

# Microbial Quality Analysis of Locally Packed and Commercially Available Bread in Chennai Region of Tamilnadu, India

## ABSTRACT

**Aims:** In today's world consumer preference towards packed products is increasing than locally available bakery shops due to unhygienic processing and improper handling. This reason became an interesting fact to analyse the microbial quality of Bread in Chennai region.

**Methodology:** This study was conducted to determine the microbial load of bread sold commercially and by the local in Chennai region. Locally made bread samples and commercially available breads were procured in shops from Chennai region. The microbial quality was analysed to check the microbial load of bread samples using standard microbial procedures such as total bacterial count, total yeast/mould counts and total coliforms. It was carried out with the help of Plate Count Agar, Sabouraud Dextrose Agar and Eosin Methylene Blue Agar through the serial dilution and plating techniques.

**Results:** The plates were incubated and results were collected through the colony counting method. Total aerobic bacterial count was found in all the bread samples while coliform count was absent in all samples. Yeast/mold/fungi was present in all commercially available breads except sample 1 & 2 of local bread.

**Conclusion:** It can be concluded that the local and commercially available breads were having the microbial load within the permissible limits.

**Keywords:** Bread, Bakery, Microbial profile, Plate Count Agar, Sabouraud Dextrose Agar, Eosin Methylene Blue Agar.

## 1. INTRODUCTION

In the past, bakery goods were thought to be a poor man's diet. The majority of the human population currently relies on them as daily nutritional staples [1]. With a yearly revenue of over 3000 crores, the bakery business in India is the largest of the food industries [2]. Bread is a common, high-energy food that is also convenient and low in calories, glycaemic index, vitamins E, B-vitamin, and minerals [3]. When paired with fillings made of meat or fish, veggies, and fat like margarine or mayonnaise, it is commonly used by people as a snack or a meal. Rich bread is made with sugar, eggs, milk, and flavouring added to the flour and raising agent; yeast bread is made with flour, liquid, fat, milk, yeast and salt [4].

Bread is a significant staple food created from baking dough comprised of flour and water [5]. In 2011, bread made from wheat provided 20% of the daily calories consumed worldwide, feeding the world's population [6]. One of the non-indigenous foods that is most commonly consumed in India is bread [7]. The demand for ready-to-eat food items has increased as a result of urbanisation [1].

In recent decades, there has been a growth in the sales of a range of breads and other bakery goods. Before adding grains to the making of bread, grains are ground to a powdery form to create flour. It has been revealed that flours give the final baked bread its primary structure. Worldwide, many flours derived from wheat, rye, barley, maize, and other types of grains are available. In both rural and urban populations, bread and wheat flour make up a sizable portion of the daily diet. The amount of minerals, lipids, and proteins in flour is reduced and the proportion of carbohydrates is higher [8].

47 The most frequent cause of bread product deterioration is water activity. Microbiological  
48 spoilage, particularly mould growth, is a major economic concern with regard to bakery products. For  
49 bakeries, mould deterioration is a severe and expensive issue [2]. Physical, chemical, and  
50 microbiological spoiling issues can affect bread and other bakery products. The latter is the most  
51 significant, notably bacterial (*Bacillus* sp.) and mould growth [9]. The main causes of microflora in  
52 bread are contamination in the baking environment, contamination from handlers, and contamination  
53 from market vendors [1]. The features of the product and how it is preserved have a big impact on the  
54 microbial flora that colonises a specific food. Intrinsic, extrinsic, and processing and preservation  
55 methods are among the factors that can affect the growth of microbes in food [10]. Due to the  
56 suggested endogenous disease and the fact that they have minimal water activity from a  
57 microbiological standpoint, flour and breads are typically considered to be safe foods [11].

58 This study was initiated to make a comparison for better quality with respect to microbial load  
59 of the commonly available breads so that consumers may get a nutritive, more hygienic and shelf  
60 stable product. Our objective is to assess the microbial load of selected bread samples.

## 61 2. MATERIALS AND METHODS

### 62 2.1 Sample Collection

63 The area selected for the collection of bread samples was Chennai, Tamilnadu, India. The  
64 popular shops nearby the laboratory were selected, so that the items were feasible to purchase. The  
65 number of shops selected were six. Among six, we purchased locally made breads from three shops  
66 and commercially available breads from three shops. Samples were labelled as per the table 1. The  
67 samples were collected aseptically in a sterile air lock bag to prevent the contact of any other source  
68 that can contaminate the sample. The collected samples are carried to laboratory further processing.

