

1
2
3
4
5
6
7
8
9
10
11

The Effect of Extract Turmeric (*Curcuma longa*) on Spermatozoa Motility in Fresh Semen of Goats

ABSTRACT

Aims: Determine the motility of spermatozoa in fresh goat semen after being given turmeric extract.

Study design: Randomized Block Design

Place and Duration of Study: Sample: Galang, Deli Serdang Regency, Indonesia, between December 2022 and January 2023.

Methodology: Giving turmeric extract to the goat once a day. The treatment included adding 0ml, 25ml, 50ml, 75ml, and 100ml turmeric extract. The parameter observed was the percentage of motility spermatozoa in fresh goats semen. The preparation of turmeric extract, Feeding goats using turmeric extract every day at 08.00, collection of fresh semen, evaluating motility spermatozoa. This research obtained conducted using a Randomized Block Design consisting of 5 treatment levels and five replications. Semen storage using 30 Bucks, which has done for three days.

Results: The addition of turmeric extract 0ml showed an motility value of 65%, 25ml = 70%, 50ml = 75%, 75ml = 75% and 100 ml = 75%. The analysis of variance showed that the results give a real impact on the motility of spermatozoa. The results showed that the addition of turmeric extract had significant effect ($P < 0.01$).

Conclusion: Adding 50 ml of turmeric extract was the best result on spermatozoa motility in fresh goat semen.

Keywords: Fresh semen, Goat, Motility, Spermatozoa, Turmeric.

1. INTRODUCTION

The goat livestock population still needs to be increased to meet the demand for meat as a source of animal protein [1]. Therefore it is necessary to make efforts to increase the population of goats. The goat breeding business is underdeveloped in several countries, and most are still considered side businesses. Business management is not good because the reproductive activity is not optimal, so the production and productivity of the resulting seeds are low [2].

Research on efforts to increase reproductive efficiency in male livestock still needs to be improved. In this study, the herbal feed comes from plant extracts. Herbal plant extracts have several advantages over fresh ingredients. The advantages of herbal plant extracts include that the body more easily absorbs the nutritional content, the chance of being contaminated with pesticides, heavy metals, and others is smaller than fresh ingredients, the handling is practical, and the dosage is easily adjusted, especially if need a large number of doses [3].

Turmeric is an herbal ingredient that is easy to find and cheap. Turmeric contains antioxidant ingredients that can improve sperm quality. Curcumin compounds contained in turmeric can increase the percentage value of spermatozoa motility. Motility is essential for

* E-mail address: sukmaaditya@dosen.pancabudi.ac.id

12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32

33 fertilization because spermatozoa can move efficiently toward the egg (ovum). Curcumin can
34 also protect sperm against oxidative damage from chemical exposure [4].

35 Turmeric's effect on animals can increase spermatozoa motility, increase
36 concentration, protect against oxidative damage, and increase levels of the hormone
37 testosterone. In addition, the curcumin compound contained in turmeric can help reduce
38 stress, which also impacts male fertility [5]. Other ingredients contained in turmeric, such as
39 vitamins C and E, are potent antioxidants that can help ward off free radicals. These
40 compounds can also help increase male sperm volume [6].

41 Based on this background, this study will observe the effect of giving turmeric extract
42 to increase the reproductive productivity of male goats, especially the motility of goat
43 spermatozoa in fresh semen. Hopefully, this research can be applied to buck, which will be
44 starters, and for producing fresh semen to produce liquid/frozen semen.

45
46

47 **2. MATERIAL AND METHODS**

48

49 This research is experimental. That is, researchers conduct experiments with several
50 treatments on the samples obtained. The scope of this research focuses on observing
51 sperm macroscopically and microscopically. The research materials used were Boer
52 goats, 2% eosin, straw, liquid N₂, aquabidestillata, warm water (45-55°C), and tissue. The
53 research tools used were a 400 ml beaker glass, deck glass, tissue, stir bar, a set of an
54 artificial vagina, test tube, water bath, electric microscope, thermometer, pipette, object
55 glass, cover glass, pH meter, Bunsen burner, denominator, holding cages and doormats.
56 The population and sample in this study were Boer Buck that had been fed turmeric
57 extract with the treatment given as follows:

- 58 P0 = Turmeric Extract 0 ml
- 59 P1 = Turmeric Extract 25 ml
- 60 P2 = Turmeric Extract 50 ml
- 61 P3 = Turmeric Extract 75 ml
- 62 P4 = Turmeric Extract 100 ml

63

64 The procedure of this research is as follows.

65 1. Preparation of Materials and Tools

66 The equipment was cleaned and sterilized using an autoclave at 121°C for 15 minutes.

67 2. Preparation of Turmeric Extract

68 Chopped turmeric of ± 5 mm, then dried in the oven at 50°C for 6 hours until constant final
69 weight. The dried meat is then mashed using a blender and filtered into powder (Simplicia).
70 The simplicial was extracted by maceration method using 70% ethanol and distilled water for
71 three days at room temperature. Then filtered and concentrated with a vacuum rotary
72 evaporator at 60°C.

