

1 Original Research Article

2 PHARMACOGNOSTIC PROFILE AND ANTI-HYPERGLYCEMIC
3 ACTIVITY OF *Jatropha tanjorensis* LINN (EUPHORBIACEAE) LEAF ON
4 ALLOXAN-INDUCED HYPERGLYCEMIC RATS

5

6 ABSTRACT

7 **Background:** *Jatropha tanjorensis* has been used traditionally for the treatment of a variety of
8 diseases, these include; renal problem, cardiovascular disease, hypertension, inflammation and in
9 moderate depression. The pharmacognostic standardization of it's can be used in the
10 development of it's monograph. Diabetes mellitus is disease that is responsible for millions of
11 death yearly. Over the years, efforts have been made in the discovery of new bioactive
12 compounds with Antidiabetic activity

13 **Objective:** The objective of the study was to establish the pharmacognostic profile and anti-
14 hyperglycemic activity of methanol leaf extract of *Jatropha tanjorensis* on alloxan-induced
15 hyperglycemic rats.

16 **Method:** The phytochemical analysis, pharmacognostic profile and acute toxicity study were
17 done using standard methods. The anti-hyperglycemic activity of the methanol leaf extract of *J.*
18 *tanjorensis* was investigated by using normal and alloxan-induced hyperglycemic rats for acute
19 and sub-acute studies.

20 **Results:** Phytochemical screening revealed alkaloids, flavonoids, carbohydrates, reducing
21 sugars, cardiac glycosides, saponins and tannins. The moisture content obtained was 5.67% w/w.
22 The total ash value was 8.39%w/w, acid-insoluble ash value was 0.72% w/w and water-soluble
23 ash value was 3.91% w/w. There was no mortality or any signs of behavioral changes or toxicity
24 observed after oral administration of *Jatropha tanjorensis* up to the dose of 5000 mg/kg body
25 weight in mice. The sub-acute anti-hyperglycemic study was carried out for 14days, there was a
26 significant ($p < 0.05$) dose-dependent anti-hyperglycemic effect of the methanol leaf extract of *J.*
27 *tanjorensis* of 11.76, 55.51 and 77.65% blood glucose level reduction for 100, 200 and 400
28 mg/kg respectively, when compared with the negative control (- 23.18 %) and 67.29 % for
29 positive control; Glibenclamide (5 mg/kg) which has less activity.

30 **Conclusion:** From this study, it shows that *J. tanjorensis* leaf have anti-hyperglycemic property,
31 which could justify its use in traditional medicine for the treatment of *diabetes mellitus*. The
32 Pharmacognostic profile can be used for a monograph of the plant for its proper identification
33 and quality control.

34 **Keywords:** Diabetes, Hyperglycemia, *Jatropha tanjorensis*, Medicinal Plant, Pharmacognostic
35 Profile, Alloxan.

36 1. INTRODUCTION

37 “Diabetes mellitus, one of the major public health problems worldwide, is a metabolic disorder
38 of multiple etiologies distinguished by a failure of glucose homeostasis with disturbances of
39 carbohydrate, fat and protein metabolism as a result of defects in insulin secretion and/ insulin
40 action” [1]. According to International Diabetes Federation (IDF) report, “elevated blood glucose
41 is the third uppermost risk factor for premature mortality, following high blood pressure and
42 tobacco use globally” [2].

43 “In 2019, according to IDF report, 463 million adults (20-79yrs) were living with diabetes, and
44 by 2045 this will rise to 700 million. **Seventy nine percent** of adults with diabetes were living in
45 low- and middle-income countries. 1 in 5 people who are above **65 yrs** have diabetes and have
46 caused 4.2 million deaths” [3]. In Nigeria, there is an overall pooled prevalence of 5.77 % with
47 **11.7 million** Nigerians (1 out of every 17 adults) are living with diabetes.

48 Diabetes can be managed through pharmacological and non-pharmacological approaches [4].

49 “Different extracts from medicinal plants have also been used traditionally to manage diabetes
50 globally, and these are considered as relatively inexpensive, less toxic and with relatively little or
51 no side effects” [5].

52 “*Jatropha tanjorensis* (J.L. Ellis and Saroja) belonging to the Euphorbiaceae family is a
53 common plant of field crops, in rainforest zones of West Africa including Nigeria” [6]. “It is
54 commonly called “Hospital too far”, “Catholic vegetable”, “Iyana ipaja” or “Ugu-Oyibo”. Its
55 primary use is for fencing while its secondary use are a source of edible leafy vegetables and
56 medicine, prepared locally in most Southern Nigeria by collecting the leaves and squeezing out

57 the juice” [7]. “*J. tanjorensis* leaf exhibit low antioxidant and very low haemagglutination titre
58 value, the latter indicating low toxicity on red blood cells. The leaf extract has hypoglycemic
59 properties and is taken as a remedy against diabetes” [8]. “It is popular as a natural remedy
60 against malaria infection and hypertension in Southern Nigeria where they drink the squeezed-
61 out juice. However, there is no scientific validation to these claims. Research has shown that
62 fresh *Jatropha tanjorensis* leaves contain a high water and low protein content. The trace
63 elements, zinc, iron and selenium are in concentrations comparable to those found in food
64 regarded as good dietary sources of these elements. The leaf extract also possesses antimicrobial
65 properties and inhibit the growth of *S. aureus* and *E. coli*” [9]. “*J. tanjorensis* plant leaves are
66 popularly consumed in Nigeria as soup and as a tonic with the claim that it increases blood
67 volume. The leaves have anti-anemic effects (blood replenishing potentials). The leaf was found
68 to contain some important biogenic principles for rapid hematopoiesis in the bone marrow” [10].
69 “*J. tanjorensis* leaf is a potent anti-HIV agent (effective against HIV-1 vector)” [11]. The leaves
70 are also employed traditionally in the treatment of renal problem, cardiovascular disease,
71 hypertension, inflammation and in moderate depression.

