

## Original Research Article

### EVALUATION OF THE THERAPEUTIC EFFECTS OF *MORINGAOLEIFERA* SEED

#### OIL ON CADMIUM AND HERBAL ALCOHOLIC BEVERAGE – INDUCED

#### PREFRONTAL CORTEX DAMAGE IN WISTAR RATS.

Comment [NI1]: No point

#### ABSTRACT

The use of *Moringaoleifera* seed oil in the prevention of neurodegenerative diseases is on increasing trend. Cadmium is one of the most toxic environmental pollutants causing many known damages to the brain, the consumption of herbal alcoholic beverages is known to cause neurodegeneration. The aim of this study was to investigate the ameliorative effects of *Moringaoleifera* seed oil on cadmium and herbal alcoholic beverage induced- damage to frontal cortex of wistar rats. Eighty Wistar rats were divided into eight groups of 10 rats each. Group A served as control which received 2.5mg/kgbw phosphate buffer intra-peritoneally, while group F served as *Moringa*-treated control and received oral administration of 2.0 mg/kgbw *Moringaoleifera* oil. Groups B1, B2, D and E were injected intra-peritoneally with 3.5mg/kgbw CdSO<sub>4</sub>.8H<sub>2</sub>O single dose. Group C1, C2 and D received oral administration of 0.5 ml Herbal Alcoholic Beverage (HAB) and group B2, C2 and E were administered orally with 2.0mg/kgbw *Moringaoleifera* oil for four weeks followed by sacrificed. Quantitative enzymes and biochemical antioxidant markers showed that cadmium and HAB administration caused significant increase in AchE, SDH, CAT, GPx and MDA levels and decrease in SOD level. Conversely, there were significant decrease in AchE, SDH, CAT, GPx and MDA and increased

SOD level upon administration of *Moringaoleifera* oil. *Moringaoleiferaseed* oil has natural antioxidant constituents that ameliorated damage caused by Cd and HAB.

**Key Words:** Cadmium, herbal alcoholic beverages, *Moringaoleiferaseed*, frontal cortex

## Introduction

*Moringaoleifera Lam (Moringaceae)* is a highly valued plant distributed in many countries of the tropics and subtropics. It is the most widely cultivated species of the genus *Moringa*, which is the only genus in the family *Moringaceae*<sup>1</sup>. English common names include moringa, benzolive tree and West Indian Ben. It is also known as drumstick tree, from the appearance of the long slender triangular seed pods, horseradish tree, from the taste of the roots which resembles horseradish or Ben oil tree<sup>1</sup>. Moringa seed oil (yield 30-40% by weight), also known as Ben oil, is a sweet non-sticking, non-drying oil that resists rancidity. It has been used in salads, for fine machine lubrication and in the manufacture of perfume and hair care products<sup>2</sup>. It is an exceptionally nutritious vegetable tree with a variety of potential uses<sup>3</sup>. This tree has in recent times been advocated as an outstanding indigenous source of highly digestible Protein, Ca, Fe, Vitamin C and carotenoids suitable for utilization in many of the so-called developing regions of the world where undernourishment is a major concern<sup>4</sup>. Moringa leaves contain more vitamin A than carrots, more calcium than milk, more iron than spinach, more vitamin C than oranges and more potassium than bananas and that the protein quality of Moringa leaves rivals that of milk and eggs<sup>5</sup>. In addition to its compelling water purifying powers and high nutritional value, *Moringa oleifera* is very important for its medicinal value.

**Comment [N12]:** In the text, citations should be indicated by the reference number in brackets [].

Various parts of this plant such as the leaves, roots, bark, flower, seed, immature pods and fruit act as cardiac and circulatory stimulants possess antitumor, antipyretic, antiepileptic, anti-inflammation, antifungal diuretic, cholesterol lowering antioxidant, anti-diabetic, hepatoprotective, antibacterial and are being employed for the treatment of different ailments in the indigenous system<sup>6</sup>.

