

Immunological Features of COVID-19 in Hodeidah, Yemen

ABSTRACT

Background: Monitoring of the immunological status of patients with coronavirus disease 2019 (COVID-19) in Yemen is practically absent. Several studies vary in study design, populations under study, serologic tests used, timing of sample collection, and quality.

Objective : Therefore , our study aimed to present the validation of immunological method namely rapid test for detection of immunoglobulin G (IgG) of COVID-19 infection immune response development in the blood of healthy participants living in the context of the COVID-19 pandemic area and of the patients who have undergone COVID-19 .

Methodology : Rapid test was validated that included the sensitivity, specificity, and accuracy parameters and used for sampling in research analysis. The research was designed in pilot study and one time cross sectional COVID-19 antibodies survey after three months of COVID-19 pandemic and implemented in four groups (N:64): the first group was severe recovered patients (n:16) admitted in COVID-19 isolation department , Center of Tropical Medicine and Infectious Diseases (CTMID), Al Thawara Public Hospital Authority, Hodeidah, Yemen, contacts of infected patients (n:16), mild and moderate cases (n:16) at home ,and asymptomatic cases (n:16). Data obtained were analyzed based on appropriate statistical tools.

Results : The results of rapid test validation showed that is sensitive (79.17 %; CI : 65.01 – 89.53 %) , specific (87.50%;CI : 61.65 – 98.45 %) , precise (95.00 %; CI : 83.75 – 98.59%) and accurate (81.25 %; CI : 69.54 – 89.92%) for detection of IgG of COVID-19 in Hodeidah, Yemen. In total, 40 of 64 participants were positive, giving a prevalence of IgG – based COVID-19 infection of 62.5 %. The IgG were detected in 16/16 cases (100 %) of recovered severe patients; 13/16 cases (81.25 %) of contacts cases. In addition, IgG were detected in 9/16 cases (56.25 %) of mild and moderate cases and 2/16 cases (12.5 %) of asymptomatic cases (healthy people). On the other hand , the higher frequency of IgG was in elderly between 50 and 59 year (n:17 ; 25.56 %) , followed by adults between 40 and 49 year (n: 15 ; 23.43 %) , and the lower frequency was in adult between 30 and 39 year (n:8 ; 12.5%).

Conclusion: The study concluded that antibodies become detectable after symptom onset of severe cases and their contacts completely and strongly based on validated immunological method . On the other hand, the antibodies were developed in mild and moderate patients partially and weakly. A little developed in asymptomatic patients. However, additional data are needed before modifying public health recommendations based on serologic test results.

Keyword: COVID -19, Immunological, IgG , Antibodies , Hodeidah , Yemen

1. INTRODUCTION

Millions in the world were infected with the coronavirus disease 2019 (COVID-19) , the virus that causes COVID-19, they develop antibodies a few weeks after infection. The infection can be evened in the

peoples who have even severe disease, mild disease, and even asymptomatic infection, do develop these antibodies [1]. Center for Diseases Prevention and Control (CDC) reported that antibodies most commonly become detectable 1–3 weeks after symptom onset, at which time evidence suggests that infectiousness likely is greatly decreased and that some degree of immunity from future infection has developed. However, additional data are needed before modifying public health recommendations based on serologic test results, including decisions on discontinuing physical distancing and using personal protective equipment [2,3].

World health organization (WHO) reported “ There are now more than 200 peer-reviewed publications, pre-prints, manuscripts and government reports of Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) seroprevalence studies. These studies vary in study design, populations under study, serologic tests used, timing of sample collection, and quality. Overall, the population-based seroprevalence reported across available studies remains low, at below 10%. Some studies conducted in areas of known high virus transmission and studies of health care workers in areas of known high transmission have reported seroprevalence estimates over 20% [4].

Despite the great interest of the scientific community in the behavior of the human body after contact with COVID-19, monitoring of the immunological status of patients with COVID-19 having varying severity degrees and of the people with a low COVID-19 viral load is practically absent [5].

The aim of this study was a detecting of COVID-19 infection immune response development using qualitative assessment of IgG in the blood of healthy donors living in the context of the COVID-19 pandemic and of the patients who have undergone COVID-19 infection (post-recovery of patients and post exposure of community).