72 The aim of this study was to find out the scientific basis of the use of *Jatropha tanjorensis* by
73 traditional medicine practitioners in the management of diabetes melitus; using the methanol leaf
74 extract on alloxan-induced hyperglycemic rats.

75

76

77

78

79 **2. MATERIALS AND METHODS**

80 **2.1 Collection and preparation of plant materials**

81 The leaves of *Jatropha tanjorensis* were freshly collected from a private garden in the locality of
82 Nsukka Local Government Area, Enugu State in July, 2020. The plant was identified by Mr.
83 **Felix Uzor**, a taxonomist in University of Nigeria, Nsukka and voucher specimen
84 (PCG/UNN/0378) deposited in the Herbarium of Department of **Pharmacognosy** and
85 Environmental Medicine of same University. The leaves were air dried at room temperature for
86 four days then further dried in an oven at 40°C for 6 hr. The crispy leaves were ground into
87 powder and filtered using a sieve aperture of 1.0 mm. The fine powder was preserved in
88 moisture-free airtight container and used for phytochemical analysis, microscopic and **anti-**
89 **hyperglycemic** evaluations.

90 **2.2 Animals**

91 Healthy mixed sexes of Wistar Albino mice and rats were purchased from the Animal Farm,
92 Department of Pharmacology and toxicology, University of Nigeria, Nsukka. The animals were
93 examined and acclimatized to the environmental conditions and were housed in aluminum cages
94 floored with saw-dust with provision of food and water a week prior the experiment.

95 **2.3 Preparation of extract**

96 A 650 g of the powdered leaf sample was macerated in **3 litres** of methanol (Analytical grade)
97 for 72 hr. The suspension was filtered and the resulting filtrate was evaporated to dryness over a
98 water bath to obtain a sticky extract. The percentage yield was then determined.

99

100

101 **2.4 Phytochemical Analysis**

102 Phytochemical analysis tests were carried out on the methanol leaf extract using standard
103 methods [12, 13] to test for the presence of secondary metabolites such as alkaloid,
104 carbohydrates, saponins, tannins, flavonoids, etc.

105 **2.5 Pharmacognostic profile**

106 **2.5.1 *Fresh Leaf Microscopy***

107 “Foliar epidermis of the adaxial (upper surface) and abaxial (lower surface) surfaces of the
108 leaves were prepared by clearing method. The leaf samples were cleared by soaking in 3.5%
109 sodium hypochlorite for 18 hr. Then, the epidermal strips of the leaf samples were scrapped
110 gently with the aid of a pair of forceps and placed on a clean slide, and then stained with Safranin
111 solution and covered with a cover slip” [14]. “The slides were viewed under a light phase
112 contrast microscope (Motic B3, Motic Carlsbad, CA, USA) at x 40, x 100 and x 400
113 magnifications and photomicrographs were taken with a Moticom 2.0 image system with
114 software (Motic Carlsbad, CA, USA) fitted to the microscope. The following parameters were
115 observed and assessed; Epidermal cells, Stomata type, Stomata size (length and width), Stomatal
116 density, Stomatal index, Trichome parameters, Vein islet number, vein islet termination number
117 and palisade ratio. All parameters were observed on both the adaxial and abaxial surfaces of the
118 leaves” [14].

119 Transverse section (TS) of the leaf was made using a Reichert sledge microtome following the
120 procedures of [15] and [16]. “The sections were microtomed at 10 – 15 unimicrons and were
121 picked with the aid of a camel hair brush from the tip of the microtome knife into separate Petri

122 dishes containing 70% absolute alcohol and labeled appropriately. Safranin and Fast green
123 served as biological stains in differentiating lignified tissues” [15,16].

124 **2.5.2 Chemomicroscopy evaluation**

125 Chemomicroscopy conducted on the powders to determine the presence of starch, calcium
126 oxalate crystals and lignified vessels. A judicious quantity of the sample was dropped on a glass
127 slide. One drop of chloral hydrate was dropped and passed over a Bunsen burner repeatedly until
128 bubbles formed. This signified the successful clearing of the tissues. The chemomicroscopy of
129 the plant constituents such as Starch, Lignin, Cellulose, Tannins, Calcium oxalate crystals, Gums
130 and Mucillages was done using appropriate reagents and following the standard
131 chemomicroscopy techniques.

132 **2.6 Acute toxicity study**

133 The acute toxicity test was accessed in mice using Lorke’s method [17]. Twelve mice were
134 selected for this study, all weighing 18 to 40g. The animals were starved for 18hrs prior to the
135 study and were only allowed access to water. The animals were divided into six treatment groups
136 labelled group ‘A to F’. All treatments were administered orally. Groups A to C had 3 animals
137 each. Group A received 10mg/kg of extract while groups B and C received 100 and 500mg/kg
138 respectively of extract. Groups D, E and F, each containing just one mice, received 1600, 2900
139 and 5000mg/kg of extract respectively. The animals were observed for signs and symptoms of
140 toxicity including mortality for 24 h after treatment and findings adequately recorded.

141 **2.7 Induction of hyperglycemia**

142 “The Albino rats were fasted overnight (12-14 hr) and their weight and fasting blood glucose
143 level recorded with a weighing balance and glucometer respectively. hyperglycemia was induced
144 by a single intraperitoneal injection (1 ml/kg) of freshly prepared alloxan monohydrate solution
145 (Sigma-Aldrich, USA), (120 mg/kg body weight). 10% glucose solution bottles were kept in
146 their cages for the next 24hrs to prevent hypoglycemia and also food was given to the animals 30

147 min after administration of alloxan. After 48hrs of alloxan injection, plasma blood glucose level
148 of each animal was determined by taking blood from the tail and animals with a fasting blood
149 glucose level above 200 mg/dl were included in the study while those that did not develop more
150 than 200mg/dl glucose levels were excluded from the study” [18, 19].