**Comment [NI3]:** typo

Cadmium (cd) is a toxic metal in the environment, found in the soil, rock phosphate fertilizer and in tobacco plant. Cd is a highly accumulative toxicant with very long biological half-life<sup>7</sup>. It is not biodegradable and its levels in the environment are increasing due to industrial activities and human exposures to cadmium are inevitable<sup>7, 8</sup>. Acute Cd exposure produced toxicities to the lung, liver, testes and brain, while chronic exposure to Cd often leads to renal dysfunction, anemia, osteoporosis and bone fractures. Cd is a potent carcinogen in a number of tissues of rodents and classified as a humancarcinogen<sup>9, 10</sup>. The neurotoxic effects of Cd have been reported in neonatal mouse brain<sup>11</sup> and young rat brain<sup>12</sup>.

**Comment [NI4]:** Cadmium (beginning of sentence)

**Comment [NI5]:** Cadmium (beginning of sentence)

Cadmium produces oxidative damage to isolated rat optic nerve<sup>13</sup> and culture rat cortical neurons<sup>14</sup>. In humans, occupational exposure to Cd is associated with nerve psychological disorders<sup>15</sup> and Parkinsonism has been reported in a 64-year-old man exposed to Cd at a high dose<sup>16</sup>. Thus, accumulating evidence clearly indicates that Cd is neurotoxic in a number of settings. The mechanisms involved in neurotoxicity of Cd are poorly understood. Oxidative stress has been proposed as a mechanism for Cd toxicity in a number of tissues such as the kidney<sup>17, 18</sup> and brain<sup>19</sup>. Due to its low permissible exposure limit (0.5 g/L to 1.0 g/L) in human, over exposure may occur even in situations where trace quantities of cadmium are found. Exposure to cadmium is addressed in specific standards for the general industry, shipyard employment, construction industry and the agricultural industry<sup>20</sup>.

**Comment [NI6]:** In the text, citations should be indicated by the reference number in brackets [16].

**Comment [NI7]:** the kidney [17,18] and brain[19].?!

Herbal alcoholic beverages commonly called Ogogoro, alomo bitter, Opaeyin in Nigeria. It is locally manufactured and packaged; it is consumed locally by the general public for sexual enhancement and as stimulants. Various investigations has revealed the deleterious effects of high percentage of alcohol (ranging from 17% to 70%) in most of the herbal alcoholic beverages<sup>21</sup>. Excess consumption of alcoholic beverages has been associated with high libido causing excess sexual enhancement and over stimulation has also been linked to excess herbal alcoholic consumption. Overall effects of herbal alcoholic consumption have been revealed to cause health hazard leading to soft tissues damage such as cardiovascular, lung, liver, kidney and brain<sup>21</sup>.

Frontal Cortex is the anterior part of the cerebrum which extends from the superior frontal sulcus to the lateral frontal sulcus of (sylvius) and aligned to the central frontal sulcus (of Ronaldo).

Frontal Cortex plays a vital role in learning, personality and in information processing<sup>21</sup>

Comment [NI8]: processing[21].

The aim of this study was to investigate the ameliorative effects of *Moringa oleifera* seed oil on cadmium and herbal alcoholic beverage induced- damage to frontal cortex of wistar rats.

### **Materials and Methods**

Eighty (80) Wistar Rats of both sexes were used for the research, *Moringa oleifera* seeds were procured from Ladoke Akintola University Farm in Ogbomoso, Nigeria and the plant specimen was identified with voucher number (No. FHI. 110266) assigned at Forestry Research Institute of Nigeria (FRIN), Jericho hill, Ibadan, where it was deposited.

Herbal Alcoholic Beverage (Local Market in Nigeria), Cadmium Sulphate (Guangzhou linhuada Chemical Reagent Co. Ltd., [GuangdongGuangdong](#), China)