2. METHODOLOGY

2.1. Study area

The study was carried out in COVID-19 isolation department, Molecular Biological Unit , Center of Tropical Medicine and Infectious Diseases (CTMID) , AL Thawara Public Hospital Authority , Hodeidah Yemen . In this study area 2020 , AL Kamarany et al reported that 386/505 (76.43%) of suspected cases, and 70/505 (13.86%) of probable cases that were isolated and treated at home. A total of 49/505 cases (9.70 %) were confirmed and admitted in isolation department. On the other mean, 386 patients (76.43%) of mild and moderate cases, 70 patients (13.86%) of severe illness were treated at home. 21 patients of severe illness (4.16%) and 28 patients (5.54%) of critical illness were treated at isolation department. 49 patients (9.7 %; severe and critical) needed admission in an intensive care unit (ICU) that were confirmed based on RT –PCR. Other cases (mild and moderate) were confirmed epidemiologically (epidemiologically linked case: a case in which the patient has/ had contact with one or

more persons who have/had the disease, and transmission of the agent by the usual modes of transmission is plausible. A case may be considered epidemiologically linked to a laboratory-confirmed case if at least one case in the chain of transmission is laboratory confirmed [6].

2.2. Study design

The research was designed in pilot study and one time cross sectional COVID-19 antibodies survey that included 64 participants , divided into four groups : the first group included 16 of recovered patients from COVID-19 who admitted and treated in isolation department ,CTMID, AL Thawara Public Hospital Authority , Hodeidah Yemen. The second group was contacts of infected severe patients (n:16), the third group was mild mild and moderate cases (n:16) and treated at home, and the fourth group was asymptomatic cases “healthy peoples “ (n:16) (Figure 1).

