

Physico-chemical properties in rhizosphere of field crop covers during growth stages in a sandy loam soil

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ABSTRACT

The effects of different cultivated plants and their growth periods on some hydraulic and chemical properties of a sandy clay loam soil at the University of Port Harcourt, Teaching and Research Farm was studied. The objective was to understand the behavior of soil properties near the plant environment during the growing period for optimum management, especially of 3fragile soils. Four (4) plants: Maize, Okra, Pepper, and Garden egg were planted to native fallowed soil and the soil properties measured at three (3) growth stages of the crops viz: Establishment, Flowering, and Maturity. Results revealed that the plants did not modify the soil textural class. Maximum activities at the rhizospheres of the plants were found during the flowering stage. Bulk density of 1.36 and 1.34 g cm⁻³ were significantly (p<0.05) low at during the flowering and maturity stages, respectively, in Garden egg soils. The highest saturated hydraulic conductivity (K_{sat}) values of 22.9 and 27 cm h⁻¹, respectively, were significant (p<0.05) during the same periods for garden egg, followed by Maize and Okra. Okra, Pepper and Garden egg significantly (p<0.05) reduced the soil acidity at flowering growth period. Maize contributed significant additions of soil organic matter to the soil at flowering and maturity periods. Maximum removal of available plant nutrients was at flowering stage for maize when more than 80% of the plant had shown tasseling and cob formation. Therefore, monitoring the growth stages of specific plant could help in both nutrient soil management and possible expected changes in soil physical and chemical properties.

Comment [NS2]: In the title you have mentioned sandy loam here you are writing sandy clay loam ..clarify and rewrite

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Key words: soil texture; bulk density; saturated hydraulic conductivity; exchangeable bases; soil acidity

1. INTRODUCTION

Rhizosphere is the region of soil in the vicinity of plant roots in which the chemistry and microbiology is influenced by their growth, respiration, and nutrient exchange. Raising of plants to the soil affects physicochemical properties and biological features of the soil environment close to the growing roots [1]. Deposition of nutrients in the rhizosphere by plant roots can increase microbial biomass, microbial activity, and microbial community different from those found in bulk soil. This phenomenon is referred to as the "rhizosphere effect" [3]. Soil physical conditions, particularly in the rhizosphere, are continually modified by the release of plant root exudates and microbial metabolites [1]. However, the effect of biological exudates on soil physical properties might depend on their physicochemical characteristics and age of the plant [4, 5]

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According to [6, 7], the quantity and physicochemical characteristics of root exudates are determined by the plant species, the age of an individual plant and external factors such as soil structure, presence of microorganisms and nutritional status. The mucilage secreted by plant root, a viscous, high molecular weight, and insoluble polysaccharide-rich material helps in binding of soil particles together to form aggregates which maintain hydraulic conductivity in the soil system [8,9], thus improving soil quality by increasing water infiltration and aeration. [10] also observed a significant increase in the aggregate stability of soil with different substrates such as polygalacturonic acid, modelled soluble exudates and maize root exudate [10].

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Formation of the soil structure occurs when soil is wet and particles are mobile; therefore, rheological measures of wet soil movement under stress are more relevant physically in understanding how mechanical stresses from root growth and exudation affect soil structural development [11]. Root networks enhance soil porosity as well as soil aggregation through

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direct binding of particles and/or secretion of mucilages that help accolade soil particles together [12]. Living roots exude a wide range of compounds into the rhizosphere soil. Some compounds such as malate in root exudates are important in chelating cations which may be phytotoxic to plants [13,14]. Organic exudates such as organic acids can affect availability Fe and P in the rhizosphere and assist in the uptake of nutrients by plants. On the other hand, plant exudates in the rhizosphere can liberate nutrients through dissolution of insoluble mineral phases or desorption from clay and organic matter surfaces into the soil solution [15]. Plant roots can exude organic acids such as malic and citric acids into the rhizosphere which have positive effect in reducing rhizosphere pH, thus, solubilizing phosphorus bound in mineral soils [15].

