3 plants prior to their recruitment into the C4 pathway. *Journal of experimental botany*. 2011 May 1;62(9):3049-59.
20. Sage RF, Zhu XG. Exploiting the engine of C4 photosynthesis. *Journal of experimental botany*. 2011 May 1;62(9):2989-3000.
21. Sadhukhan A, Debangshi U. Millets in Meeting the Nutrition Security: A Review. *Just Agriculture e-magazine*. 3(5), 35-40.
22. Ajithkumar IP, Panneerselvam R. ROS scavenging system, osmotic maintenance, pigment and growth status of *Panicum sumatrense* roth. under drought stress. *Cell biochemistry and biophysics*. 2014 Apr;68:587-95.
23. Yang P, Luo Y, Gao Y, Gao X, Gao J, Wang P, Feng B. Soil properties, bacterial and fungal community compositions and the key factors after 5-year continuous monocropping of three minor crops. *PLoS One*. 2020 Aug 24;15(8):e0237164. doi: 10.1371/journal.pone.0237164.
24. Babu C, Sheelu G, Reddy B.V.S, Rao V.M and Sudhakar C. Millet-Based Farming System: An Approach for Improving Soil Organic Matter Content and Soil Health. *Communications in Soil Science and Plant Analysis*. 2015;49(13), 1655-1663.
25. Muthukumarasamy R, Prasad R, Hegde DM, Bindraban PS, & Sreenivas G. Millets for soil and nutritional security. *International Journal of Agricultural Sustainability* 2010; 8(3), 170-184.
26. Chakraborty D, Bhattacharyya R, Pramanick S. The role of minor millets in sustainable nutrient recycling and improving soil health in tropical agroecosystems. *Archives of Agronomy and Soil Science*. 2006;62(1), 80-94.
27. Behera UK, Singh R. Assessment of soil erosion in relation to land use/land cover changes in the northeastern hilly region of India using remote sensing and GIS. *Environmental Earth Sciences*. 2017;76(5), 214.
28. Vetriventhan, M., Azevedo, V.C., Upadhyaya, H.D., Nirmalakumari, A., Kane-Potaka, J., Anitha, S., Ceasar, S.A., Muthamilarasan, M., Bhat, B.V., Hariprasanna, K. and Bellundagi, A., 2020. Genetic and genomic resources, and breeding for accelerating improvement of small millets: current status and future interventions. *The Nucleus*, 63, pp.217-239.
29. Maharajan, T., Ceasar, S.A., Krishna, T.P.A. and Ignacimuthu, S., 2022. Mining genes and markers across minor millets using comparative genomics approaches. In *Omics of Climate Resilient Small Millets* (pp. 185-203). Singapore: Springer Nature Singapore.
30. SL K, Kumar S, Kothari A, Watanabe KN. Applications of biotechnology for improvement of millet crops: review of progress and future prospects. *Plant Biotechnology*. 2005;22(2):81-8.
31. Padulosi S, Mal B, King OI, Gotor E. Minor millets as a central element for sustainably enhanced incomes, empowerment, and nutrition in rural India. *Sustainability*. 2015 Jul 8;7(7):8904-33.
32. Nwanze KF, Harris KM. Insect pests of pearl millet in West Africa. *Review of Agriculture Entomology*. 1992;80(12):1132-55.
33. Kishore P. Evolving management strategies for pests of millets in India. *Journal of Entomological Research*. 1996;20(4):287-97.
34. Kumar DA, Channaveerswami AS. Pre and post emergence control measures for shootfly incidence and its influence on seed yield of little millet (*Panicum sumatrense*). *Journal of Experimental Zoology, India*. 2015;18(2):811-4.
35. Bekoye, B. M., and A. Dadie. 2015. Evaluation des pertes en grains de mil dues aux insectes. *Eur. Sci. J.* 11: 266–27.
36. Gahukar RT, Jotwani MG. Present status of field pests of sorghum and millets in India. *International Journal of Pest Management*. 1980 Jun 1;26(2):138-51.