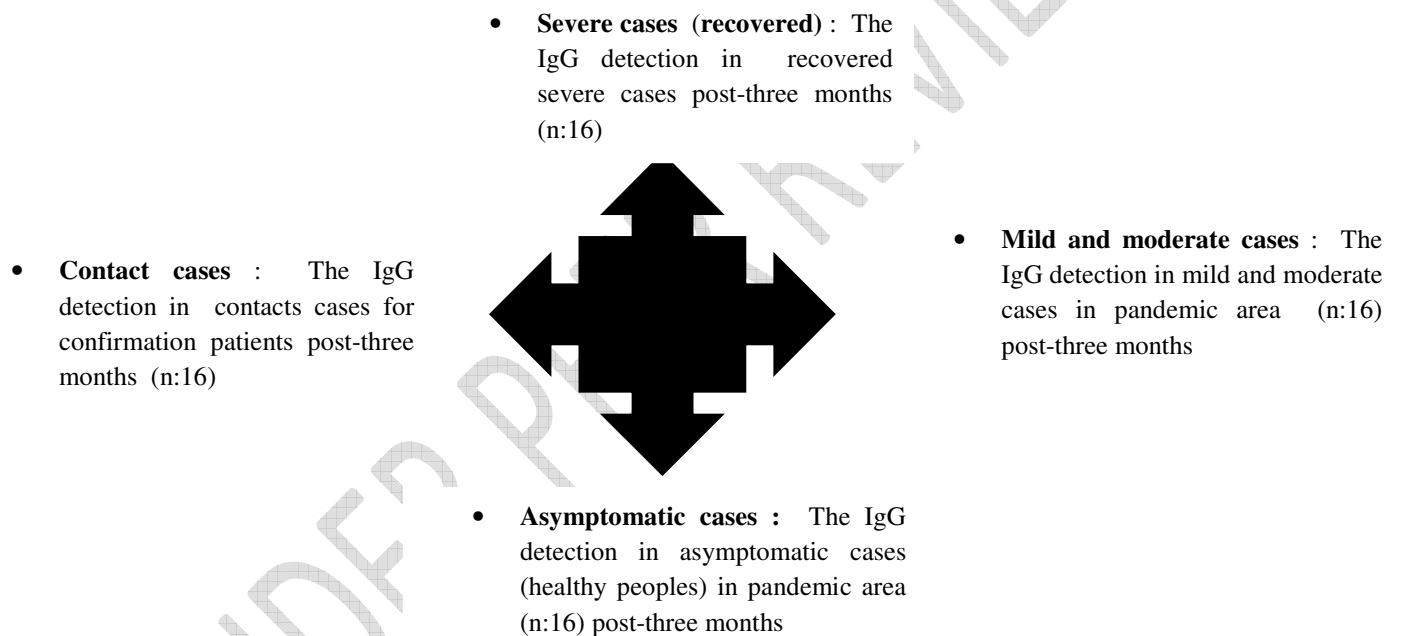


Figure 1. Study design for diagnostic test evaluation of rapid test method for IgG detection in Hodeidah peoples, Yemen : Note : Mild Cases : Symptoms of respiratory infection (fever, cough , pharyngitis, headache, ... etc) Symptomatic, meeting the case definition for COVID-19, without evidence of viral pneumonia or hypoxia . Moderate Cases: Clinical signs of non-severe pneumonia (cough or difficulty breathing and fast breathing and/or chest indrawing) and no signs of severe pneumonia. Severe cases : Clinical signs of pneumonia (fever, cough, dyspnea, fast breathing) plus one of the following: respiratory rate > 30 breaths/min; severe respiratory distress; or SpO₂ < 90% on room air. Contacts cases: The persons contacted with confirmed cases [7 - 9]. Note: Major symptom in mild and moderate cases were acute smell and/or taste loss

2.3. Real Time – Polymerase Chain Reaction (RT-PCR) for Detection the COVID-19 infection

The RT-PCR of COVID-19 detection was re-validated partially in Molecular Biological Unit of CTMID, AL Thawara Public Hospital Authority of Hodeidah, Yemen . The assay for molecular detection of COVID-19 on nasopharyngeal swabs was performed using the RT-PCR Bio-System. The Norgen's COVID-19 TaqMan RT-PCR kit that was designed for the detection of COVID-19 specific RNA [10].

2.4. Rapid Test for Detection of Antibodies of COVID-19 infection

6 samples have been tested positive using RT-PCR, and all of them were also positive based on rapid test IgG .The rapid test (indirect infection detection) was re-validated based gold bio-analytical method namely RT-PCR that detects the infection directly by detecting the viral RNA. The IgG were detected in 6 recovered severe patients that were confirmed with COVID-19 RT-PCR. The analytical efficiency including sensitivity, specificity, precision, accuracy, positive predictive value , negative predictive value and confidence interval (CI) . Finally, the the study applied the rapid test for immunological response (detection of IgG only) post - recovery of patient from COVID-19 infection and post exposure of community for COVID-19 pandemic (antibodies detection in patients and community). On the other mean, IgG was detected after 3 months from infection (Figure 1) [11-13].

2.5. Data Management

The simple statistical process was used to partial validation of RT-PCR for detection of COVID-19 and rapid test assay for IgG detection. Firstly , MedCalc Software Ltd was used for validation and evaluation of diagnostic test [13]. Secondly, data were collected ,checked and entered in an Excel Software 2013 and then the data was analyzed using tables , graphs, percentages , median , range , and average were the main descriptive tools.

3. RESULTS

3.1. Partial validation of RT-PCR

The RT-PCR (Bio-system 7500) was validated partially for assessing the accuracy, precision and quantification with limit of different nasopharyngeal samples. Participated volunteers of this study were provided written consent and the results of re-validated method were precise to each analyze with percent relative standard deviations (RSD %) that was 3.36 % (< 5.0%) . Furthermore, the accuracy of validated method exhibit well recovery values of 98 % - 102 % ($\pm 5\%$) (Table 1).

Table 1. Partial validation of RT-PCR method (N:6)

Parameters	Limit of Quantification	Limit of Detection	Precision (RSD %)	Accuracy (Recovery %)
Cycle Threshold (Ct)	10 – 39	7	3.36 %	98 - 102 %.

3.2. Diagnostic test evaluation of rapid test method for IgG detection

The results of rapid test validation showed that was sensitive (79.17 %; CI : 65.01 – 89.53 %) , specific (87.50%;CI : 61.65 – 98.45 %) , precise (95.00 %; CI : 83.75 – 98.59%) and accurate (81.25 %; CI : 69.54 – 89.92%) for detection of IgG of COVID-19 in Hodeidah, Yemen .(Table 2 and 3)

Table 2. Re-validation of rapid test method based on RT-PCR (N:6)

	RT-PCR	Rapid Test
* Recovered patients	Confirmed	IgG detected strong (100%)
* Recovered patients : The IgG was detected after three months from discharge of patients		

