

Review Article

CHITOSAN APPLICATION IN AQUATIC FEED AND ITS IMPACT ON FISH AND SHRIMP PRODUCTIVITY

ABSTRACT

The culture of crustaceans, such as shrimp and crabs, in Indonesia has resulted in a significant increase in waste production, particularly in the form of shells and heads. This waste contains valuable components like chitin, chitosan, and carotenoids, which can be extracted and utilized in aquaculture activities. Some research results show that the use of chitosan, derived from chitin, in fish feed, is a functional polysaccharide with various biological activities, including antimicrobial, anti-inflammatory, and immune-stimulating effects. It also acts as a growth promoter in shrimp and improves the performance of farmed fish. Additionally, the inclusion of chitin and chitosan in fish feed formulations enhances digestibility, growth, and nutrient absorption. This paper provides a comprehensive review of the potential, nutritional value, and benefits of processed crustacean waste products, focusing on chitosan, and their impact on the productivity of fish in aquaculture.

Keywords: aquaculture, crustacean waste, chitosan, fish feed, performance, shrimp

1. INTRODUCTION

The cultivation of crustaceans, including shrimp and crabs, has witnessed significant growth in Indonesia. However, this growth has led to an increase in waste generated from the crustacean industry, primarily in the form of shells, heads, and shells. Crayfish carapace is by-product from seafood processing with hundreds of thousands of tons it contains 40% calcium carbonate, 30% protein and 30% chitin [1]. In the case of shrimp, heads alone contribute around 35-45% of the total shrimp production [2]. Failure to utilize this waste not only contributes to environmental pollution but also poses challenges to maintaining environmental sanitation due to the decomposition-associated unpleasant odor.

Shrimp and crab waste possess considerable potential as sources of chitin, chitosan, and carotenoids, all of which can be utilized in aquaculture activities. Chitin can be extracted from shrimp and crab shells through demineralization and deproteination processes. Subsequently, chitin can be further processed into chitosan through a deacetylation process. Furthermore, the shells of shrimp, which contain carotenoids, particularly astaxanthin, can be processed to obtain carotenoid-rich compounds using non-polar solvents, edible oils, or enzymatic methods employing protease enzymes like trypsin. Consequently, the waste-derived products obtained from these processes can be effectively utilized as additives in fish feed. This paper aims to comprehensively review the potential, nutritional value, and benefits of processed crustacean waste products, with a particular focus on chitosan, and their impact on the performance of farmed fish.

2. CHITOSAN: STRUCTURE, SOURCES, AND NUTRITIONAL CONTENT

Chitosan, a result of deacetylation of chitin, is a long-chain polymer of glucosamine (2-amino-2-deoxy-glucose) [3] (Figure 1). While insoluble in organic solvents like dimethyl sulfoxide and at pH 6.5, chitosan readily dissolves in organic acid solutions, with acetic acid being an effective solvent [4].

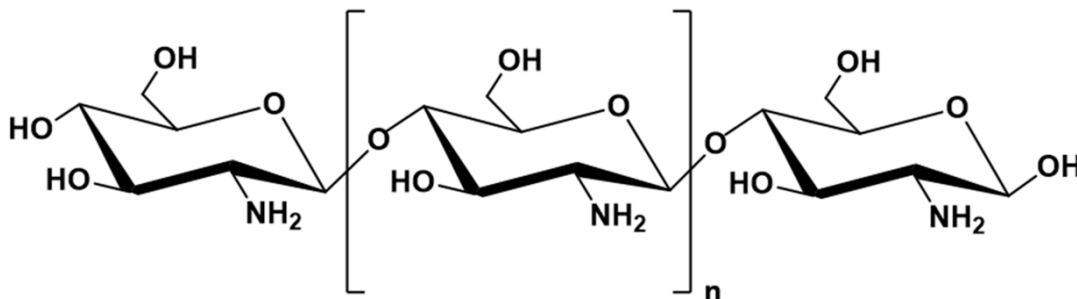


Fig. 1. Chitosan chemical structure
Source: [4]

Chitosan, a polyglucosamine, exhibits solubility in various acids such as acetic, lactic, and organic acids (adipic, malic), as well as mineral acids like HCl and HNO₃, at a concentration of 1%. However, it has limited solubility in phosphoric acid and is insoluble in sulfuric acid. With its amine group as a functional group, chitosan displays a high degree of chemical reactivity [5]. [6] noted that chitosan carries a positive charge in solution due to the presence of amine groups, distinguishing it from other polysaccharides that are typically negatively charged or neutral. The quality of chitosan varies according to its intended use, with higher purity required for pharmaceutical applications compared to wastewater treatment processes [7].