73 3. Provision of Turmeric Extract

74 100 ml of turmeric extract is given orally every morning using a bottle

75 4. Fresh Semen Collection

76 Semen collection uses the artificial vagina method, in which males collect semen using a
77 method similar to natural mating activities. The shelter is carried out in the morning using an
78 artificial vagina (temperature 40°C – 42°C). The ejaculate results were immediately brought
79 to the laboratory for microscopic evaluation [7].

80 5. Motility Evaluation

81 Observations using a microscope. Motility by looking at the spermatozoa, which move
82 progressively forward. The evaluation used observing the spermatozoa in eight different
83 fields of view with a 400x magnification light microscope [8].

84

85

86 **3. RESULTS AND DISCUSSION**

87

88 **Table 1. Spermatozoa motility before equilibration with turmeric extract**
89 **supplementation**

Parameter	Treatment	Percentage (%)
Motilitas	0 ml	65±0.00
	25 ml	70±0.00
	50 ml	75±0.00
	75 ml	75±0.00
	100 ml	75±0.00

90 *Note: Different superscripts in the column show a significant difference (P<0.01)*

91

92 The motility test results of fresh goat semen spermatozoa showed that the lowest
93 percentage value was without treatment, namely 65%, while the highest was with the
94 addition of 50 ml, 75 ml, and 100 ml of turmeric extract, namely, motility 75%. From the data
95 obtained, adding turmeric extract increased the motility percentage of fresh goat semen
96 spermatozoa. The higher the administration level of turmeric extract will further increase the
97 percentage value of spermatozoa motility.

98 The analysis of variance showed that adding turmeric extract feed had a significant
99 (P<0.01) effect on spermatozoa motility in fresh goat semen. The results of the BNT follow-
100 up test showed the highest motility in the 50ml, 75ml, and 100ml treatments, namely 75%
101 motility.

102 The percentage of motility in spermatozoa or the motility of spermatozoa is one of
103 the determinants of the success of spermatozoa in reaching the ovum in the fallopian tubes.
104 It is the simplest way of evaluating sperm for artificial insemination [9].

105 The results showed that the average observed sperm motility in goat semen of all
106 treatments met the standards for use as goat semen for Artificial Insemination because the
107 above motility was 40%, and this figure met the requirements for Artificial Insemination.
108 Requirements for proper semen Artificial Insemination has a minimum motility of 40% [10].
109 The best spermatozoa motility that can be used and meets the standards in this study is in
110 the treatment of 50 ml of turmeric extract. Semen quality will decrease if storage is not with
111 the correct diluent [11].

112 Hydroxynonenal is a lipid peroxidation that can inhibit glycolysis and motility of
113 spermatozoa. In addition to the damage caused by lipid peroxidation, decreased motility can
114 also occur due to several factors [12]. Factors that can reduce motility are changes in pH
115 medium, osmotic pressure, and electrolyte and non-electrolyte effects [13].

116 The motility percentage decreased due to reduced food reserves after being stored
117 for four days, significantly affecting livestock reproduction [14]. Semen contains many
118 compounds needed for the metabolism and movement of spermatozoa [15]. In cement, the
119 primary carbohydrate source is fructose [8]. Spermatozoa will utilize fructose in plasma for
120 survival and movement. After fructose runs out, GPC (Glycerol Phosphoryl Choline) will
121 break slowly to resupply fructose [16].

122

123

124 **4. CONCLUSION**

125

126 The best research result is the administration of 50 ml of turmeric extract per day to get the
127 highest motility. Recommended to research by evaluating spermatozoa microscopically
128 using other parameters.

129
130

131 **ACKNOWLEDGEMENTS**

132

133 The authors would like to thank the University of Pembangunan Panca Budi for funding this
134 research.

135
136

137 **COMPETING INTERESTS**

138

139 Authors have declared that no competing interests exist.

140
141

142 **AUTHORS' CONTRIBUTIONS**

143

144 This work was carried out in collaboration between all authors. Sukma Aditya Sitepu
145 designed the study, performed the statistical analysis, wrote the protocol and wrote the first
146 draft of the manuscript. Julia Marisa managed the analyses of the study. Muhammad Miftah
147 Farid managed the literature searches. All authors read and approved the final manuscript