37. Kalaisekar A, Padmaja PG, Bhagwat JV, Patil JV. Insect pests of millets: systematics, bionomics, and management. Academic Press; 2016 Dec 23.
38. Gahukar RT, Reddy GV. Management of economically important insect pests of millet. *Journal of Integrated Pest Management*. 2019;10(1):28.
39. Goron TL, Raizada MN. Genetic diversity and genomic resources available for the small millet crops to accelerate a New Green Revolution. *Frontiers in plant science*. 2015 Mar 24;6:157.

40. Rao BL, Husain A. Presence of cyclopiazonic acid in kodo millet (*Paspalum scrobiculatum*) causing 'koda poisoning' in man and its production by associated fungi. *Mycopathologia*. 1985 Mar;89(3):177-80.
41. Ragini S, Subramanian A, Nirmalakumari A, Babu JK. Genetic diversity in kodo millet (*Paspalum scrobiculatum*) relative to culm strength and yield. *Journal of Crop Improvement*. 2015 Jul 4;29(4):420-31.
42. Pramitha JL, Ganesan J, Francis N, Rajasekharan R, Thinakaran J. Revitalization of small millets for nutritional and food security by advanced genetics and genomics approaches.
43. Kumar B, Kumar J. Evaluation of small millet genotypes against endemic diseases in mid-western Himalayas. *Indian Phytopathology*. 2009;62(4):518-21.
44. Yadav VK, Yadav R, Kumar B, Malik N. Success story: PRJ 1: Disease resistant, high yielding variety of barnyard millet. College of Forestry and Hill Agriculture, GBPUAT, Hill Campus, Ranichauri, Tehri Garhwal, Uttarakhand. 24p. 2010.
45. Kumar BI. Management of grain smut disease of barnyard millet (*Echinochloa frumentacea*). *Indian Phytopathology*. 2013;66(4):403-5.
46. Rajesh M, Sudha A, Nirmalakumari A, Parasuraman P. Identification of resistant sources for blast and rust in foxtail millet incited by *Pyricularia setariae* and *Uromyces setariae-Italica*. *Int J Curr Microbiol App Sci*. 2019;8:1796-800.
47. Munirathnam P, Venkataramanamma K, Anusha A. Evaluation of foxtail millet genotypes for blast and rust diseases under field conditions. *Curr Biot*. 2015;9:263-8.
48. Singh BP, Pad BS, Nema AG, Deshkar MV. Varietal reaction to blast of Kangni. *Curr Res*. 1976;1:13.
49. Makwana K, Tiwari S, Tripathi MK, Patel V. Selection of Blast Resistant Lines from Diverse Germplasm Set of Foxtail Millet. *Biological Forum - An International Journal*. 2023 January 06;15(1).
50. Patro TS, Meena A, Divya M, Anuradha N. Diseases reaction of donor screening nursery (DSN) of Proso millet against *Rhizoctonia solani*, the cause of Sheath blight. *Journal of Pharmacognosy and Phytochemistry*. 2018;7(3):2684-5.
51. Patro TS, Anuradha N, Madhuri J, Suma Y, Soujanya A. Identification of resistant sources for blast disease in finger millet (*Eleusine coracana* Gaertn.). In *Varietal Improvement of Small Millets*. National seminar on "Recent Advances of Varietal Improvement in Small Millets 2013 (pp. 5-6).
52. Kumar B, Srivastava JN. Kodo Millet or Kodo (*Paspalum scrobiculatum* L.) Diseases and Their Management Strategies. In *Diseases of Field Crops: Diagnosis and Management 2020* May 16 (pp. 273-290). Apple Academic Press.
53. Pall BS, Nema AG. Screening of kodo (*Paspalum scrobiculatum* L.) varieties against udbatta disease. *JNKVV Res. J*. 1976;10(2):194.
54. Kumar B, Srivastava JN. Common Millet or Proso Millet or Cheena Millet or French Millet (*Panicum miliaceum* L.) Diseases and Their Management Strategies. In *Diseases of Field Crops: Diagnosis and Management 2020* May 16 (pp. 205-217). Apple Academic Press.
