

Original Research Article

Ammonia Pre-Treated Cotton Stalks for Bioethanol Production

Comment [j1]: Look like you have already used book chapter or review make it like experimental

ABSTRACT

Cotton stalks are good raw material for bioethanol production due to its availability throughout the world, abundance, high carbohydrate content and not involved in any food chain. Due to recalcitrant nature of cotton stalks pre-treatment, hydrolysis were not effective. In the present study pre-treatment with ammonia at room temperature for 1 week period and 121°C for 60min protocol ?? were attempted and it is compared with the standard 0.2M NaOH treatment time and removal efficiency?. A 1.5% ammonia pre-treatment at room temperature for 1 week found to remove 86% of lignin and subsequently undergone 91% is it 91% acid taken?? sulphuric acid hydrolysis. The acid hydrolysate obtained consists less lignin and furfurals and fermented to 5.75% ethanol with 91% fermentation efficiency with *saccharomyces cerevisiae*.

Comment [j2]: Protocol not matched with M&M

Key words: Ammonia; Pre-Treatment; Cotton Stalk; Bioethanol; Yeast.

1. INTRODUCTION

Bioethanol is recognized as a clean-burning, non-petroleum liquid fuel. Countries dependence on imported oil, environmental issues, and employment in rural areas has been reasons for the consideration of the replacement of fossil fuels with bioethanol. But only a few countries are successful to use ethanol as fuel, where excessive ethanol is produced from additional or specifically grown raw materials like maize and sugar cane. As a substrate, conventional crops such as corn and sugarcane are unable to meet the global demand for bioethanol production due to their primary value of food and feed therefore, lignocellulosic substances such as agricultural wastes have emerged as an attractive feedstock for bioethanol production (1). Cotton stalk (CS), remain in the field after harvesting the cotton, as an agricultural residue. It needs to be removed from the field (2, 3). It is estimated that for every hectare of cotton production, 2MT of cotton stalks are generated (4). Cotton stalks, which mainly contain lignocellulose, have the potential to serve as a low-cost feedstock to increase the production of fuel ethanol. However, direct saccharification or biotransformation of the cotton stalk is extremely difficult because of the recalcitrant nature of lignocellulosics (5, 6, 7). The pretreatment is perhaps the single most crucial step as it has a large impact on all the other steps in the process, e.g. hydrolysis, fermentation, downstream processing, and wastewater handling. Many pretreatment strategies focus on lignin removal from biomass to

Comment [j3]: Rewrite sentence

Comment [j4]: The repeated

34 achieve a more efficient substrate hydrolysis process. Alkaline pretreatment limits the
35 degradation of hemicellulose polymers (8). Alkaline pretreatment was used on cotton stalks
36 for generating value-added products (9, 10). In the present study, the cotton stalk was used as
37 feedstock with ammonia pretreatment at room temperature.

38 2. MATERIALS AND METHODS:

39 2.1.Raw Material and Reagents:

40 Cotton stalks (CS) of spp. *Gossypium hirsutum* were obtained from the cotton crop land of
41 Mahabubnagar district, Telangana India. Before compositional analysis, the biomass consisted
42 primarily of stalks, which were collected, dried, debarked, and ground to 2mm particle size and
43 stored at room temperature. All chemicals were analytical grade, obtained from MERK.

Comment [j5]: Brief detail on procedure

Comment [j6]: Check spelling

44 2.2.Composition analysis:

45 The composition of cotton stalks was analyzed for holocellulose, cellulose, pentosens, klason
46 lignin, and ash content. The bark-free cotton stalk was taken and fractioned using a laboratory
47 knife mill to attain a particular size (4-10 mm). The obtained wood dust was passed through by
48 40 mesh and retained on 60 mesh and was used for proximate chemical analysis and further
49 chemical hydrolysis experiments. The chemical analyses were performed by following the
50 TAPPI test methods (11).

Comment [j7]: Check spelling

Comment [j8]: 2 mm or else?

Comment [j9]: Recheck??