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Plant secretes 10-30% of photosynthates through the root system into the rhizosphere soil [16], which help to modify the soil physico-chemical properties. Organic matter derived from plant

debris and mucilages and extracellular polymeric substances of plants influences the soil environment, modifying local soil chemistry and properties to provide conditions conducive for specific needs [17] Polysaccharides contribute to mineral particle adhesion, resulting in large transiently stable aggregates [18]. Plant residue is also an important source of carbon, providing soil microbes and soil fauna with substrates for the production of stabilizing material as well as providing physical protection of the soil surface against structure-altering processes like rainfall.

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Understanding relationship between soil rhizosphere, soil microbial community and nutrient cycling is increasingly considered as essential for the ecosystem sustainability. Previous studies have reported that the species, age and abundance of host plants can shape the rhizosphere soil properties and the ability of plant to tolerate stress [19]. However, very few studies have investigated the effect of plant growth stages on properties of soil within the rhizosphere [21]. As a result of this, we speculate that the rhizosphere of field crops at different growth stages can modulate soil physical and chemical properties. Therefore, the objective of this study was to determine the physical and chemical properties of rhizosphere soils under four different plants at seedling, flowering and maturity growth stages. This will improve our knowledge on the effective management of certain crops in order to sustain soil productivity.

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2. MATERIALS AND METHODS

2.1 The Study Area

The study was conducted at the Faculty of Agriculture Teaching and Research Farm, University of Port Harcourt, Choba, Rivers State, Nigeria (Lat. 4° 45'N, Long. 7° 30'E) (Fig. 1). The mean annual rainfall in the area is more than 2700 mm in the months of July and October [22]. Mean annual minimum and maximum temperatures are 22°C and 31°C; while relative humidity is 85-90%. The soil is sandy clay loam, low in total nitrogen, developed from the recent alluvium, in the

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low-lying coastal plains and soil of the Niger Delta [23]. The dry bulk density is about 1.53 g cm^{-3} . Some physical and chemical properties of the site are shown in Table 1.

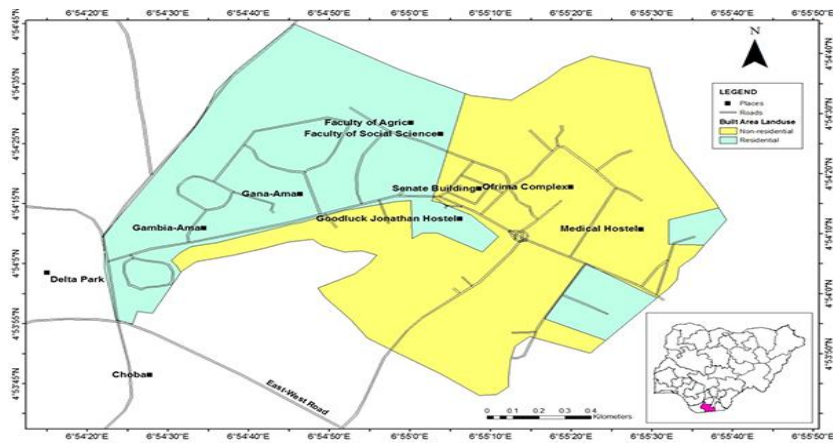


Fig. 1. Map of University Park showing the sampling location

2.2 Experiment Layout, Planting and Sampling

The experiment was laid out in a Randomized Complete Block Design (RCBD) consisting of 4 treatments in three (3) replications giving a total of twelve (12) plots on a 0.225 ha farm with each plot of.....ha. Four (4) crops (maize, okra, pepper and garden egg each) were planted as sole crops during the early rains. The pepper and garden egg were first raised in the nursery for 4 weeks before transplanting to the field, while the maize and okra were planted directly to the field. Seed rateand planting distance (row to row and Plant to plant of Peper and garden egg and duration of all the crops and how many years crops were raised?

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Soil samples were collected in triplicates at 0-20cm depth at the different growth stages of the crops viz: establishment, flowering and maturity. The growth stages were determined at 80%

physiological appearance. Soil samples at the plant rhizosphere were collected in triplicates from two plant stands in each plot, by carefully uprooting and shaking the soil adhering to the roots into a labeled polybag at each growth stages, and bulked to obtain composite samples. Twelve (12) soil samples were collected at each growth stages, placed in labeled polythene bags, and transferred to the laboratory for analysis. Cylindrical metal cores of 6cm x 5 cm were used to collect undisturbed soil samples for determining some of the physical properties.

