

Analysis The Effect of Different Levels of Inorganic and Bio Fertilizers on Physico - Chemical Properties of Soil in Mung Bean

Abstract

The research conducted at the Soil Science Research Farm of Sam Higginbottom University of Agriculture, Technology and Sciences, Prayagraj, U. P. during the *Summer* season (May to July 2022). The experiment was laid out in randomized block design with nine treatment and three replications with three levels of inorganic fertilizer (0,50 and 100 % NPK), and three level of biofertilizer (0, 50 and 100 % Rhizobium and Azotobacter) that leads to the non-significant findings *i. e.* bulk density, particle density, pH and EC and remaining % pore space, WHC, OC, and NPK were found significantly low to medium range, which comprises yellowish brown colour, sandy loam textured soil and neutral to alkaline soil that is non- saline in nature among all the nine treatments combination applied in treatment T₉ [NPK @ 100 % + Rhizobium@ 100 % + Azotobacter @ 100 %] has shown the best results in improving the soil nutritional status that leads to increased crop yield and also increased morphological parameters as compare with in treatment T₁ [NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %] Application of inorganic and biofertilizer increased improved physical and chemical properties of soil.

Key word: Mung bean, NPK, Rhizobium, Azotobacter, Physio-chemical Properties of Soil, etc.

Introduction

“Soils supply the essential nutrients, water, oxygen and root support that our food-producing plants need to grow and flourish. They also serve as a buffer to protect delicate plant

roots from drastic fluctuations in temperature. A healthy soil is a living, dynamic ecosystem, teeming with microscopic and larger organisms that perform many vital functions including converting dead and decaying matter as well as minerals to plant nutrients (nutrient cycling); controlling plant disease, insect and weed pests; improving soil structure with positive effects for soil water and nutrient holding capacity, and ultimately improving crop production. A healthy soil also contributes to mitigating climate change by maintaining or increasing its carbon content". [16]

Green gram (*Vigna radiata* L.) is one of the important pulse crops in India. Green gram commonly known as "mung bean" has been cultivated in India since ancient times. "It belongs to *Fabaceae* family. Green gram is originated from India and central Asia. It is the third most popular pulse crop cultivated throughout in India" (Miachieo *et al.*, 2019). "Green gram is a protein rich staple food. It has enormous potential for the future needs to be capitalized. It has an edge over other pulses because of its high nutritive value, digestibility and non-flatulent behavior. It is grown principally for protein rich edible seeds which contain 24% crude protein, 56.7% carbohydrates, 1.3% fats, 3.5% minerals, 0.43% lysine, 0.1% methionine and 0.04% tryptophan". (Miachieo *et al.*, 2019)

Green gram also known as moong or mung, is the third most important pulse crop in India after gram and red gram (Miachieo *et al.*, 2019). It belongs to the "Leguminosae" family and sub family "Papillionaceae". Green gram is thought to have originated in India and Central Asia. It extends from India to China, Iraq, Japan, Africa, and other countries. Green gram is primarily growing Rajasthan, Maharashtra, Andhra Pradesh, Orissa, Gujarat, Madhya Pradesh, Punjab, and Utter Pradesh in India.

The inorganic fertilizers, no doubt, are the important source of nutrients in crops which can meet the nutrient requirement but their imbalance and continuous use causes environmental pollution and deterioration of soil health (Zafar *et al.*, 2013). Another issue for the farmer is the availability of fertilizer at reasonable rates. Under these circumstances, farmers should not depend on single source of plant nutrients like inorganic fertilizers. A balanced use of inorganic fertilizers, organic manures and bio-fertilizers are required to develop an integrated plant nutrition supply system.

"Increasing the application of N fertilizer during the early growth period promotes vegetative growth and creates conditions favoring high yield. P fertilizer promotes root growth, disease resistance, drought tolerance, and enhances nutrient and water absorption in

the seedlings after they have depleted their endosperm reserves. K fertilizer improves sugar metabolism, enhances osmotic cell concentration, maintains stomatal guard cell turgor, helps regulate stomatal opening, participates in photosynthesis, enhances drought resistance, and increases yield". [17]

