

***In vitro* compatibility of *Trichoderma* and *Bacillus* biocontrol agents with different fungicides**

ABSTRACT

Biocontrol agents are beneficial for plant and soil health and are effective in controlling many plant diseases. These biocontrol agents in combination with fungicides at reduced doses can be more effective than using alone. Keeping this in view, an *in vitro* study was carried out to test the compatibility of commonly used fungicides viz., captan 50 WP, thiram 75% DS, tebuconazole 5.36% FS, carboxin 37.5 % + thiram 37.5 % DS, prochloraz 24.4% + tebuconazole 12.1% w/w EW and thiophanate methyl 450 g/l + pyraclostrobin 50 g/l with two fungal (*Trichoderma asperellum* and *T.viride*) and three bacterial biocontrol agents (*Bacillus subtilis* S4KB5, *B. subtilis* S8KB2 and *B. subtilis* B3). All the five fungicides were found to be compatible with all the biocontrol agents at 100 and 250 ppm. All the fungicides except thiram, showed complete inhibition of *T. asperellum* at 1500 and 2000 ppm and with *T.viride* 100 per cent inhibition is shown by all the fungicides at 1500 and 2000 ppm except carboxin + thiram. With biocontrol isolates, *B. subtilis* S4KB5, *B. subtilis* S8KB2 and *B. subtilis* B3, highest inhibition zone were recorded by prochloraz + tebuconazole (2000 ppm), tebuconazole (2000 ppm) and thiophanate methyl + pyraclostrobin (2000 ppm). Present findings suggest that compatible fungicides can be used with biocontrol agents in an integrated disease management practices for the control of seed and soil borne pathogens.

Key words: Biocontrol agents, Sustainable Agriculture, Compatibility, Fungicides, *Trichoderma* and *Bacillus*.

INTRODUCTION

The productivity of crops is limited by various biotic and abiotic constraints such as incidence of pests and diseases, high/low temperatures, drought, erratic rainfall and unfavourable soil factors etc. Among various biotic stresses, disease caused by both fungal and bacterial phytopathogens are causing major yield losses. Seed and soil borne diseases caused by the different fungal genera such as *Alternaria*, *Colletotrichum*, *Aspergillus*, *Pencillium*, *Helminthosporium*, *Fusarium*, *Verticillium*, *Sclerotium* and *Macrophomina*, etc., are considered as a major limitation causing 50 – 75 per cent yield losses in field and horticultural crops

(Baysal-Gurel *et al.*, 2018) ^[4]. To control these seed and soil borne pathogens, conventional synthetic chemical fungicides and fumigants need to be applied at regular intervals throughout the growing season of the crop. Though the use of fungicides against these pathogens can manage some of these diseases, frequent and indiscriminate use can adversely affect the environment and health and can also lead to development of fungicide resistance (Panth *et al.*, 2020) ^[17].

In the recent years, use of biocontrol agents for the management of seed and soil borne diseases has been advocated widely. Control of plant diseases by the use of antagonistic microorganisms can be an effective means. Microorganisms, such as *Trichoderma* spp. and *Bacillus* spp. have emerged as most powerful biocontrol agents against plant diseases. *Trichoderma* spp. have long been recognized for their biocontrol properties and have become an important tool in agricultural disease management. Similarly, *Bacillus* spp. found in soil are considered as safe alternatives to harmful chemicals, as they exhibit antagonistic activities against various fungal and bacterial phytopathogens (Pandey *et al.*, 2015 ^[16]; Zhao *et al.*, 2015 ^[22]; Kumar *et al.*, 2018 ^[9]). Since the biocontrol agents are applied either to seed or soil or both, there is a possibility of interaction and interference that would arise if fungicides are also applied to the crops. Combined application of biocontrol agents with commonly used fungicides may result either in synergism/antagonism between them. In several disease management practices, the addition of fungicide at a reduced rate in combination with biocontrol agents has significantly inhibited the pathogen compared to biocontrol agents alone (Buck, 2004) ^[5]. Combining antagonists with fungicides eliminates the chance of resistance development and reduces the fungicides application (Kumar *et al.*, 2018) ^[9].

By finding compatible combinations of biocontrol agents and fungicides, sustainable and environmentally friendly strategies can be developed to mitigate the impact of these diseases on crop productivity. The ultimate goal of the research mentioned is to develop effective combination of biocontrol agents and fungicides for controlling seed and soil borne plant diseases. In view of this, laboratory experiments were conducted to test the possibility of compatibility of fungal and bacterial bioagents with fungicides.

MATERIAL AND METHODS

The current investigation was carried out at Department of Plant Pathology, College of Agriculture, Professor Jayashankar Telangana State Agricultural University, Rajendranagar, Hyderabad. Two fungal (*Trichoderma asperellum* and *T.viride*) and three

bacterial biocontrol agents (*Bacillus subtilis* S4KB5, *B. subtilis* S8KB2 and *B. subtilis* B3) were collected from the Department of Plant Pathology. They were tested for their compatibility with six fungicides.

Fungicides

Six fungicides viz., captan 50 WP, thiram 75% DS, tebuconazole 5.36% FS, carboxin 37.5 % + thiram 37.5 % DS, prochloraz 24.4% + tebuconazole 12.1% w/w EW and thiophanate methyl 450 g/l + pyraclostrobin 50 g/l were tested at different concentrations under *in vitro* conditions.

***In vitro* evaluation of compatibility of fungicides with fungal biocontrol agents**

The compatibility of six fungicides at six concentrations i.e., 100 ppm, 250 ppm, 500 ppm, 1000 ppm, 1500 ppm and 2000 ppm with fungal biocontrol agents viz., *Trichoderma asperellum* and *T.viride* was tested using poisoned food technique. Stock solution of 1,00,000 ppm concentration was prepared using 10 ml of sterilized diluted water. Desired concentration of fungicide was obtained by diluting the stock solution using the following formula.

$$C_1V_1 = C_2V_2$$

Where, C_1 = concentration of the stock solution (ppm), V_1 = volume of the stock solution to be added (ml), C_2 = desired concentration (ppm) and V_2 = volume of Potato Dextrose Agar (PDA) in which fungicide is to be amended (ml).