The animals were divided into the following groups control group (A) received 2.5 mg/kgbw of phosphate buffer intraperitoneally single dose and the induced control groups (B1 and B2) received 3.5 mg/kgbw of  $3\text{CdSO}_4 \cdot 8\text{H}_2\text{O}$ <sup>22</sup>. intraperitoneally and left for 72 hours. B<sub>1</sub> rats were maintained under normal laboratory condition for period of four weeks and B<sub>2</sub> rats received 2.0 mg/kgbw of *Moringaoleifera* oil extract single dose daily for the period of four weeks. C<sub>1</sub> rats received 0.5 ml, 40% Herbal-gin via gavage, single dose daily four the period of four weeks while C<sub>2</sub> rats received 0.5 ml, 40% Herbal-gin and 2.0 mg/kgbw of *Moringaoleifera* oil extract simultaneously via gavage, single dose daily for the period of for weeks. Group D rats were injected intraperitoneally with 3.5 mg/kgbw of Cadmium sulphate ( $3\text{CdSO}_4 \cdot 8\text{H}_2\text{O}$ ) single dose and maintained for 72hrs<sup>22</sup>. Following oral administration of 0.5 ml, 40% Herbal-gin single dose daily for the period of four weeks. Group E animal were also injected intraperitoneally with 3.5 mg/kgbw of Cadmium sulphate ( $3\text{CdSO}_4 \cdot 8\text{H}_2\text{O}$ ) single dose and maintained for 72hrs<sup>22</sup>. Following oral administration of 0.5 ml, 40% Herbal-gin and 2.0 mg/kgbw *Moringaoleifera* seed oil extract single dose daily for the period of four weeks. Group F rats received 2.0 mg/kgbw of *Moringaoleifera* seed oil via gavage, single dose per day for the period of four (4) weeks. After the treatment, the animals were sacrificed via cervical dislocation.

Comment [NI9]: Reference?

### Biochemical Analysis

The rats were sacrificed and the brain from all the rats were carefully excised from the skull using brain forcep, weighed and the frontal cortex from all the rats were homogenized in 5% sucrose solution at about 4°C for quantitative enzyme assay for Succinate Dehydrogenase (SDH) and Acetyl cholinesterase (AChE) and oxidative stress markers Catalase (CAT), Superoxide Dismutase (SOD), Malondialdehyde (MDA) and Glutathione Peroxidase (GPx).

Comment [NI10]: what method to measure it?

## Statistical analysis

The data collected was subjected to statistical analysis, One- Way ANOVA was used with computer software package model of graph pad prism version 5 and multiple comparison test was carried out and significant difference was observed and tested at  $p < 0.05$ , at percentage confidence interval of 95 (95% CI).

**Comment [NI11]:** See "p" in Guideline for Reporting P values

## Ethical Committee Approval

**Comment [NI12]:** The manuscript has not been equipped with Ethical Committee Approval!

## Results and discussions

**Comment [NI13]:** In the text, citations should be indicated by the reference number in brackets [].

- Table 1 showing Effect of MO seed oil on the activities of Ache & SDH in rats.

**Comment [NI14]:** The table form? look at on the journal issue

Groups	A	B	B2	C	C2	D	E	F
AChE ( $10^{-5}$ moles/min/gm)04	2.87±0.08*	0.56±0.095	4.63±0.02*	0.54±0.07	8.12±3.75	4.57±0.25	11.46±0.26*	13.50±0.52*
SDH ( $10^{-3}$ $\mu$ moles/min.mg)04	4.13±0.099	5.41±0.109	5.39±0.008	1.81±0.01*	1.50±0.01*	8.11±0.08*	5.65±0.06	1.99±0.09*

Data are expressed as mean  $\pm$  SEM

\* It indicates significant level at  $p < 0.05$  when compared with each other (multiple comparison test) and with the control group A.

**Formatted:** Font: Italic

**Legend:** AChE: Acetylcholinesterase, SDH: Succinate Dehydrogenase, **A:** Control, **B1:** Cadmium only, **B2:** Cadmium and Moringa oil, **C1:** HAB only, **C2:** HAB and Moringa oil, **D:** Cd and HAB, **E:** Cd + HAB + MO, **F:** Moringa oleifera seed oil

In table 1, an increase in the level of SDH activity was observed in groups B<sub>1</sub>, B<sub>2</sub>, D and E ( $5.41 \pm 0.19$ ,  $5.39 \pm 0.07$ ,  $8.11 \pm 0.08$ ,  $5.65 \pm 0.05$ )  $\mu\text{moles}/\text{min}/\text{mg}$  respectively and this increase (p-value = 0.0001), was significant when compared with each other and with the control group A at  $P < 0.05$ . Decrease in SDH level was also observed in groups C<sub>1</sub>, C<sub>2</sub> and F ( $1.81 \pm 0.01$ ,  $1.50 \pm 0.01$  and  $1.99 \pm 0.89$ )  $\mu\text{moles}/\text{min}/\text{mg}$  respectively, as against the control group A and as compared with other groups at  $P < 0.05$ .