Rhizobium are known to form colonies on the root surface stimulating biological nitrogen fixation and providing nitrogen to the leguminous crops and hence considered as a significant process for improving yield and soil fertility. Azotobacter spp. is sensitive to acidic pH, high salt concentration and temperature. They pose advantageous impacts on the crop growth and yield through the biosynthesis of biologically active substances, instigation of rhizospheric microbes, production of phytopathogenic inhibitors, alteration of nutrient uptake and eventually magnifying the biological nitrogen fixation. **The objective of this study is to analysis the Effect of Different Levels of Inorganic and Bio Fertilizers on Physico - Chemical Properties of Soil in Mung Bean.**

Materials and Methods

A field experiment conducted at the Soil Science Research Farm, Sam Higginbottom University of Agriculture, Technology and Sciences, Prayagraj, during *summer* season May to July 2022 growing mung bean *Var.* RMG-975 applied 3 levels of NPK and Rhizobium + Azotobacter respectively 0 %, 50 % and 100 % including RDF for mung bean = 20:60:40 kg ha⁻¹ experiment is lead to observe the physical and chemical parameters. In physical parameters like that bulk density, particle density, pore space and water holding capacity through method by 100 ml graduated measuring cylinder and process by Muthuvel *et al.*, 1992.

In chemical parameters through method by-

- a) Soil pH – method given by (Jackson, M. L. 1958) through using digital pH meter.
- b) Soil EC (dSm⁻¹) - method given by (Wilcox, 1950) through using digital EC meter.
- c) Organic Carbon (%) - Wet oxidation method given by (Walkley and Black, 1947)
- d) Available Nitrogen (kg ha⁻¹) - Kjeldhal Method (Subbiah and Asija, 1956)
- e) Available Phosphorus (kg ha⁻¹) - Colorimetric method by using Jasper single beam U.V. Spectrophotometer at 660 nm wavelength given by (Olsen *et al.*, 1954)
- f) Available Potassium (kg ha⁻¹) - Flame photometric method by using Metzger Flame Photometer given by (Toth and Prince, 1949)

Result and Discussion

Physical Properties of Soil

Bulk density (Mg m^{-3})

The data presented in table and fig. 1 show the influence on bulk density (Mg m^{-3}) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in bulk density of soil was found non-significant due to level of inorganic and bio fertilizers. The maximum bulk density of soil 1.38 and 1.45 Mg m^{-3} at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) and minimum 1.26 and 1.30 Mg m^{-3} at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) respectively. Similar result has been recorded by Venkatarao *et al.*, 2017.

Particle density (Mg m^{-3})

The data presented in table and fig. 1 shows the influence on particle density (Mg m^{-3}) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in particle density of soil was found non-significant due to level of inorganic and bio fertilizers. The maximum particle density of soil 2.50 and 2.62 Mg m^{-3} at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) and minimum 2.32 and 2.37 Mg m^{-3} at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) respectively. Similar result has been recorded by Chaudhari *et al.*, 2016 and Venkatarao *et al.*, 2017.

Percent pore space

The data presented in table and fig. 1 depicted the influence in percent pore space of soil after crop harvest due to application of inorganic and bio fertilizers. The response in percent pore space of soil was found significant due to level of inorganic and bio fertilizers. The maximum percent pore space of soil 44.86 and 42.18 % at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) and minimum 38.28 and 35.62 % at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) respectively. Similar result has been recorded by Bhavya *et al.*, 2018.

Water holding capacity (%)

The data presented in table and fig. 1 depicted the influence in water holding capacity (%) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in water

holding capacity (%) of soil was found significant due to level of inorganic and bio fertilizers. The maximum water holding capacity of soil 38.65 and 35.80 % at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) and minimum 32.63 and 29.26 % at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) respectively. Similar result has been recorded by Bhavya *et al.*, 2018.

Chemical Properties of Soil

Soil pH (1:2.5) w/v

The data presented in table and fig. 2 depicted the influence in pH of soil after crop harvest due to application of inorganic and bio fertilizers. The response in pH of soil was found non-significant due to level of inorganic and bio fertilizers. The maximum pH of soil 7.56 and 7.62 at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) and minimum 6.80 and 6.92 at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) respectively. Similar result has been recorded by Ghanshyam *et al.*, 2010 Rathour *et al.*, 2015; Bhavya *et al.*, 2018; and Rekha *et al.*, 2018.