Comment [j10]: Name the compounds /groups analyzed with bit detail

51

52 2.3.Pretreatment of Cotton Stalks:

53 Aqueous solutions of NH₃ at concentrations 0, 0.5, 1, and 1.5% (w/v) were used to pretreat CS
54 samples at a solid loading of 15% (w/v). preTreatments were performed in duplicate in an
55 autoclave at 121-°C with 15 psi (103.4 kPa) pressure for 60_min holding time and at room
56 temperature for a week. Clarify whether after autoclaving kept in room temperature?or vice
57 versa?

58 The pretreated solids were filtered, washed thoroughly with deionized water, dried in an air-
59 circulated oven for 16 h at 85°C, and used for the subsequent hydrolysis and fermentation
60 experiments.

61 2.4.Acid hydrolysis and detoxification of cotton stalk:

62 The pre-treated Cotton stalk CS was subjected to sulphuric acid hydrolysis. In 0.5N sulphuric
63 acid solution, pre-treated biomass (20% w/v) was treated with steam under pressure at 121°C in
64 an autoclave for 30 minutes and four-hour heat treatment at 90°C in the water bath (12). The
65 obtained acid hydrolysate was detoxified by the addition of dried lime up to pH 10 for an hour
66 and then filtered and pH was readjusted up to 6 with acid. This is followed by 2% (w/v) charcoal

67 treatment for half an hour with stirring and then filtering (12). The obtained filtrate solution was
68 used as a sole carbon source for fermentation studies.

69 **2.5.Fermentation:**

70 | The yeast *Saccharomyces cerevisiae* CP11 [paper reference or gene bank number???](#) strain
71 isolated and maintained in our laboratory was used in the study. The inoculum was prepared by
72 | growing yeast on YPD (Yeast, Peptone and Dextrose) media, for 24h at 30°C [reference??](#). The
73 prepared cultures of *Saccharomyces cerevisiae* CP11 were used as inoculum in fermentation.

74 | To acid hydrolysate (100 [ml](#)), the following were added to make fermentation media: 1.5 g yeast
75 extract, 1 g each of peptone and (NH₄)₂SO₄, 0.5 g each of K₂HPO₄, MgSO₄.H₂O and MnSO₄ at
76 pH 5.5. The medium was sterilized for 25 min at 110°C. After cooling the media, 3% inoculum
77 was added to the flask containing sterilized media. Fermentation was carried out for 96 hours at
78 30°C. Initially, shaking of 100rpm was provided for 4 hours followed by static anaerobic
79 conditions for 92 hours. The samples were collected at 24h intervals throughout the fermentation
80 | process and analyzed for ethanol content and reducing sugars [reference paper followed??](#).

81 **2.6.Analytical methods:**

82 Total Reducing sugars were estimated by DNS method of Miller (13).

83 Hydroxymethylfurfural was determined based on absorbance in spectrophotometer. An aliquot
84 of 5ml of hydrolysate dissolved in 25ml of distilled water and added to Carrez I solutions
85 | [company](#)(0.5ml) and Carrez II (0.5ml) the solution was filtered and the first 10ml was discarded.
86 From the filtrate, absorbance at 284 and 336 nm was read with an aliquot of solution filtered with
87 0.2% sodium bisulfite as blank. The HMF is determined by the equation: HMF/ 100ml of
88 | hydrolysate = (Abs284 – Abs336) × 14.97 × 5ml of the sample [reference??](#).

89 Total content of phenolic compound in hydrolysate was determined by FolinCiocalteus (FC)
90 | method (14). [Bit detail?](#)

91 Ethanol estimation:

92 Ethanol estimation in fermented broth was carried out by gas chromatography. The method uses
93 a SHIMADZU GC 2010 with a flame ionization detector. GC was carried out according to
94 | NREL procedure LAP # 011, using ZB-Wax column (30mm × 0.25mm) [flow rate of gases??](#).

95

96

97

98

Comment [j11]: Make all L caps

99 **3. RESULTS**

100 **3.1. Chemical composition of raw cotton stalks:**

101 The results revealed that the cellulose content in the raw (untreated) cotton stalk was 44.8 ± 0.55
 102 % and hemicellulose was 13.25 ± 0.50 %; whereas, the lignin content was found to be $29.6 \pm$
 103 0.75 %. Cellulose and hemicellulose content in a defined combination makes the holocellulose,
 104 which was found to be 58.05%.

