

### **Bioaccessibility of trace elements in different oyster mushroom varieties grown in Kenya**

#### **ABSTRACT**

Trace elements, especially chromium (Cr), vanadium (V) and selenium (Se) have potential beneficial effects on glucose metabolism in people with type 2 diabetes. Food products incorporating mushrooms are not only a good source of such nutrients but are thought to have readily bioavailable nutrients. Nutritional efficacy of food products may be ensured by accessing bioaccessibility of nutrients, which provides valuable information on matrix and appropriate dosage. The study determined bioaccessible Cr, V and Se in four varieties of oyster mushrooms *Pleurotus ostreatus* (PO), *Pleurotus sajor-caju* (PS), *Pleurotus pulmonarius* (PP) and *Pleurotus citrinopileatus* (PC) grown in Kenya. Bioaccessibility was estimated using in vitro simulated gastrointestinal procedure, while nutrient levels were determined using atomic absorption procedure. Bioaccessible levels of chromium ranged from 26.56% in PS to 78.50% in PC; selenium from 92.52% in PC to ND in PS and PP; and vanadium from 92.46% in PC to 69.95% in PP. Vanadium was the most bio accessible than the other elements in the four oyster mushrooms, while chromium was the most bio accessible in the PC variety.

**Keywords:** Bioaccessibility, Oyster mushrooms varieties, Vanadium, Chromium, Selenium

## Introduction

“Bioaccessibility” can be defined as quantity or fraction which is released from the food matrix in the gastrointestinal tract and becomes available for absorption” (Heaney, 2001; Onyambu *et al.*, 2021; Nambafu *et al.*, 2021). “Bioavailability on the other hand is the ingested fraction available at the site of action for utilization in normal physiological functions; usually determined through *in vivo* assays” (Guerra *et al.*, 2012; Lusi *et al.*, 2013). “It is the result of three main steps: digestibility and solubility of the element in the gastrointestinal tract; absorption of the element by the intestinal cells and transport into the circulation; and incorporation from the circulation to the functional entity or target” (Wienk *et al.*, 1999; Etcheverry *et al.*, 2012). “Bioaccessibility” is usually evaluated by *invitro* digestion procedures generally simulating gastric and small intestinal digestion, sometimes followed by Caco-2-cell uptake” (Courraund *et al.*, 2013). “*In vitro* methods are developed to simulate the physiological conditions (temperature, agitation, pH, enzyme, and chemical composition) and the sequence of events that occur during digestion in the human gastrointestinal tract” (Fernandez Garcia *et al.*, 2009). The bioaccessible level of trace elements after consumption is not necessarily related to the levels in foods since food processing and cooking affect intestinal transit time of nutrients as well as their enteric formation of mixed micelles (Nambafu *et al.*, 2021, Nawiri *et al.*, 2013). In this study a static method was used which involved *invitro* enzymolysis procedure simulating human gastro intestinal involving two steps (gastric and intestinal).

Mushrooms offer tremendous applications as they can be used as food and medicines besides their key ecological roles. Bano (1976) suggested that “food value of mushrooms lies between meat and vegetables”. “The fruiting bodies of mushrooms are characterized by a high level of well assimilated mineral elements. Major mineral constituents in mushrooms are K, P, Na, Ca,

Mg and elements like Cu, Zn, Fe, Mo, Cd form minor constituents” (Samsudin and Abdullah, 2019; Kalac, 2013; Li *et al.*, 2021). “The present use of mushrooms is totally different from the traditional use because a lot of research has been done on the chemical composition of mushrooms, which has revealed that mushrooms can be used as a diet to combat diseases. The early history regarding the use of mushrooms in different countries has been reviewed by a number of workers” (Samsudin and Abdullah, 2019; Murugesan, 2017; Chang and Wassers, 2017; Ho *et al.*, 2020; Kalac, 2013; Li *et al.*, 2021). “The mineral proportions vary according to the species, age, and the diameter of the fruiting body. It also depends upon the type of the substratum” (Kalac and Svoboda, 2000). “The mineral content of wild edible mushrooms has been found higher than cultivated ones” (Aletor, 1995; Mattilla *et al.*, 2001; Rudawska and Leski, 2005). “The minerals have been found to boost the immune system, have anti-cancerous properties, and act as anti-hypercholesterolaemic and hepatoprotective properties. Some mushrooms such as *Pleurotus* species are excellent food for the people suffering from hypertension and cardiovascular diseases due to high potassium and sodium content” (Ganesan and Xu, 2018). “Chromium is generally recognized to play an important role in glucose and lipid metabolism. One effect of chromium is that it is very helpful in preventing and reversing type 2 diabetes because plasma glucose is more effectively regulated in the presence of chromium” (Anderson *et al.*, 1997).

