

EFFECT OF CO-ADMINISTRATION OF VITAMIN A AND ARTESUNATE IN MALARIA-INFECTED ALBINO MICE

ABSTRACT

Malaria is a potentially fatal illness that results from infection by parasitic protozoa, which are spread to humans via the bites of mosquitoes that have been infected with the parasite. This research examines the impact of the simultaneous administration of Vitamin A and Artesunate on *Plasmodium berghei*-infected albino mice. A cohort of sixteen (16) albino mice, with a mean weight of 28g and indeterminate gender, were subjected to *p.berghei* inoculation. The mice were subsequently separated into four groups, each consisting of four animals. Group A received an artesunate treatment, Group B received distilled water, Group C received vitamin A as an antioxidant, and Group D received an artesunate and vitamin A combo treatment. Following the administration of several therapies, including an antimalarial medicine (Artesunate), distilled water, Vitamin A, and a combination therapy of both Artesunate and Vitamin A, the study examined the parasitemia level, hematocrit (PVC), percentage mortality rate, and change in body weight. The effectiveness of Artesunate in treating malaria infection may be reduced when vitamin A and Artesunate are used together. The research illustrates the intricate relationship between Artesunate and antioxidant (vitamin A) in mice infected with malaria. The efficacy of Artesunate in clearing the parasite was demonstrated by the reduction of Parasitaemia levels from +2(++) on the 5th day to +3(+++) on the 7th day, followed by a decrease to +2(++) on the 9th day, and ultimately to +1(+) Parasitaemia on the 10th day. The co-administration of Vitamin A and Artesunate resulted in a reduction of the latter's efficacy in treating malaria, particularly in cases where the parasitemia level was at +3(+++) on the 5th, 7th, and 11th days and +2(++) on the 9th day. This suggests that Vitamin A acted as an antagonist to the antimalarial effects of Artesunate.

Furthermore, the mortality rate was 50%, and the PCV was reduced compared to Group A. Artesunate, a pro-oxidant that generates free oxygen radicals that act against the parasite. However, the presence of antioxidants like Vitamin A potentially counteracted the effects of pro-oxidants, indicating some form of pharmacological antagonism. The research findings indicate that the co-administration of vitamin A and Artesunate may reduce therapeutic effectiveness. It is recommended that clinical interventions be implemented to regulate the simultaneous administration of Artesunate and Vitamin A, as this may impact the therapeutic objective of malaria treatment.

Keywords: *Plasmodium berghei, Parasitaemia, Artesunate, Vitamin A, Mice.*

INTRODUCTION

Despite the significant efforts invested in controlling *Plasmodium* spp. in endemic countries,

malaria remains a disease of public health significance, as per the World Health Organization's

report in 2014. The onset of the COVID-19 pandemic in early 2020 has presented a complex challenge to the fight against infectious diseases in numerous African nations, where public health services were already grappling with significant obstacles (Monroe et al., 2022). Significant advancements have been made on a global scale in the reduction of malaria cases in Africa. The epidemiology of malaria infection has persisted or even escalated in some areas of the African continent, necessitating immediate inquiry and measures to reinforce its management, as per the World Malaria Report of 2021. According to a recent malaria report by the World Health Organization (WHO), estimated 240 million cases of malaria and 620,000 deaths were attributed to the disease globally in 2021 (Monroe et al., 2022). The present dataset indicates a rise of approximately 14 million instances and 69,000 fatalities compared to the previous year, 2020. According to Monroe et al. (2022), the COVID-19 pandemic has impaired health systems, which may have contributed to a 10-case increase in incidence per population of 1000 at risk in 2020.

The etiology of severe malaria in humans is predominantly attributed to *Plasmodium Falciparum*. In contrast, other species of *Plasmodium*, such as *P. vivax*, *P. ovale*, and *P. malaria*, are also known to cause the disease. The presence of human *Plasmodium* among domestic animals may account for malaria's notable prevalence and persistence in certain endemic regions, as reservoir

hosts can exhibit varying presentation patterns (Munirah et al., 2023). The successful acclimatization of *Plasmodium berghei*, a malaria-causing parasite, to various rodent hosts has been established as a reliable model for studying the disease (Joachim & Taco, 2015).

Of the various *Plasmodium* species, *P. falciparum* is distinguished by its ability to infiltrate all red blood cells and induce multiple infections within a single cell. This characteristic leads to rapid proliferation and exacerbation of the illness. Red blood cells undergo structural and functional alterations, including sequestration, inflammation, and endothelial dysfunction, which are associated with the development of severe malaria. The study was conducted by Gao and colleagues in 2019.