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2.3 Laboratory Analyses

Particle size distribution was determined by hydrometer method [24] after the removal of gravel from the fine earth material. Total organic carbon was determined by the wet oxidation dichromate method with $H_2SO_4 - K_2Cr_2O_7$, followed by residual titration with 1 N HCl [25], and was converted to organic matter by multiplying the organic carbon values by the Van Bemmelen factor of 1.724. Total nitrogen was determined by the modified macro Kjeldahl digestion method [26]. Soil pH in water was measured with glass electrode using a 1:2.5 soil/water aqueous suspension. Bulk density was determined with core samples using the method of [27]. Total porosity was calculated with core samples using the method [28]. Saturated hydraulic conductivity was measured using the constant head soil core method [29], and calculated by rearranging the Darcy's equation for constant head condition as:

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$$K_{sat} = \frac{V}{AT} \times \frac{L}{\Delta H} \quad (1)$$

where, V is the volume of water collected at steady state (cm^3), L is the length of the soil core (cm), A is the cross-sectional area (cm^2), T is the time (h) and ΔH is the hydraulic head difference (cm).

2.4 Data Analysis

Data collected were analyzed by general analysis of variance using while means were separated by Fisher least significant difference (LSD) at 5% probability level.

Table 1. Some physical and chemical properties of experimental soil before execution of treatments

Soil properties	Plots			
	Maize	Okra	Pepper	Garden egg
Sand (g kg ⁻¹)	564	552	582	582
Silt (g kg ⁻¹)	310	320	310	320
Clay (g kg ⁻¹)	126	128	108	98
Texture	SCL	SCL	SCL	SCL
Total Nitrogen (g kg ⁻¹)	2.2	2.1	2.1	2.1
Organic matter (g kg ⁻¹)	24.8	25.8	24.6	25.6
pH (H ₂ O)	4.3	4.0	4.2	4.0
Available P (m kg ⁻¹)	51.2	55.2	41.8	55.2
Ca ²⁺ (C mol kg ⁻¹)	4.0	4.0	4.1	4.1
Mg ⁺ (C mol kg ⁻¹)	2.81	2.80	2.80	2.81
K ⁺ (C mol kg ⁻¹)	0.26	0.28	0.27	0.21
Base saturation (%)	85.56	85.88	85.33	85.85
Bulk density (g cm ⁻³)	1.53	1.52	1.53	1.53
Total porosity (%)	42.3	42.1	42.1	42.3
Ksat (cm h ⁻¹)	9.61	10.10	9.68	9.77

Ksat- saturated hydraulic conductivity, SCL- sandy clay loam

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3. RESULTS

3.1 Effects on Physical Properties

The mean values of the physical properties of the soil at different growth stages showed that there was no significant ($p > 0.05$) different in the soil textural class during the growing periods.

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The marginal changes in sand, silt and clay contents did not influence the soil texture (Table 2). The sand content ranged between 567 and 582, 575 and 592, and 542 and 592 g kg⁻¹ at establishment, flowering, and maturity growth periods, respectively. Clay content ranged between 118-128 g kg⁻¹. Bulk densities were not significantly different at the establishment growth stage of the crop, but was significantly low ($p < 0.05$) being 1.36 and 1.34 g cm⁻³ during flowering and maturity, respectively in Garden egg plots (Table 1). Total porosity was consistently higher during flowering and maturity growth stages of Okra, with values of 50.2% at flowering and 51.7% at the crop maturity. Saturated hydraulic conductivity (K_{sat}) ranged from 9.94 cm h⁻¹ Okra plots at establishment period to 27 cm h⁻¹ Garden egg at maturity. Significant increase in K_{sat} was consistent in Garden egg plots during the growth periods.