69 **Table 1: Sample Information and their codes.**

Sample Code	Bread Type
Sample 1	Locally made
Sample 2	Locally made
Sample 3	Locally made
Sample 4	Commercially available
Sample 5	Commercially available
Sample 6	Commercially available

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### 71 2.2 MEDIA PREPARATION

72 Microbiological assay of the bacteria, yeast and mold counts were performed as per  
73 the methods outlines in the compendium of methods for microbiological examination of foods  
74 with few modifications [12].

#### 75 2.2.1 PLATE COUNT AGAR (PCA)

76 Plate Count Agar (PCA) media was used to detect the microorganisms. 2.35g of PCA  
77 and 2 gm of agar-agar was added to the conical flask containing 100 ml of distilled water and  
78 homogenized using heating mantle. The mouth of the conical flask is plugged with cotton  
79 wool and covered with aluminium foil and then autoclaved at 121°c temperature 15 minutes.  
80 Autoclaved media was poured on the sterile petri plate in laminar air flow chamber and

81 solidified.

## 82 **2.2.2 SABOURAUD DEXTROSE AGAR (SDA)**

83 Sabouraud dextrose agar (SDA) media was used to detect the yeast or mould or  
84 fungus. 6.5 gm of SDA and 2 gm of agar-agar was added to the conical flask containing 100  
85 ml of distilled water and dissolved using heating mantle. The mouth of the conical flask is  
86 plugged with cotton wool and covered with aluminium foil. The media was autoclaved at  
87 121°C temperature for 15 minutes. Autoclaved media is poured on the sterile petri plate in  
88 laminar air flow chamber and solidified.

## 89 **2.2.3 EOSIN METHYLENE BLUE AGAR (EMB)**

90 Eosin methylene blue agar (EMB) media was used to detect the coliforms. 7.18g of  
91 EMB and 2 gm of agar-agar was added to the conical flask containing 100 ml distilled water.  
92 The media was dissolved using heating mantle. The mouth of the conical flask is plugged  
93 with cotton wool and covered with aluminium foil and then autoclaved at 121°C for 15  
94 minutes. Autoclaved media is poured on the sterile petri plate in laminar air flow chamber  
95 and solidified.

## 96 **2.3 Plating of samples on media**

97 Serial dilution method was used in this experiment. 1g of each sample was added into  
98 test tubes containing 9 ml sterile water and homogenized to prepare standard stock  
99 solutions. 1ml was removed from each of the stock solution and added to another set of test  
100 tubes containing 9ml of sterile water which makes  $10^{-1}$  dilution. The same procedure was  
101 repeated to make  $10^{-6}$  dilution. Like this sample solution was prepared for all the 6 samples.  
102 From  $10^{-6}$  dilution, 0.1ml of each sample solution was taken in micropipette and poured on  
103 solidified plates of PCA and EMB labelled accordingly with the sample code for each  
104 sample. Similarly for SDA plates of each sample, 0.1ml of  $10^{-3}$  dilution was taken in  
105 micropipette and spread on SDA plates. Each plate was gently swirled to mix the 0.1µl of  
106 diluted sample over the agar media using sterile L rod, clockwise and anticlockwise, to and  
107 for thrice and taking care that the contents do not touch the lid. The plates were left without  
108 moving for at least 15 minutes to allow the agar to set. PCA and EMB Plates were incubated  
109 at  $35^{\circ}\text{C}\pm 2^{\circ}\text{C}$  for 48 hours while SDA plates were incubated at  $28^{\circ}\text{C}\pm 2^{\circ}\text{C}$  for 72 hours.  
110 Following incubation, the number of colonies on each media were counted and the results  
111 were recorded in CFU/g [1]. Colony Forming Unit was calculated using the following formula:

112 
$$\text{CFU/g} = (\text{average no. of colonies} \times \text{total dilution factor}) / \text{volume plated}$$

## 113 **3. RESULT AND DISCUSSION**

114 'Table 2' shows the outcome of microbial analysis of local and commercial bread samples. The total  
115 bacterial counts of the bread samples ranged from  $2.7 \times 10^3$  to  $8.3 \times 10^3$  cfu/g with samples, 1 and 2;4  
116 having the lowest and highest values respectively. Yeast/Mold/Fungi counts ranged from  $3.9 \times 10^1$  to  
117  $5.5 \times 10^1$  cfu/g with the lowest and highest counts recorded for sample 4 and sample 3 respectively.  
118 There was no detection of Coliforms in the bread samples evaluated. The microbial counts were  
119 within the permissible limit which is set by the Standard Organization of Nigeria, which states that the  
120 counts of aerobic bacterial must not exceed  $10^3$  cfu/g and coliform growth must not be detected in  
121 bread samples [13].