“Vanadium compounds have been demonstrated to mimic the action of insulin in isolated cell systems, animal models and diabetic patients. This has brought the use of V compounds as potential sources of diabetes therapy into focus” (Shechter, 1990 and Shamberger, 1996).

“Selenium have been shown to reduce the risk of developing dysglycemia- a broad term that

refers to any abnormalities in blood glucose levels leading to disease” (Akbaraly *et al.*, 2010). “It is also thought to work by exerting insulin-like actions *in vitro*” (Becker, *et al.*, 1996).

In Kenya hypertension and diabetes are becoming the major causes of morbidity and mortality with their prevalence surpassing that of HIV/AIDs (WHO, 2012). Various mushrooms species are used in the treatment and prevention of diabetes, due to their rich nutritional and medicinal properties such as the elements (K, Ca, Na, P, Fe, Cr, Se, and V) (Rajarithnam and Shashirekha, 2011). The management of diabetes through use of nutraceuticals requires that the substrate not only contain high levels nutrients but also bioavailable nutrients. Since the levels of the nutrients depend on growing medium, variety, maturity of tissue and climatic condition it is important to quantify the levels in various species of mushrooms. Mushrooms have a chitin structure that is likely to have effect on the micronutrients bioavailable. Bioavailability of nutrients depends on various factors such as food tissue and on host related factors and is studied through intervention (*in vitro* procedures), or algorithms and simulated gastrointestinal digestion (*in vivo* methods). Bioavailability is improved by formulating a food product containing high levels of nutrients such Cr, V and Se that are important in management of diabetes. This study was conducted to determine the bioaccessibility of trace elements (chromium, vanadium, and selenium) in the fresh mushrooms commonly consumed in Kenya.

## **2. Material and Methods**

### **2.1 Experimental**

#### **2.1.1 Samples and sample preparation**

Four oyster mushroom that is *Pleurotus ostreatus* (PO), *Pleurotus sajor-caju* (PS), *Pleurotus pulmonarius* (PP) and *Pleurotus citrinopileatus* (PC) varieties were collected from nearby farms in Jomo Kenyatta University and identified by a taxonomist. The freshly harvested mushrooms were cleaned, cut into pieces, and dried in the shade for four days and then powdered before keeping in a labeled special glass container awaiting analysis.

### **2.1.2 Reagents and Apparatus**

All reagents were of analytical reagent grade. Double deionized water was used in dilution of reagent solutions. HNO<sub>3</sub> and H<sub>2</sub>O<sub>2</sub> were of supra pure quality (Merck). Gastric juice, Pepsin (1% w/v), Pcreatin (3% w/v), Amylase (1% w/v), Bile salt, selenium, chromium and vanadium as standards were purchased from Sigma Andrich. All plastics and glassware were cleaned, soaked in dilute HNO<sub>3</sub> (10%) and rinsed with distilled water prior to use.

### **2.1.3 Instrumentation**

Atomic Absorption Spectrophotometer (AAS), Thermo Jarell Ash (model AAS S11) was used for analysis of trace elements. The most appropriate wavelength, flow rate, slit width and other AAS instrument parameters for metals, minerals were selected as given in the instrument user's manual and background correction was used during determination of metals/minerals.

## **2.2 Procedures of analysis**

### **2.2.1 Method validation**

Calibration of AAS was done using the working standard prepared from commercially available metal/mineral standard solutions (1000 $\mu$ g/ml. The stock solutions were kept under refrigeration conditions to be used for analysis. The working solutions of different concentrations were prepared daily by serial dilution of the standard. Various volumes of the stock solutions in each case (2, 4, 6, 8 ml) were further diluted to 100 ml to obtain the working solutions of concentrations 50 ppm, 100 ppm, 150 ppm and 200 ppm. After filtering absorbencies were obtained and used to draw calibration curves. All analyses were done in triplicates.

### **2.2.2 Determination of trace elements**

Each mushroom sample was air-dried at 105 °C overnight and crushed using a mortar and pestle into powder. Digestion of the mushroom samples was performed using a mixture of HNO<sub>3</sub>:H<sub>2</sub>O<sub>2</sub> (10:1, 12 ml g<sup>-1</sup> of sample) and heated at 100 °C for between 10-15 min. After cooling, the solution was made to 50 ml with deionized water after filtration. The amounts of Cr, Se and V were determined using an atomic absorption spectrophotometer. All analyses were done in triplicates.