The World Health Organization has suggested the utilization of artemisinin-based combination therapy as a means to impede or postpone the emergence of resistance. Therefore, the acquisition of novel medications necessitates the utilization of Artemisinin as the most extensively employed framework for the synthesis of artificial compounds. The present study provides an overview of artemisinin-based drugs and their derivatives, encompassing hybrid derivatives and dimers, trimers, and tetramers containing an endoperoxide bridge (Luiz et al., 2018). These drugs have demonstrated efficacy against *P. falciparum* strains that have developed resistance to other antimalarial agents. However, they are ineffective in treating malaria caused by *P. vivax*, *P.*

ovale, and *P. malaria*. The cleavage of the endoperoxide bond at the infected cell, which generates radical intermediates and leads to subsequent chemical rearrangements, is a crucial factor in the antimalarial activities of Artemisinin, as noted by Shikha et al. in their recent study (2022).

In recent decades, research studies have demonstrated the efficacy of Artesunate against malaria as well as its potential anticancer effects on different tumor cell lines, both *in vitro* and *in vivo*, as reported by Zhang D. et al. in 2017. Furthermore, Artesunate is significant in treating lupus, as indicated by Ismail M., Du Y., Ling L., and Li X. in 2019. Additionally, Shenoy R.K. et al. in 2019 and Kumaran U. et al. in 2021 have reported the efficacy of Artesunate against several viruses. Moreover, Artesunate exhibits anti-diabetic properties, as Li Z. et al. reported in 2019.

The anti-tumor properties of Artesunate have been observed to induce non-apoptotic cell death through various mechanisms, including autophagy, oncosis, and ferroptosis. Several studies have reported these findings (Zhu W. et al., 2019; Ooko E. et al., 2015; Shi X. et al., 2017; Jiang F. et al., 2018). The compound, known as Artesunate, has been observed to impact various critical processes in cancer development. This includes inhibiting cancer cell proliferation and invasion, induction of cell cycle arrest, disruption of cancer signaling pathways, induction of oxidative damage, and stimulation of cell apoptosis. The compound in question exerts its effects through

the inhibition of angiogenesis and as a metastasis agent, as reported by Xu et al. in 2020 and Zhao et al. in 2013.

Retinyl esters are the primary dietary source of vitamin A, a crucial micro-essential nutrient. Furthermore, β -carotene, a pro-vitamin A, is a precursor to vitamin A in green and yellow vegetables. Upon ingestion, it converts to retinol within the body (Noriko et al., 2022).

The essentiality of Vitamin A lies in its role in promoting skeletal growth, facilitating vision, enabling cell differentiation, and supporting reproduction. In comparison, some research studies have indicated the potential benefits of Vitamin A in disease prevention. The co-administration of Artemisinin with certain nutritional supplements, particularly Vitamin A, may impact its efficacy in treating malaria infection.

MATERIALS AND METHODS

Acquisition of parasite: The parasite was obtained from the Research Laboratory of the College of Medicine, University of Lagos, and was authenticated by a microbiologist in Nigeria.

Acquisition of drugs: The drugs used (Artesunate and Vitamin A) were obtained and authenticated by a University chemist in Nigeria.

Preparation of drugs: The powdered drug of Artesunate was diluted with distilled water to a level of 160mg/80mls which was kept as the stock. One millimeter (1ml) of the stock was later diluted with 9mls of distilled water making the concentration 0.2mg/ml.

Parasite Inoculation: The mice were infected with parasites from parasitized blood by cardiac puncture of an infected mouse. 0.2ml of infected blood was diluted in 9.8mls of normal saline. The mice were inoculated on day 1 intraperitoneally with 0.2ml parasitized saline suspension. Development of Parasitemia was monitored by microscopic examination until day 5 when it was established.

Experimental Animals: Before the start of the trials, sixteen (16) mice were kept in the Animal House of the College of Medicine at Ambrose Ali University in Ekpoma, Edo State, and given two weeks to acclimate. All of the animals received unlimited access to food and water ad libitum. The mice were placed into four (4) treatment groups, each with four (4) mice, and from day one (4) on, they received an intraperitoneal injection with 0.2 ml of a parasitized saline suspension. The microscopic inspection was used to track the progression of parasitemia until day 5 when it became entrenched. Both before and after the experiment, the experimental mice were weighed.