Table 3. Diagnostic test evaluation of rapid test method for IgG detection

Parameters	Value (%)	95 % CI (%)
Sensitivity	79.17	65.01 – 89.53
Specificity	87.50	61.65 – 98.45
Precision	95.00	83.75 – 98.59
Positive likelihood ratio	5.11	1.72 – 23.34
Negative likelihood ratio	0.24	0.13 – 0.43
Positive predictive value	95.00	83.75 – 98.59
Negative predictive value	58.33	43.90 – 71.47
Accuracy	81.25	69.54 – 89.92

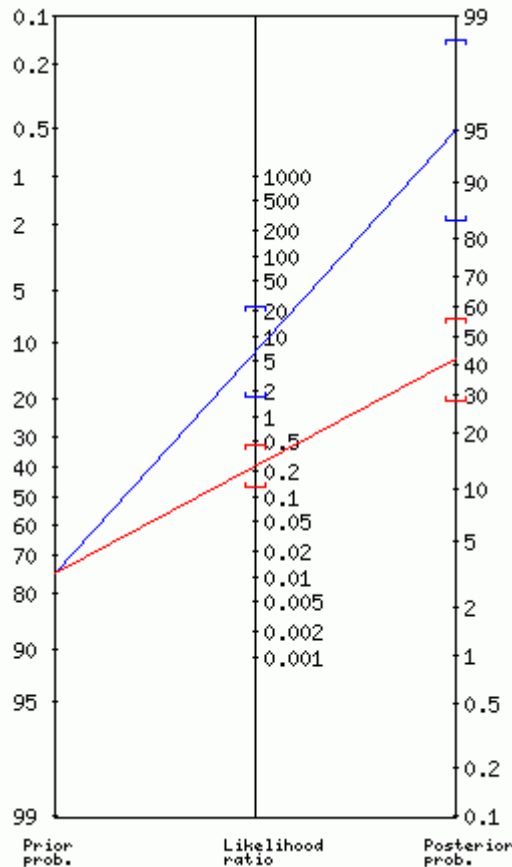


Figure 2. Prior probability (odd) : 75 % (3.0) ; POSITIVE TEST : Positive Likelihood ratio: 6.33 with 95% confidence interval: [1.72,23] . Posterior probability (odds): 95% (19.0) with 95% confidence interval: [84%,98%] (~ 1 in 1.1 with positive test are sick) . NEGATIVE TEST: Negative Likelihood ratio: 0.24 with 95% confidence interval: [0.13,0.43]. Posterior probability (odds): 42% (0.7) with 95% confidence interval: [28%,56%] (~ 1 in 1.7 with negative test are well)

3.3. Detection of IgG in the patients and community

3.3.1. Age and sex

Serum samples were collected from 64 participants during the period from September 2020 to November 2020. The age range of patients was from 35 year to 59 year and the median age of subjects was 41 year, where 45% of participants occurred between 40 and 49 year. The study was represented in the males as 92.18 % while in the females it was represented as 7.81 %.

3.3.2. Detection of IgG in different groups

In total, 40 of 64 participants were positive, giving a prevalence of COVID-19 infection – based IgG of 62.5%. The IgG were detected in different groups, 16/16 cases (100 %) of recovered patients (severe ills that admitted in isolation department) had strong IgG; 13/16 cases (81.25 %) of contact for severe patients had strong IgG. In addition, IgG were detected in 9/16 cases (56.25 %) of mild and moderate cases and treated at home that had weak IgG. In addition , IgG were detected in 2/16 cases (12.5 %) of healthy people (asymptomatic cases) that had weak IgG. On the other hand, the higher frequency of IgG was in elderly

between 50 and 59 year (n:17 ; 25.56 %), followed by adults between 40 and 49 year (n: 15 ; 23.43 %), and the lower frequency was in adult between 30 and 39 year (n:8 ; 12.5%) (Figure 3 and 4).