Soil Electrical Conductivity (dSm⁻¹)

The data presented in table and fig. 2 show the influence in electrical conductivity (dSm⁻¹) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in electrical conductivity of soil was found non-significant due to level of inorganic and bio fertilizers. The maximum electrical conductivity of soil 0.51 and 0.55 dSm⁻¹ at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) and minimum 0.38 and 0.41 dSm⁻¹ at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) respectively. Similar result has been recorded by Bhavya *et al.*, 2018; and Rekha *et al.*, 2018.

Organic Carbon (%)

The data presented in table and fig. 2 depicted the influence in organic carbon (%) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in organic carbon (%) of soil was found significant due to level of inorganic and bio fertilizers. The maximum organic carbon of soil 0.54 and 0.51 % at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) and minimum 0.41 and 0.37

% at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) respectively. Similar result has been recorded by Rekha *et al.*, 2018.

Available Nitrogen (kg ha⁻¹)

The data presented in table and fig. 3 depicted the influence in available nitrogen (kg ha⁻¹) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in available nitrogen (kg ha⁻¹) of soil was found significant due to level of inorganic and bio fertilizers. The maximum available nitrogen of soil 260.45 and 264.18 at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) and minimum 242.15 and 245.32 kg ha⁻¹ at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) respectively. Similar result has been recorded by Chaudhari *et al.*, 2016 and Venkatarao *et al.*, 2017.

Available Phosphorus (kg ha⁻¹)

The data presented in table and fig. 3 depicted the influence in available phosphorus (kg ha⁻¹) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in available phosphorus (kg ha⁻¹) of soil was found significant due to levels of inorganic and bio fertilizers. The maximum available phosphorus of soil 29.14 and 25.82 kg ha⁻¹ at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) and minimum 16.42 and 14.26 kg ha⁻¹ at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) respectively. Similar result has been recorded by Chaudhari *et al.*, 2016 and Venkatarao *et al.*, 2017.

Available Potassium (kg ha⁻¹)

The data presented in table and fig. 3 depicted the influence in available potassium (kg ha⁻¹) of soil after crop harvest due to application of inorganic and bio fertilizers. The response in available potassium (kg ha⁻¹) of soil was found significant due to levels of inorganic and bio fertilizers. The maximum available potassium of soil 211.29 and 207.62 kg ha⁻¹ at 0-15 and 15-30 cm was recorded in treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) and minimum 182.32 and 178.25 kg ha⁻¹ at 0-15 and 15-30 cm was recorded in treatment T₁ (NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %) respectively. Similar result has been recorded by Chaudhari *et al.*, 2016; Venkatarao *et al.*, 2017.

Table 1: Influence in bulk density (Mg m^{-3}), particle density (Mg m^{-3}), pore space (%) and water holding capacity (%) of soil after crop harvest due to application of inorganic and bio fertilizers.

Treatment		Bulk density (Mg m^{-3})		Particle density (Mg m^{-3})		Pore space (%)		Water holding capacity (%)	
		0 – 15 cm	15 – 30 cm	0 – 15 cm	15 – 30 cm	0 – 15 cm	15 – 30 cm	0 – 15 cm	15 – 30 cm
T₁	NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %	1.38	1.45	2.50	2.62	38.28	35.62	32.63	29.26
T₂	NPK @ 0 % + Rhizobium @ 50 % + Azotobacter @ 50 %	1.36	1.44	2.46	2.58	39.35	35.92	33.02	29.85
T₃	NPK @ 0 % + Rhizobium @ 100 % + Azotobacter @ 100 %	1.35	1.40	2.43	2.54	40.62	36.29	33.78	30.12
T₄	NPK @ 50 % + Rhizobium @ 0 % + Azotobacter @ 0 %	1.37	1.42	2.42	2.52	40.92	37.06	34.15	30.72
T₅	NPK @ 50 % + Rhizobium @ 50 % + Azotobacter @ 50 %	1.33	1.39	2.40	2.49	41.22	37.82	34.78	31.82
T₆	NPK @ 50 % + Rhizobium @ 100 % + Azotobacter @ 100 %	1.30	1.36	2.37	2.45	42.71	39.20	35.60	32.48
T₇	NPK @ 100 % + Rhizobium @ 0 % + Azotobacter @ 0 %	1.32	1.38	2.35	2.42	43.08	40.36	36.06	33.24
T₈	NPK @ 100 % + Rhizobium @ 50 % + Azotobacter @ 50 %	1.30	1.35	2.33	2.38	44.12	41.32	36.82	34.26
T₉	NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %	1.26	1.30	2.32	2.37	44.86	42.18	38.65	35.80
F-Test		NS	NS	NS	NS	S	S	S	S
S.Ed. (\pm)		-	-	-	-	0.65	0.48	0.42	0.35
C.D. at 0.5%		-	-	-	-	1.32	0.98	0.87	0.73

Table 2: Influence in pH (1:2.5) w/v, electrical conductivity (dSm⁻¹) and organic carbon (%) of soil after crop harvest due to application of inorganic and bio fertilizers.