METHODOLOGY

Experimental Design

A total number of 16 mice (male and female) were distributed randomly into **four** groups (four mice/group). The experiment lasted for a period of ten days during which the inoculated parasitized mice were grouped as follows:

Group A (Artesunate control group) received Artesunate at 0.7mls/kg for day 6 and 0.35mls from day 7 to day 10.

Group B (The negative control group) received 0.5ml/kg of distilled water daily from day 6 to day 10.

Group C (The positive control group) received 0.03ml/kg of Vitamin A daily from day 6 to day 10.

Group D (Artesunate and Vitamin A control group) received 0.03ml/kg of Vitamin A and Artesunate at 0.35mls/kg from day 6 to day 10.

Animals Sacrifice:

At the end of the experiment (day 11), all the animals were humanely sacrificed. Blood was collected through the tail vein puncture of the animals in a heparinized centrifuge tube under deep anesthesia with chloroform. The blood collected was centrifuged using a centrifuge machine at 10,000 rpm for five minutes and the serum collected was subjected to packed Cell volume analysis and the assessment of Malaria Parasite count.

Assessment of Malaria Parasite Count:

A semi-quantitative parasite count was done through the tail vein puncture, a drop of blood was collected on a clean slide to get a thick film and allowed to dry and a drop of gemisa stain which was diluted with distilled water at a ratio of 1:10 was dropped on the smear. The smear was left for 5 minutes which enabled the blood cells and the malaria parasite to be viewable under the microscope and later flooded with distilled water and allowed to dry, a drop of emulsion oil was dropped on the smear and the smear was viewed under the microscope and parasite seen was recorded.

Packed Cell Volume:

This procedure was done by collecting blood from the tail vein into heparinized microhematocrit tubes to 1cm from one end of the tube and the other end of the tube was sealed with a flame. The preparation is centrifuged in a timed micro hematocrit centrifuge for 5 minutes. With a special microhematocrit reader employed for this purpose, the ratio of the Packed Cell to the volume of blood in the heparinized tube can be read and converted to the percentage of the Packed Cell to the whole blood.

RESULTS

Parasite Level:

The clearance level of parasitemia was measured on days 5, 7, 9, and 11 of post-inoculation. According to Table 1, **Group A** had a clearance level of +2, which rose to +3 on day seven (7) before falling to +2 and then to +1. This could be related to the administration of artesunate.

Compared to Group A, C, and D without any medications preventing its growth, **Group B's** clearance level for days 5 and 7 was +3, and on days 9 and 11, it climbed to +4, which could be linked to the parasitemia growth.

On day 5, the clearance level for **Group C** was +2; on days 7, 9, and 11, it increased to +3, which may indicate that vitamin A had no effect in halting the already present parasitemia. Instead, the parasitemia continued to spread, albeit more slowly than in Group B.

On days 5 and 7, the **Group D** clearance level was +3, which was average and did not increase or drop, but on day 9, it was +2, which was a slight decline and likely caused by the presence of artesunate. A possible explanation for the +3 clearance level on day 11 is the antagonistic impact of vitamin A on artesunate.

Table 1: Effect of co-administration of Vitamin A and Artesunate on parasite level in *P.berghei* infected mice.

GROUP	5 TH DAY	7 TH DAY	9 TH DAY	11 TH DAY
A	++	+++	++	+
B	+++	+++	++++	++++
C	++	+++	+++	+++
D	+++	+++	++	+++

n=4(mean rounded to nearest whole number)