151

152 **2.8 Experimental design**

153 **2.8.1 Acute anti-hyperglycemic study**

154 The animals weighing (138.90 -190.19 kg) were divided into 5 groups (A-E) clearly
155 differentiated using coloured permanent markers for the evaluation of fasting blood glucose level
156 with 5 animals in each group. They were treated with the plant extracts two days after
157 administration of alloxan, excluding the hyperglycemic control groups. At intervals of 0, 1, 3, 6,
158 9, 12 and 24hr after treatment administration, blood samples were drawn from the tail of each
159 animal, and the blood glucose levels checked. Groups A, B and C were given 100, 200 and 400
160 mg/kg respectively of the plant extracts, while Group D and E were administered 5 mg/kg
161 Glibenclamide (standard) and 5ml/kg distilled water (negative control) respectively. All
162 treatments were administered orally. Results were recorded and analyzed.

163 **2.8.2 Sub-acute anti-hyperglycemic study**

164 Fresh animals weighing (154.02-186.95kg) were used and also divided into 5 groups with 5
165 animals in each group. Same procedure as followed in acute anti-hyperglycemic study was used
166 except that fasting blood glucose level were checked on 0, 3rd, 7th, 10th and 14th day. The results
167 were carefully recorded and analyzed.

168

169 **2.9 Statistical analysis**

170 Numerical data obtained from the study were expressed as the mean values \pm Standard Error of
 171 Mean (N=5). Differences among means of control tested groups were determined using one-way
 172 ANOVA, followed by Dunnett's multiple comparison test. A probability level of less than 5%
 173 ($p < 0.005$) was considered significant.

174 3. RESULTS

175 3.1 Preliminary phytochemical Screening

176 The preliminary phytochemical screening tests carried out on the methanol leaf extract of
 177 *Jatropha tanjorensis* showed the presence of carbohydrates, reducing sugars, alkaloids, cardiac
 178 glycosides, saponins, tannins, flavonoids, and fixed oils [Table1].

179

180 **Table 1: Result of the phytochemical analysis of methanol leaf extract of *J. tanjorensis*.**

Test	Class of compounds	Results
Molisch	Carbohydrate	+
Fehling's	Reducing sugar	+
Dragendorf	Alkaloids	+
Frothing	Saponins	+
Ferric chloride	Tannins	+
Ammonium solution	Flavonoids	+
Paper translucency	Oils	+
Salkowski	Aglycone and steroidal cardiac glycosides	+

181 KEY: + ----- Present - ----- Absent

182

183

184 3.2. Fresh Leaf Microscopy and Chemomicroscopy

185

186 **Table 2: Summary of the fresh leaf microscopy of *J. tanjorensis***

Epidermal cell	The epidermal cells are polygonal to irregular in shape on the upper surface but irregularly shaped with wavy/undulated anticlinal cell walls on the lower surface.
----------------	---

Stomata type	The leaf is amphistomatic (stomata occur both on the upper and lower surfaces but more on the lower surface) with anomocytic type of stomata (lack of subsidiary cells)
Trichome	Covering unicellular trichomes are scarcely present.
Stomata density (mm^{-2})	Upper surface: 5.88 ± 0.00 ; Lower surface: 60.29 ± 1.47
Stomata length (μm)	Upper surface: 24.91 ± 0.00 ; Lower surface: 28.75 ± 0.74
Stomata width (μm)	Upper surface: 17.18 ± 0.00 ; Lower surface: 18.13 ± 0.26
Stomata index (%)	Upper surface: 1.18 ± 0.00 ; Lower surface: 14.10 ± 0.47
Stomata size (μm^2)	Upper surface: 427.95 ± 0.00 ; Lower surface: 521.55 ± 18.45
Palisade ratio	8.00 ± 0.41 (7 – 9)

187 Values are mean \pm SEM, n=4

188

189 **Table 3: Result of Chemomicroscopy of *J. tanjorensis* Leaf**

Parameter	Reagent(s)	Result
Starch grains	Iodine solution	Present
Lignified tissues	Conc. Hydrochloric acid + Phloroglucinol	Present
Calcium oxalates	Iodine solution Conc. Sulphuric acid	Present; Prism and druse shape
Tannin	Ferric chloride	Present
Cellulose	Zinc chloride; Conc. Sulphuric acid	Present
Gum/Mucilage	Ruthenium red	Absent
Protein	Biuret reagent; Nihydrin	Present
Oil globules	Sudan IV reagent	Present