Poisoned medium (20ml) was poured into sterilized Petri plate under aseptic conditions in laminar air flow and were allowed to solidify. Each plate was inoculated in the centre with a five mm diameter of five day old fungal culture disc cut from the periphery of actively growing culture under aseptic conditions and incubated at 28 ± 1 °C in a BOD incubator. Fungal cultured on Potato Dextrose Agar plates with non-poisoned medium served as control. Radial growth of the fungus was recorded daily in the control plate starting from the initiation of the fungal growth in correspondence to treatment plates till the fungal growth was full in control. Per cent inhibition of the biocontrol agent growth over control was calculated using the formula given by Vincent (1947) ^[21].

$$I = \frac{C - T}{C} \times 100$$

Where, I = Per cent growth inhibition,

C = Growth of biocontrol agent in control plate and

T = Growth of biocontrol agent in treatment plate

***In vitro* evaluation of compatibility of fungicides with bacterial biocontrol agents**

The compatibility of six fungicides at six concentrations i.e., 100 ppm, 250 ppm, 500 ppm, 1000 ppm, 1500 ppm and 2000 ppm with bacterial biocontrol isolates viz., *Bacillus subtilis* S4KB5, *B. subtilis* S8KB2 and *B. subtilis* B3 was tested using well diffusion technique (Magaldi, 2004^[10]; Valgas, 2007^[20]). The sterilized Petri plates is poured with nutrient agar medium and allowed to solidify. Overnight culture of bacteria was evenly spread over the media surface by means of sterilized spreader. Thereafter, 5 mm diameter well was made in each agar plate by using sterilized cork borer. The required concentration of fungicide was loaded into the each well (50µl/well) in Petri plates separately with the help of micropipette. The petri plates without fungicidal suspension served as control. The plates were then incubated at 28± 1°C for 48hours and observed for the inhibition zone. Experiment was replicated thrice. The fungicidal suspension diffuses in the agar medium creating a zone of inhibition in case of incompatibility of bacterial bioagents with the fungicide. The inhibition zone was measured in mm with the help of a scale after 48 h of incubation. Absence of inhibition zone indicated the compatibility with respective bacterial strains.

RESULTS AND DISCUSSION

The present experiment was aimed to find the compatibility of biocontrol agents viz., *T. asperellum*, *T. viride*, *Bacillus subtilis* S4KB5, *B. subtilis* S8KB2 and *B. subtilis* B3 with different fungicides.

***In vitro* evaluation of compatibility of fungicides with fungal biocontrol agents**

Compatibility of six fungicides at different concentrations with fungal biocontrol agent *T. asperellum* are indicated in Table 1 and Figure 1. Among the six fungicides evaluated with *T. asperellum*, captan and thiram showed significantly lowest inhibition on mycelial growth of 88.29 and 90.00 mm respectively and is on par with control. On the contrary, tebuconazole showed significantly highest inhibition of 46.35 per cent with a radial mycelial growth of 48.28

mm at 100 ppm. Fungicide thiram was found to be significantly compatible with *T. asperellum* with minimum per cent inhibition ranging from 0 per cent to 92.15 per cent at 100 to 2000 ppm concentrations respectively, followed by captan with minimum per cent inhibition ranging from 1.9 per cent to 87.98 per cent at all concentrations except at 1500 and 2000 ppm and it was on par with carboxin + thiram at all tested concentrations. Complete inhibition on radial mycelial growth of *T. asperellum* was shown by all the fungicides at 1500 and 2000 ppm except carboxin + thiram. Fungicides, prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin showed 100 per cent inhibition from 500 ppm. Overall, fungicides captan and thiram were more compatible with *T. asperellum* from 100 to 2000 ppm and prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin were least compatible.

At 100 and 250 ppm, both captan and thiram were highly compatible with *T. viride* and showed significantly lowest inhibition with highest mycelial growth of 90 mm and is on par with control. While tebuconazole showed significantly highest inhibition with lowest mycelial growth of 27.44 mm and 24.19 mm at 100 and 250 ppm respectively followed by prochloraz + tebuconazole (39.24 and 27.42 mm). The fungicide carboxin + thiram was significantly compatible with *T. viride* with minimum per cent inhibition ranging from 0 per cent to 93.75 per cent at 100 to 2000 ppm concentrations respectively. All the remaining fungicides viz., captan, thiram, tebuconazole, prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin showed 100 per cent inhibition at 1500 and 2000 ppm (Table 2 and Fig. 2). Overall, fungicides captan and thiram were more compatible with *T. viride* from 100 to 2000 ppm and tebuconazole and prochloraz + tebuconazole were least compatible.

Earlier various studies have been conducted on compatibility of biocontrol agents with fungicides. Similar results have been obtained by other workers. Mclean *et al.* (2001) ^[12] reported that *T. harzianum* was least sensitive to procymidone and captan and most sensitive to mancozeb, tebuconazole and thiram. Bagwan (2010) ^[3] reported that thiram (0.2%) copper oxychloride (0.2%) and mancozeb (0.2%) were found comparatively safer against *T. harzianum* and *T. viride*. Kay and Stewart (1994) ^[7] reported that four fungal antagonists (*Chaetomium globosum*, *Trichoderma harzianum*, *T. viride*, *Trichoderma* spp.) were found insensitive to captan, mancozeb and thiram but were sensitive to benomyl and procymidone. More or less similar results have been found by other workers also (Nallathambi *et al.*, 2009) ^[15]. Different workers have reported chlorothalonil and captan as tolerant for *T. harzianum*

even at higher concentrations up to 2000 mg/ml in spore germination tests (Abdel-Moity *et al.*, 1982 ^[1]; Papavizas *et al.*, 1982 ^[15]; Mishra *et al.*, 2004 ^[13]).