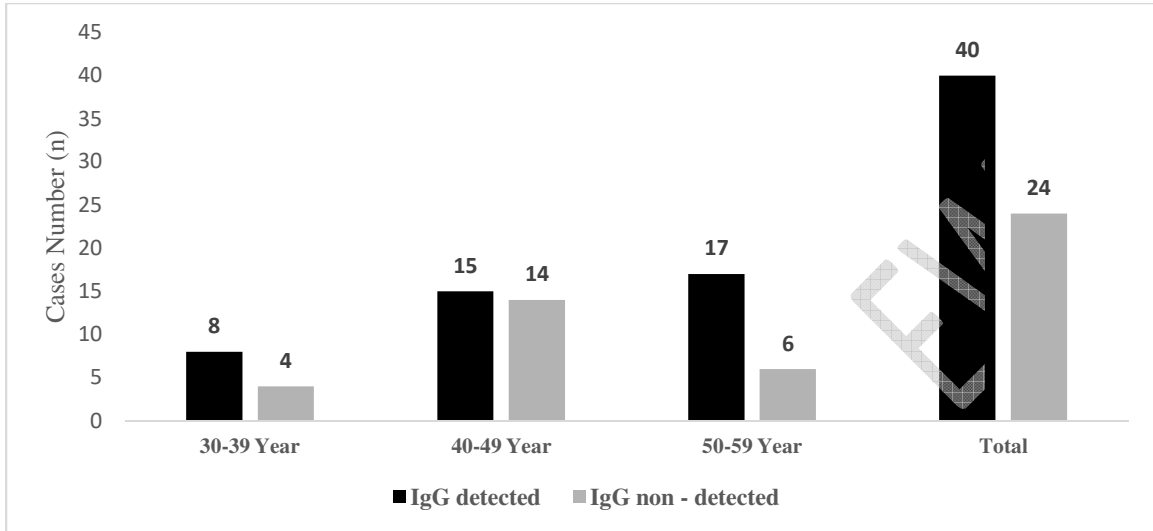


Figure 3: Detection of IgG against COVID-19 in Hodeiadh, Yemen according to age

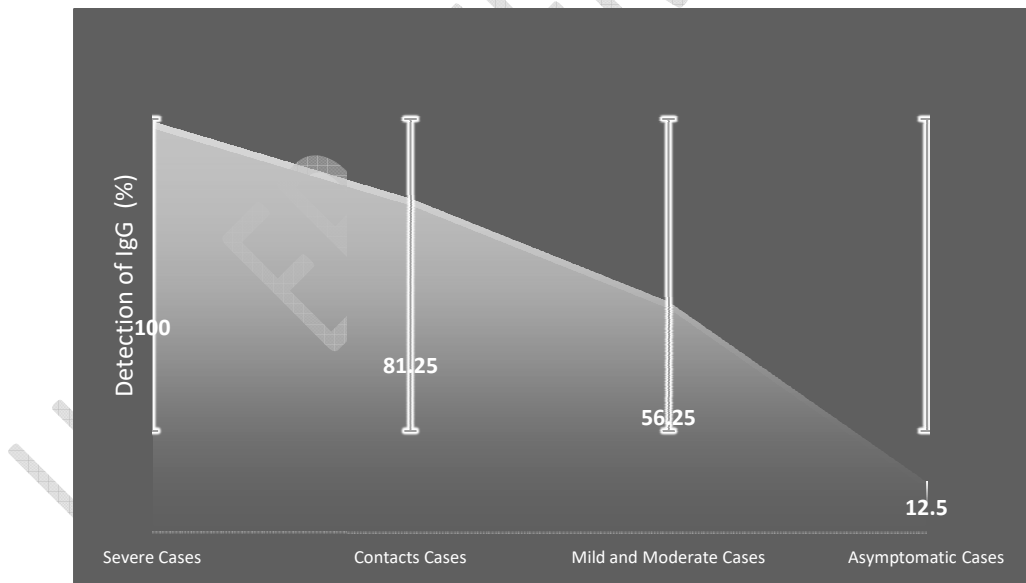


Figure 4: Detection of IgG against COVID-19 in symptomatic and asymptomatic of Hodeidah peoples, Yemen

4. DISCUSSION

Firstly, due to lack of formal guidance or regulatory requirements, several approaches are possible to select the experimental design, for choosing the statistical data treatment and hence for the decision process namely the rapid test of COVID-19 to detect of antibodies. The success of an analytical method validation of rapid test for detection of antibodies of COVID-19 is tested by comparing results of RT-PCR of COVID-19 as standard confirmation method. On the other mean, the objective of this work is to demonstrate the applicability of the simple approaches with certain statistical models to a more variability domain bio-analytical methods namely rapid test (immune -chromatographic technique) and in the interpretation of acceptance criteria of validation of rapid test present the immunological response (detection of IgG) in Hodeidah peoples based on validated bio-analytical method.