Treatment		Soil pH (1:2.5) w/v		Electrical Conductivity (dSm ⁻¹)		Organic carbon (%)	
		0 – 15 cm	15 – 30 cm	0 – 15 cm	15 – 30 cm	0 – 15 cm	15 – 30 cm
T₁	NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %	7.56	7.62	0.38	0.41	0.41	0.37
T₂	NPK @ 0 % + Rhizobium @ 50 % + Azotobacter @ 50 %	7.42	7.58	0.40	0.43	0.42	0.38
T₃	NPK @ 0 % + Rhizobium @ 100 % + Azotobacter @ 100 %	7.40	7.52	0.44	0.46	0.43	0.40
T₄	NPK @ 50 % + Rhizobium @ 0 % + Azotobacter @ 0 %	7.34	7.46	0.39	0.42	0.42	0.39
T₅	NPK @ 50 % + Rhizobium @ 50 % + Azotobacter @ 50 %	7.27	7.38	0.41	0.45	0.44	0.41
T₆	NPK @ 50 % + Rhizobium @ 100 % + Azotobacter @ 100 %	7.18	7.26	0.42	0.49	0.47	0.44
T₇	NPK @ 100 % + Rhizobium @ 0 % + Azotobacter @ 0 %	7.01	7.15	0.45	0.51	0.48	0.45
T₈	NPK @ 100 % + Rhizobium @ 50 % + Azotobacter @ 50 %	6.88	6.97	0.49	0.53	0.51	0.47
T₉	NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %	6.80	6.92	0.51	0.55	0.54	0.50
	F-Test	NS	NS	NS	NS	S	S
	S.Ed. (±)	-	-	-	-	0.08	0.05
	C.D. at 0.5%	-	-	-	-	0.20	0.12

Table 3: Influence in available nitrogen (kg ha^{-1}), available phosphorus (kg ha^{-1}) and available potassium (kg ha^{-1}) of soil after crop harvest due to application of inorganic and bio fertilizers.

Treatment		Available nitrogen (kg ha^{-1})		Available phosphorus (kg ha^{-1})		Available potassium (kg ha^{-1})	
		0 – 15 cm	15 – 30 cm	0 – 15 cm	15 – 30 cm	0 – 15 cm	15 – 30 cm
T₁	NPK @ 0 % + Rhizobium @ 0 % + Azotobacter @ 0 %	242.15	245.32	16.42	14.26	182.32	178.25
T₂	NPK @ 0 % + Rhizobium @ 50 % + Azotobacter @ 50 %	244.68	246.54	17.36	14.68	183.54	179.42
T₃	NPK @ 0 % + Rhizobium @ 100 % + Azotobacter @ 100 %	245.42	248.35	19.27	15.65	186.05	181.46
T₄	NPK @ 50 % + Rhizobium @ 0 % + Azotobacter @ 0 %	246.72	249.28	20.52	17.02	188.38	184.02
T₅	NPK @ 50 % + Rhizobium @ 50 % + Azotobacter @ 50 %	248.46	251.60	22.48	17.80	192.65	187.80
T₆	NPK @ 50 % + Rhizobium @ 100 % + Azotobacter @ 100 %	252.08	254.32	23.96	19.18	197.82	191.56
T₇	NPK @ 100 % + Rhizobium @ 0 % + Azotobacter @ 0 %	253.36	255.45	24.05	20.32	201.25	196.25
T₈	NPK @ 100 % + Rhizobium @ 50 % + Azotobacter @ 50 %	256.17	259.62	26.82	22.65	206.38	202.74
T₉	NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %	260.45	264.18	29.14	25.82	211.29	207.62
	F-Test	S	S	S	S	S	S
	S.Ed. (\pm)	2.30	1.95	0.75	0.60	1.40	1.15
	C.D. at 0.5%	4.63	3.98	1.52	1.24	2.82	2.34