190

191

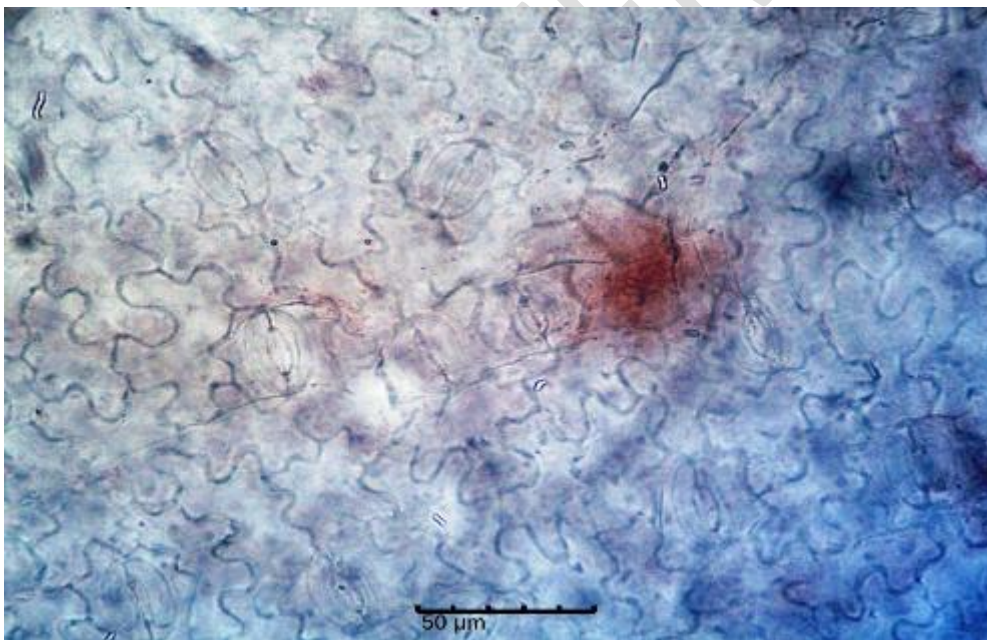
192

193



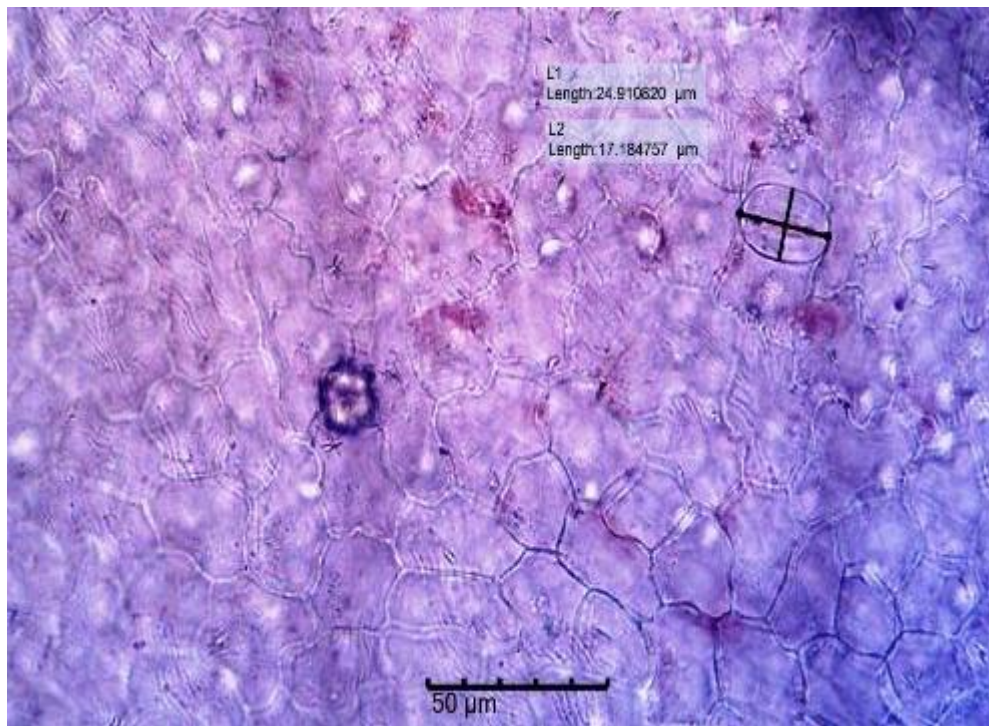
194

195 **Figure 1:** Upper surface of te leaf of *J. tanjorensis* showing polygonal irregularly
196 shaped epidermal cells. Stomata are scarcely distributed and are of anomocytic type (X 400).
197



198

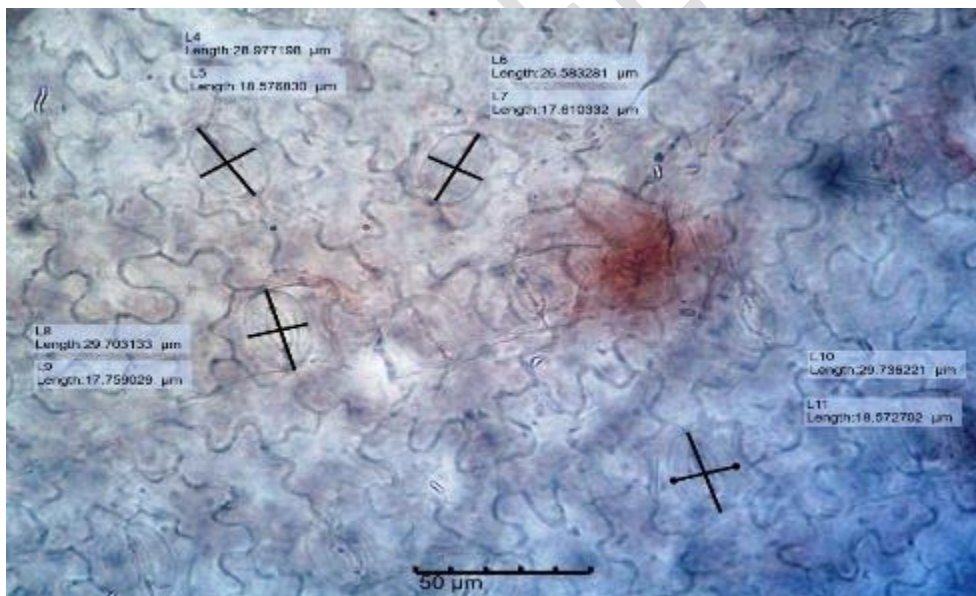
199 **Figure 2:** Lower epidermal surface of the leaf of *J. tanjorensis* showing irregularly
200 shaped epidermal cells with wavy cell walls and anomocytic stomata (X 400)
201