Secondly, it was used to identify past COVID -19 infection in Yemeni people who were infected at 3 months previously. The present study showed that antibodies namely IgG were detected in Hodeidah, Yemen (62.5 %), the IgG was detected in recovered severe patents (100 %), contacts (81.25 %), mild and moderate (56.25 %) "acute loss in their sense of smell and/or taste in a community and asymptomatic peoples (12.5 %). In comparing with other study carried out in Aden, Yemen, the prevalence of IgG was 25% and the prevalence of asymptomatic COVID-19 in the entire study group was 7.9%, the prevalence of COVID-19 was significantly higher among females, housewives and subjects with a history of contact with a COVID-19 patient: 32%, 31% and 39%, respectively [14].

On the other hand, in comparing with other studies carried out in different countries of the world, the present study results agreed with a study done by Makaronidis et al. in London, UK (77.6%) with acute smell and/or taste loss had SARS-CoV-2 antibodies [15]. IgG seroprevalence was recorded randomly by Stringhini et al. in Geneva, Switzerland " in the first week was 4.8%, the estimate increased to 8.5% in the second week, to 10.9% in the third week, 6.6% in the fourth week, and 10.8% in the fifth week [16]. Antibody prevalence in England fell from 6.0% to 4.4% over three months [17]. In Wuhan, IgG prevalence was 89.8% in COVID-19 patients, 4.0% in healthcare providers, 4.6% in general workers, and 1.0% in other patients [18]. 8.3% tested positive for IgG in an asymptomatic population in Sergipe, Brazil [19]. In Italy, a prevalence in symptomatic individuals and their family contacts was 23.1% and the highest prevalence was found in the age class 40 - 49 years. Overall, 34.4% of the participants reported at least one symptom and among the symptoms, anosmia and ageusia were strongly associated with seropositivity [20]. In previous study included prospective longitudinal cohort study entitled "dynamics of IgG-avidity and antibody levels after Covid-19" where Löfström E et al found a significant ongoing increase in avidity maturation after Covid-19 whilst the levels of antibodies were declining, suggesting a possible aspect of long-term immunity [21].

Finally, the question, what is the degree of susceptibility of previously infected individuals to reinfection by SARS-CoV-2 ? Alzaabi et al. indicated a sustained and prolonged positive immune response in COVID-19 recovered patients. The consistent rise in antibody and positive levels of IgG titers within the first 5 months suggest that immunization is possible, and the chances of reinfection minimal [22]. In Brazil “dynamics of anti-SARS-CoV-2 IgG antibodies post-COVID-19 was studied “ and the authors showed a high frequency of loss of anti-SARS-CoV-2 IgG antibodies within 3 months after COVID-19 [23]. Previous study entitled “ SARS-CoV-2 reinfection in patients negative for immunoglobulin G following recovery from COVID-19 “ concluded patients who recover from COVID-19 with no detectable anti-nucleocapsid IgG concentration appear to remain more susceptible to reinfection by SARS-CoV-2, with no apparent immunity. Also, the authors suggested the chance is lower, the possibility for recovered patients with positive anti-nucleocapsid IgG findings to be reinfected similarly exists [24].

CDC reported “ COVID-19 vaccination causes a more predictable immune response than infection with the virus that causes COVID-19. Getting a COVID-19 vaccine gives most people a high level of protection against COVID-19 and can provide added protection for people who already had COVID-19. One study showed that, for people who already had COVID-19, those who do not get vaccinated after their recovery are more than 2 times as likely to get COVID-19 again than those who get fully vaccinated after their recovery [25].

Limitations of the study: There are some limitations in this study that need to be considered. The small samples size in this study and other immunological features are not included.

5. CONCLUSION

The study concluded that antibodies become detectable after symptom onset of of severe cases and their contacts completely and strongly based on validated immunological method. On the other hand, the antibodies were developed in mild and moderate patients partially and weakly. A little developed in asymptomatic people. However, additional data are needed before modifying public health recommendations based on serologic test results.

CONSENT

As per international standard or university standard, Participants’ written consent has been collected and preserved by the authors. The raw data are secured in the Center of Tropical Medicine and Infectious Diseases (CTMID), Al-Thawara Public Hospital Authority, Hodeidah, Yemen.

ETHICAL APPROVAL

The studies involving human participants were reviewed and approved by Ethics Committee of CTMID, Al-Thawara Public Hospital Authority, Hodeidah, Yemen.