Chitosan can be obtained from various crustaceans, such as shrimp, crabs, and crayfish, in the form of shells, heads, and exoskeletons. Additionally, chitin, the precursor of chitosan, is found in diverse sources. It is widely distributed in nature and serves as a major component of exoskeletons in marine invertebrates. Chitin is also present in the structural composition of insects, arthropods, and molluscs. Figure 2 illustrates the primary sources of chitin production and extraction. From a nutritional standpoint, chitosan exhibits favorable nutritional value (Table 1), making it a viable ingredient in fish feed formulations.

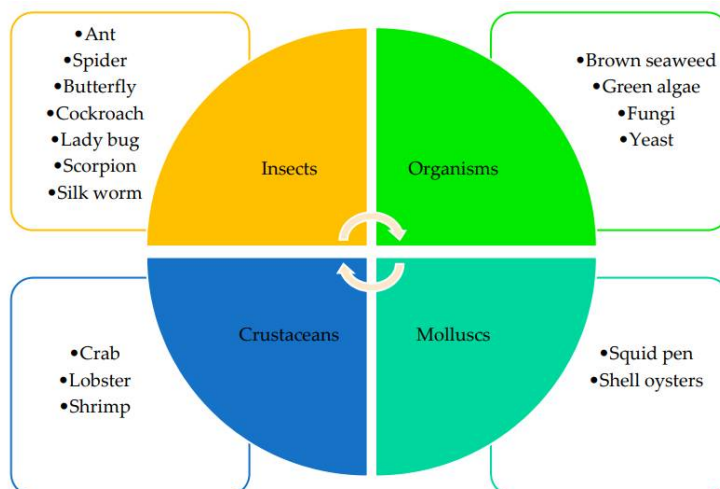


Fig. 2. Sources of chitin production

Table 1. Characteristics of shrimp shell chitosan

Specification	Analysis Results	% dry basis	
		ESFA 2010	GRAS 2012
Color	Brownish white	-	White to off white powder
Odor	Odorless	-	Neutral
Flake	Form	-	18-120 mesh
Moisture content (%)	12.29	≤ 10	≥ 10
Ash content (%)	0.99	≤ 3	≥ 0.5
Fat content (%)	3.13	≤ 1	-
Total nitrogen (%)	2.20	≤ 6	0.02g/100g
Carbohydrates (%)	81.39*	-	-
Viscosity (cPs)	1713.04	-	25-5000
Degree of deacetylation	98.65	≥ 90	75-95

Description: *) by difference

3. ROLE OF CHITOSAN IN FISH AND SHRIMP FEED

Chitin and chitosan, carbohydrate polymers found in crustacean waste, offer diverse applications, including their use as additives in fish and shrimp feeds. Additionally, crustacean waste contains valuable carotenoids, which can be extracted from chitosan and utilized as additives in aquaculture and ornamental fish feeds, enhancing color intensity.

In aquaculture, chitosan plays a crucial role in improving the performance of farmed fish. As a functional polysaccharide, chitosan exhibits various biological activities, such as haemostatic properties [8], anti-inflammatory effects [9], antitumor potential [10],[11], antimicrobial activity [12], hypoglycemic and hypocholesterolemic effects [13],[14], as well as immune-stimulating properties [15],[16]. Chitosan, containing glucosamine derived from chitin, can serve as a growth promoter in shrimp, primarily by accelerating carbohydrate metabolism [17],[18],[19],[20]

Moreover, [21] reported that the inclusion of chitin at levels of 1% to 4% in shrimp feed enhances digestibility and growth. This finding suggests that shrimp have an improved ability to absorb and utilize glucose from the feed source. Similarly, it is anticipated that glucosamine from chitin sources can stimulate the growth of other aquatic organisms. In addition, chitosan supplementation in feed demonstrates immune-stimulating effects, enhances nutrient digestion and absorption, and increases protein content in Indian major carp (*Labeorohita*), consequently improving growth performance [22].

Furthermore, [23] found that chitosan protects α -fold acid antioxidant compounds from damage caused by heat, light, and acidic conditions. Chitosan has emerged as a promising growth promoter in animal feeds. [24] further revealed that the addition of chitosan to milkfish feed mitigates the odor of mud attributed to geosmin, as chitosan binds with the geosmin compound and facilitates its excretion through feces, thereby reducing the characteristic odor.

4. FISH AND SHRIMP PERFORMANCE BASED ON CHITOSAN FEED

Numerous research studies have investigated the effects of chitosan-based feed on the productivity of fish and shrimp in the aquaculture industry. Table 2 provides a concise overview of the key findings from these studies, highlighting the outcomes of employing chitosan in feed formulations.