202

X400

203 **Figure 3:** Quantitative measurement of the stomata on upper epidermal surface
 204 of the leaf of *J. tanjorensis* (X 400)
 205



206

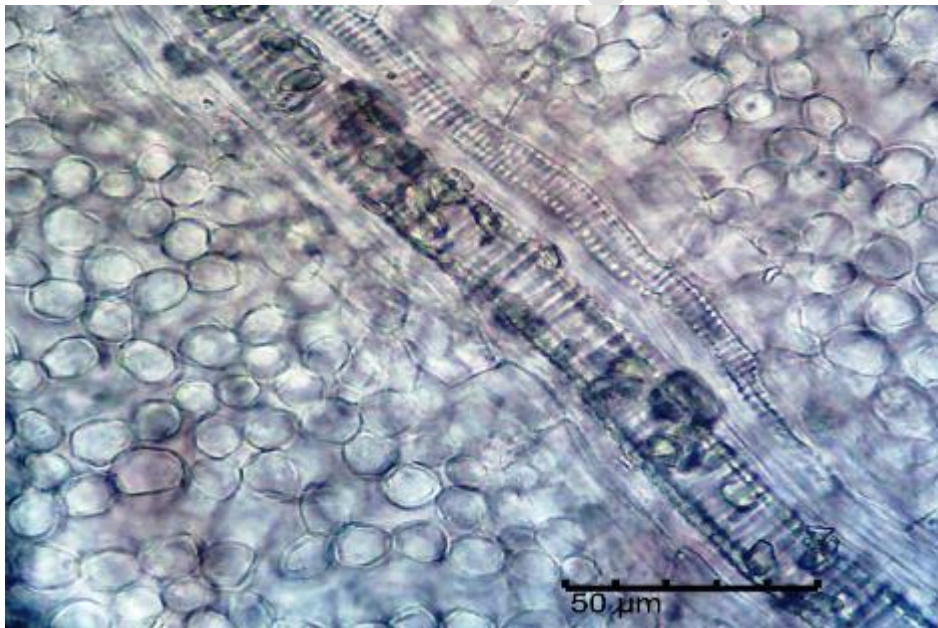
207 **Figure 4:** Photomicrograph of the Quantitative measurement of the stomata
 208 on lower epidermal surface of the leaf of *J. tanjorensis* (X 400)
 209



210

211 **Figure 5:** Chemomicroscopy of the leaf powder showing a bundle of lignified
212 vessel and fibre elements (X 400)

213

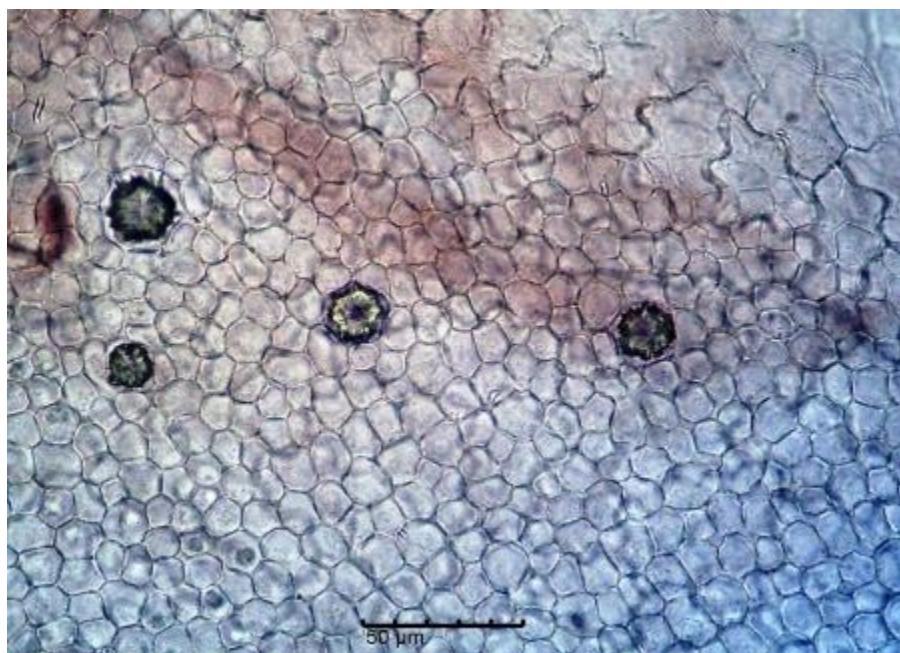


214

215 **Figure 6:** Photomicrograph of the leaf fragment showing arrangement of the
216 palisade cells and vascular bundle within the veins of *J.tanjorensis* (X 400)

217

218



219

220 **Figure 7:** Photomicrograph of the leaf fragment showing the palisade cell,
221 epidermal cells and some sphaeraphides (calcium oxalate) (X 400)