### **2.2.3 *In vitro* simulated gastrointestinal digestion**

The *in vitro* enzymolysis procedure simulating human gastrointestinal digestion was carried out in triplicate. Procedural blanks were run to check the presence of Se, Cr and V in the reagents. Mushroom samples (0.5 g) in a flask were incubated with 5 ml of gastric juice (1% w/v pepsin in 0.15 M NaCl, adjusted to pH 2 with HCl (37% v/v) and, after 1 min of vigorous shaking for initial degassing, the flask was placed in a mixing water bath at 37<sup>0</sup> C for 4 h (Crews *et al.*, 1996). The solution was then adjusted to pH 6.8 with NaHCO<sub>3</sub>. After adding 5 ml of intestinal juice (3% w/v pancreatin, 1.5% w/v amylase, 1% w/v bile salts in 0.15 M NaCl), solution was

vigorously shaken for 1 min, degassed, and further incubated for 4 h at 37 °C under gentle shaking. The solution was then centrifuged at 8000 g and 4 °C for 15 min, the supernatant collected, filtered through 0.45 µm membranes, and stored at -80°C until analysis. The amount of solubilized chromium, selenium, and vanadium in the supernatant was measured using atomic absorption spectrophotometer as a measure of their bioaccessibility and data analyzed using SPSS.

#### **2.2.4 Stastical analysis**

The data was analysed using SPSS program. ANOVA was used to determine whether there is significant difference in the bioaccessibility of the trace elements. In this study the between sample estimate of variance is greater than within sample that is Chromium between group (BG) variation was 2.808 while the within group variation (WG) was 0.472; Vanadium BG(0.013) and WG(0.001) and selenium BG(116.691) and WG(44.222) to be explained by random error hence a one-tailed F-test was calculated to test whether it is significantly greater. From the table of critical values of F for a one-tailed test ( $p=0.05$ ) the critical value of F is 8.845. The calculated value of F (5.946 for chromium) and 2.639 is smaller than this the null hypothesis is accepted the sample means do not differ significantly while that for Vanadium is 9.094 is greater than the critical value. A significant result in one way ANOVA can arise for several reasons: one mean may differ from others, all the means may differ from each other, and the means may fall into two distinct groups. A simple way of deciding the reason for a significant result is to arrange the means in increasing order and compare the difference between adjacent values with a quantity called least significant difference (Miller and Miller, 1988). "The least significant difference method is not entirely rigorous, it can be shown that it leads to rather too many significant difference. However it is a simple follow up test when ANOVA has indicated that there is a

significant difference between means” (Harris, 2007). In this study the lowest mean was 0.450 and the highest was 3.501. The least significant difference test is used in the context of ANOVA when the F-ratio suggests rejection of the null hypothesis. Turkey, Students-Newman-Keuls (SNK) and Bonferroni Post Hoc test are used for reporting the range tests and pairwise multiple comparisons to determine means that differ in ANOVA. Range tests identify homogeneous subsets of means that are not different from each other. Pairwise multiple comparisons test the difference in each pair of means and yield a matrix where asterics indicates significantly different group means at alpha level of 0.05. In this study Bonferroni post Hoc test was used (Table 1). The PO variety in chromium showed a significant difference (Table 1).

**Table 1: Bioaccessible levels of Chromium, Vanadium and Selenium**

Mushroom Variety	Level of Chromium Mean (Number) ug/g		Level of Vanadium Mean (Number) ug/g		Level of Selenium Mean (Number) ug/g	
	Bio	Raw	Bio	Raw	Bio	Raw
PC	1.1221(3)	0.9506(3)	0.4159(3)	0.4498(3)	14.8466(3)	1.1102(3)
PO*	1.0025(3)*	3.510(3)*	0.4138(3)	0.4880(3)	8.91368(3)	1.1652(3)
PP	0.6789(3)	0.7478(3)	0.4237(3)	0.6057(3)	-1.3883(3)	0.7757(3)
PSC	0.4119(3)	0.9559(3)	0.4011(3)	0.4710(3)	-5.1000(3)	2.3377(3)

\*. The mean difference is significant at the 0.05 level. Based on *post hoc* Turkey HSD

### 3 Results and Discussion

#### 3.1 Levels of trace elements in oyster mushrooms

The mean trace element concentrations ( $\mu\text{g/g}$ ) in dry weight in the mushroom samples were Chromium PSC(1.551), PP(1.798), PO(3.501), and PC(1.430). Vanadium concentrations in the four species were PS(2.338), PP(0.776), PO(1.165) and PC(1.110). Selenium levels were PSC (0.471),

PP(0.606),PO(0.488) and PC(0.450)(Table 2).The precision of the results was evaluated based on the standard deviation of the results of triplicate samples(n=3) analyzed under the same conditions mean±SD. The mean concentration of chromium was highest in PO and lowest in PP.Selenium concentration was highest in PSC and lowest in PP.Vanadium concentration was highest in PC and lowest in PP. The values are comparable to those discussed by authors (Patil *et al.*, 2010;Yang *et al.*, 2001;Ahmed *et al.*, 2009;Alam *et al.*, 2008;Gosh and Chakraborty,1990;Gasecka *et al.*, 2016;Kortei and Wiafe-kwangan,2015;Bano *et al.*,1981;Jegadeesh *et al.*,2018,Ijeh *et al.*,2009).