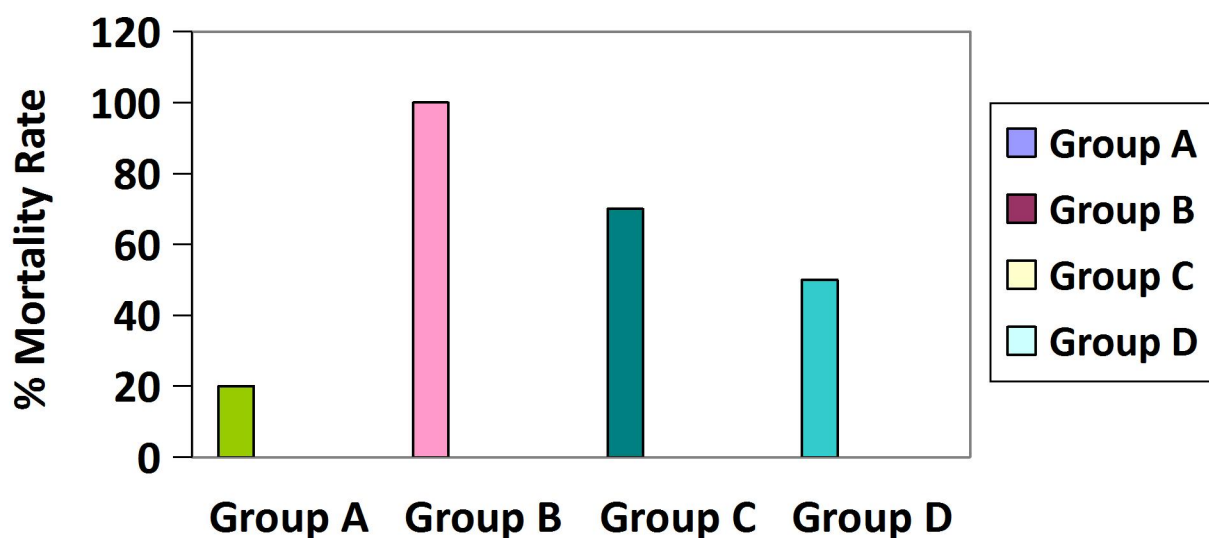


Fig. 1: Percentage (%) mortality of co-administration of Vitamin A and Artesunate in *P. berghei* infected mice.

Where: Group A = Artesunate + Parasite

Group B = Distilled water + Parasite

Group C = Vitamin A + Parasite

Group D = Vitamin A + Artesunate + Parasite

Packed cell volume:

In **Group A**, the PVC level was high on Day 1 of the pre-inoculation period. On Day 5, there was a large decline, and on Day 11, there was a tiny increase in the PVC, but not as much as on Day 1. This might be explained by the administration of Artesunate raising the PCV level.

When compared to Group A, **Group B** experienced a significant decline in PVC levels on day 5 and a lesser decline in PCV levels on day 11—days when the mice had lost more PCV—which may have been caused by the absence of administration following the mice's attack of parasitemia.

Although **Group C's** PCV is a little higher than Group B's but less than Group A's, there was a significant decrease in PVC level on day 5 and a decrease in PCV level on day 11, which was less pronounced than day 5, which showed more loss of PCV in the mice. This could be because Vitamin A had no effect on the parasitemia.

Group D experienced a large drop in PVC levels on day 5 and a lesser drop in PCV levels on day 11 than on day 5, when more PCV was lost in the mice. This could be the result of Vitamin A's antagonistic impact on artesunate. Group D's PCV level is marginally greater than Group B's, but much lower than Groups A and C's.

Table 2: Packed Cell Volume for *P.berghei*-infected mice.

Group	1 st Day (Pre-inoculation)	5 th Day (Post-Inoculation)	11 th Day (Post-Inoculation)
A	52.25±2.63	31.25±1.26	42.00±1.83
B	53.25±2.22	28.75±2.22	18.00±1.41
C	50.75±0.96	25.00±0.82	21.00±0.96
D	51.00±1.41	23.75±0.82	19.50±2.63