222

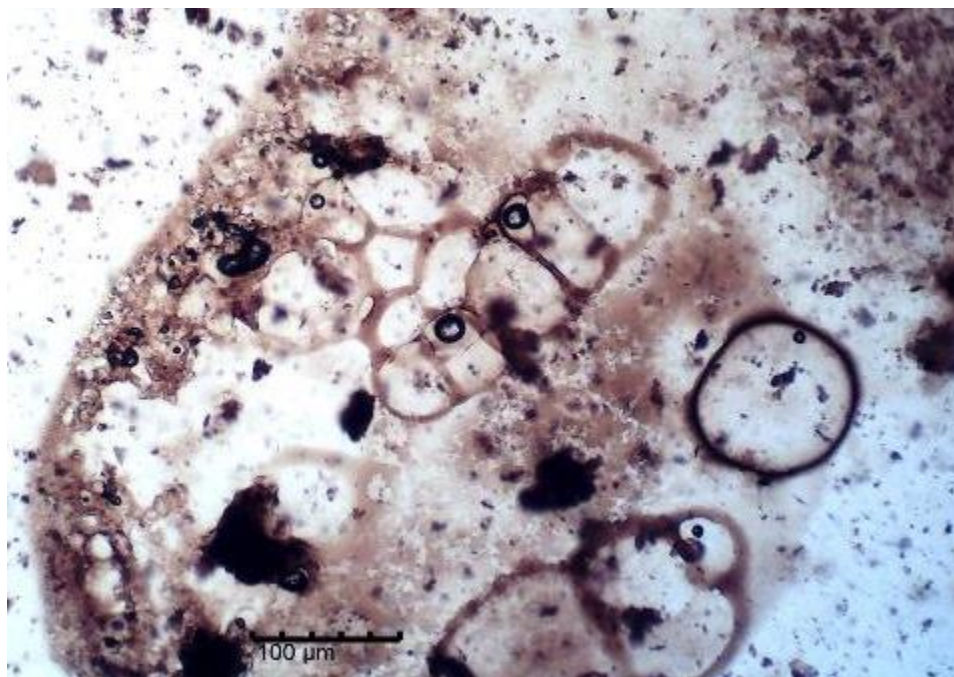
223



224

225 **Figure 8:** Photomicrograph of the leaf powder showing a fragment of unicellular
226 Trichome (X 400)

227



229

230 **Figure 9:** Photomicrograph of the leaf powder showing groups of secretory
231 tissues (most likely laticiferous tissues) (X 400)

232



233

234 **Figure 10:** Transverse section of the leaf of *J. tanjorensis* (X 400)

235

236 3.3. Physicochemical Studies

237 Table 4 shows the values obtained for the analytical standards tested for. These include;
238 Moisture content, ash values and Extractive values.

239

240 **Table 4: Result of Analytical standards of the leaf powder of *J. tanjorensis***

S/N	Parameter	Value (% w/w)
1.	Moisture content (LOD)	5.67 ± 0.15
Ash values		
2.	Total ash	8.39 ± 0.04
3.	Water soluble ash	3.91 ± 0.05
4.	Acid insoluble ash	0.72 ± 0.00
Extractive values		
5.	Hexane soluble extractive value	8.50 ± 0.06
6.	Ethylacetate soluble extractive value	6.19 ± 0.04
7.	Alcohol soluble extractive value	13.51 ± 0.07
8.	Water soluble extractive value	15.75 ± 0.03

241 Values expressed in mean ± SEM, n= 3.

242

243

244

245 3.4 Results for anti-hyperglycemic Studies of *J.tanjorensis*

246 The results of the acute anti-hyperglycemic study [Tables 5 and 6] and Sub-acute studies [Tables
247 7 and 8] are shown below.

248 **Table 5: Results of acute anti-hyperglycemic activity of methanol extract of *Jatropha***
249 ***tanjorensis***

Treatment group	Dose (mg/kg)	Blood glucose level (mg/dl)
-----------------	--------------	-----------------------------

		0h	1h	3h	6h	9h	12h	24h
Extract	100	310.25±66.70	181.00±41.80	162.50±51.50*	145.25±27.60*	119.25±18.03*	170.00±71.70*	78.75±10.50*
	200	432.75±92.80	277.25±11.4	205.50±59.10*	146.00±51.40*	98.00±19.20*	84.75±8.24*	65.75±5.51*
	400	486.00±50.70	353.25±86.80	336.25±92.80	99.25±7.32*	90.50±11.20*	67.25±2.75*	62.75±2.43*
Gb	5	552.00±33.25	394.25±79.50	282.50±33.40*	185.00±75.72*	103.75±23.14*	83.00±14.50*	69.00±4.18*
Distilled water (5 ml/kg)	-	518.00±64.69	488.00±68.60	532.25±41.20	580.25±19.75	574.50±25.50	593.00±7.00	600.00±.00

250 Values are mean ± Standard Error of Mean (N=4) with level of significance, *p<0.05 level
251 compared with the negative control (one-way ANOVA followed by Dunnett's multiple
252 comparison test)

253
254

255 **Table 6: Results of the percentage blood glucose reduction of methanol extract of *J.***
256 ***tanjorensis* in the acute anti-hyperglycemic study**

257

Treatment group	Dose mg/kg	Blood glucose level reduction (%)					
		1h	3h	6h	9h	12h	24h
Extract	100	41.66	47.62	53.18	61.56	45.21	74.62
	200	35.93	52.51	66.26	77.35	80.42	84.81
	400	27.31	30.81	79.58	81.38	86.16	87.09
Gb	5	28.52	48.82	66.49	81.20	84.96	87.50
Distilled water (5 ml/kg)	-	5.79	-2.75	-12.02	-10.91	-14.48	-15.83

258 Gb – Glibenclamide

259 **Table 7: Results of the sub-acute anti-hyperglycemic effect of the crude methanol extract of**
 260 ***J. tanjorensis***

Treatment group	Dose mg/kg	Blood glucose level (mg/dl)				
		Day 0	Day 3	Day 7	Day 10	Day 14
Extract	100	365.67±37.17	334.00±76.45	322.67±44.71*	107.00±05.50*	110.00±08.16*
	200	492.75±58.67	318.50±01.00	218.75±58.25*	121.00±29.54*	229.25±57.06*
	400	432.20±87.65	237.75±68.56*	96.60±05.71*	86.40±07.57*	84.20±09.25*
Gb	5	600.00±.00	380.50±86.62	196.25±65.53*	85.25±12.43*	87.00±16.52*
Distilled water (5 ml/kg)	-	459.50±48.08	494.50±41.36	566.00±28.32	578.00±16.55	600.00±.00