Table 2. Several research related to chitosan-based feed

Treatment	Result	References
Use of chitosan in milkfish	In this study, the optimal results were achieved when incorporating 10 ppm of chitosan into the feed formulation, which effectively reduced the mud odor in milkfish. The feeding duration of 4 days was found to be the most effective in reducing the mud smell. Proximate analysis revealed that the fat and carbohydrate contents decreased in milkfish treated with the best formulation (10 ppm chitosan for 4 days) compared to the control. Conversely, there was an increase in ash content and protein percentage in the treated group compared to the control	[25]
The use of chitosan for carp growth	Supplementing fish feed with 8.0% chitin has shown a positive impact on growth response and feed utilization in carp. The inclusion of chitin at this concentration resulted in improved growth performance 27.3% higher than control and enhanced feed utilization efficiency	[26]
Use of chitosan for tilapia growth	The optimal treatment for chitosan supplementation in tilapia was found to be at a dose of 100 ppt. This treatment resulted in a feed conversion ratio (FCR) of 3.48 ± 2.25 , average growth rate (SGR) of 0.85 ± 0.84 , average protein gain (APG) of 28.70 ± 16.54 . Statistical analysis (ANOVA) indicated that all chitosan treatments did not have a significant effect ($P > 0.05$) on the number of erythrocytes and leukocytes in tilapia	[27]
Use of chitosan for shrimp growth	The addition of chitosan to the feed during a 30-day rearing period exhibited a significant effect on the specific growth rate (SGR) and protein retention in vannamei shrimp. The highest SGR was observed with the addition of 4% chitosan, which was not significantly different from the inclusion of 3% chitosan. Similarly, increased protein retention was achieved by incorporating 4% chitosan, without significant difference from the addition of 3% chitosan	[28]

5. CONCLUSION

The utilization of crustacean waste, particularly shells and heads, presents an opportunity to harness valuable components such as chitin, chitosan, and carotenoids for the enhancement of aquaculture activities. Chitosan, derived from chitin, has shown significant potential as a functional polysaccharide with various biological activities and as a growth promoter in shrimp. Its inclusion in fish feed formulations improves digestibility, growth, and nutrient absorption. **Chitosan would be a good immunostimulant/ growth promoter in shrimp diets.** Further research and development in this area are warranted to optimize the utilization of crustacean waste and maximize its potential in aquaculture practices.

REFERENCES

1. Meyers, S.P., H.M. Chen, H.K. No and K.S. Lee, 1990. An Integrated Approach to Recovery and Utilization of Louisiana Crawfish Processing Wastes. In: Making Profits Out of Seafood Wastes, Keller, S. (Ed.). Alaska Sea Grant College Program, Anchorage, AK., pp: 161-171
2. Jeyaprakash Sabari S.1 and Aanand S. 2021. Shrimp Waste - A Valuable Protein Source for Aqua Feed. *AgriCos e-Newsletter*, 2(7):64-67
3. Knorr D. Functional properties of chitin and chitosan, *J. Food Sci. and Technol.* 1982; 47: 593
4. Ornum VJ. Shrimp Waste-Must it be Wasted. In *INFOFISH International* 1992; 6: 48-52.
5. Mahfouz ME and Leporatti S. Chitosan as a natural copolymer with unique properties for the development of hydrogels. *Applied sciences*, 1019; 9(11): 2193
6. Johnso, EL and Peniston QP. Utilization of Shellfish Waste for Producing of Chitin and Chitosan Production In: *Chemistry and Biochemistry of Marine Food Product*. Westport Connecticut: The AVI Pub. Co. Inc. 1982.
7. Muzzarelli RAA. Chitin. Pergamon Press. New York. 1977
8. Bastaman S. Studies on Degradation and Extraction of Chitin and Chitosan from Prawn Shells. Belfast: The Queen's University. 1989
9. Pusateri AE, Holcomb JB, Kheirabadi BS, Alam HB, Wade C and Ryan KL. Making sense of the preclinical literature on advanced hemostatic products. *The Journal of Trauma*. 2006; 60(3): 674-682.
10. Dai T, Tegos GP, Burkatovskaya M, Castano AP and Hamblin MR. Chitosan Acetate Bandage as a topical Antimicrobial Dressing for Infected Burns. *Antimicrob. Agents Chemother.* 2009; 53: 393-400
11. Tsukada K, Matsumoto T, Aizawa K, Tokoro A, Naruse R, Suzuki S, and Suzuki M. Antimetastatic and Growth-Inhibitory Effects of N-Acetylchitohexaose in: Mice Bearing Lewis Lung Carcinoma. *Japanese Journal Cancer Research*. 1990; 81: 259-265.
12. Koide H. Pseudodementia and delirium in depression: A contribution to psychosomatic medicine. 1992; 46(4)
13. Benhabiles MS, Salah R, Lounici H, Drouiche N, Goosen MFA and Mameri N. Antibacterial Activity of Chitin, Chitosan and Its Oligomers Prepared from Shrimp Shell Waste. *Food Hydrocoll.* 2012; 29: 48-56
14. Yao HT, Huang SY and Chiang MT. Effect of Chitosan on Plasma Cholesterol and Glucose Concentration in Streptozotocin-Induced Diabetic Rats. *Taiwan. J. Agric. Chem. Food Sci.* 2006; (44):122-132.
15. Yao HT, Huang SY and Chiang MT. A Comparative Study on Hypoglycemic and Hypocholesterolemic Effects of High and Low Molecular Weight Chitosan in Streptozotocin-Induced Diabetic Rats. *Food Chemistry, Toxicology*. 2008; 46:1525-1534.
16. Moon JS, Kim HK, Koo HC, Joo YS, Nam HM, Park YH and Kang MI. The antibacterial and immunostimulative effect of chitosan-oligosaccharides against infection by *Staphylococcus aureus* isolated from bovine mastitis. *Appl. Microbiol. Biotechnol.* 2007; 75: 989-998.
17. Yin YL, Tang ZR, Sun ZH, Liu ZQ, Li TJ, Huang RL, Ruan Z, Deng ZY, Gao B, Chen LX, Wu GY and Kim S. Effect of Galacto-Mannan-Oligosaccharides or Chitosan Supplementation on Cytoimmunity and Humoral Immunity Response in Early-Weaned Piglets. *Asian-Aust. Journal Animal Science*. 2008; 21:723-731
18. Kanazawa A, Shimaya M, Kawasaki M and Kashiwada K. Nutritional requirements of prawn, *L. feeding on artificial diet*. *Bull. Jap. Soc. Sci. Fish*, 1970; 36: 949-954
19. Kitabayashi K, Kurata H, Shudo K, Nakamura K and Ishikawa S. Studies on formula feed for kuruma prawn. I. On relationship among glucosamine, phosphorus and calcium. *Bull. Tokai Reg. Fish. Res. Lab.*, 1971; 65: 91-207