**Table 2: Mean levels of trace elements in Oyster mushrooms**

Mushroom variety	Concentration (µg/g)		
	Chromium Mean±SD	Vanadium Mean±SD	Selenium Mean
PSC	1.551±1.661	0.471±1.481	2.338±0.055
PP	1.798±1.472	0.606±3.341	0.776±0.066
PO	3.501±0.857	0.488±0.946	1.165±0.01
PC	1.430±0.657	0.450±2.472	1.110±0.06

PO *Pleurotus ostreatus*, PSC *Pleurotus Saju-sajur Caju*, PP *Pleurotus pulmonarius*, PC *Pleurotus cintrinopileatus*

### 3.2 Trace elements Bioaccessibility

“Absorption of selenium occurs throughout gastrointestinal tract, with duodenum as a major site” (Humaloja and Mykkänen, 1986). Selenium absorption is very efficient, normally in the range of 26.3% - 97% (Durcos *et al.*, 2005; Thompson *et al.*, 1978), with higher retention of organic selenium compound than inorganic selenium compounds (Finely, 1999). “Very little is known

about mechanism of selenium absorption, it may involve both carrier mediated process for organic compound and diffusion-controlled process for inorganic selenium compound” (Arduser *et al.*, 1985).

The amounts of selenium bioaccessible in the four oyster mushrooms are as shown in Table 3 are PO-86.94%, PC-92.52% which is high and is comparable to literature value (Durcos *et al.*, 2005; Thompson *et al.*, 1978) and thus is potentially bioavailable. The other varieties, PS and PP had undetected levels. The soluble extract obtained after *in vitro* simulated gastrointestinal digestion of the selenized mushroom contained about 106gSe dry weight. This means that 75% of the Se taken up by the mushroom was solubilized in conditions simulating human gastrointestinal digestion and thus was potentially bioavailable (Hur *et al.*, 2011). Se content of mushrooms is generally higher than that of most vegetables (Rayman *et al.*, 2008) but it is very variable.

“Se that is not solubilized after gastrointestinal digestion might be present in form of indigestible Se-containing polysaccharides. For instance, it has been shown that part of the Se in Se-enriched mycelia of *Pleurotus ostreatus* is associated with chitin-containing structures in cell walls” (Munoz *et al.*, 2006). “Formation of Se-containing polysaccharides might explain the low Se bioavailability found elsewhere for other species” (Chansler *et al.*, 1986; Mutanen, 1986). Limited bioavailability of Se in these species might be the result of a low bioaccessibility due to a larger incorporation of Se in mushroom polysaccharides compared to *Pleurotus ostreatus* in the conditions of the present study. Bioavailability of selenium from PO, PF, PSC, and PE was found to range between 0.011 and 0.512mg/100 g (Gasecka *et al.*, 2016; Tang *et al.*, 2006).

The bioaccessible vanadium in the four oyster mushrooms in this study was PS-85.16%, PP-69.95%, PO-84.80% and PC-92.46% (Table 3). Many studies confirm that of the total dietary

vanadium ingested, less than 5% is absorbed by the gastrointestinal tract(GI)(Curran *et al.*, 1990;Byrne, 1978;Nielsen, 1988).Other studies claim greater than 10% of ingested vanadium may have resulted in greater than normal absorption efficiencies (Nielsen, 1990).The majority of the 5% dietary vanadium absorbed is taken up by the upper GI tract(Patterson,1986).

**Table3: Bioaccessibility of elements in four oyster mushrooms**

Mushroom variety	Bioaccessibility (%)		
	Chromium	Vanadium	Selenium
PSC	26.56	85.16	ND
PP	37.59	69.95	ND
PO	28.63	84.80	86.94
PC	78.50	92.46	92.52

ND: *Not Detected*

The bioaccessible chromium levels in the four oyster mushrooms were PS-26.56%,PP-37.59%,PO-28.63% and PC-78.50% (Table 3).These values are comparable to those in literature *invitro* and in vivo studies in rats have shown that about 80% Cr in the blood is associated with transferring (Feng, 2003). “Chromium is absorbed together with other metal ions in the gut through the unsaturated passive transport.The absorption process depends on the Cr content in the diet and on the chemical form of this element and other food components” (Dowling *et al.*,1989). “The efficiency of this process is very low with the average absorption ranging from 0.4-2.5%” (European Commission, 2003). “The absorption process depends on the Cr content in the diet and on the chemical form of this element and other food components” (Anderson, 1996). “Organic sources of Cr (picolinate, or propionate-methionine salt) are much better absorbed than inorganic forms (oxides), and lead to the increase of these compounds’ concentration in tissues” (Ohh and Lee, 2005; Wang *et al.*, 2009; Zha *et al.*,2007). “However, other factors present in the

diet show a significant impact on the amounts of Cr absorbed from the gastrointestinal tract. Starch, simple sugars, ascorbic acid, oxalic acid, nicotinic acid, some amino acids, aspirin increase absorption of this element” (Chen *et al.*, 1973; Davis *et al.*,1995; Offenbacher,1994; Samanta,2008) while “high concentrations of phosphate, calcium, magnesium, titanium, zinc, vanadium and iron reduce the rate of this process” (Chen *et al.*, 1973; Hill,1976).