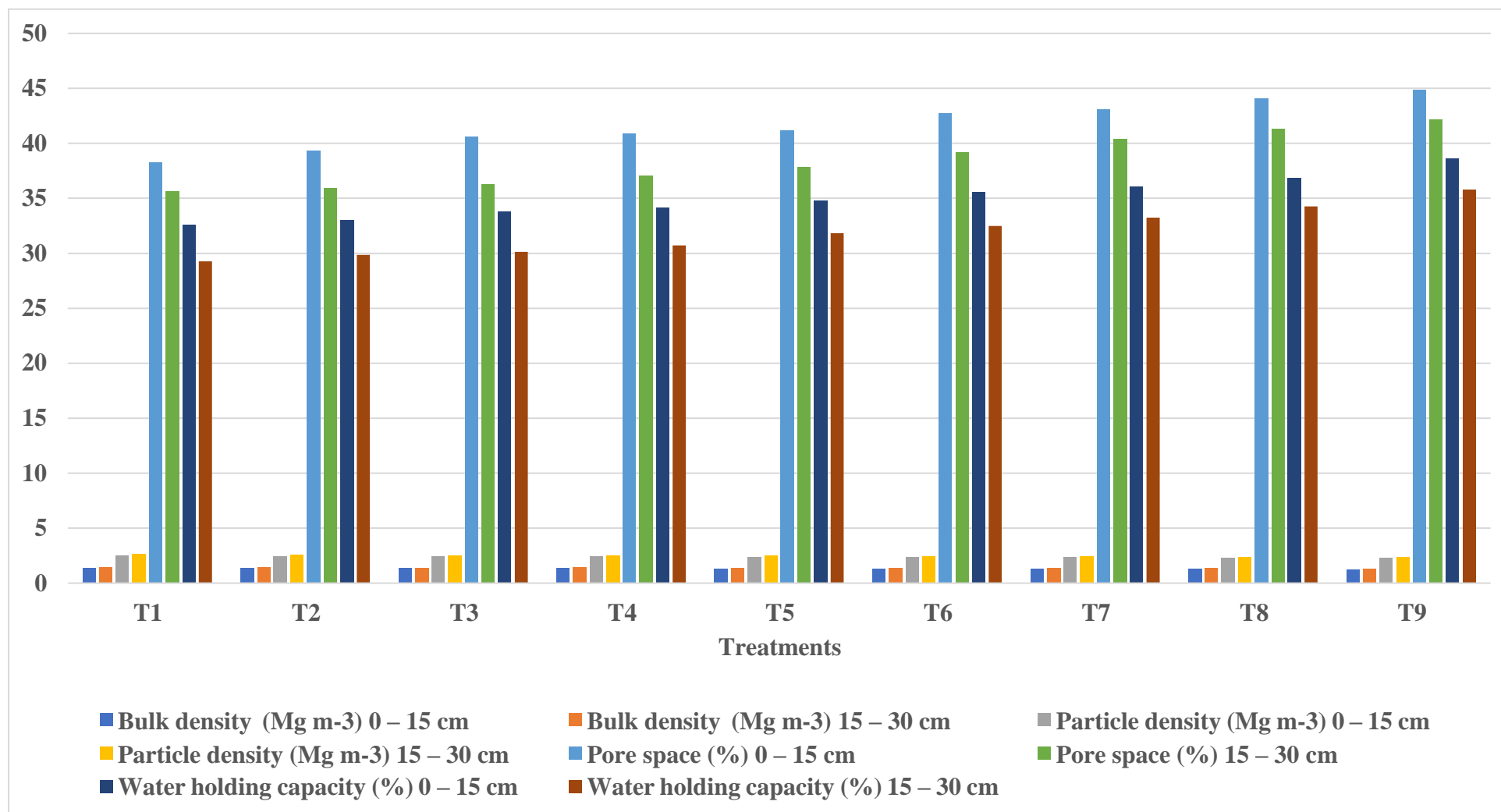


Fig. 1: Influence in bulk density (Mg m⁻³), particle density (Mg m⁻³), pore space (%) and water holding capacity (%) of soil after crop harvest due to application of inorganic and bio fertilizers.