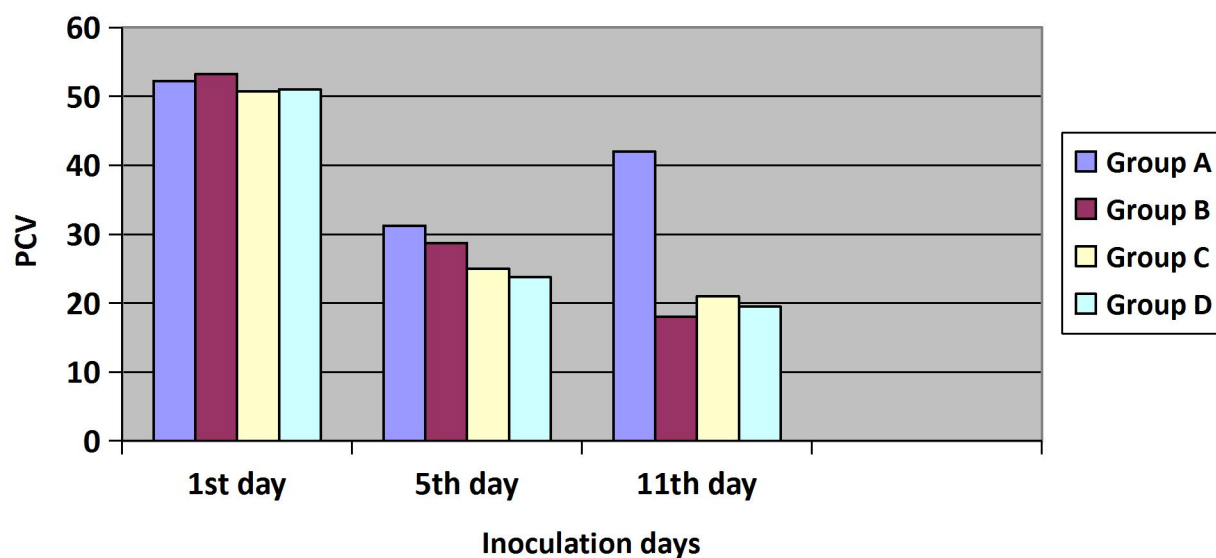


Fig. 2: Showing the Packed Cell Volume for the pre-inoculation day (1st day) and post-inoculation days (5th, 10th day) for *P.berghei* infected mice.

Where: Group A = Artesunate + Parasite

Group B = Distilled water + Parasite

Group C = Vitamin A + Parasite

Group D = Vitamin A + Artesunate + Parasite

Body weight:

During the period of pre-inoculation of the Parasitaemia of *P. berghei*-infected mice, all the experimental animals were observed to have normal physical activity. Weight changes were observed in the experimental animals, day 1 before Inoculation, and day 11 Post-Inoculation after administration of the experiment, with their weight differences. During the period of administration, **Group A** had a slight decrease in physical activities and ate a bit less than normal. There was a significant decrease in their weight when compared to the mean weight at pre-inoculation. **Group B** has the least weight when compared to other group and it could be a result

of the parasitemia, **Group C and D** has a slight decrease in weight but not as severe as **Group B** and not better than **Group A** and it could be due to the administration administered.

Table 3: Weight for pre and post-inoculation for *P. berghei* infected mice.

Group	Mean ± standard deviation
All groups: Pre inoculation	28.00±1.71
Group A: Post Inoculation	26.25±1.23
Group B: Post Inoculation	15.25±0.50
Group C: Post Inoculation	19.50±0.58
Group D: Post Inoculation	17.25±1.50

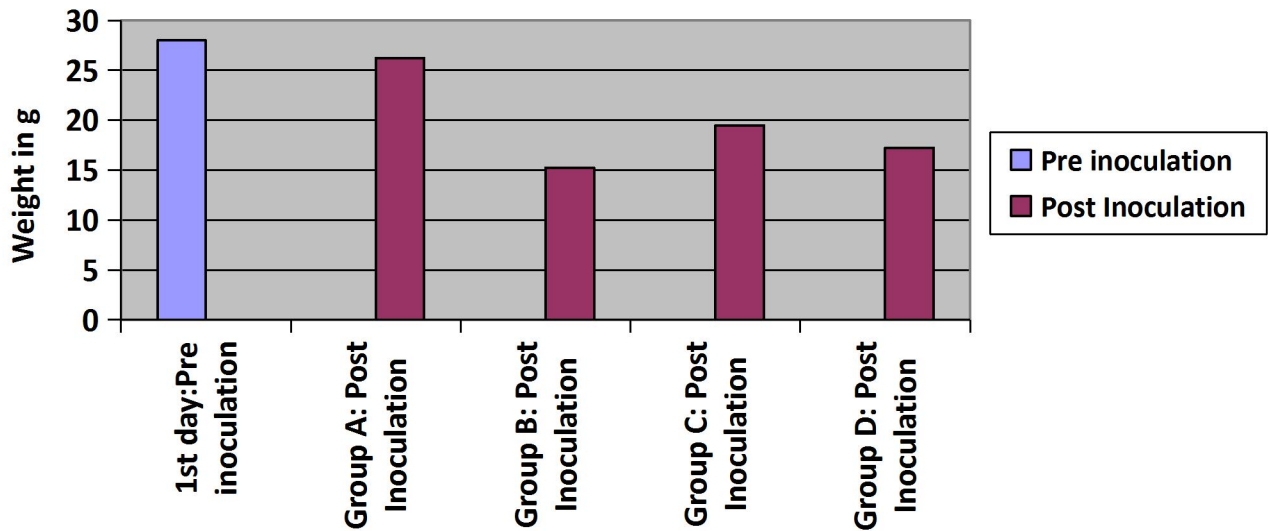


Fig. 3: Weight for pre and post-inoculation for *P.berghei*-infected mice.