“More than 80% of Cr is removed from the body in the form of urine, while the remaining part of this element is excreted via faeces and sweat” (Ducros, 1992). “In humans, consumption of large amounts of sugar, exhaustive physical exercise, pregnancy, and lactation leads to increased Cr excretion in the urine” (Anderson, 1989).

#### **4 CONCLUSIONS**

The study indicates that the oyster mushroom contains fairly high levels of the elements, but their bio accessibility differs in all the varieties. The extent of bio accessibility of the elements was affected by the variety although selenium remained least bio accessible in all varieties. Vanadium was better bio accessible in the four oyster mushrooms, while chromium was better bio accessible in the PC variety.

#### **REFERENCES**

- Ahmed, S.A., Kadam, J.A., Mane, V.P. (2009). Biological efficiency and nutritional contents of Pleurotus Florida. Singer cultivated on different agro-wastes. *Nature Sci.* **7(1)**:44-48.
- Akbaraly, T.N., Arnaud, J., Rayman, M.P., Hininger-Favier, I., Roussel, A.M., Berr, C., Fontbonne.A. (2010). Plasma selenium and risk of dysglycemia in an elderly French population: Results from the prospective epidemiology of Vascular Ageing Study. *Nutrition Metabolic London.* **18**:71-121.
- Alam, N., Amin, R., Khan, A. (2008). Nutritional analysis of cultivated mushrooms in Bangladesh -Pleurotusostreatus, Pleurotussajorcaju, Pleurotusflorida and Calocybeindica. *Mycobiology.* **36(4)**:228-232.

Aletor, V.A and Aladetimi, O.O. (1995). Compositional studies on edible tropical species of mushrooms. *Food Chem.***54**: 265-268.

Anderson, R.A., Bryden, N.A., Polansky, M.M., Gautschi, K. (1996). Dietary chromium effects on tissue chromium concentrations and chromium absorption in rats. *J Trace Elem Exp Med.***9(1)**: 11-25.

Anderson, R.A., Bryden, N.A., Polansky, M.M., Richards, M.P. (1989). Chromium supplementation of turkeys: effects on tissue chromium. *J Agric Food Chem.* **37**: 131-134.

Anderson, R.A., Cheng, N., Bryden, N.A., Polansky, M.M., Cheng, N., Chi, J., Feng, J. (1997). Elevated intakes of supplemental chromium improve glucose and insulin variables in individuals with type 2 diabetes. *Diabetes.***46**:1786-91.

Arduser, F., Wolfram, S. and Scharrer, E. (1985). Active Absorption of Selenate by Rat Ileum. *J. Nutr.***115**:1203-1208.

Bano, Z. (1976). Nutritive value of Indian mushrooms and medicinal practices. *Eco. Bot.***31**: 367-371.

Becker, D.J., Reul, B., Ozcelikay, A.T., Buchet, J.P., Henquin, J.C., Brichard, S.M. (1996). Oral selenate improves glucose homeostasis and partly reverses abnormal expression of liver glycolytic and gluconeogenic enzymes in diabetic rats. *Diabetologia.***39**:3-11.

Byrne, A.R and Kosta, L. (1978) Vanadium in foods and in human body fluids and tissues. *Sci. J. Nutr.***115**:1203-1208

Chang, S., Wasser, S. (2017). *The cultivation and environmental impact of mushroom*. New York. Oxford University press. P43.

Chansler, M. W., Mutanen, M., Morris, V. C., Levander, O. A. (1986). Nutritional bioavailability to rats of selenium in Brazil nuts and mushrooms. *Nutrition Research.***6(12)**.1419-1428.

Chasteen, N.D. (1983). The biochemistry of vanadium. *Struc Bonding.***53**:105.

Chen, J., Gaikwad, V., Holmes, M. (2011). Development of a simple model device for in vitro gastric digestion investigation. *Food Funct.***2**:174-182.

Chen, N.S., Tsai, A., Dyer, I.A. (1973). Effect of chelating agents on chromium absorption in rats. *J Nutr.***103(8)**: 1182-1186.

Combs, J.R. (2001). Selenium in global food systems. *Br J Nutri* .**85**:1517-547.