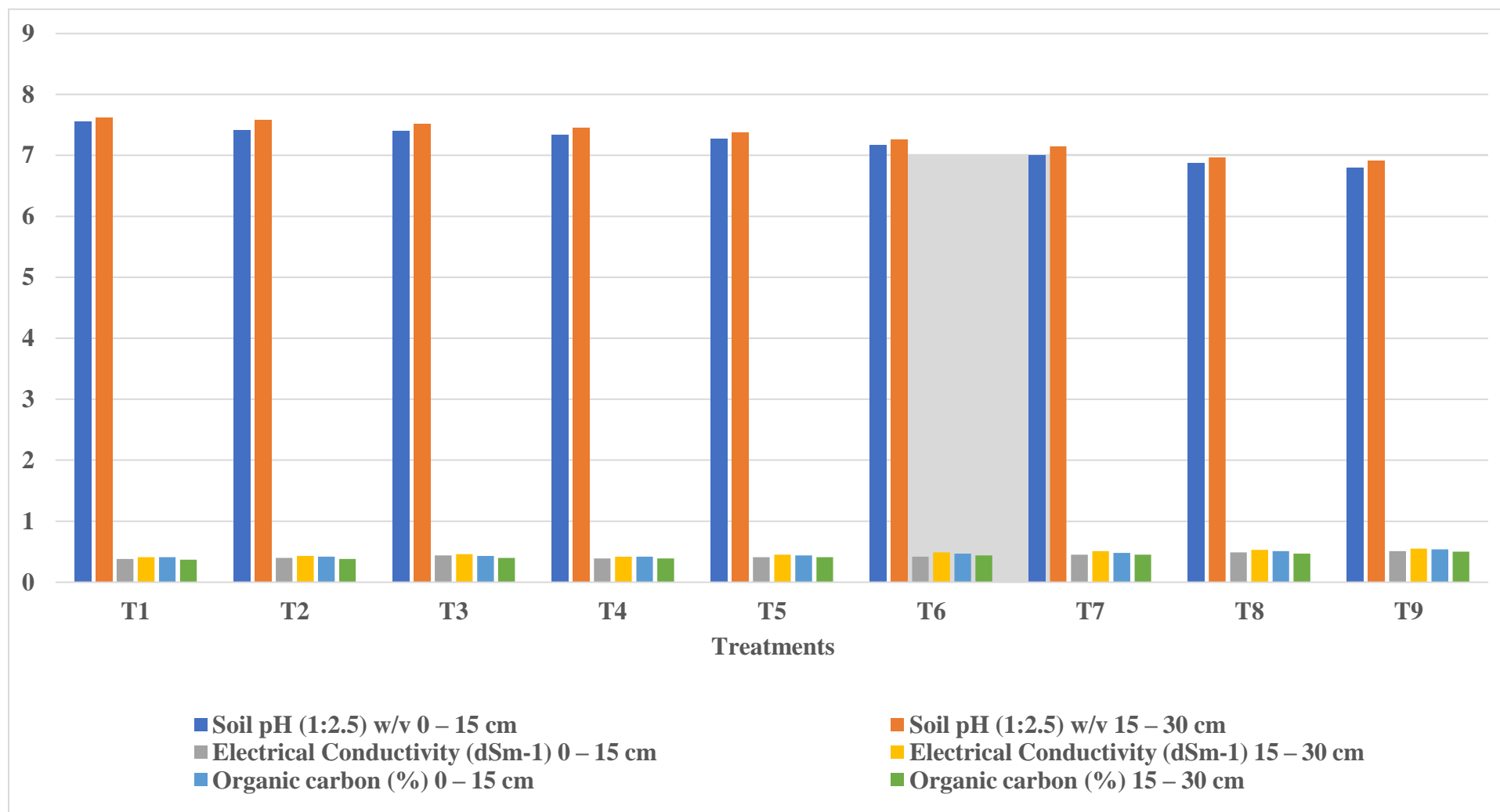


Fig. 2: Influence in pH (1:2.5) w/v, electrical conductivity (dSm⁻¹) and organic carbon (%) of soil after crop harvest due to application of inorganic and bio fertilizers.