Where: Group A = Artesunate + Parasite

Group B = Distilled water + Parasite

Group C = Vitamin A + Parasite

Group D = Vitamin A + Artesunate + Parasite

The above results show that Group A had the lowest Parasitaemia level, percentage mortality rate, increased weight, and PVC from the treatment of Artesunate as compared to the responses from the treatment of other Groups.

DISCUSSION

The findings of this investigation illustrate the intricate interaction between Artesunate and the antioxidant Vitamin A in mice infected with malaria. The study results indicate that the Parasitaemia level was significantly elevated as early as the 5th day following inoculation of the mice. The Parasitaemia level was predominantly within the range of +3(+++) prior to administering the drug. The life span of the mice was determined to be between the 6th and 11th-day post-inoculation.

The drug was administered to the mice on the 6th day following confirmation of parasitemia in this particular study. Upon administration of Artesunate and Artesunate in combination with Vitamin C to mice, parasitemia exhibited a decline and subsequent fluctuations across multiple groups beginning on day 5. As an established antimalarial drug with proven and validated efficacy, the study's findings were anticipated.

The parasitemia level of Group A observed to be +2 (++) on the 5th day, which gradually decreased to +1 (+) on the 11th day following the initiation of treatment with Artesunate. According to a paper by Atanu FO et al. in 2021, which showed the effectiveness of artemisinin derivatives as effective antimalarial medicines, it was linked to a lower mortality rate. According to Dibessa TT et al., in 2020, a slight decrease in the mice's body weight was noticed, which may have been caused by the presence of paracetamol in their systems. It has been noted that the administration of Artesunate improves the host's immunological response against the parasite. It is thought that the production of free oxygen radicals in vivo is how this action is mediated. Figure 3's representation of our results shows that animals treated with Artesunate had significantly higher body weights than those in group D. Additionally, we saw that these animals had an increase in Packed Cell Volume. Due to the administration of Artesunate, a noticeable

decrease in packed cell volume was seen on the fifth day, followed by a modest increase on the eleventh day. Severe malaria anemia (SMA) is a condition that often leads to a significant reduction in hemoglobin levels, which can be life-threatening. Artesunate as a sole treatment for SMA may exacerbate this condition, as it does not promote an increase in hemoglobin levels and may even cause a mild reduction. The studies conducted by Jauréguiberry and colleagues in 2014 and White and colleagues in 2018 are of relevance to the field of medical research.

On the 11th day, Group B exhibited a significantly elevated level of Parasitaemia ranging from +3(+++) to +4(++++) and experienced mortality, potentially attributable to oxidative stress induced by the parasite. A marked reduction in Packed Cell Volume was also observed, which may indicate hemolysis and resultant anemia. The study was conducted by Lopes LN and colleagues in 2016. The observed symptoms included a significant decrease in body weight, suggesting a substantial impact on the animals. This ultimately resulted in a mortality rate of 100%, leading to the mice's demise.

The study observed that Group C elevated Parasitaemia level from +2(++) on the 5th day to +3(+++) on the 11th-day post-administration of Vitamin A. Additionally, the group displayed a high Packed Cell Volume on the 1st day of pre-inoculation, which subsequently decreased on the 11th day. This suggests that the antioxidant effect of Vitamin A on malaria oxidative stress may not be discernible during the infection. It is important to note that Vitamin A is not an antimalarial drug, and the study conducted by Chen G et al. in 2019 reported a mortality rate of 75%. According to Jørgensen MJ et al., 2011, it was observed that the body weight of the animals exhibited a significant decrease. This decrease in body weight may be attributed to the fact that vitamin A, a non-antimalarial drug, does not bring about any changes in this regard.

The findings indicate that Group D experienced a modest decrease in Parasitaemia level, from +3(+++) on the 5th day to +2(++) on the 9th day, followed by an increase to +3(+++) on the 11th day. These results suggest that Vitamin A may have counteracted the antimalarial properties of Artesunate, resulting in a 50% mortality rate and decreased PCV compared to Group A (refer to Figure 1). According to Shija KM et al.'s research conducted in 2020, it has been proposed that Artesunate, a pro-oxidant, exhibits antiparasitic activity by producing free oxygen radicals. The

study also suggests that Vitamin A, an antioxidant, may counteract the effects of pro-oxidants, indicating the presence of pharmacological antagonism.

The comparative analysis indicates that the efficacy of Artesunate in combating the malaria parasite is superior in group A compared to the combined administration of Artesunate and Vitamin A in group D.

CONCLUSION

The study demonstrated that Vitamin A can reduce the efficacy of Artesunate when used in combination. Therefore, clinical efforts should be made to control the concurrent use of Artesunate and Vitamin A as this may affect the treatment goal in malaria therapy.