261 Values are mean ± SEM (N=4) *p<0.05 level compared with the negative control (one-way
 262 ANOVA followed by Dunnett's multiple comparison test), Gb (Glibenclamide)

263

264

265

266

267 **Table 8: Results of the percentage blood glucose reduction of the methanol extract of *J.***
 268 ***tanjorensis* in sub-acute anti-hyperglycemic study**

Treatment groups	Dose mg/kg	Blood glucose level reduction (%)			
		Day 3	Day 7	Day 10	Day 14
Extract	100	8.66	11.76	70.74	69.92
	200	35.36	55.61	75.44	53.48
	400	44.99	77.65	80.01	80.52
Gb	5	36.58	67.29	85.79	85.50
Distilled water (5 ml/kg)	-	-7.62	-23.18	-25.79	-30.58

269 Gb - Glibenclamide

270

271

272

273

274

275

276 4. DISCUSSION

277 The bioactive constituents present in the plant (Table 1) may be responsible for the observed
278 anti-hyperglycemic and other therapeutic effects of the plant. However, the alkaloids, saponins
279 and flavonoids contained in the plant extract have been verified to possess the anti-
280 hyperglycemic effects by enhancing the activity of hexokinase and phosphofructokinase,
281 resulting in glucose transport, carbohydrate digestion and absorption and also involved in Insulin
282 secretion respectively [20].

283 In recent times, however, there has been an increase in consciousness of the need for
284 standardization of medicinal plant extracts, especially for those with potential therapeutic uses
285 [21-23]. Microscopic features (Table 2 and 3, Figures 1-10) could be used pharmacognostically
286 for identification and differentiation. The water-soluble extractive value (Table 4) indicated the
287 presence of water-soluble matters such as sugars, amino acids and vitamins derived from plants
288 while the alcohol soluble extractive values indicate the presence of polar compounds, also
289 indicating that this plant sample can best be extracted with an alcohol-based solvent. The
290 moisture content obtained was 5.67 % w/w which is low when compared to the African
291 Pharmacopoeia limit of moisture content for vegetable drug (8 – 14 % w/w) and may less likely
292 degrade due to hydrolytic reactions and enzymatic activation during storage. The ash value
293 indicates the presence of inorganic ions and used to determine the quality and purity of crude
294 drug. The values show that there was neither adulteration nor substitution. High ash value
295 indicates the presence of impurities.

296 There was no mortality or any signs of behavioral changes or toxicity observed after oral
297 administration of *Jatropha tanjorensis* up to the dose of 5000 mg/kg body weight in mice used
298 for the study.

299 The result of the acute anti-hyperglycemic study shows that low dose (100 mg/kg) of the
300 methanol extract of *Jatropha tanjorensis* has quick on set anti-hyperglycemic action (41.66 %)
301 compared to the other doses and the standard drug (28.52 %) when compared with the negative
302 control [Tables 5 and 6]. At the sixth hour, 100, 200 and 400 mg/kg of the extract and the
303 standard drug have significant anti-hyperglycemic effect of 53.18, 66.26, 79.58 and 66.49 %
304 respectively. The result of the sub-acute anti-hyperglycemic study shows that there is a dose-
305 dependent anti-hyperglycemic effect of the methanol extract of *Jatropha tanjorensis* when
306 compared with negative control group which received distilled water. The 400 mg/kg gave the
307 highest anti-hyperglycemic effect (44.99 %) with the statistical significance of 0.035 when
308 compared to the standard drug (36.58) on day three (Tables 7 and 8). On day fourteen, all the
309 doses studied had outstanding significant effect. The hyperglycemic model was used to screen
310 the anti-hyperglycemic activity of plant extracts. “An excessive amount of glucose in the blood
311 induces the insulin secretion. This secreted insulin will stimulate peripheral glucose consumption
312 and control the production of glucose through different mechanisms” [24].
313 “The effect of Glibenclamide, a standard oral anti-hyperglycemic used in this study on blood
314 glucose reduction, has been attributed to enhanced beta cells of the pancreas resulting in
315 secretion of larger amounts of insulin. So, the mechanism behind this anti-hyperglycemic activity
316 of plant extracts involves an insulin-like effect, probably through peripheral glucose
317 consumption or enhancing the sensitivity of beta cells to glucose, resulting in increased insulin
318 release” [25]. “Alloxan produces hyperglycemia by a selective cytotoxic effect on pancreatic beta
319 cells. One of the intracellular phenomena for its cytotoxicity is through generation of free
320 radicals demonstrated both *in vivo* and *in vitro*” [26]. Administration of glucose solution after

321 inducing hyperglycemia with alloxan was to prevent the hypoglycemic shock associated with
322 alloxan.

323 5. CONCLUSION

324 The methanol leaf extract showed that *Jatropha tanjorensis* possesses anti-hyperglycemic
325 properties at various doses studied. This research support the inclusion of this plant in traditional
326 anti-diabetic preparations and shows appreciable results to support the traditional claims of the
327 plant extract. The pharmacognostic standards obtained will assist in the preparation of a
328 monograph of *Jatropha tanjorensis* for proper identification of the plant.