**Comment [NI 15]:** This way of writing is better: B<sub>1</sub>( $5.41 \pm 0.19$ ), B<sub>2</sub>( $39 \pm 0.07$ ), D ( $8.11 \pm 0.08$ ), and E ( $5.65 \pm 0.05$ ).

**Comment [NI 16]:** See guidelines for author

**Formatted:** Font: Italic

**Comment [NI 17]:** Writing corrected

**Formatted:** Font: Italic

Table 1 shows an increase in the level of AChE activity in groups B<sub>2</sub>, C<sub>2</sub>, D, E and F ( $4.63 \pm 0.95$ ,  $8.12 \pm 3.77$ ,  $4.57 \pm 0.26$ ,  $11.46 \pm 0.26$  and  $13.50 \pm 0.52$ )  $\mu\text{moles}/\text{min}/\text{gm}$  respectively and this was significant at  $p < 0.0001^{***}$  when compared with each other in the experimental group at  $P < 0.05$ . A decrease in the level of AChE was observed in groups B<sub>1</sub> and C<sub>1</sub> ( $0.56 \pm 0.08$  and  $0.54 \pm 0.02$ )  $\mu\text{moles}/\text{min}/\text{gm}$  respectively at  $P < 0.0001^{***}$  when compared with group A and with other groups at  $P < 0.05$ .

**Comment [NI 18]:** More clearly if separately

**Formatted:** Font: Italic

**Formatted:** Font: Italic

**Formatted:** Font: Italic

The preservation of sufficient levels of acetylcholine (ACh) at neurotransmission sites is the basic plan of action involved in the medical treatment of neurodegenerative disorders such as Alzheimer's disease<sup>23</sup>. Consequently, the inhibition of acetyl cholinesterase (AChE) prevents the hydrolysis of ACh thereby maintaining normal memory function. Frequency of neurodegenerative disorders reduces in correlation to consumption of antioxidants<sup>24</sup>. Due to this,

diets including natural compounds with high levels of antioxidants have been advocated as useful remedial approach for neurodegenerative disorders<sup>24</sup>.

AChE ceases the action of acetylcholine post-synaptically<sup>25</sup>. The inhibition or introduction of AChE activity will result in alterations in the concentration of acetylcholine which may have a negative effect on the body. When AChE is inhibited in the body, high concentration of acetylcholine will accumulate in the body while hydrolysis of the acetylcholine into acetate and choline occurs due to introduction of AChE in the body, this will reduce the concentration of the acetylcholine in the body<sup>24</sup>. The toxicity of acetylcholine is manifested by muscarinic and nicotinic signs and symptoms such as ocular pain, confusion, ataxia, slurred speech, loss of reflexes, generalized convulsions and coma<sup>24</sup>.

There was a significant decrease in AChE activity in group B1 and group C1 animals' frontal cortex tissue. This finding supports previous research findings that the rate of AChE activity decreased significantly when compared with the normal control group A<sup>25</sup>. Another study reported that cadmium and alcoholic herbal beverages increases AChE activity in response to a dose per body weight of rat<sup>26</sup>. There was an increase in acetyl cholinesterase enzyme activity of group B2, group C2, group E and group F rats when compared to the cadmium induced group A rats, though it increased compared to the group B1 and group C1 rats. This shows the antioxidants properties of Moringa oleifera oil extract in restoring the action of AChE activity from very low levels to a significant high level. This is supported by previous researches that antioxidants are powerful therapeutic agents that can prevent neurodegenerative disorders, and a combination, rather single use of any of them is more effective<sup>27</sup>. This may also be the reason why there was a significant reduction in AChE activity in group D rats when compared to the normal group A animals. There was a significant increase in the level of activity of AchE in

frontal cortex of group C2 when it was compared to animals in the control group A, this observation was in line with report of<sup>27</sup>. In group F rats, that are the *Moringa oleifera* seed oil control group, there was a significant increase in Acetyl cholinesterase activity when compared with the control rats. This probably might be due to the antioxidant activity in the *Moringa* plant<sup>28</sup>.