105 **3.2. Pre-treatment of cotton stalks:**

106 The cotton stalks were delignified with different concentrations of NH_3 0, 0.5,1, and 1.5 in an
 107 autoclave and at room temperature. The results after pretreatment showed that the lignin content
 108 decreased with the increase in the concentration of NH_3 . The cotton stalks soaked at a
 109 concentration of 1.5% NH_3 at room temperature showed the highest delignification rate (86%) and
 110 high cellulose content (72.7%). This was subjected to further processes.

111 **Table 1: Cotton stalks composition after pretreatment at room temperature for a week**
 112 **only or?? abt autoclave??**

Concentration of NH_3 (%)	Cellulose (%)	Hemicellulose (%)	Holocellulose (%)	Lignin (%)	Delignification (%)	Furfurals(mg/L)
Control NaOH	54.2	15.20	69.4	21.48	27.43	2.1
0	44.8	13.25	58.05	29.6	0	2.8
0.5	52.5	14.8	67.3	15.4	47.97	3.4
1	60.6	15.8	76.4	9.30	68.58	4.9
1.5	72.7	16.2	84.1	4.14	86.01	5.3

113

114 **Table 2: Cotton stalks composition after pretreatment in autoclave.**

Concentration of NH_3 (%) Pretreatment strategy or else	Cellulose (%)	Hemicellulose (%)	Holocellulose (%)	Lignin (%)	Delignification (%)	Furfurals(mg/L)
Control NaOH	50	14.9	64.1	21.49	27.39	40.1
0% NH_3	48.4	14.7	63.2	23.5	20.6	42.4
0.5	55.4	15.2	69.6	18.5	37.5	44.7
1	67.8	15.6	72.2	12.6	57.43	50.2
1.5	70.1	15.9	78.6	9.4	68.24	55.2

115

116

Formatted: Subscript

117
118
119
120
121

3.3. Acid hydrolysis and detoxification of cotton stalk:

122 Cotton stalks after pre-treatment at room temperature for a week:

123 The reducing sugars were 67.51 ± 0.21 g and the total sugar were 77.21 ± 0.5 g in acid
124 hydrolysate obtained with 100g pre-treated substrate with a maximum saccharification efficiency
125 of 91.1% And the total phenols were found to be 5.3mg/100g substrate. Calculate in per gram??
126 Or %

127 Cotton stalks after pre-treatment in autoclave:

128 The reducing sugars were 59.11 ± 0.2 g and the total sugars were $61.05 \pm .75$ g in acid
129 hydrolysate obtained with 100g make /g pre-treated substrate with a maximum saccharification
130 efficiency of 77.6% and the total phenols were found to be 55.2mg/100g substrate.

131 3.4. Fermentation:

132 Cotton stalks after pre-treatment at room temperature for a week:

133 Total reducing sugars in 100 ml fermentation medium was 13.5 grams. ???? Maximum ethanol
134 concentration and reducing sugar consumption was found at 48 h of fermentation. The leftover
135 sugar was 1.26g and the consumed sugar was 12.24g/100ml ?? g?? fermentation medium. The
136 maximum ethanol concentration produced was 5.75 % with an ethanol yield of 0.469g/g. A
137 fermentation efficiency of 91.78% was achieved.

138 Cotton stalks after pre-treatment in autoclave:

139 Total reducing sugars in 100 ml fermentation medium was 11.82 grams. Maximum ethanol
140 concentration and reducing sugar consumption rate was found at 48 h of fermentation. The
141 leftover sugar was 3.02g and the consumed sugar was 8.8 g/100ml fermentation medium. The
142 maximum ethanol concentration produced was 4% with an ethanol yield of 0.454g/g. A
143 fermentation efficiency of 88.84% was achieved.