329 6. Ethical Approval:

330 Animal Ethic committee approval has been collected and preserved by the author(s)

331 REFERENCES

- 332 1. Ross MST, Brain KR. (1977) An introduction to phytopharmacy. 1st Ed. London: Pitman
333 Medical Publishing.
- 334 2. Barcelo A, Rajpathak S. (2001) Incidence and prevalence of diabetes mellitus in the
335 Americas. Pan Am J Public Health. 10(5):300-8.
- 336 3. International Diabetes Federation (IDF) Diabetes Atlas-7th Edition, 2015.
337 <http://www.diabetesatlas.org/resources/2015-atlas.html>
- 338 4. International Diabetes Federation (IDF) Diabetes Atlas-9th Edition, 2019.
339 <http://www.diabetesatlas.prg/resources/2019-atlas.html>
- 340 5. Koski RR. (2006) Practical review of oral antihyperglycemic agents for type 2 diabetes
341 mellitus. The Diabetes Educator. 32(6):869-76.
- 342 6. Gupta R, Bajpai GK, Johri S, Saxena AM. (2008) An overview of Indian novel
343 traditional medicine plants with anti-diabetic potentials. African Journal of Traditional,
344 Complementary and Alternative Medicine. 5(1): 1-17.
- 345 7. Iwalewa EO, Agbani EO. (2005) Proantioxidant effects and cytoprotective potentials of
346 nine edible vegetables in southwest Nigeria. J. Med. Food. 8:539_544.
- 347 8. Prabakaran AJ, Sujatha M. (1999) *Jatropha tanjorensis* Ellis and Saroja, a natural
348 interspecific hybrid occurring in tamilnadu. India Gen. Res. Crop. 46:213-218.
- 349 9. Oboh FOJ, Masodje HI. (2009) Nutritional and antimicrobial properties of *Jatropha*
350 *tanjorensis* leaves. American-Eurasian Journal of Scientific Research. 4(1): 7-10.

- 351 10. Omoregie ES, Osagie AV. (2007) Phytochemical screening and anti-anemia effect of
352 *Jatropha tanjorensis* leaf in protein malnourished rat. Plant Archives, 7 No. 2, 509-516.
- 353 11. Esimone CO, Rita-Marie L, Omobuwa JO. (2008) Single cycle vector based antiviral
354 screening assays for evaluation of potential anti-HIV medicinal plants.
355 Phytopharmacology & Therapeutic Value 1. RPMP. Vol 19: 49-60.
- 356 12. Harbourne J. (1984) Phytochemical Methods, A Guide To Modern Techniques of Plant
357 Analysis. 3rd ed. Chapman and Hall;
- 358 13. Trease GE, Evans WC (1989). Textbook of Pharmacognosy. 13th ed. London: Bailiere-
359 Tindall.343-83.
- 360 14. Nwafor FI, Nwosu MO, Nwafor AZ. (2019) Taxonomic and Ecological Significance of
361 Foliar Epidermal Characters in Four Taxa of *Mussaenda L.* (Rubiaceae) in Nigeria.
362 Annual Research & Review in Biology.32(5): 11_12.
- 363 15. Johansen DA. (1950) Plant Microtechnique. 1st Ed. New York: McGraw-Hill Book
364 Company.230.
- 365 16. Nwosu MO. (2006) Preparation of Botanical Slides. In: Inyang NM, Nwosu MO. and
366 Ivoke N. (eds). Manual of Laboratory Techniques in Biology. University of Nigeria Press
367 Ltd Nsukka, Nigeria.131-163.
- 368
- 369 17. Lorke DA (1983) new approach to practical acute toxicity testing. Arch. Toxicol.
370 ,54: 275-287.
- 371 18. Sabu MC, Subburaju T. (2002) Effects of *Cassia auriculata* Linn. On serum glucose
372 level, glucose utilization by isolated rat hemi-diaphragm. J Ethnopharmacol. 80: 203-206.
- 373 19. Jamal AAB, Issa AAH, Mohammed HHA. (1997) Hypoglycemic and Antihyperglycemic
374 Effects of *Trigonella foenium-graecium* leaf in normal and alloxan-induced diabetic rats. J
375 Ethnopharmacol. 58: 149-155.
- 376 20. van de Venter M, Roux S, Bungu LC, Louw J, Crouch NR, Grace OM, Maharaj V, Pillay
377 P, Sewnarian P, Bhagwandin N, Folb P. (2008) Antidiabetic screening and scoring of 11
378 plants traditionally used in South Africa. J Ethnopharmacol.119: 81_86.
- 379 21. Chanda S. (2014) Importance of pharmacognostic study of medicinal plants: An
380 Overview. Journal of Pharmacognosy and Phytochemistry.2(5):69_73.
381
- 382 22. Thomas S, Patil DA, Patil AG, Naresh C. (2009) Pharmacognostic evaluation and
383 physiochemical analysis of *Averrhoa carambola*. Journal of Herbal Medicine and
384 Toxicology.2:51-59.
385
- 386 23. Kunle OF, Egharevba HO, Ahmadu PO. (2012) Standardization of herbal medicines –A
387 review. Journal of Biodiversity and Conservation. 4(3):101_112.

- 388 24. Andrew JK. (2000) Diabetes. New York: Churchill Livingstone. 1-9.
- 389 25. Muniappan L, Leelavinothan P, Sandhya S, Ramesh B. (2004) Insulin-secretagogue
390 activity and cytoprotective role of the traditional antidiabetic plant *Scoparia dulcis*
391 (Sweet Broomweed). Life Sci. 75.
- 392 26. Yadav S, Vats V, Dhunnoo Y, Grover JK. (2002) Hypoglycemic and antihyperglycemic
393 activity of *Murray koenigii* leaves in diabetic rats. J Ethnopharmacol. 82: 111-116.
- 394
- 395
- 396
- 397
- 398
- 399
- 400
- 401
- 402
- 403
- 404
- 405

UNDER PEER REVIEW