From the results of this study, there was an increase in the level of SDH activity in group B1, group B2 and group E rats which were significant at  $P < 0.05$  when compared with control group A animals. This observation was in line with investigation carried out by<sup>29</sup>. There was a decrease in the level of SDH activity in the frontal cortex homogenates in group C1, group C2 and group F, this decrease in SDH activity is significant at  $P < 0.05$  when compared with animal frontal cortex homogenate in group A which explained the importance of SDH at decreased level in Kreb's cycle to generate high- requiring ATP needed for brain cells.

Formatted: Font: Italic

Formatted: Font: Italic

### Result of the activities of oxidative stress markers

**Table 2 showing Effects of MO seed oil on oxidative stress markers in brain tissues homogenates of rats**

Comment [NI 19]: The table form? look at on the journal issue

GROUPS	A	B1	B2	C1	C2	D	E	F
CAT (μmoles/mg)	1158±14. 47	1331±23. 12*	1456±49. 45*	1355±60. 68*	1397±0.7 4*	1209. ±39.29	1797±41. 17*	1807±67. 84*
MDA (10-6 units/mg)	5.40±0.08 3	7.30±0.17 *	42.36±1.4 9	20.79±0. 89	15.37±0.1 1	22.94±0. 82	55.40±1. 19*	24.16±1.2 0*

GPx ( $\mu$ moles/min/ mg)	366.50 $\pm$ 4.87	444.40 $\pm$ 9.08*	466.80 $\pm$ 16.1*	461.40 $\pm$ 19.9	446.40 $\pm$ 8.08	461.7 $\pm$ 15.7	639.40 $\pm$ 14.3	560.90 $\pm$ 21.4
SOD (units/mg)	2796.00 $\pm$ 6.11	83.33 $\pm$ 0.00	666.70 $\pm$ 0.00	116.70 $\pm$ 0.00	785.70 $\pm$ 3.07	78.57 $\pm$ 3.07	83.33 $\pm$ 0.00	3476.00 $\pm$ 2.02

Data are expressed as means  $\pm$  SEM \* It indicates significant level at  $p < 0.05$  when compared with each other (multiple comparison test) and with the control group A.

**Legend:** CAT: Catalase, MDA: Malondialdehyde, GPx: Glutathione Peroxidase, SOD: Superoxide Dismutase, A: Control, B1: Cadmium only, B2: Cadmium and Moringa oil, C1: HAB only, C2: HAB and Moringa oil, D: Cd and HAB, E: Cd + HAB + MO, F: Moringa oleifera seed oil.

There was a progressive increase in the level of catalase activity in groups B1, B2, C1, C2, D, E and F as shown in Table 2, this progressive increase was significant at  $p \leq 0.0001$ \*\*\* when compared within the groups and with the control group A at  $p < 0.05$ .

Formatted: Font: Italic

Formatted: Font: Italic

A progressive increase in the level of malondialdehyde was observed in groups B1, B2, C1, C2, D, E and F as shown in table 2, the increase was significant at  $p \leq 0.0047$ \*\* when compared with the control group A and with each other across the groups at  $p < 0.05$ .

Formatted: Font: Italic

Formatted: Font: Italic

There was an increase in the level of glutathione peroxidase in groups B1, B2, C1, C2, D, E and F as described in table 2, this increase was significant at  $p \leq 0.0001$ \*\*\* when compared with each other across the experimental groups and with the normal control group A at

Formatted: Font: Italic

$p < 0.05$ .

Formatted: Font: Italic

### **Result of the level of SOD in Brain Homogenate**

Formatted: Font: Bold

There was a progressive decrease in the level of superoxide dismutase in groups B1, B2, C1, C2, D and E and an increase in the level of SOD in group F ( $3476.00 \pm 2.02$ )units/min which was significant at  $P \leq 0.0001^{***}$  when compared with control group and with each other across the experimental groups at  $P < 0.05$ .

Formatted: Font: Italic

Formatted: Font: Italic

The activities of antioxidant enzymes such as superoxide dismutase (SOD), ascorbate peroxidase (APX) and catalase have been affected by cadmium. Studies have demonstrated these changes in enzyme activities<sup>30, 31</sup>. Under normal conditions, free radicals are formed in minute quantities and are rapidly scavenged by natural cellular defense mechanisms comprising of enzymes like superoxide dismutase (SOD), glutathione peroxidase (GPx), glutathione reductase (GR), and catalase (CAT) and others. The activity of antioxidant enzymes such as CAT, SOD, and GST, can vary depending on the intensity and duration of the chemical stress applied to the organism in addition to the susceptibility of the exposed species<sup>32</sup>.