REFERENCE:

Atanu FO, Idih FM, Nwonuma CO, Hetta HF, Alamery S, El-Saber Batiha G. Evaluation of Antimalarial Potential of Extracts from *Alstonia boonei* and *Carica papaya* in *Plasmodium berghei*-Infected Mice. *Evid Based Complement Alternat Med*. 2021 Oct 6;2021:2599191. doi: 10.1155/2021/2599191. PMID:

Cai L., Tang H., Zhou M., Ding Y., Li X., Shi Z. Artesunate Attenuated the Progression of Abdominal Aortic Aneurysm in a Mouse Model. *J. Surg. Res*. 2021;267:404–413. doi: 10.1016/j.jss.2021.05.001.

Chen G, Du YT, Liu JH, Li Y, Zheng L, Qin XS, Cao YM. Modulation of anti-malaria immunity by vitamin A in C57BL/6J mice infected with heterogenic plasmodium. *Int Immunopharmacol*. 2019 Nov;76:105882. doi: 10.1016/j.intimp.2019.105882. Epub 2019 Sep 11. PMID: 31520991.

Dibessa TT, Engidawork E, Nedi T, Teklehaymanot T. Antimalarial activity of the aqueous extract of the latex of *Aloe pirottae* Berger. (*Aloaceae*) against *Plasmodium berghei* in mice. *J Ethnopharmacol*. 2020 Jun 12;255:112763. doi: 10.1016/j.jep.2020.112763. Epub 2020

Gao F., Zuo Q., Jiang T., Song H., Zhou J. A Newly Synthesized Oleanolic Acid Derivative Inhibits the Growth of Osteosarcoma Cells in Vitro and in Vivo by Decreasing C-MYC-Dependent Glycolysis. *J. Cell. Biochem.* 2019;120:9264–9276. doi: 10.1002/jcb.28202.

Ismail M., Du Y., Ling L., Li X. Artesunate-Heparin Conjugate Based Nanocapsules with Improved Pharmacokinetics to Combat Malaria. *Int. J. Pharm.* 2019:162–171. doi: 10.1016/j.ijpharm.2019.03.031.

Jauréguiberry S., Ndour P.A., Roussel C., Ader F., Safeukui I., Nguyen M., Biligui S., Ciceron L., Mouri O., Kendjo E., et al. Post-artesunate Delayed Hemolysis Is a Predictable Event Related to the Lifesaving Effect of Artemisinins. *Blood.* 2014;124:167–175. doi: 10.1182/blood-2014-02-555953.

Jiang F., Zhou J.Y., Zhang D., Liu M.H., Chen Y.G. Artesunate induces apoptosis and autophagy in HCT116 colon cancer cells, and autophagy inhibition enhances the artesunate-induced apoptosis. *Int. J. Mol. Med.* 2018;42:1295–1304.

Joachim MM, Taco WK. Towards genome-wide experimental genetics in the *in vivo* malaria model parasite *Plasmodium berghei*. *Pages* 46-60 | Published online: 19 Mar 2015.

Jørgensen MJ, Hein-Kristensen L, Hempel C, Ravn H, Wiese L, Kurtzhals JA, Benn CS. The effect of vitamin A supplementation and diphtheria-tetanus-pertussis vaccination on parasitaemia in an experimental murine malaria model. *Scand J Infect Dis.* 2011 Apr;43(4):296-303. doi: 10.3109/00365548.2010.535845. Epub 2010 Nov 25. PMID: 21105844.

Kumaran U., Gaonkar S., Chaudhuri M., Sheriff A.K., Rao N., Raja M., Shenoi A. Chromosomally Integrated Human Herpes Virus 6A-Associated Myocarditis in a Neonate Treated with Artesunate. *J. Paediatr. Child Health.* 2021;1:6–8. doi: 10.1111/jpc.15391.

Li Z., Shi X., Liu J., Shao F., Huang G., Zhou Z., Zheng P. Artesunate Prevents Type 1 Diabetes in NOD Mice Mainly by Inducing Protective IL-4—Producing T Cells and Regulatory T Cells. *FASEB J.* 2019;33:8241–8248. doi: 10.1096/fj.201900146R.