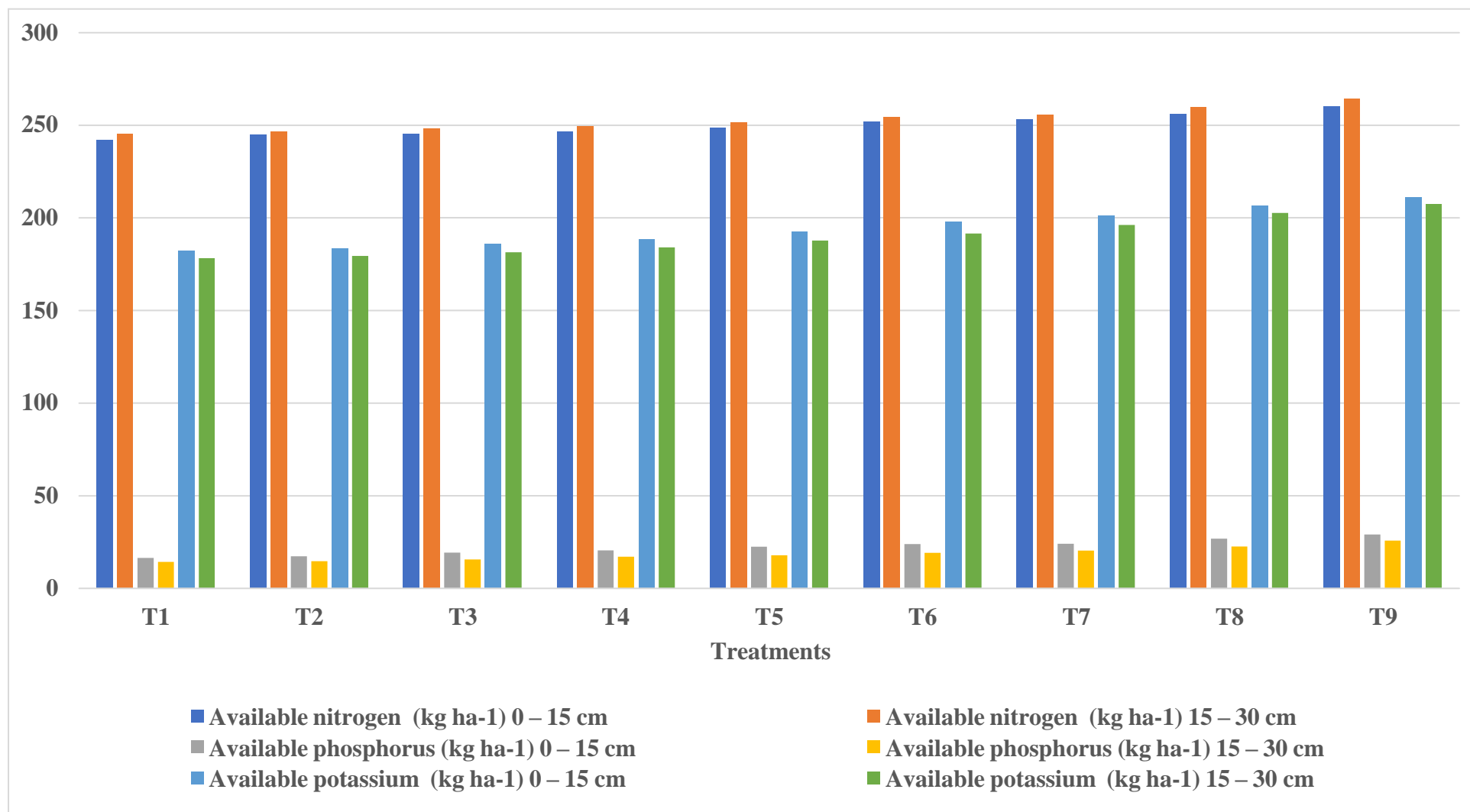


Fig. 3: Influence in available nitrogen (kg ha⁻¹), available phosphorus (kg ha⁻¹) and available potassium (kg ha⁻¹) of soil after crop harvest due to application of inorganic and bio fertilizers.

Conclusion

The results of the experiment were concluded as the effect of inorganic and bio fertilizers on Nitrogen, Phosphorus and Potassium (kg ha^{-1}), % pore space and water holding capacity (%) of soil after crop harvest was found significant except on bulk density (Mg m^{-3}), particle density (Mg m^{-3}), pH, EC (dSm^{-1}) and organic carbon (%) of soil after harvest. The treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) was recorded as best treatment for major soil parameters. The treatment T₉ (NPK @ 100 % + Rhizobium @ 100 % + Azotobacter @ 100 %) also shows the significantly.