Cadmium caused a significant increase in malondialdehyde (MDA) activity in Group B1 animals' brain tissue homogenates. It also produced a resultant increase in glutathione peroxidase (GPx) and catalase (CAT) activities, while superoxide dismutase (SOD) activity decreased, though these increments were not significant and decrease in SOD activities was significant when compared with control group A animals<sup>33</sup> reported a significant increase in lipid peroxidation in animals exposed to cadmium. Superoxide dismutase (SOD) and catalase (CAT) activities were also reported to be decreased ( $P \leq 0.05$  and  $P < 0.005$ ) in the same experiment, whereas glutathione peroxidase (GPx) increased.

Formatted: Font: Italic

Formatted: Font: Italic

The enzyme activities of GPx, MDA and CAT increased in group B2, compared to their activities in groups of rats induced with cadmium only. This suggests that the extracts have protective action against free radicals. Previous researches have shown that non enzymatic antioxidant components which consists of molecules such as alpha tocopherol (Vit. E), ascorbic acid (Vit. C), glutathione and beta-carotene that react with activated oxygen species and thereby prevent the propagation of free radical chain reactions. Whereas, there was a significant decrease in SOD activity.

In group C1, group C2, group D and group E animals showed a significant increase in the activities of MDA, GPx and CAT, while there was a significant decrease in enzyme activity of SOD. This is in agreement with previous studies which reported that, *Moringa oleifera* contains antioxidant phytochemicals, such as vitamin C, betacarotene, lycopene and vitamin E all of which acts as antioxidant and subsequently decrease the consumption of these antioxidant enzymes to combat oxidative stress<sup>34, 35, 36, 37</sup>. In an earlier study, the presence of alkaloids, flavonoids, saponin, tannin, anthraquinones, and anthacyanosides in *Moringaoleifera seed oil* extract was reported<sup>37</sup>. Also, previous independent studies have reported that the protective actions of hepatoprotective medicinal plants are mediated by their flavonoids or alkaloids components or by their combination via antioxidant and free radicals scavenging activities<sup>37</sup>. The presence of these active biological principles may thus be accounting for the biological effect of *Moringa oleifera* extract and could be via antioxidant and/or free radicals scavenging activities. Group F rats treated with *Moringa oleifera* seed oil extract also showed significant increase in SOD, GPx and CAT activities and a significant decrease in MDA activity as compared with control group rats at  $p < P=0.05$ . In conclusion, this study has shown that *Moringa oleifera* oil

Formatted: Font: Italic

extract has antioxidant properties that might have ameliorated morphological damage caused by cadmium and herbal alcoholic beverages.

**Comment [NI20]:** The conclusion is written separately with the results and discussion

## Conclusion

## References

**Comment [NI21]:** The writing of some references does not follow the guidelines

1. Jed, W. Fashey, S.C.D. *Moringaoleifera*; A Review of the Medical Evidence for its Nutritional, Therapeutic and Prophylactic properties, 2005; Part 1. December 1.
2. Lalas, Tsaknis, J. Extraction and identification of natural antioxidant from the seeds of the *Moringaoleifera* tree variety of Malawi. *Journal of Academy of science*, 2002, 79:677-683.
3. Anwar, F., Latif, S., Ashraf, M., Gilani, A. H. *Moringaoleifera*: A Food Plant with Multiple Medicinal Uses. *Journal of Phototherapy Research*, 2007; (21) 17-25.
4. Omotoso, O.D., Owolabi, J.O., Samanja, Y.J., Dare, B.J., Ahamu, E.A., Adelokun, S.A. Histological and Histochemical Evaluation of Anticadmium Toxicity effects of *Moringaoleifera* seed oil and Anacardium occidentale Nut oil in the Hippocampus of Juvenile male Wistar Rats. *Journal of Advances in medical and pharmaceutical sciences*, 2015;5:1-13.
5. Friberg, L., Nordberg, G.F., Vouk, V.B. eds. *Handbook of the toxicology of metals*. Vol.II. Amsterdam, Elsevier, 1986; 130–184.