55. Kumar B, Srivastava JN. Little Millet or Gundali or Goudli or Gondola (*Panicum miliare* L.) Diseases and Their Management Strategies. In *Diseases of Field Crops: Diagnosis and Management 2020* May 16 (pp. 291-297). Apple Academic Press.
56. Sood S, Khulbe RK, Saini N, Gupta A, Agrawal PK. Interspecific hybrid between *Echinochloa esculenta* (Japanese barnyard millet) and *E. frumentacea* (Indian barnyard millet)—a new avenue for genetic enhancement of barnyard millet. *Electronic Journal of Plant Breeding*. 2014 Jun 30;5(2):248-53.
57. Thulasinathan T, Jain P, Yadav AK, Kumar V, Sevanthi AM, Solanke AU. Current Status of Bioinformatics Resources of Small Millets. In *Omics of Climate Resilient Small Millets 2022* Jul 16 (pp. 221-234). Singapore: Springer Nature Singapore.
58. Zhu XG, Long SP, Ort DR. Improving photosynthetic efficiency for greater yield. *Annual review of plant biology*. 2010 Jun 2;61:235-61.
59. Flood PJ, Harbinson J, Aarts MG. Natural genetic variation in plant photosynthesis. *Trends in plant science*. 2011 Jun 1;16(6):327-35.
60. Pavithra KS, Senthil A, Babu Rajendra Prasad V, Ravikesavan R, Djanaguiraman M. Variations in photosynthesis associated traits and grain yield of minor millets. *Plant Physiology Reports*. 2020 Sep;25:418-25.
61. SL K, Kumar S, Kothari A, Watanabe KN. Applications of biotechnology for improvement of millet crops: review of progress and future prospects. *Plant Biotechnology*. 2005;22(2):81-8.
62. Dong Y, Duan S. Establishment of embryogenic cell suspension culture and plant regeneration of millet and gene transfer. *J Basic Sci Eng*. 1999;7(1):34-40.
63. Wang MZ, Pan YL, Li C, Liu C, Zhao Q, Ao GM, Yu JJ. Culturing of immature inflorescences and *Agrobacterium*-mediated transformation of foxtail millet (*Setaria italica*). *African Journal of Biotechnology*. 2011;10(73):16466-79.
64. Sathish R, Manjunatha M, Rajashekarappa K. Effect of intercropping on incidence of shoot fly, *Atherigona pulla* (Wiedemann) in little millet. *International Journal of Pure and Applied Bioscience*. 2017;5(4):1845-9.
65. Sridhar K, Sridharan S, Muthukumar M. Management of shoot fly *Atherigona soccata* (Rondani) with different seed dressing chemicals. *International Journal of Plant Protection*. 2016 Apr;9(1):193-8.

66. Gahukar RT. Conservation of parasitoids as management strategy for lepidopterous stem borers in tropical cereals. *Journal of Entomological Research*. 2005;29(4):293-301.
67. Deshmukh SS, Prasanna BM, Kalleshwaraswamy CM, Jaba J, Choudhary B. Fall armyworm (*Spodoptera frugiperda*). *Polyphagous pests of crops*. 2021:349-72.
68. Rana A, Chandel RS, Verma KS, Joshi MJ. Termites in important crops and their management. *Indian Journal of Entomology*. 2021;83.
69. Befikadu D. Factors affecting quality of grain stored in Ethiopian traditional storage structures and opportunities for improvement. *International Journal of Sciences: Basic and Applied Research*. 2014;18(1):235-57.
70. Anonymous. Millet Production, Press Information Bureau, Ministry of Agriculture & Farmers Welfare, (Release ID: 1796559) Accessed 08 FEB 2022. Available: <https://pib.gov.in/PressReleaselframePage.aspx?PRID=1796559>
71. International Year of Millets (IYoM)-2023, National Conference on Kharif Campaign, 2022 19th April 2022, Ministry of Agriculture & Farmers Welfare.
72. Verma HP, Verma A. Chhattisgarh Millet Mission. *Just Agriculture e-magazine*. January 2023;67-68.

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