***In vitro* evaluation of compatibility of fungicides with bacterial biocontrol agents**

Six fungicides *viz.*, captan, thiram, tebuconazole, carboxin + thiam, prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin at six concentrations *viz.*, 100 ppm, 250 ppm, 500 ppm, 1000 ppm, 1500 ppm and 2000 ppm were evaluated for their compatibility with three *Bacillus subtilis* isolates (S4KB5, S8KB2 and B3). The results revealed an absence of a distinct inhibition zone around the wells indicating compatibility of the three bacterial biocontrol agents with different concentrations of fungicides. The inhibition zone values significantly differed in all the fungicides evaluated at different concentrations.

At 100 ppm, all the fungicides were significantly compatible with all the *B. subtilis* isolates (S4KB5, S8KB2 and B3) showing zero inhibition except prochloraz + tebuconazole (8.95 mm) and on par with control. Fungicides, captan and thiram showed zero inhibition and were significantly compatible with all three isolates and on par with control. With *B. subtilis* isolate, S4KB5 prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin were recorded significantly highest inhibition zone of 39.18 mm and 36.20 mm respectively at 2000 ppm as indicated in Table 3 and Fig. 3. Overall, fungicides captan and thiram were more compatible with *B. subtilis* isolate, S4KB5 from 100 to 2000 ppm and tebuconazole and prochloraz + tebuconazole were least compatible.

Tebuconazole and thiophanate methyl + pyraclostrobin were recorded significantly highest inhibition zone of 35.74 mm and 35.11 mm respectively and were highly incompatible with *B. subtilis* isolate, S8KB2 at 2000 ppm. Overall, fungicides thiram and carboxin + thiram were more compatible with *B. subtilis* isolate, S8KB2 from 100 to 2000 ppm and prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin were least compatible (Table 4 and Fig. 4).

With *B. subtilis* isolate, B3 prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin were recorded significantly highest inhibition zone of 33.33 mm and 31.71 mm respectively. Overall, fungicides captan and thiram were more compatible with *B. subtilis* isolate, B3 from 100 to 2000 ppm and prochloraz + tebuconazole and carboxin + thiram were least compatible (Table 5 and Fig. 5).

Similar bacteriostatic as well as bactericidal effects of the fungicides with bacterial antagonists were reported earlier by several workers. Mohiddin and Khan (2013) ^[14] reported that fungal and bacterial bioagents are tolerant to fungicides viz., carbendazim, mancozeb, metalaxyl, captan, thiram, and nemacur at lower concentrations and as the concentration increases the bioagents become more sensitive. Singh and Pandey (2020) ^[19] observed that maximum tolerance concentration of mancozeb was 15 µg/ml, vitavax was 0.1 µg/ml, metalaxyl was 1000 µg/ml for *Bacillus* spp. Harsha *et al.* (2023) ^[6] reported that *Bacillus* spp. was found safer with thiophanate methyl + pyraclostrobin, carboxin + thiram and thiamethoxam. It also found that bacterial biocontrol agents were found more tolerant to fungicides than fungi. Kishore and Jacob (1987) ^[8], Aislabie and Jones (1995) ^[2], Mohiddin and Khan (2013) ^[14] made similar kind of observations and concluded that it may be due to the reason that, some bacteria can use chemicals as nutrients and hence can tolerate higher concentrations of chemicals.

CONCLUSION

The present study on fungicide compatibility clearly indicates that all the six fungicides viz., captan, thiram, tebuconazole, carboxin + thiam, prochloraz + tebuconazole and thiophanate methyl + pyraclostrobin during the course of investigation were found to be comparatively more compatible at 100 ppm and 250 ppm against fungal (*T. asperellum* and *T. viride*) and bacterial (*Bacillus subtilis* S4KB5, *B. subtilis* S8KB2 and *B. subtilis* B3) biocontrol agents. It also came to known that lesser concentration of the fungicides had lesser inhibitory effect as compared to higher concentration. However, all the fungicides were comparatively more toxic against all the five biocontrol agents at 1500 ppm and 2000 ppm. It also clearly indicates the selective response of biocontrol agents to fungicides. The variation in the sensitivity of biocontrol agents to fungicides might be due to their inherent ability to degrade them. Further the data on fungicide tolerance helps to select suitable selective fungicides that are compatible with biocontrol agents. Thus, it can be concluded that the use of fungicides at lower concentrations with biocontrol agents for achieving sustainable plant diseases and agroecosystem management is highly recommended.

REFERENCES

- [1] Abd-El Moity, T.H., Papavizas, G.C and Shatla, M.N. 1982. Induction of new isolates of *Trichoderma harzianum* tolerant to fungicides and their experimental use for control of white rot of onion. *Phytopathology*. 72(4): 396-400.

- [2] Aislabie, J and Lloyd-Jones, G. 1995. A review of bacterial-degradation of pesticides. *Soil Research*. 33(6): 925-942.
- [3] Bagwan, N.B. 2010. Evaluation of *Trichoderma* compatibility with fungicides, pesticides, organic cakes and botanicals for integrated management of soil borne disease of soybean [*Glycine max* (L.) Merrill]. *International Journal of Plant Protection*. 3(2): 206-209.
- [4] Baysal-Gurel, F and Kabir, N. 2018. Comparative performance of fungicides and biocontrol products in suppression of *Rhizoctonia* root rot in viburnum. *Journal of Plant Pathology and Microbiology*. 9: 451.
- [5] Buck, J.W. 2004. Combination of fungicides with Phylloplane yeasts for improved control of *Botrytis cinerea* on geranium seedlings. *Phytopathology*. 94: 196-202.
- [6] Harsha, M.K., Daunde, A.T., Bhalerao, P.B and Sakhare, S.S. 2023. Compatibility studies of *Bacillus* spp. with commonly used agrochemicals. *Pharma Innovation Journal*.
- [7] Kay, S.J and Stewart, A. 1994. The effect of fungicides on fungal antagonists of onion white rot and selection of dicarboximide-resistant biotypes. *Plant Pathology*. 43(5): 863-871.
- [8] Kishore, G.M and Jacob, G.S. 1987. Degradation of glyphosate by *Pseudomonas* sp. PG2982 via a sarcosine intermediate. *Journal of Biological Chemistry*. 262(25): 12164-12168.
- [9] Kumar, R., Singh, S.K., Yadav, S., Kumar, R., Choubey, A.K and Kumari, A. 2018. Compatibility of *Trichoderma viride* with different fungicide and organic cake. *Journal of Pharmacognosy and Phytochemistry*. 7(2): 2398-2401.
- [10] Magaldi, S., Mata-Essayag, S., De Capriles, C.H., Pérez, C., Colella, M.T., Olaizola, C and Ontiveros, Y. 2004. Well diffusion for antifungal susceptibility testing. *International journal of infectious diseases*. 8(1): 39-45.
- [11] Malathi, P., Viswanathan, R., Padmanaban, P., Mohanraj, D and Sunder, A.R. 2002. Compatibility of biocontrol agents with fungicides against red rot disease of sugarcane. *Sugar Tech*. 4: 131-136.