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3.2 Effects on Chemical Properties

The pH value was significantly higher ($p < 0.05$) at 5.9, 5.6 and 5.7 in Okra, Pepper and Garden egg, respectively during establishment growth periods (Table 3). Available P, total N and organic matter (OM) were consistently higher ($p < 0.05$) under Maize plots. At harvesting, available P was in the order of Maize > Okra > Garden egg > Pepper (Table 3). However, low mobility of P is revealed in their very marginal changes during the growing periods. Organic matter was significantly higher in Maize plot at 21.5 g kg⁻¹ during flowering period and reduced by about 25% at harvesting for the same crop. Total N was generally not significantly different across the crops. Exchangeable Ca²⁺ was significantly higher ($p < 0.05$) in pepper plots at 4.5 C mol kg⁻¹ soil during the establishment growth period, whereas, exchangeable K⁺ was generally not significant across the four crops and growth stages. Exchangeable acidity was significantly higher at 1.90 C mol kg⁻¹ in Maize plots, followed by Pepper and Okra, while garden egg plots had the lowest exchangeable acidity value of 1.48 C mol kg⁻¹ (Table 3). Percent base saturation indicating the

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proportion of the exchangeable bases significantly higher ($p < 0.05$) at 80.3, 81.4 and 80.4 in maize, Okra, and Pepper plots, respectively, during the establishment growth period. At harvesting, percent base saturation were in the order of Okra > Maize > Garden egg > Pepper.

Table 2 Some physical properties of the soil at different growth stages of the crops

Soil properties	Maize	Okra	Pepper	Garden egg	LSD (0.05)
Establishment					
Sand (g kg^{-1})	567	582	582	570	NS
Silt (g kg^{-1})	305	290	290	300	NS
Clay (g kg^{-1})	128	118	128	128	NS
Bulk density (g cm^{-3})	1.52	1.50	1.54	1.54	NS
Total porosity (%)	43.7	43.1	44.2	43.6	NS
K_{sat} (cm h^{-1})	10.08	9.94	10.11	11.2	NS
Flowering at 80%					
Sand (g kg^{-1})	592	592	575	582	NS
Silt (g kg^{-1})	280	280	310	300	NS
Clay (g kg^{-1})	128	128	113	118	NS
Bulk density (g cm^{-3})	1.49	1.49	1.50	1.36	0.13*
Total porosity	41.6	50.2	49.2	42.5	4.29*
K_{sat} (cm h^{-1})	20.4	18.1	18.3	22.9	3.18*
Maturity/Harvesting					
Sand (g kg^{-1})	592	562	542	564	NS
Silt (g kg^{-1})	290	310	340	310	4.12*
Clay (g kg^{-1})	118	128	118	126	NS
Bulk density (g cm^{-3})	1.41	1.42	1.45	1.34	0.11*
Total porosity	41.2	51.7	51.4.5	43.6	6.21*
K_{sat} (cm h^{-1})	26.9	20.4	20.2	27.0	3.11*

NS- non-significant at $p > 0.05$, *Significant at $p < 0.05$, K_{sat} - saturated hydraulic conductivity