Courraud, J., Berger, J., Cristol, J.P., Avallone, S. (2013). Stability and Bioaccessibility of different forms of carotenoids and vitamin A during In vitro digestion. *Food Chem.***136**:871-7.

Crews, H.M., Clarke, P.A., Lewis, D. J., Owen, M., Strutt, P. R., Izquierdo, A. (1996). Investigation of selenium speciation in vitro gastrointestinal extracts of cooked cod by high-performance liquid chromatography-inductively coupled plasma mass spectrometry. *Journal of Analytical Atomic Spectrometry.***11**:1177-1182.

- Curran, G.L., Azarnoff, D.L., Bolinger, R.E. (1990). *J Clin Invest.***38**:1251-126.
- Davis, M.L., Seaborn, C.D., Stoecker, B.J.(1995).Effects of over-the-counter drugs on Chromium retention and urinary excretion in rats.*Nutr Res.* **15**(2): 201-210.
- Dowling, H.J., Offenbacher, E.G., Pi-Sunyer, F.X.(1989). Absorption of inorganic trivalent chromium from the vascular perfused rat small intestine.*J Nutr.***119** (8): 1138-1145.
- Ducros, V. (1992).Chromium metabolism, a literature review.*Biol Trace Elem Res.* **32**: 65-77.
- Durcos, V., Arnanda, J., Tahiri, M., Coudray, C., Baratt.F.,Bouteloipdemange, C., Brown, F., Rayssiguier, Y. and Roussel, A. M. (2005). Influence of Short- Chain Fracto Oligosaccharides on Absorption of Cu, Zn and Se in Healthy Postmenopausal Women.*J. Am. Coll. Nutr.***24**:30-37.
- Etcheverry, P., Grusak, M.A., Fleige, L.E. (2012). Application of in vitro bioaccessibility andbioavailability methods for calcium, carotenoids, folate, iron, magnesium, polyphenols, zinc, and vitamins B6, B12, D, and E. *Front Physiol.* **3**:1-21.
- European Commission. (2003). *Opinion of the Scientific Committee on Food on the Tolerable Upper Intake Level of Trivalent Chromium* (expressed on 4 April 2003)<http://ec.europa.eu/food/fs/sc/scf/out197-en.pdf> (access: 2010.04.29).
- Feng, W., Li, B., Liu, J., Chai, Z., Zhang, P., Gao, Y. (2003). Study of chromium-containing proteins in sub cellular fractions of rat liver by enriched stable isotopic tracer technique and gel filtration chromatography. *Anal Bioanal Chem.* **375**(3): 363-368.
- Fernandez Garcia, E., Carvajal-Lérida, I., Perez Galvez, A. (2009).In vitro bioaccessibility assessment as a prediction tool of nutrient efficiency.*Nutr Res.* **29**:751-760.
- Finley, J. W. (1999). The Retention and Distribution by Healthy Young Men of Stable Isotopes of Selenium Consumed as Selenite, Selenate or Hydroponically, Brown Broccoli Dependent on the Isotopic Form. *J. Nutr.***129**:865- 871.
- Ganesan, K., Xu, B. (2018).Anti-obesity effects of medicinal and edible mushrooms.*Molecules.***23**:2880.
- Gao, Y., Walder, K., Sunderland, T., Kantham, L., Feng, H.C., Quick, M., Bishara, N., de Silva, A., Augert, G., Tenne-Brown, J., Collier, G.R. (2003). Elevation in Tanis expression alters glucose metabolism andinsulin sensitivity in H4IIE cells. *Diabetes.* **52**: 929-934. [PubMed: 12663463]
- Gasecka, M., Mleczek, M., Siwulski, M. (2016). Phenolic composition and antioxidant propertiesof *Pleurotusostreatus* and *Pleurotuseryngii* enriched with selenium and zinc.*Eur Food Res Technol.***242**(5):723-732.

Gosh, N., Chakravarty, D.K. (1990). Predictive analysis of the protein quality of *Pleurotus citrinopileatus*. *J Food Sci Tech.* **27(4)**:236-238.

Guerra, A., Etienne-Mesmin, L., Livrelli, V. (2012). Relevance and challenges in modeling human gastric and small intestinal digestion. *Trends Biotechnology.* **30**:591-600.

Heaney, R.P. (2001). Factors influencing the measurement of bioavailability taking calcium as a model. *J Nutri.* **131**:1344-8.

Hill, C.H. (1976). Mineral interrelationships. Trace Elements in Human Health and Disease. Academic Press. New York. p.281-300.

Ho, L.H., Zulkifli, N. A, and Tan, T.C. (2020). Edible mushrooms: nutritional properties potential nutraceutical values in food product development. An introduction to mushroom. London: In tech Open. P19-36.