144 Better show all in table???

145 4. DISCUSSION

146 In the present study, the chemical composition of the cotton stalk was cellulose (44.8%),
147 hemicellulose (13.25%), and lignin (29.6%). The almost similar composition was found in

148 | earlier studies from Greece (15), Pakistan (16), and India (17) [their composition??](#). The
149 | hardwoods had similar lignin content (18%-30%) as cotton stalks, whereas the herbaceous
150 | plants had lower lignin content (10%-20%) than cotton stalks. [When](#) The cotton stalks were
151 | delignified with different concentrations of NH₃ 0, 0.5, 1, and 1.5 in an autoclave and at room
152 | temperature in our study. Cotton stalks were pretreated with different concentrations of
153 | NaOH ranging from 0 to 10% (w/w, g of NaOH/100 g CS) at 15% (w/v) substrate
154 | concentration in autoclave at 121°C /15 psi for 60 min (7). Cotton stalks were soaked for 1
155 | hour in 1L (2%) NaOH solution in 3 flasks and autoclaved at the residence times of 30, 60,
156 | and 90 min at constant temperature of 121°C with 15 psi pressure (16). The cotton stalk was
157 | subjected to dual-stage sulfuric acid treatment (12). Ammonia pretreatment at room
158 | temperature was found effective in delignification. In this study, the content of reducing
159 | sugars in acid hydrolyzate was 67.51 ± 0.21 g/L with the maximum saccharification
160 | efficiency of 91.8%, the total sugars content was 77.21 ± 2.37 g/L and phenol content was
161 | 5.3mg/100g substrate. Maximum values of glucose obtained at enzymatic hydrolysis at
162 | different enzyme loads were 9.89g/L to 68.19g/L (18). The highest reducing sugar values
163 | were 67.25 ± 1.62 g/L obtained after 72 h of hydrolysis with a saccharification efficiency of
164 | 77.39 % (13). The detoxified hydrolysate formed, contained a sugar concentration of 11 g/L,
165 | and corresponds to a yield of 0.396 g/g of biomass (12). The maximum ethanol
166 | concentration of 5.75% with the ethanol yield of 0.469 g/g after 48 h of incubation at 30°C
167 | with pH 5.5 was achieved. Maximum values of ethanol and ethanol yield according to
168 | prehydrolysis time and substrate's concentration ranging 15.95-34.8 and 10.63-17.4 (15). The
169 | ethanol concentration and yield increased at first 48 h and started to decrease after 48 h. The
170 | highest ethanol concentration was 22.93 ± 1.74 g/L with 0.36 g/g ethanol yield at 62.2 %
171 | reducing sugars consumption rate (16). A Peak ethanol concentration of 3.94 g/L
172 | (corresponds to a yield of 0.355 g/g of available sugar) was achieved after 36 h of
173 | fermentation as reported by Wendhausen et al. (19). Fermentation efficiency of cotton stalks
174 | pretreated at room temperature and in the autoclave was reported as 91% and 88%
175 | respectively. The fermentation efficiency was 55.4% in (NSSF) Non-isothermal
176 | Simultaneous Saccharification and Fermentation followed after 14h pre-hydrolysis (15). A
177 | fermentation efficiency of 69.53% was reported by Mirza Zaheer Baig and Smita M.
178 | Dharmadhikari (12). 78.06% of Saccharification efficiency was reported by K Shahzad et al.
179 | (16). Cotton stalks pretreated at room temperature (5.75%) produced more ethanol than
180 | cotton stalks pretreated in an autoclave (4%) because there is a formation of furfurals at
181 | larger amounts in the case of autoclave pretreated cotton stalks. Traditional methods of pre-
182 | treatment at higher temperatures are releasing more furfurals. These furfurals are known to
183 | inhibit or reduce fermentation efficiency. So in this connection, it is aimed to perform pre-
184 | treatment at ambient room temperature with ammonia. This has yielded high efficiency of
185 | treatment, acid hydrolysis, and high ethanol fermentation efficiency.

186 | 5. CONCLUSION

187 | Ammonia pre-treatment at room temperature for extended time can a suitable pre-treatment
188 | method for lignocellulosic materials in general and cotton stalks in particular for bioethanol
189 | production. 1.5% ammonia pre-treatment at room temperature for 1 week showed 86%
190 | delignification and subsequently undergone 91% acid hydrolysis. The acid hydrolysate
191 | fermented to 5.75% ethanol with 91% fermentation efficiency