6. Goering, P.L., Waalkes, M.P., Klaassen, C.D. Toxicology of cadmium. In Toxicology of metals: Biological Aspects. Hand book of experimental pharmacology, 1995 vol.115 pp. 189-213. Springer-verlag, New York.
7. Klaassen, C.D., Liv, J. Choudhri, S. Metallothionien An intracellular protein to protect against cadmium toxicity. Annual Rev. Pharmacological Toxicology, 1999; 39: 267-294.
8. Waalkes, M.P. Cadmium carcinogenesis. Mutation Res, 2003; 533: 107-120.
9. Webster, W.S. Valois, A.A. The toxic effect of cadmium on the neonatal mouse central nervous system. Journal of Nevropathology. Experimental neuroscience, 1981;4: 247-257.
10. Wong, K.L. Klaassen, C.D. Neurotoxic effects of cadmium in young rats. Toxicology applied pharmacology, 1982 63: 330-337.
11. Lopez, E., Figuereroa, S., Oset-Gasque, M.J. Gonzalez, M.P. Apoptosis and necrosis: two distinct events induced by cadmium in cortical neurons in culture. British Journal of Pharmacology, 2003; 138:901-911.
12. Hart, B.A., Lee, C.H., Shukla, G.S., Shukla, A., Osier, M., Eneman, J.D. Characterization of cadmium –induced apoptosis in rat lung epithelial cells Evidence for the participation of oxidant stress. Journal of Toxicology, 1999. 133:43-58.
13. Okuda, R., Iwamoto, Y., Tachibana, H. Sugita, M. Parkinsonism after acute cadmium poisoning. Clinical Neurology Neurosurgery, 1997; 99:263-265.
14. Bagchi, D., Vuchetich, P.J., Bagchi, E.A., Tran, M.X., Tang, L. Stohs, S.J. Induction of oxidative stress by chronic administration of sodium dichromate and cadmium chloride to rats. Free radicals. Biomedical Journal, 1997; 22, 471-478.

Comment [NI22]: ???

15. Liu, J., Corton, C., Dix, D.J., Liu, Y., Waalkes, M.P. Klaassen, C.D. Genetic background but not metallothionein phenotype dictates sensitivity to cadmium-induced testicular injury in mice. *Toxicology. Applied Pharmacology*, 2001; 176: 1–9.
16. Kumar, R., Asic, K., Agarural, K., Seth, P.K. Oxidative stress mediated neurotoxicity of cadmium. *Journal of toxicological letters*, 1996; 89:65-69.
17. WHO, 2007 Health risks of heavy metals from long-range trans boundary air pollution. *Copenhagen, World Health Organization Regional Office for Europe*
18. Eisenberg, D.M., Davis, R.B., Ettner, S.L. Trends in alternative medicine use in the United States. *JAMA*, 1998;280:1569–75.
19. Eroschenko, V.P. difiore's Atlas of Histology with Functional Correlations (11th Ed.) Philadelphia: Lippincott Williams & Wilkins, 2008; Pg 146-148.
20. Ige S.F., Salawu, E.O., Olaleye, S.B., Adeeyo, O.A., Badmus, J. Adeleke, A.A. Onion (*Allium cepa*) extract prevents cadmium induced renal dysfunction. *Indian Journal of Nephrology*, 2010. 19 (4):140-144.
21. Ndhlala, A.R., Aremu, A.O., Moyo, M., Amoo, S.O., Van, S.J. Acetylcholinesterase inhibitors from plant sources: friends or foes? In *Cholinesterase: Production, Uses and Health Effects*. Edited by White CJ, Tait JE. New York: Nova, 2012; 67-98.
22. Howes, M.J.R., Houghton, P.J., Plants used in Chinese and Indian traditional medicine for improvement of memory and cognitive function. *Pharmacology of Biochemical Behavior*, 2003; 75:513–527.