Lopes LN, Folha Santos FA, Marques Oliveira LC, Ferreira Araujo MT, Sequeira CG, Libonati RM, Revoredo da Silva Ventura AM. An analysis of the influence of sex hormones on Balb/c

mice infected with *Plasmodium berghei*. *Microb Pathog.* 2016 Jan;90:7-12. doi: 10.1016/j.micpath.2015.10.020. Epub

Luiz CS, PinheiroliviaM., Feitosaflavia F, Da silveiranubia B. Current antimalarial Therapies and Advances in the Development of Semi-Synthetic Artemisinin Derivatives. *Articles • An. Acad. Bras. Ciênc.* 90 (01 Suppl 2) • 2018 • <https://doi.org/10.1590/0001-3765201820170830>

Monroe, A.; Williams, N.A.; Ogoma, S.; Karema, C.; Okumu, F. Reflections on the 2021 World Malaria Report and the future of malaria control. *Malar. J.* 2022, 21, 154. World Malaria Report 2021 [Internet]. Available online: <https://www.who.int/teams/global-malaria-programme/reports/world-malaria-report-2021> (accessed on 13 August 2022).

Munirah M, Wahyuni S, Wahid I and Hamid F. The discovery of human *Plasmodium* among domestic animals in West Sumba and Fakfak, Indonesia [version 2; peer review: 2 not approved]. *F1000Research* 2023

Noriko T, Daisuke S, Shinya H, Masahiro Y, Masahiko I. Vitamin A in health care: suppression of growth and induction of differentiation in cancer cells by vitamin A and its derivatives and their mechanisms of action. *Pharmacology & Therapeutics* 230, 107942, 2022

Ooko E., Saeed M.E., Kadioglu O., Sarvi S., Colak M., Elmasaoudi K., Janah R., Greten H.J., Efferth T. Artemisinin derivatives induce iron-dependent cell death (ferroptosis) in tumor cells. *Phytomedicine.* 2015;22:1045–1054. doi: 10.1016/j.phymed.2015.08.002.

Shenoy R.K., Gokarn A., Toshniwal A., Kalantri S.A., Chichra A., Punatar S., Bonda A., Nayak L., Mathew L.J., Bhat V., et al. Efficacy of Artesunate for Treatment of Cytomegalovirus Reactivations in Allogeneic Haematopoietic Stem Cell Transplant Recipients Who Are Intolerant/Unsuitable for Ganciclovir Therapy. *Blood.* 2019;134:4506. doi: 10.1182/blood-2019-128766.

Shi X., Wang L., Li X., Bai J., Li J., Li S., Wang Z., Zhou M. Dihydroartemisinin induces autophagy-dependent death in human tongue squamous cell carcinoma cells through DNA double-strand break-mediated oxidative stress. *Oncotarget.* 2017;8:45981. doi: 10.18632/oncotarget.17520.

Shija KM, Nondo RSO, Mloka D, Sangeda RZ, Bwire GM. Effects of lemon decoction on malaria parasite clearance and selected hematological parameters in Plasmodium berghei ANKA infected mice. BMC Complement Med Ther. 2020 Jan 30;20(1):24. doi: 10.1186/s12906-020-2820-1. PMID: 32020885; PMCID:

Shikha S, Md Ehesan A. Nonreductive homolytic scission of endoperoxide bond for activation of artemisinin: A parallel mechanism to Heterolytic cleavage, Journal of Physical Organic Chemistry 35 (9), e4392, 2022

White N.J. Anaemia and Malaria. Malar. J. 2018;17:371. doi: 10.1186/s12936-018-2509-9.

World Health Organization. (2014). Malaria: fact sheet (No. WHO-EM/MAC/035/E). World Health Organization. Regional Office for the Eastern Mediterranean. [2] Antinori, S., Galimberti, L., M

Xu C., Zhang H., Mu L., Yang X. Artemisinins as anticancer drugs: Novel therapeutic approaches, molecular mechanisms, and clinical trials. Front. Pharmacol. 2020;11:1608. doi: 10.3389/fphar.2020.529881.

Zhang D., Zhou C., Lv P., Zhao Y., Liang J., Liao X., Yang B. Preparation and Characterization of a Novel Host-Guest Complex Based on Folate-Modified β -Cyclodextrin and Artesunate. Mater. Sci. Eng. C. 2018;86:48–55. doi: 10.1016/j.msec.2017.12.009.

Zhu W., Li Y., Zhao D., Li H., Zhang W., Xu J., Hou J., Feng X., Wang H. Dihydroartemisinin suppresses glycolysis of LNCaP cells by inhibiting PI3K/AKT pathway and downregulating HIF-1 α expression. Life Sci. 2019;233:116730. doi: 10.1016/j.lfs.2019.116730.