Hulamoja, T. and Mykkänen, H. M. (1986). Intestinal Absorption of <sup>75</sup>Se-Labeled Sodium Selenite and Selenomethionine in Chicks: Effects of Time Segment Selenium Concentration and Method of Measurement. *J. Nutr.* **116**:142-148.

Hur, S. J., Lim, B. O., Decker, E. A., McClements, D. J. (2011). *In vitro* human digestion models for food applications. *Food Chemistry.* **125(1)**:1-12.

Ijeh, I., Okwujiako, I. A., Nwosu, P.C. (2009). Phytochemical composition of *Pleurotus tuber regium* and effect of its dietary incorporation on body weights and serum triacylglycerols in albino mice. *J Med Plants Res.* **3 (11)**:939-943.

Jegadeesh, R., Lakshmanan, H., Kabyeul, J. (2018). Cultivation of pink oyster mushroom *Pleurotus djamor* var. *roseus* on various agro-residues by low cost technique. *J Mycopathol Res.* **56(3)**:213-220.

Kalac, P and Svoboda, L. (2000). A review of trace element concentrations in edible mushrooms. *Food Chem.* **69**: 273-281.

Kalac, P. (2013). A review of chemical composition and nutritional value of wild growing and cultivated mushrooms. *J Sci Food Agric.* **93(2)**:20-218.

Kortei, N.K., Wiafe- Kwagyan, M. (2015). Comparative appraisal of the total phenolic content, flavonoids, free radical scavenging activity and nutritional qualities of *Pleurotus ostreatus* (EM-1) and *Pleurotus ostreatus* (P-31) cultivated on rice (*Oryza sativa*) straw in Ghana. *J Adv Biol Biotechnol.* **3 (4)**:153-164.

Kustin, K., Macara, I.G. (1982). The new biochemistry of vanadium. *Comments Inorg Chem.* **2**:1

Li, H., Tian, Y., Menolli, J., Ye, L., Karunarathna, S.C., Perez-moreno, J., Rahmn, M.M., Rashid, M.H., Phengsintham, P., Rizal, L., Kasuya, T. (2021). Reviewing the world's edible mushroom species: a new evidence-based classification system. *Compre Rev Food Sci.* **20(2)**:1982-2014.

Lusi, P., Nyambaka, H., Mbakaya, C.F., Masetta, E., Bwete, V., Murungi, J. (2013). Bioavailability studies of trace elements in a potential food formulation for use in the management of HIV and AIDS. *International Journal of Pure and Applied Chemistry*.**8(1)**: 47-53.

Mattila, P., Konko, K., Eurola, M., Pihlawa, J.M., Astola, J., VahteristoLietaniemi, V., Kumpulainen, J., Valtonen, M., Piironen, V. (2001). Contents of vitamins, mineral elements, and some phenolic compounds in cultivated mushrooms. *J. Agric. Food Chem.***49**: 2343-2348.

Miller J.C., Miller, J.N. (1988). *Statistic for Analytical chemistry*. 2<sup>nd</sup> edition, Ellis Horwood limited publisher. Chichester.

Munoz, A. H. S., Kubachka, K., Wrobel, K., Corona, J. F. G., Yathavakilla, S. K. V., Caruso, J. (2006). Se-enriched mycelia of *Pleurotus ostreatus*: Distribution of selenium in cell walls and cell membranes/cytosol. *Journal of Agricultural and Food Chemistry*.**54**: 3440-3444.

Murugesan, S. (2017). *Sustainable food security-edible and medicinal mushroom. Sustainable Agriculture towards food security*. Singapore. Springer. p185-196.

Mutanen, M. (1986). Bioavailability of selenium in mushrooms, *Boletus edulis*, to young women. *International Journal for Vitamin and Nutrition Research*.**56**:297-301.

Nambafu, R., Swaleh, S., Nyambaka, H. (2021). Bioavailability Studies of Vitamin A and E in Indigenous Vegetables and their Potential Use in the Management of HIV and AIDS. *Advances in Research*.**22 (2)**: 36-44.

Nawiri, M.P., Nyambaka, H.N., Murungi, J.I. (2013). Sundried cowpeas and amaranthus leaves recipe improves beta-carotene and retinol in serum and hemoglobin concentration among preschool children. *European Journal of Nutrition*. **52 (2)**:582-589

Nielsen, F.H. (1988). *Trace Minerals in foods*. Marcel Dekker. New York.357-428

Nielsen, F.H., Uthus, E.O. (1990). *Vanadium in biological systems. Physiology and biochemistry*. Kluwer. Academic, London.51-62.

Offenbacher, E.G. (1994). Promotion of chromium absorption by ascorbic acid. *Trace Elem. Elect.***11**: 178-181.