COMPETING INTERESTS DISCLAIMER:

Authors have declared that no competing interests exist. The products used for this research are commonly and predominantly use products in our area of research and country. There is absolutely no conflict of interest between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

REFERENCES

1. Maria Van Kerkhove. Episode #18 - COVID-19 - Immunity after recovery from COVID-19 , World Health Organization (WHO). . <https://www.who.int/emergencies/diseases/novel-coronavirus-2019/media-resources/science-in-5/episode-18---covid-19---immunity-after-recovery-from-covid-19> [Accessed December 23, 2020]
2. Center for Diseases Prevention and Control (CDC) – US. Interim Guidelines for COVID-19 Antibody Testing, <https://www.cdc.gov/coronavirus/2019-ncov/lab/resources/antibody-tests-guidelines.html> [Accessed March 17, 2021]
3. Klingler J, Weiss S, Itri V, Liu X, Oguntuyo K Y., Ikegame S, Hung C-T, Enyindah-Asonye G, Amanat F, Baine I, Arinsburg S, Bandres J C, Milunka Kojic E, Stoever J, Jurczynszak D, Bermudez-Gonzalez M, Nádas A, Liu S, Lee B, Zolla-Pazner S, and Hioe C E. Role of IgM and IgA Antibodies in the Neutralization of SARS-CoV-2 . *medRxiv*. 2020; 08.18.20177303. doi:10.1101/2020.08.18.20177303.
4. World Health Organization (WHO), Coronavirus disease (COVID-19): Serology, antibodies and immunity. <https://www.who.int/news-room/questions-and-answers/item/coronavirus-disease-covid-19-serology> [Accessed December 31, 2020]
5. Andrei Ivanov , Elena Semenova. Long-term monitoring of the development and extinction of IgA and IgG responses to SARS-CoV-2 infection. *J Med Virol* . 2021;93(10):5953-5960. doi:10.1002/jmv.27166.
6. AL-Kamarany, M. A., Suhail, K. A., Majam, A. S., Abdulabari Alabsi, E., Hamoud Dowbalah, M., and Mohammed Zohairy, A. Epidemiological and Clinical Features of COVID-19 in Hodeidah, Yemen. *International Journal of TROPICAL DISEASE & Health*, 2021; 42:21. DOI <https://doi.org/10.9734/ijtdh/2021/v42i2130550>
7. World Health Organization (WHO), WHO COVID-19: Case Definitions. file:///C:/Users/nt/Downloads/WHO-2019-nCoV-Surveillance_Case_Definition-2020.2-eng.pdf [Accessed December 16, 2020]
8. World Health Organization (WHO). Operation Consideration for Case Management of COVID - 19 Health Facility , https://apps.who.int/iris/bitstream/handle/10665/331492/WHO-2019-nCoV-HCF_operations-2020.1-eng.pdf?sequence=1&isAllowed=y [Accessed March 19, 2020]