- [12] McLean, K.L., Hunt, J and Stewart, A. 2001. Compatibility of the biocontrol agent *Trichoderma harzianum* C52 with selected fungicides. *New Zealand Plant Protection*. 54: 84-88.
- [13] Mishra, A., Sharma, S.D and Patel, S.I. 2004. Cross-tolerance of *Trichoderma harzianum* rifai to fungicides. *Indian Journal of Agricultural Research*. 38(3): 207-211.
- [14] Mohiddin, F.A and Khan, M.R. 2013. Tolerance of fungal and bacterial biocontrol agents to six pesticides commonly used in the control of soil borne plant pathogens. *African Journal of Agricultural Research*. 8(43): 5272-5275.
- [15] Nallathambi, P., Umamaheswari, C., Thakore, B.B.L and More, T.A. 2009. Post-harvest management of ber (*Ziziphus mauritiana* Lamk) fruit rot (*Alternaria alternata* Fr. Keissler) using *Trichoderma* species, fungicides and their combinations. *Crop Protection*. 28(6): 525-532.
- [16] Pandey, N., Singh, M and Gupta, S. 2015. Characterization of *Bacillus* isolates from Pea fields based on IAA production and growth promotion activities. *Annals of Plant Protection Sciences*. 23(2): 313-318.
- [17] Panth, M., Hassler, S.C and Baysal-Gurel, F. 2020. Methods for management of soil borne diseases in crop production. *Agriculture*. 10(1): 16.
- [18] Papavizas, G.C., Lewis, J.A and Moity, T.H.A.E. 1982. Evaluation of new biotypes of *Trichoderma harzianum* for tolerance to benomyl and enhanced biocontrol capabilities. *Phytopathology*. 72(1): 126-132.
- [19] Singh, M and Pandey, N. 2020. Evaluation of antifungal activity of *Bacillus* spp. against *Fusarium oxysporum* and *Rhizoctonia solani* in Chick pea (*Cicer arietinum* L.). *Annals of Plant Protection Sciences*. 28(1): 33-37.
- [20] Valgas, C., Souza, S.M.D., Smânia, E.F and Smânia Jr, A. 2007. Screening methods to determine antibacterial activity of natural products. *Brazilian Journal of Microbiology*. 38: 369-380.
- [21] Vincent, J.M. 1947. Distortion of fungal hyphae in the presence of certain inhibitors. *Nature*. 159(4051): 850-850.

- [22] Zhao, L., Xu, Y., Lai, X.H., Shan, C., Deng, Z and Ji, Y. 2015. Screening and characterization of endophytic *Bacillus* and *Paenibacillus* strains from medicinal plant *Lonicera japonica* for use as potential plant growth promoters. *Brazilian Journal of Microbiology*. 46: 977-989.

UNDER PEER REVIEW

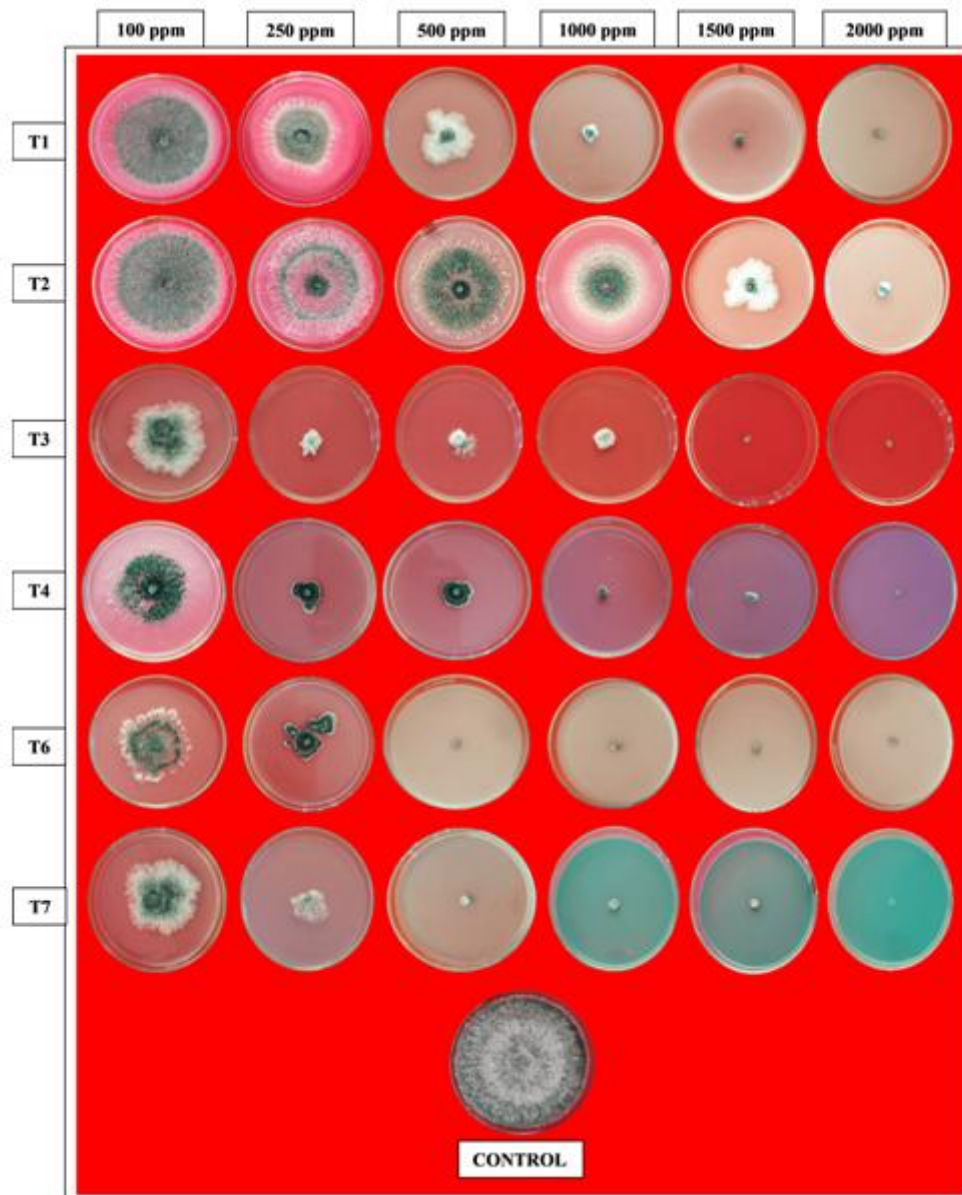
Table 1. Compatibility of fungicides with *T. asperellum* under *in vitro* conditions