148
149

150 **REFERENCES**

151

- 152 1. Marisa, J., Sitepu, S. A., dan Rianto, A. A. 2022. ANALYSIS OF ACTORS AND
153 ACTIVITIES IN VALUE CHAIN BUSINESS SHEEP FAULTING. In *Proceeding*
154 *International Conference of Science Technology and Social Humanities* (Vol. 1, pp. 69-
155 76).
- 156 2. Hastono and Masbulan E. 2001. Performance of people's sheep reproduction in Garut
157 Regency, In: *Proceedings. National Seminar on Animal Husbandry and Veterinary*
158 *Technology*. Bogor, Indonesia, 17-18 September 2001. pp 100-105.
- 159 3. Farzaneh, V., & Carvalho, I. S. (2015). A review of the health benefit potentials of herbal
160 plant infusions and their mechanism of actions. *Industrial Crops and Products*, 65, 247-
161 258.
- 162 4. Yan, W., Kanno, C., Oshima, E., Kuzuma, Y., Kim, S. W., Bai, H., ... & Kawahara, M.
163 (2017). Enhancement of sperm motility and viability by turmeric by-product dietary
164 supplementation in roosters. *Animal reproduction science*, 185, 195-204.
- 165 5. Djaelani, M.A. (2010). Spermatozoa Concentration of Mice (*Mus Musculus*) Swiss
166 Webster L. after Administration of Turmeric (*Curcuma Domestica*) Rhizome Powder with
167 Chronic Doses. *PHYSIOLOGICAL ANATOMY*, 18(2), 56-63.
- 168 6. Angulo, C., Rauch, M. C., Droppelmann, A., Reyes, A. M., Slebe, J. C., Delgado-López,
169 F., ... & Concha, I. I. (1998). Hexose transporter expression and function in mammalian
170 spermatozoa: cellular localization and transport of hexoses and vitamin C. *Journal of*
171 *cellular biochemistry*, 71(2), 189-203
- 172 7. Sitepu, S. A., and Marisa, J. 2019. Percentage value of membrane integrity and
173 acrosome integrity of spermatozoa in simmental liquid semen with addition of penicillin

- 174 and sweet orange essential oil. In IOP Conference Series: Earth and Environmental
175 Science (Vol. 327, No. 1, p. 012027). IOP Publishing.
- 176 8. Gaffney, E. A., Gadêlha, H., Smith, D. J., Blake, J. R., & Kirkman-Brown, J. C. (2011).
177 Mammalian sperm motility: observation and theory. *Annual Review of Fluid*
178 *Mechanics*, 43, 501-528.
- 179 9. Sitepu, S. A., Marisa, J., dan Rianto, A. A. 2022. SPERMATOZOA MOTILITY IN BOER
180 BUCK LIQUID SEMEN WITH ADDITION OF SWEET ORANGE ESSENTIAL OIL AND
181 PENICILLIN. In *Proceeding International Conference Keputeraan Prof. H. Kadirun*
182 *Yahya* (Vol. 1, No. 1, pp. 98-103).
- 183 10. Lehti, M. S., & Sironen, A. (2017). Formation and function of sperm tail structures in
184 association with sperm motility defects. *Biology of reproduction*, 97(4), 522-536.
- 185 11. Sitepu, S. A., & Udin, Z. (2020). Spermatozoa Motility of Boer Buck Semen with Addition
186 of Sweet Orange Essential Oil to Tris Yolk Extender. *Asian Journal of Biotechnology and*
187 *Genetic Engineering*, 3(3), 35-38.
- 188 12. Hafez, E. S. E., dan B. Hafez. 2005. Reproduction in Farm Animal 7 th ed. Lippincott
189 Williams and Walkins, South Carolina.
- 190 13. Affandy, L., U. Umiyasih, D. Pamungkas and D.E. Wahyono, 1997. Reproductive
191 Response of Fat Ekr Sheep To Differences in Feed Energy Levels. GratiPasuruan
192 Agricultural Technology Research and Assessment Installation. National Seminar on
193 Animal Husbandry and Veterinary
- 194 14. Marisa, J., and Sitepu, S.A. 2020. Relationship analysis between production factors and
195 business production of beef cattle livestock in West Binjai District, Indonesia. *Asian*
196 *Journal of Advanced Research and Reports*, 1-7.
- 197 15. Hasan, F., Sitepu, S. A. P., and Alwiyah, A. 2017. The Influence of Parity Against a
198 Percentage of Estrus of Ekor Tipis Sheep that are Synchronized Estrus Using
199 Prostaglandin F_{2α} (PGF_{2α}). *Jurnal Ilmu Produksi dan Teknologi Hasil Peternakan*, 5(1),
200 46-48.
- 201 16. Ashari, M., Suhardiani, R. A., and Andriati, R. 2018. Analysis of Reproductive Efficiency
202 of Fat-tailed Sheep in East Lombok District. *Indonesian Journal of Animal Husbandry*
203 *Science and Technology Volume*, 4(1), 207-213