Reference

1. **Bhavya, G., Shaker, K. C., Jayasree, G. and Reddy, M. M. (2018)** Effect of Integrate Use of Phosphorus, PSB and Vermicompost on Acid and Alkaline Phosphatase Activity and Yield of Green gram (*Vigna radiata* L.). *Int. J Curr. Microbiol. App. Sci.*, 7(1): 1465- 1468.
2. **Bouyoucos, G.L, (1927)** The hydrometer as a new method for the mechanical analysis of soils.
 - a. *Soil Sci.* 23: 343-353.
3. **Chaudhari, S., Thanki, J. D., Chaudhari, V. and Verma, C. (2016)** Yield attributes, yield and quality of black green gram (*Vigna radiata* L.) as influenced by organic manures, biofertilizer and phosphorus fertilization. *The Bio Scan.*, 11(1): 431-433.
4. **Fisher, R. A. and Yates (1960)** *Statistical method for research worker Oliver and Boyd Ltd.*
 - a. Edin. burgh and London.10.
5. **Jackson, M. L. (1958)** Soil chemical analysis Prentice Hall of India Ltd. New Delhi. 219-221.
6. **Miachio, P., Ardakani, M.R., Farzad, P. and Saied, V. (2019)** Effects of vermicompost, mycorrhizal symbiosis and biophosphate solubilizing bacteria on seed yield and quality of chickpea as autumn plantation in rainfed conditions. *Bulletin of Environment, Pharmacology and Life Sciences* 3(2): 53-58.
7. **Munsell, A. H. (1971)** Munsell's description of his colour system, from a lecture to the American Psychological Association. *American Journal of Psychology* 23(2): 236-244.
8. **Muthuvel, P., Udayasoorian, C., Natesan, R. and Ramaswamy, P. P. (1992)** Introduction to Soil Analysis, *Tamil Nadu Agricultural University Coimbatore-641002.*
9. **Olsen, S. R., Cole, C. V., Watanabe, F. S. and Dean, L. A. (1954)** Estimation of

available phosphorus in soils by extraction with sodium bicarbonate (NaHCO_3), *U.S.D.A. Circular*. 939: 1-19.

10. **Rekha, K., Pavaya, R. P., Malav, J. K., Chaudhary, N., Patel, I. M. and Patel, J. K. (2018)** Effect of FYM, phosphorus and PSB on yield, nutrient content and uptake by green gram [*Vigna radiata* (L.)] on loamy sand. *Indian Journal of Chemical Studies*. 6(2): 1026-1029.
11. **Subbiah, B. V. and Asijja, E. C. (1956)** A rapid procedure for estimation of available nitrogen in soil. *Current Science* 25(8): 259-260.
12. **Toth, S. J. and Prince, A. L. (1949)** Estimation of cation exchange capacity and exchangeable Ca, K and Na content of soil by flame photometer technique. *Soil Sci.*, 67: 439-445.
13. **Venkatarao, C. V., Naga, S. R., Yadav, B. L., Koli, D. K. and Rao, I. J. (2017)** Effect of Phosphorus and Biofertilizers on Growth and Yield of Mungbean [*Vigna radiata* (L.) Wilczek]. *Int. J. Curr. Microbiol. App. Sci.*, 6(7): 3992-3997.
14. **Walkley, A. and Black, I. A., (1947)** Estimation of soil organic carbon by the chromic acid titration method. *Soil Science*. 47: 29-38.
15. **Wilcox, L.V. (1950)** Electrical conductivity. *Am. Water Works Assoc. J.* 42: 775-776.
16. **Zafar, Z. U. and Athar, H. U. R. (2013)** Influence of different phosphorus regimes on disease resistance in two cotton (*Gossypium hirsutum* L.) cultivars differing in resistance to cotton leaf curl virus (clcuV). *Pakistan Journal Botany*. 45(2): 617-627.
Ighodaro UB, Idumah FO, Mangodo C, Isese MO. Prospects of Sustainable Soil Management to a Green Economy. *Greener Journal of Agricultural Sciences*. 2017;7(8):197-202.
17. Yin Z, Guo W, Xiao H, Liang J, Hao X, Dong N, Leng T, Wang Y, Wang Q, Yin F. Nitrogen, phosphorus, and potassium fertilization to achieve expected yield and improve yield components of mung bean. *PloS one*. 2018 Oct 25;13(10):e0206285.