Fungicide/ Concentration (ppm)	Radial growth (mm)							Per cent inhibition						
	100	250	500	1000	1500	2000	Mean	100	250	500	1000	1500	2000	Mean
Captan	88.29 ^a	60.33 ^c	30.56 ^c	10.82 ^c	0.00 ^c	0.00 ^c	31.67 ^c	1.90 ^c (1.62)*	32.97 ^b (5.83)	66.04 ^c (8.19)	87.98 ^c (9.43)	100.00 ^a (10.05)	100.00 ^a (10.05)	64.82 ^c (7.53)
Thiram	90.00 ^a	86.55 ^b	79.61 ^b	66.55 ^b	34.26 ^b	7.07 ^b	60.67 ^b	0.00 ^c (1.00)	3.83 ^d (2.14)	11.55 ^d (3.47)	26.06 ^d (5.19)	61.93 ^b (7.93)	92.15 ^b (9.65)	32.59 ^d (4.90)
Tebuconazole	48.28 ^d	19.54 ^f	17.46 ^d	8.94 ^c	0.00 ^c	0.00 ^c	15.70 ^{de}	46.35 ^a (6.88)	78.29 ^a (8.90)	80.60 ^b (9.03)	90.06 ^b (9.54)	100.00 ^a (10.05)	100.00 ^a (10.05)	82.55 ^b (9.08)
Carboxin + Thiram	54.62 ^b	24.19 ^e	21.19 ^d	6.53 ^c	0.00 ^c	0.00 ^c	17.75 ^d	39.31 ^b (6.35)	73.13 ^b (8.61)	76.46 ^{bc} (8.80)	92.74 ^b (9.68)	100.00 ^a (10.05)	100.00 ^a (10.05)	80.27 ^b (8.92)
Prochloraz + Tebuconazole	52.38 ^{bc}	33.88 ^d	0.00 ^e	0.00 ^d	0.00 ^c	0.00 ^c	14.38 ^e	41.80 ^b (6.54)	62.36 ^c (7.96)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	84.03 ^a (9.12)
Thiophanate methyl + Pyraclostrobin	49.54 ^{cd}	23.85 ^e	0.00 ^e	0.00 ^d	0.00 ^c	0.00 ^c	12.23 ^e	44.96 ^{bc} (6.78)	73.50 ^b (8.63)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	86.41 ^a (9.27)
Control	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	0.00 ^c (1.00)	0.00 ^d (1.00)	0.00 ^e (1.00)	0.00 ^e (1.00)	0.00 ^c (1.00)	0.00 ^c (1.00)	0.00 ^e (1.00)

Values expressed are mean of three replications;

*Figures in the parenthesis are square root transformed values.

Figure 1. Compatibility of fungicides with *T. asperellum* under *in vitro* conditions



Where, T1 – Captan; T2 – Thiram; T3 – Tebuconazole; T4 – Carboxin + Thiram; T5 – Prochloraz + Tebuconazole and T6 – Thiophanate methyl + Pyraclostrobin.

Table 2. Compatibility of fungicides with *T. viride* under *in vitro* conditions