9. Ministry of Public Health and Population (MOPHP), Therapeutic Sector, Administration of Service and Emergency , National Guideline for Case Management of Mild and Moderate at Home of Coronavirus Disease 2019 (COVID-19), 2020 .
10. Norgen Biotek Corp. Norgen’s 2019-nCoV TaqMan RT-PCR Kit, 2020.
11. Moy J. Validation of Rapid COVID-19 Antibody Test Kits , Evaluation of COVID-19 Antibody Titers in Plasma of Individuals Who Have Recovered from COVID-19 Infection , Rush University, 2020.
12. Mercado M, Malagón-Rojas J, Delgado G, Rubio VV, Muñoz Galindo L, Parra Barrera EL et al. Evaluation of nine serological rapid tests for the detection of SARS-CoV-2. *Rev Panam Salud Publica*. 2020;44:e149. DOI: <https://doi.org/10.26633/RPSP.2020.149>
13. MedCalc Software Ltd. Diagnostic test evaluation calculator. https://www.medcalc.org/calc/diagnostic_test.php (Version 20.027; accessed January 30, 2022)
14. Bin-Ghouth AS, Al-Shoteri S, Mahmoud N, Musani A, Baoom NM, Al-Waleedi AA, Buliva E, Aly EA, Naiene JD, Crestani R, Senga M, Barakat A, Al-Ariqi L, Al-Sakkaf KZ, Shaef A, Thabit N, Murshed A, Omara S. SARS-CoV-2 seroprevalence in Aden, Yemen: a population-based study. *Int J Infect Dis*. 2022. 115:239-244. doi: 10.1016/j.ijid.2021.12.330.
15. Makaronidis J, Mok J, Balogun N, Magee CG, Omar RZ, Carnemolla A, et al. Seroprevalence of SARS-CoV-2 antibodies in people with an acute loss in their sense of smell and/or taste in a community-based population in London, UK: An observational cohort study. *PLoS Med* . 2020; 17:10 . <https://doi.org/10.1371/journal.pmed.1003358>
16. Stringhini S, Wisniak A , Piumatti G, Azman A S, Lauer A S, Baysson H, Ridder D, Petrovic D, Schrempft S, Marcus K, Yerly S, Arm V I, Keiser O, Hurst S, Posfay-Barbe K M, Trono D, Pittet D, Gétaz L, Chappuis F, Eckerle I, Vuilleumier N, Meyer B, Flahault A, Kaiser L, Guessous I.(2020). Seroprevalence of anti-SARS-CoV-2 IgG antibodies in Geneva, Switzerland (SEROCoV-POP): a population-based study. *The lancet* . 2020; 396:1 , 313-319 , [https://doi.org/10.1016/S0140-6736\(20\)31304-0](https://doi.org/10.1016/S0140-6736(20)31304-0)
17. Mahase E. Covid-19. Antibody prevalence in England fell from 6.0% to 4.4% over three months, study finds . *BMJ* 2020. 371:m4163 | <http://doi:10.1136/bmj.m4163>
18. Liu T, Wu S, ; Tao H, Zeng G, Zhou F, Guo F, and Wang X. Prevalence of IgG antibodies to SARS-CoV-2 in Wuhan – implications for the ability to produce long-lasting protective antibodies against SARS-CoV-2 . *medRxiv* . 2020. DOI: <https://doi.org/10.1101/2020.06.13.20130252>
19. Borges LP, Martins AF, Melo MS, Oliveira MGB, Neto JMR, Dósea MB et al. Seroprevalence of SARS-CoV-2 IgM and IgG antibodies in an asymptomatic population in Sergipe, Brazil. *Rev Panam Salud Publica*. 2020;44:e108. DOI: <https://doi.org/10.26633/RPSP.2020.108>
20. Stefanelli P, Bella A, Fedele G, Pancheri S, Leone P, Vacca P, Neri A, Carannante A, Fazio C, Benedetti E, Fiore S, Fabiani C, Simmaco M, Santino I, Zuccali MG, Bizzarri G, Magnoni R, Benetollo PP, Merler S, Brusaferrò S, Rezza G, and Ferro A. Prevalence of SARS-CoV-2 IgG

antibodies in an area of northeastern Italy with a high incidence of COVID-19 cases: a population-based study. *Clin Microbiol Infect.* 2021; 27:4. 633e1-633e7 . DOI: <https://doi.org/10.1016/j.cmi.2020.11.013>

21. Löfström E, Eringfält A, Kötz A, Wickbom F, Tham J, Lingman M Nygren, J M., and Undén J. Dynamics of IgG-avidity and antibody levels after Covid-19. *Journal of Clinical Virology.* 2021: 144:104986. DOI: <https://doi.org/10.1016/j.jcv.2021.104986>
22. Alzaabi AH, Ahmed LA, Rabooy AE, Zaabi AA, Alkaabi M, AlMahmoud F, Hamed MF, Bashaeb KO, Bakhsh AR, Adil S, Elmajed N, Abousalha AN, Uwaydah AK, Mazrouei KA. Longitudinal changes in IgG levels among COVID-19 recovered patients: A prospective cohort study. *PLoS One.* 2021 ;16(6):e0251159. DOI: 10.1371/journal.pone.0251159.
23. Bichara C.D.A. , Amoras E.S. G, and Vallinoto A. C. R, SARS-CoV-2 IgG antibodies post-COVID-19 in a Brazilian Amazon population. *BMC Infectious Diseases.* 2021; 21:443. DOI: <https://doi.org/10.1186/s12879-021-06156-x>
24. Ali A.M, Ali. K.M, Fatah M.H. Tawfeeq H.M and Rostam H.M. SARS-CoV-2 reinfection in patients negative for immunoglobulin G following recovery from COVID-19. *New Microbes and New Infections* 2021. 43:100926. DOI : <https://doi.org/10.1016/j.nmni.2021.100926>
25. Center for Diseases Control and Prevention (CDC) – US. Myths & Facts about Vaccine. <https://www.cdc.gov/coronavirus/2019-ncov/vaccines/facts.html> [Accessed December 15, 2021]