122 Total aerobic bacterial count was found in all the bread samples. Yeast/mold/fungi was present in all  
123 commercially available breads and samples 3 of local bread. Sample 1 & 2 did not show fungal  
124 presence. All bread samples breads maintained the absence of coliforms. In the present investigation  
125 the microbial profile of several packed local bread and commercial bread was analysed by comparing  
126 the colony forming units (CFU) as in 'Table 2'.  
127

## 128 **TABLE 2: MICROBIAL ANALYSIS OF LOCAL AND COMMERCIAL BREAD**

Sample	Total Bacterial Count	Total Coliform Count	Total Fungi Count
	PCA CFU/gm	EMB CFU/gm	SDA CFU/gm
Sample 1	2.7x10 <sup>3</sup>	ND	ND
Sample 2	8.3 x10 <sup>3</sup>	ND	ND
Sample 3	5.5 x10 <sup>3</sup>	ND	5.5 x10 <sup>1</sup>
Sample 4	8.3 x10 <sup>3</sup>	ND	3.9 x10 <sup>1</sup>
Sample 5	4.1 x10 <sup>3</sup>	ND	4.1 x10 <sup>1</sup>
Sample 6	4.1 x10 <sup>3</sup>	ND	4.2 x10 <sup>1</sup>

129 ND- NOT DETECTED

130 As per PCA microbiological data of packed local bread and commercial bread, Sample 1 showed the  
131 lowest values and sample 2 is showed highest values. In sample 5 and 6 probably got equal values.  
132 SDA (sabouraud dextrose agar) Among the studied, in local bread sample 1 and 2 no microbial count  
133 was found in all the samples, in sample 3 showed the highest values indicating high number of  
134 bacteria in the sample. When compared to commercial bread, local bread sample 3 had the highest  
135 microbial load. Among the studied coliform counting in EMB, no colony was determined in all the  
136 samples.

137 The findings of our investigation indicate that the bread samples are safe for ingestion by humans  
138 because they do not present any significant health risks. The samples' microbial growth may have  
139 developed during processing from the raw materials (such as flour, sugar, and yeast) or possibly from  
140 the environment.

141 Typically, bread rotting is caused by moulds such Mucor, Rhizopus, Aspergillus, Penicillium, and  
142 Fusarium. While the products are manually cut, packaged and delivered, bacteria including Bacillus,  
143 Escherichia and Salmonella, Streptococcus aureus, etc. may also infect and cause ropiness of the  
144 bakery items [14]. Any bread item that is kept open or partially sealed can become contaminated and  
145 degraded by airborne fungus, moulds, and bacteria because relatively high moisture content  
146 promotes the growth and development of mould and bacteria on the bread [15].

147 It was discovered that correct vending practises, such as cleanliness of the retailer or storekeeper,  
148 were related to the microbiological status of various food products; this relationship is also firmly  
149 thought to be related to the level of education of the vendors [16]. In the current study it was found  
150 that fungal count was high in commercially available breads than locally prepared breads. Whereas  
151 the coliforms were absent in all samples which is safe for consumers.

## 152 4. CONCLUSION

153 From this study, it was observed that the microbial load was within the permissible limit for  
154 human consumption. It is preferable to consume bread within the expiry date as mentioned by the  
155 suppliers. The determination of micro flora carried out in this study is necessary in safeguarding public  
156 health. This study therefore provides basic information about the micro flora in bread likely to cause  
157 food-borne disease when present in bread which is a ready-to-eat type of food.

## 158 COMPETING INTERESTS

159 Authors have declared that no competing interests exist.

160 **AUTHORS' CONTRIBUTIONS**

161

162 Dr. Sasikala designed the study, performed the statistical analysis, Madhanisha S wrote the protocol,  
163 and wrote the first draft of the manuscript. Bhuvaneshwari K and Kavarshi K managed the analyses of  
164 the study. Dr. Sasikala managed the literature searches. All authors read and approved the final  
165 manuscript.

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