Ohh, S.J., Lee, J.Y. (2005). Dietary chromium-methionine chelate supplementation and animal performance. *Asian-Aust. J Anim Sci*. **18(6)**: 898-907.

Onyambu, Z.M., Nawiri, M.P., Nyambaka H.N., Noah N.M. (2021). In Vitro Bioaccessibility of the Vitamin B Series from Thermally Processed Leafy African Indigenous Vegetables *Journal of Food Quality* Vol. 1, pp 1-8. <https://doi.org/10.1155/2021/5540724>

Patil, S.S., Ahmed, S.A., Telong S. M., Baig, M.M. V. (2010). The nutritive values of *pleurotus ostreatus* cultivars of different agro wastes. *Innovative Romanian Food Biotechnology*.**7**: 66-76.

Patterson, B.W., Hansard II, C.B., Ammerman, R., Henry, L.A., Zech, Fisher, W.R. (1986). *Am. J. Physiol.* **251**:325-332.

Rajarithnam, S and Shashirekha, M. N. (2011). "Mushroom nutraceuticals," in *Advances in preservation and processing technologies of fruits and vegetables*. New India Publishing Agency, New Delhi, India.

Rayman, M. P., Infante, H. G., Sargent, M. (2008). Food-chain selenium and human health: Spotlight on speciation. *British Journal of Nutrition.* **100**:238-253.

Rudawska, M and Leski, T. (2005). Macro and micro elemental contents in fruiting bodies of wild mushrooms from the Netecka forest in west - central Poland. *Food Chem.* **92**: 499-502.

Samanta, S., Haldar, S., Ghosh, T.K. (2008). Production and carcass traits in broiler chickens given diets supplemented with inorganic trivalent chromium and an organic acid blend. *Br Poultry Sci.* **49**(2): 155-163.

Samsudin, N. P and Abdullah, N. (2019). Edible mushrooms from Malaysia: a literature review on their nutritional and medicinal properties. *Intl Food Res J.* **26**(1):11-31.

Shamberger, R.J. (1996). The insulin-like effects of vanadium. *Journal Advance Medicine* **9**: 121-131.

Shechter, Y. (1990). Insulin-mimetic effects of vanadate. Possible implications for future treatment of diabetes. *Diabetes.* **39**: 1-5.

Silva, S.O., Costa, S.M.G., Clemente, E. (2002). Chemical composition of *Pleurotus pulmonarius*, substrates and residue after cultivation. *Braz Arch Biol Technol.* **45**(4):531-535.

Suzuki, K.T., Doi, C., Suzuki, N. (2006). Metabolism of <sup>76</sup>Se-methylselenocysteine compared with that of <sup>77</sup>Se-selenomethionine and <sup>82</sup>Se-selenite. *Toxicol Applied Pharmacol.* **217**:185-195.

Tang, C., Hoo, P.C., Tan, L.T. (2006). Golden needle mushroom: a culinary medicine with evidenced-based biological activities and health promoting properties. *Front Pharmacol.* **7**:474.

Thompson, C. D., Burton, C. E. and Robinson, M. F. (1978). On Supplementing the Selenium Intake of New Zealander. Short Experiments with Large Doses of Selenite or Selenomethionine. *Br. J. Nutr.*, **39**:587- 597.

Thomson, C.D. (1998). Selenium speciation in human body fluids. *Analyst.* **123**: 827-831.

Wang, M.Q., He, Y.D., Lindemann, M.D., Jiang, Z.G. (2009). Efficacy of Cr (III) supplementation on growth, carcass composition, blood metabolites, and endocrine parameters in finishing pigs. *Asian-Aust. J Anim Sci.* **22**(10): 1414-1419.

Whanger, P.D. (2002). Selenocompounds in plants and animals and their biological significance. *J Am Col Nutri.* **21**:223-252.

Whanger, P.D. (2004). Selenium and its relationship to cancer: an update. *Br J Nutri.* **91**:1-28.

WHO. (2012). Guideline: sodium intake for adults and children. Geneva: World Health Organization.

Wienk, K., Marx, J., Beynen, A.C. (1999). The concept of iron bioavailability and its assessment. *Eur J Nutr.* **38**(4):51-75.

Yang, J.H., Lin, H.C., Mau, J.L. (2001). Non-volatile taste components of several commercial mushrooms. *Food Chem.* **72**(4):465-471.

Yoneda, S., Suzuki, K.T.(1997). Detoxification of mercury by selenium by binding of equimolar Hg-Se complex to a specific plasma protein. *Toxicol Applied Pharmacol.***143**:274-280.

Zha, L.Y., Xu, Z.R., Wang, M.Q., Gu, L.Y.(2007). Effects of chromium nanoparticle dosage on growth, body composition, serum hormones and tissue chromium in Sprague-Dawley rats. *J Zhejiang UnivSci B.* **8**(5): 323-330.