Fungicide/ Concentration (ppm)	Radial growth (mm)							Per cent inhibition						
	100	250	500	1000	1500	2000	Mean	100	250	500	1000	1500	2000	Mean
Captan	90.00 ^a	90.00 ^a	79.34 ^b	17.64 ^b	0.00 ^c	0.00 ^c	46.16 ^b	0.00 ^e (1.00)*	0.00 ^e (1.00)	11.84 ^d (3.55)	80.40 ^b (9.02)	100.00 ^a (10.05)	100.00 ^a (10.05)	48.71 ^c (5.78)
Thiram	90.00 ^a	90.00 ^a	81.07 ^b	18.99 ^b	0.00 ^c	0.00 ^c	46.68 ^b	0.00 ^e (1.00)	0.00 ^e (1.00)	9.92 ^d (3.26)	78.90 ^b (8.94)	100.00 ^a (10.05)	100.00 ^a (10.05)	48.14 ^c (5.72)
Tebuconazole	27.44 ^e	24.19 ^d	15.71 ^d	0.00 ^c	0.00 ^c	0.00 ^c	11.22 ^d	69.51 ^a (8.40)	73.12 ^a (8.61)	82.55 ^b (9.14)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	87.53 ^a (9.38 ^a)
Carboxin + Thiram	84.35 ^b	35.66 ^b	22.72 ^c	16.25 ^b	8.43 ^b	5.63 ^b	28.84 ^c	6.28 ^d (2.67)	60.37 ^d (7.83)	74.76 ^c (8.70)	81.94 ^b (9.11)	90.63 ^b (9.57)	93.75 ^b (9.73)	67.96 ^b (7.94)
Prochloraz + Tebuconazole	39.24 ^d	27.42 ^{cd}	0.00 ^e	0.00 ^c	0.00 ^c	0.00 ^c	11.11 ^d	56.40 ^b (7.58)	69.54 ^{bc} (8.40)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	87.66 ^a (9.36)
Thiophanate methyl + Pyraclostrobin	48.24 ^c	31.73 ^{bc}	0.00 ^e	0.00 ^c	0.00 ^c	0.00 ^c	13.33 ^d	46.40 ^c (6.88)	64.75 ^b (8.11)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	100.00 ^a (10.05)	85.19 ^a (9.20)
Control	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	90.00 ^a	0.00 ^e (1.00)	0.00 ^e (1.00)	0.00 ^e (1.00)	0.00 ^c (1.00)	0.00 ^c (1.00)	0.00 ^c (1.00)	0.00 ^d (1.00)

Values expressed are mean of three replications;

*Figures in the parenthesis are square root transformed values.

Captan	0.00 ^b	0.00 ^b	13.61 ^b	18.62 ^b	23.40 ^d	25.18 ^e	13.47 ^b
Thiram	0.00 ^b	0.00 ^b	11.98 ^b	13.31 ^c	18.27 ^e	28.38 ^d	11.99 ^b
Tebuconazole	0.00 ^b	14.26 ^a	26.43 ^a	28.40 ^a	30.35 ^b	33.19 ^c	22.11 ^a
Carboxin + Thiram	0.00 ^b	16.10 ^a	22.40 ^a	24.49 ^a	26.59 ^c	35.50 ^b	20.85 ^a
Prochloraz + Tebuconazole	8.95 ^a	13.49 ^a	20.34 ^a	25.35 ^a	32.42 ^a	39.18 ^a	23.29 ^a
Thiophanate methyl + Pyraclostrobin	0.00 ^b	12.02 ^a	22.80 ^a	27.46 ^a	29.88 ^b	36.20 ^b	21.40 ^a
Control	0.00 ^b	0.00 ^b	0.00 ^c	0.00 ^d	0.00 ^f	0.00 ^f	0.00 ^c

*Values expressed are mean of three replications.

Table 4. Compatibility of fungicides with *B. subtilis* isolate S8KB2 under *in vitro* conditions

Fungicide/ Concentration (ppm)	Inhibition zone (mm)						
	100	250	500	1000	1500	2000	Mean
Captan	0.00 ^b	0.00 ^c	0.00 ^e	7.76 ^e	18.55 ^f	21.44 ^b	7.96 ^c
Thiram	0.00 ^b	10.79 ^b	19.06 ^b	24.51 ^b	33.14 ^a	35.74 ^a	20.54 ^a
Tebuconazole	0.00 ^b	0.00 ^c	11.94 ^d	17.42 ^d	25.64 ^d	29.43 ^{ab}	14.07 ^b
Carboxin + Thiram	11.40 ^a	13.58 ^a	21.71 ^a	27.33 ^a	31.24 ^b	31.26 ^{ab}	22.75 ^a
Prochloraz + Tebuconazole	0.00 ^b	15.44 ^a	21.18 ^a	27.08 ^a	29.33 ^c	35.11 ^a	21.35 ^a
Thiophanate methyl + Pyraclostrobin	0.00 ^b	0.00 ^c	0.00 ^e	0.00 ^f	0.00 ^g	0.00 ^c	0.00 ^d
Control	0.00 ^b	0.00 ^c	14.00 ^c	20.33 ^c	23.71 ^e	26.52 ^{ab}	14.09 ^b

*Values expressed are mean of three replications.

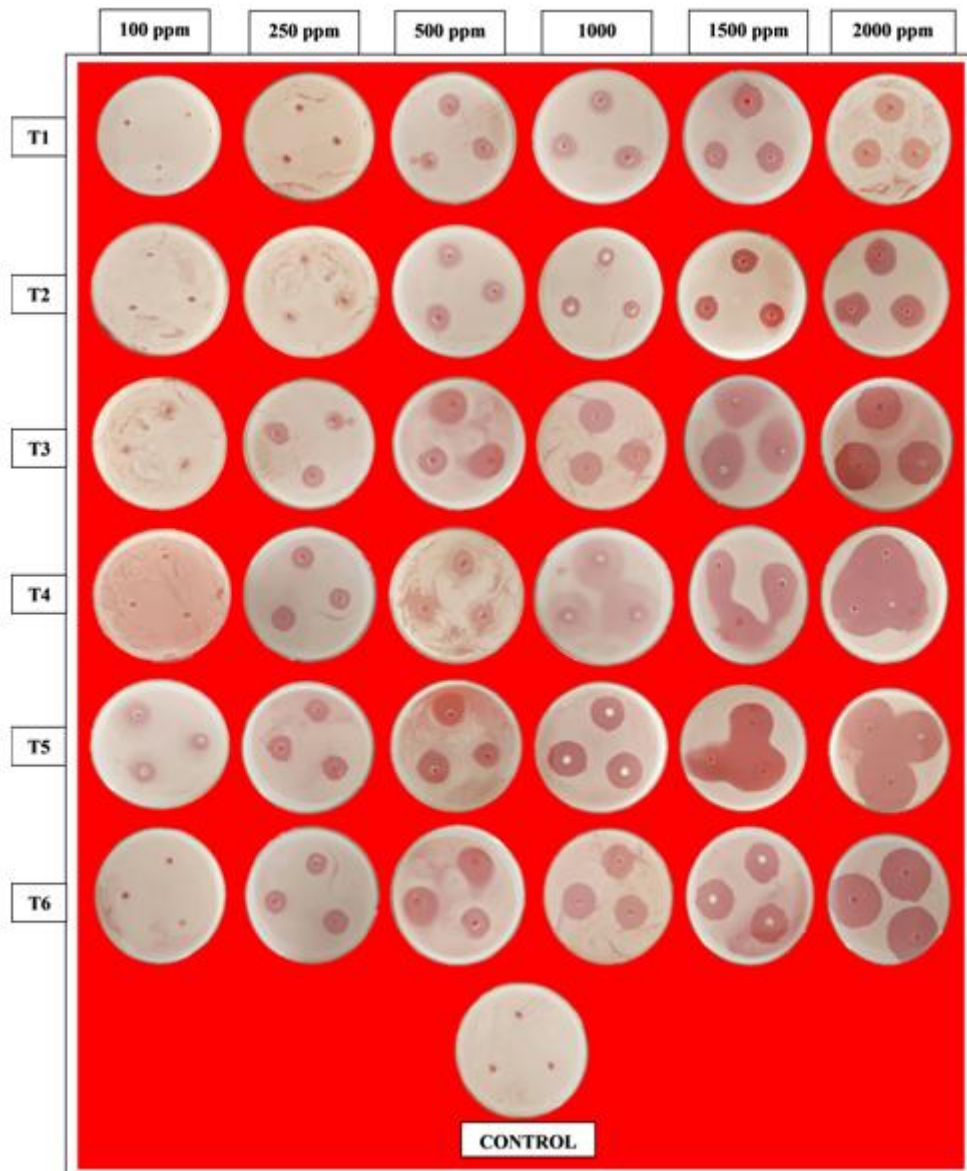
Table 5. Compatibility of fungicides with *B. subtilis* isolate B3 under *in vitro* conditions