Comment [NI23]: page writing is inconsistent with others

23. Garcia-Ayllon, M.S., Silveyra, M.X., Candela, A., Compan, A., Claria, J. Jover, R. Changes in Liver and Plasma Acetylcholinesterase in Rats with Cirrhosis Induced by Bile Duct Ligation. *Hepatology*, 2006. 43(3): 444-453.
24. Lassiter, T.L., Marshall, R.S., Jackson, L.C., Hunter, D.L., Vu, J.T. & Padilla, S. Automated measurement of acetylcholinesterase activity in rat peripheral tissues. *Toxicology*, 2003; 186: 241-253
25. Belabed, S., Soltani, N. Acute toxicity of cadmium on *Donax trunculus*: acetyl cholinesterase, glutathione S-transferase activities and pattern of recovery. *Pelagia Research Library, European Journal of Experimental Biology*, 2013; 3 (2):54-61.
26. Carageorgiou, H., Tzotzes, V., Sideris, A., Zarros, A., Tsakiris, S. Cadmium effects on brain acetyl cholinesterase activity and antioxidant status of adult rats: modulation by zinc, calcium and L-cysteine co-administration. *Basic Clinical Pharmacology of Toxicology*, 2005 97 (5): 320-324.
27. Kontush, A., Mann, U., Arlt, S., Vjeyl, A., Lihrs, C., Thomson, T.M. Beisiegel, V. Influence of vitamin E and C supplementation on lipoprotein oxidation in patients with Alzheimer's disease. *Free Radiology and Biology in Medicine*, 2001; 31: 345.
28. Adeneye A.A., Olagunju, J.A, Elias, S.O., Olatunbosun, D.O., Mustafa, A.O., Adeshile, O.I., Ashaolu, A.O., Laoye, T.A., Bamigboye, A.O., Adeoye, A.O. Protective activities of the aqueous root extract of *Harunganamadagascariensis* in acute and repeated acetaminophen hepatotoxic rats. *International Journal of Applied Res in Natural Products*, 2008; 3: 29-42.
29. Shaw, B.P. Effects of mercury and cadmium on the activities of antioxidative enzymes in the seedlings of *Phaseolusaureus*. – *Biology of Plants*, 1995; 37: 587-596.

Comment [NI24]: List the first six authors followed by et al.

30. Lozano-Rodriguez, E., Hernandez, L.E., Bonay, P., Carpena- Ruiz, R.O. Distribution of cadmium in shoots and root tissues of maize and pea plants: physiological disturbances. – *Journal of experiments in Botany*, 1997; 48: 123-128.
31. Ballesteros, M.L., Wunderlin, D.A., Bistoni, M.A. Endosulfan Induces changes in spontaneous swimming activity and acetylcholinesterase activity of *jenysiamultidentata* (*Anablepidaecyprinnodontiformes*). *Ecotoxicology of Environmental Safety*, 2009;72:199.
32. Ognjanovic, B.I., Pavlovic, S.Z., Maletic, S.D., Zikic R.V., Stajn A.S., Radojicic R.M., Saicic, Z.S., Petrovic, V.M. Protective influence of vitamin E on antioxidant defense system in the blood of rats treated with cadmium. *Physiology Res*, 2003; 52:563-70.
33. Azarkan, M., Wintjens, R., Looze, Y. Detection of three wound-induced proteins in papaya latex. *Phytochemistry*, 2004;65 (5): 525-534.
34. Aruoma, O.I. Methodological considerations for characterizing potential antioxidant actions of bioactive components in food plants. In *Mutation Research*, 2003;523: 9-20.
35. Gouado, I., Schweigert, F.J., Ejoh, R.A., Tchouanguep, M.F., Camp, J.V. Systemic levels of carotenoids from mangoes and papaya consumed in three forms (juice, fresh and dry slice). *European Journal of Clinical Nutrition*, 2007; 61(10): 1180-1188
36. Amer, J., Goldfarb, A., Rachmilewitz, E.A., Fibach, E. Fermented papaya preparation as redox regulator in blood cells of beta-thalassemic mice and patients. *Journal of Phototherapy Research*. 2008; 22(6): 820-828.

37. Adeneye A.A., Olagunju, J.A., Banjo, A.A.F., Abdul, S.F., Sanusi, O.A., Sanni, O.O., Osarodion, B.A., Shonoiki, O.E. The Aqueous Seed Extract of *Carica papaya* Linn. Prevents Carbon Tetrachloride Induced Hepatotoxicity in Rats. *International Journal of Applied Res. in Natural Products*, 2009; 2(2): 19- 32.

UNDER PEER REVIEW