Table 3 Chemical properties of the soil at different growth stages of the crops

Soil properties	Maize	Okra	Pepper	Garden egg	LSD(0.05)
Establishment					
pH (H ₂ O)	4.9	5.9	5.6	5.7	0.84*
Available P (mg kg ⁻¹)	48.9	31.5	34.1	35.5	9.8*
Total N (g kg ⁻¹)	1.7	1.5	1.3	1.2	NS
Organic matter (g kg ⁻¹)	21.9	17.8	15.4	18.5	4.1
Ca ²⁺ (C mol kg ⁻¹)	3.61	3.72	4.00	3.80	NS
Mg ²⁺ (C mol kg ⁻¹)	2.47	1.86	1.80	1.81	0.35*
K ⁺ (C mol kg ⁻¹)	0.22	0.20	0.20	0.20	NS
Na ⁺ (C mol kg ⁻¹)	0.18	0.18	0.17	0.18	NS
EA (C mol kg ⁻¹)	1.60	1.36	1.52	1.56	NS
Base saturation (%)	80.3	81.4	80.4	77.3	3.51*
Flowering at 80%					
pH (H ₂ O)	4.7	5.6	5.7	5.2	0.45*
Available P (mg kg ⁻¹)	30.52	37.65	37.70	33.85	2.11*
Total N (g kg ⁻¹)	1.5	1.4	1.5	1.5	NS
Organic matter (g kg ⁻¹)	21.5	17.5	19.2	18.7	3.1*
Ca ²⁺ (C mol kg ⁻¹)	4.05	3.80	4.50	4.40	0.57*
Mg ²⁺ (C mol kg ⁻¹)	2.20	2.55	2.22	2.22	NS
K ⁺ (C mol kg ⁻¹)	0.17	0.18	0.45	0.18	NS
Na ⁺ (C mol kg ⁻¹)	1.17	0.17	0.17	0.17	0.90*
EA (C mol kg ⁻¹)	1.90	1.80	1.87	1.48	0.41*
Base saturation (%)	77.65	78.93	79.50	82.38	NS
Maturity/Harvesting					
pH (H ₂ O)	5.7	5.9	5.7	5.8	NS
Available P (mg kg ⁻¹)	36.75	41.00	36.90	39.70	2.75*
Total N (g kg ⁻¹)	1.6	1.6	1.6	1.5	NS
Organic matter (g kg ⁻¹)	17.2	18.0	16.7	17.4	NS
Ca ²⁺ (C mol kg ⁻¹)	3.50	3.70	3.40	3.35	NS
Mg ²⁺ (C mol kg ⁻¹)	2.45	2.85	1.80	2.85	1.08*
K ⁺ (C mol kg ⁻¹)	0.18	0.20	0.20	0.45	NS
Na ⁺ (C mol kg ⁻¹)	0.20	0.17	0.19	0.17	NS
EA (C mol kg ⁻¹)	1.61	1.54	1.88	1.64	0.25*
Base saturation (%)	78.17	81.23	74.87	77.71	4.25*

NS- non-significant at $p > 0.05$, *significant at $p < 0.05$, EA- exchangeable acidity

4.0 DISCUSSION

The non-significant difference in the particle-size distribution of the soil across the different crops and growth periods was not surprising. This further confirmed that soil texture is usually a reflection of the parent materials from which the soil was formed [30]. The crop type and growth stages did not influence the soil texture, despite minor contributions to clay content by organic matter from the crop residues [31]. The soil is generally acidic; due to possible leaching of the salts like Ca and Mg and weathered soils as a result of high rainfall associated with the environment as earlier reported [32]. The highest pH value found at the establishment stage of Okra could be due to the composition of the exudates released by the root at this physiological development. It is possible that this plant roots can exude organics which have positive effect in reducing rhizosphere acidity, and stabilize P bound in soil mineral matrix [15]. The significant reduction in soil bulk density found in Garden egg plots at flowering and harvesting periods could be attributed to the root characteristics of the plant with accompanying organic matter which opened up the soil and allowed for aggregation of the soil [33]. Saturated hydraulic conductivity was rapid in Garden egg soils and further collaborate the works of [34] who found significant influence of root characteristics and density on water movement into and within the soil. The effect of the biological exudates secreted by plant root on soil physical and chemical properties may depend on their physico-chemical characteristics and the quantity of the exudates [4].

Available P, total nitrogen and organic matter were higher in pre-planting plots than at the various growth stages of the crops. The decrease may be attributed to loss of soil materials during cropping and removal by the cultivated crops [35]. The variations in the levels of exchangeable Ca^{2+} , Mg^{2+} , Na^+ and K^+ across the different growth stages could be as a result of the exudates by each plant roots which enhanced the available nutrients for plant uptake. This

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assertion is in agreement with [15] who reported that organic exudates such as organic acids can affect nutrient availability in the rhizosphere and assist in nutrient uptake by plants.

5. CONCLUSIONS

It may be concluded from the present study that different plants variably influenced the estimated physico chemical properties and exhibited markedly different pattern of nutrient removal from the rhizosphere at specific growth stages. Soil bulk density was not significantly reduced until at the flowering and harvesting periods, when the plant roots especially that of Garden egg sufficiently modified the plant environment. Saturated hydraulic conductivity, as well as the total porosity were also influenced by the type of plant and at specific growth stages. In this study, the maize plant specifically removed greater soil nutrients at the flowering stage while exchangeable acidity was lowered at harvesting. Garden egg improved bulk density and saturated hydraulic conductivity of the soil. Therefore, for crop and soil management in highly weather soils of the tropics such as the one used in this study, greater activities should be expected at the plant rhizosphere during the flowering stage of the crop, especially maize and garden egg.

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