Fungicide/ Concentration (ppm)	Inhibition zone (mm)						
	100	250	500	1000	1500	2000	Mean

Captan	0.00 ^b	0.00 ^c	0.00 ^c	17.23 ^b	21.39 ^e	27.71 ^c	11.06 ^c
Thiram	0.00 ^b	0.00 ^c	0.00 ^c	12.41 ^c	23.04 ^{cd}	28.35 ^c	10.63 ^c
Tebuconazole	0.00 ^b	0.00 ^c	14.27 ^b	21.22 ^{ab}	24.01 ^{bc}	31.25 ^b	15.12 ^b
Carboxin + Thiram	0.00 ^b	9.53 ^b	13.05 ^b	20.07 ^{ab}	22.23 ^{de}	28.56 ^c	15.57 ^b
Prochloraz + Tebuconazole	12.35 ^a	15.34 ^a	19.58 ^a	23.31 ^a	27.88 ^a	33.33 ^a	21.96 ^a
Thiophanate methyl + Pyraclostrobin	0.00 ^b	0.00 ^c	11.25 ^b	18.44 ^b	24.95 ^b	31.71 ^b	14.39 ^b
Control	0.00 ^b	0.00 ^c	0.00 ^c	0.00 ^d	0.00 ^f	0.00 ^d	0.00 ^d

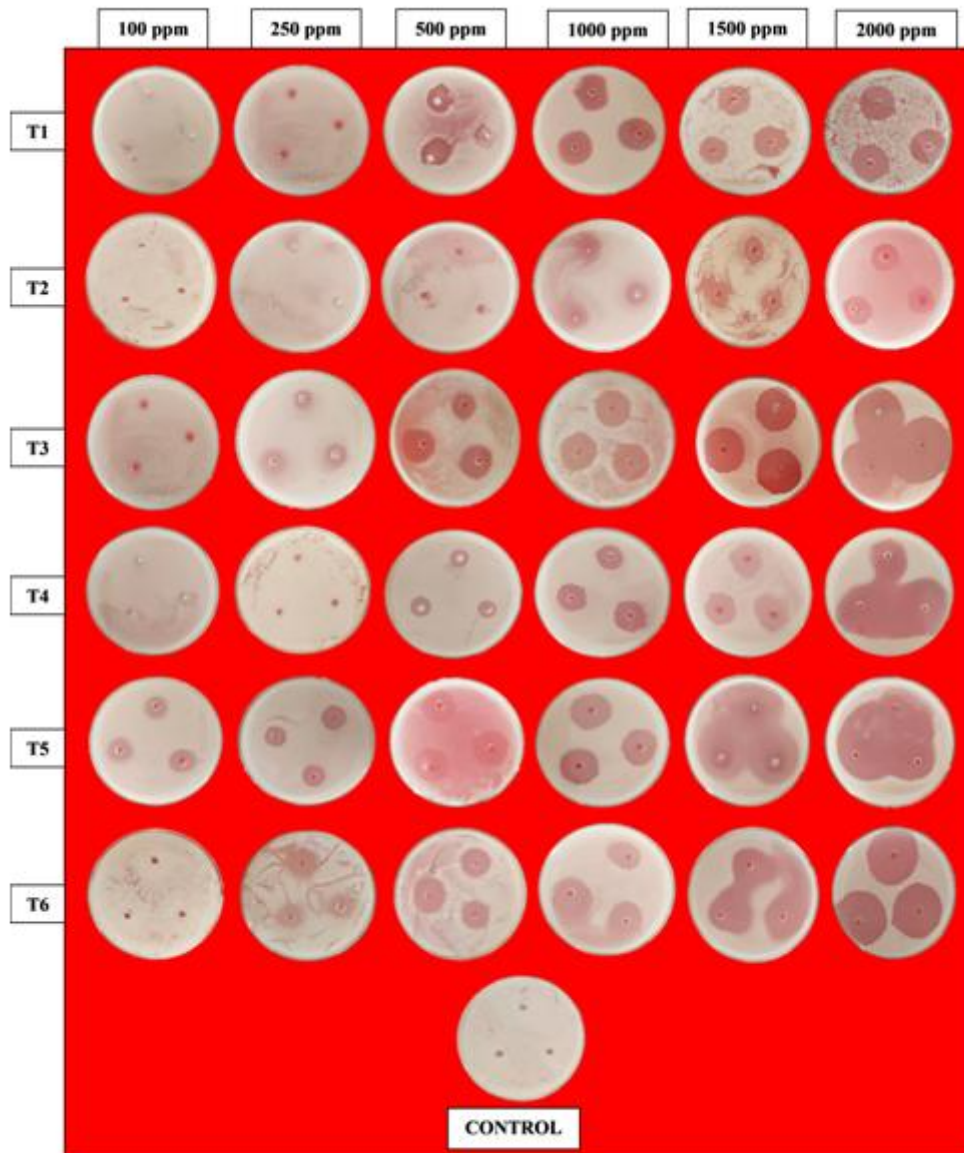
*Values expressed are mean of three replications.