20. Clark AG, Barrat TM. Steroid responsive nephrotic syndrome. In: Barratt TM, Avner ED, Harmon WE, editors. *Pediatric Nephrology*. 4th ed. Philadelphia: Lippincot Williams & Wilkins. 731-48. 1993
21. Fox CJ. The effect of dietary chitin on growth, survival and chitinase levels in the digestive gland of juvenile *Penaeus monodon*. *Aquaculture*, 1993; 109: 39-49
22. Aathi K, Ramasubramanian V, Uthayakumar V and Munirasu S. Effect of Chitosan Supplemented Diet on Survival, Growth, Hematological, Biochemical and Immunological Responses of Indian Major Carp *Labeo rohita*. *International Research Journal of Pharmacy*, 2013; 4(5): 141-147.
23. Kofuji K, Qian CJ, Nishimura M, Sugiyama I, Murata Y and Kawashima S. Relationship between physicochemical characteristics and functional properties of chitosan. *Eur. Polym. J.* 2005; 41: 2784-2791.
24. Wahyudi B. Pengaruh khitosan dalam formulasi pakan ikan terhadap kandungan lemak terkontaminasi geosmin dan pertumbuhan ikan nilahitam (*Oreochromis niloticus*). Bachelor Thesis. Teknologi Hasil Perikanan. Bogor Agricultura Institute. 2001
25. Hafiluddin and Triajie H. Penambahan khitosan pada ikan Bandeng (*Chanoschanos*) sebagai penurun cita rasa lumpur (Geosmine). *EMBRYO*, 2011; 8(2): 126-132.
26. Hariati AM, Kartikaningsih H, Wiadnya, DGR., Suryanti Y, and Subagyo. Pengaruh Kadar Kitin dalam Pakan Terhadap Laju Pertumbuhan dan Konsumsi Pakan Harian Ikan Gurami *Osphronemus gouramy* LAC. *Penelitian Perikanan Indonesia*, 2017; 6: 8-12
27. Rozi, Mukti AT, Samara H, and Santanumurti MB. Pengaruh Pemberian Khitosan dalam Pakan terhadap Pertumbuhan, Sintasan dan Efisiensi Pemanfaatan Pakan Nila (*Oreochromis niloticus*). *Perikanan Universitas Gadjah Mada*, 2018; 20(2): 103-111.
28. Ekaputri RA, Arief M, Rahardja S, and Kurniasih N. Pengaruh Penambahan Khitosan pada Pakan Komersial terhadap Laju Pertumbuhan Spesifik dan Retensi Protein Udang Vaname (*Litopenaeus vannamei*) Effect of Chitosan Supplementation in Commercial Feed For Specific Growth Rate and Protein Retention of *Litopenaeus*. *Marine and Coastal Science*, 2018; 7(2): 39-50