Fig. 3. Compatibility of fungicides with *B. subtilis* isolate S4KB5 under *in vitro* conditions



Where, T1 – Captan; T2 – Thiram; T3 – Tebuconazole; T4 – Carboxin + Thiram; T5 – Prochloraz + Tebuconazole and T6 – Thiophanate methyl + Pyraclostrobin.

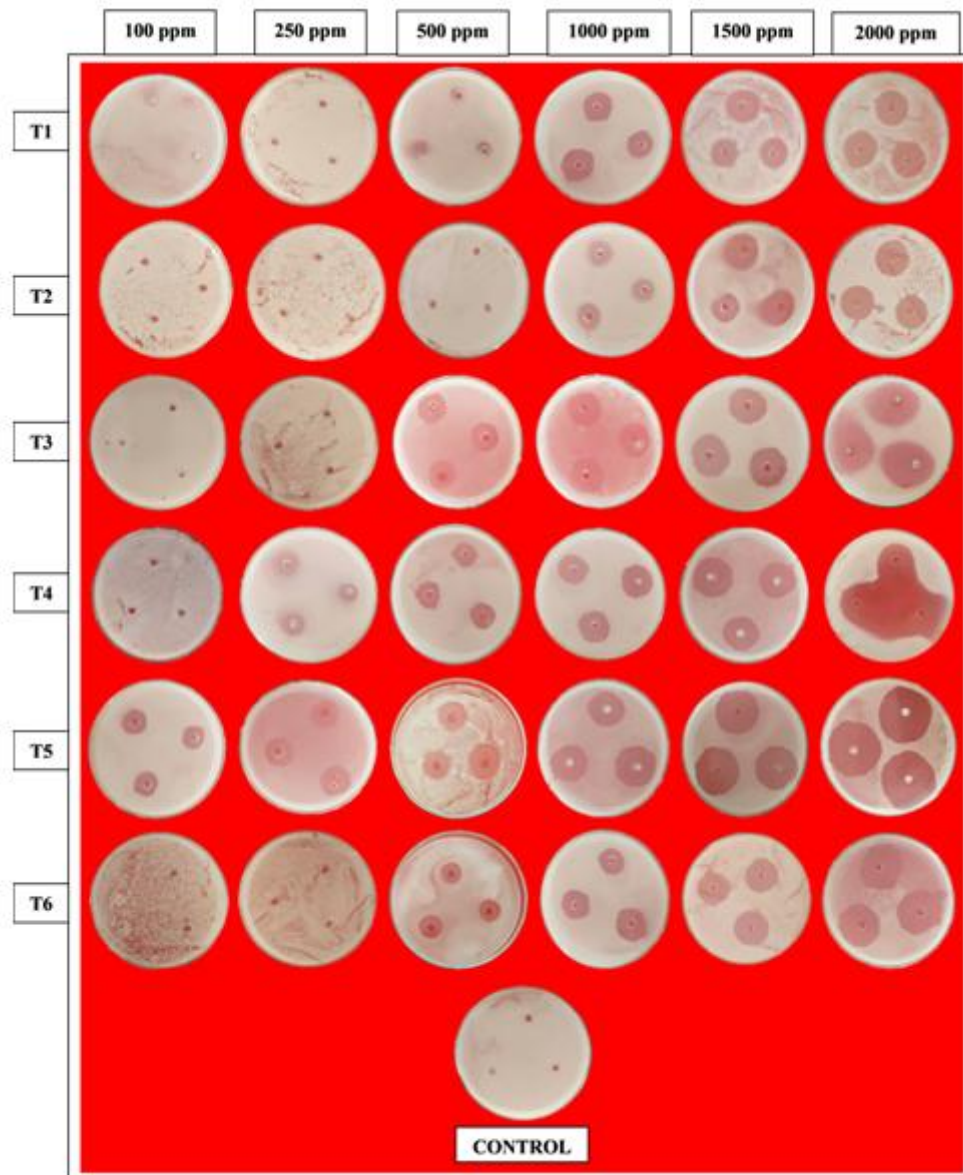
Fig. 4. Compatibility of fungicides with *B. subtilis* isolate S8KB2 under *in vitro* conditions



Where, T1 – Captan; T2 – Thiram; T3 – Tebuconazole; T4 – Carboxin + Thiram; T5 – Prochloraz + Tebuconazole and T6 – Thiophanate methyl + Pyraclostrobin.

Where, T1 – Captan; T2 – Thiram; T3 – Tebuconazole; T4 – Carboxin + Thiram; T5 – Prochloraz + Tebuconazole and T6 – Thiophanate methyl + Pyraclostrobin.

Fig. 5. Compatibility of fungicides with *B. subtilis* isolate B3 under *in vitro* conditions



Where, T1 – Captan; T2 – Thiram; T3 – Tebuconazole; T4 – Carboxin + Thiram; T5 – Prochloraz + Tebuconazole and T6 – Thiophanate methyl + Pyraclostrobin.

Where, T1 – Captan; T2 – Thiram; T3 – Tebuconazole; T4 – Carboxin + Thiram; T5 – Prochloraz + Tebuconazole and T6 – Thiophanate methyl + Pyraclostrobin.