193 REFERENCES

- 194 1. Dien BS, Iten LB, Bothast RJ. Conversion of corn fiber to ethanol by recombinant *E.coli*
195 strain FBR3. *J Ind Microbiol.* 1999; 22:575-581.
- 196 2. Haykir NI, Bakir U. Ionic liquid pretreatment allows utilization of high substrate loadings
197 in enzymatic hydrolysis of biomass to produce ethanol from cotton stalks. *Ind Crop Prod.*
198 2013; 51:408-414.
- 199 3. Kaur U, Oberoi HS, Bhargav VK, Sharma-Shivappa R, Dhaliwal SS. Ethanol production
200 from alkali- and ozonetreated cotton stalks using thermotolerant *Pichia kudriavzevii*
201 HOP-1. *Ind Crop Prod.* 2012; 37:219-226.
- 202 4. Binod P, Kuttiraja M, Archana M, Janu KU, Sindhu R, Sukumaran RK, Pandey A. High
203 temperature pretreatment and hydrolysis of cotton stalk for producing sugars for
204 bioethanol production. *Fuel.* 2012; 92:340-345.
- 205 5. Du SK, Su X, Yang W, Wang Y, Kuang M, Ma L, Fang D, Zhou D. Enzymatic
206 saccharification of high pressure assist alkali pretreated cotton stalk and structural
207 characterization. *Carbohydr Polym.* 2016; 140:279-286.
- 208 6. Du SK, Zhu X, Wang H, Zhou D, Yang W, Xu H. High pressure assist-alkali
209 pretreatment of cotton stalk and physicochemical characterization of biomass. *Bioresour*
210 *Technol.* 2013; 148:494-500.
- 211 7. Silverstein RA, Chen Y, Sharma-Shivappa RR, Boyette MD, Osborne JA. comparison of
212 chemical pretreatment methods for improving saccharification of cotton stalks. *Bioresour*
213 *Technol.* 2007; 98:3000-3011.
- 214 8. Singh R, Shukla A, Tiwari S, Srivastava M. A review on delignification of
215 lignocellulosic biomass for enhancement of ethanol production potential. *Renew Sust*
216 *Energy Rev.* 2014; 32:713-728.
- 217 9. Bahcegul E, Toraman HE, Ozkan N, Bakir U. Evaluation of alkaline pretreatment
218 temperature on a multi-product basis for the co-production of glucose and hemicellulose
219 based films from lignocellulosic biomass. *Bioresour Technol.* 2012; 103:440-445.
- 220 10. Vani S, Binod P, Kuttiraja M, Sindhu R, Sandhya SV, Preeti VE, Sukumaran RK, Pandey
221 A. Energy requirement for alkali assisted microwave and high pressure reactor
222 pretreatments of cotton plant residue and its hydrolysis for fermentable sugar production
223 for biofuel application. *Bioresour Technol.* 2012; 112:300-307.
- 224 11. TAPPI (Technical Association of Pulp and Paper Institute, Atlanta, Georgia, USA. 1992)
- 225 12. Mirza Zaheer Baig, Smita M Dharmadhikari. Bioethanol Production from Cotton Stalk
226 Hydrolysate using Immobilized Co culture of *Saccharomyces cerevisiae* and *Pachysolen*
227 *tannophilus*. *Int J Curr Microbiol App Sci.* 2016; 5(12): 389-397.
- 228 13. Miller GL. Use of dinitro salicylic acid reagent for determination of reducing sugar. *Anal*
229 *Chem.* 1959; 31:426-428.

- 230 **14.** Singleton VL, Rossi JA. Colorimetric of total phenolics with phosphomolybdic-
231 phosphotungstic acid reagents. *Am. J. Enol. Viticult.* 1965; 16:144-158.
- 232 **15.** Despoina Chilari, Konstantinos Dimos, Georgia Georgoula, Thomas Paschos,
233 Diomi Mamma, Argiro Louloudi, Nikolaos Papayannakos, Dimitris Kekos. Bioethanol
234 Production from Alkali-Treated Cotton Stalks at High Solids Loading Applying Non-
235 isothermal Simultaneous Saccharification and Fermentation. *Waste Biomass Valor.* 2017;
236 8:1919–1929.
- 237 **16.** Shahzad K, Sohail M, Hamid A. Green ethanol production from cotton stalk. *IOP Conf*
238 *Series: Earth and Environmental Science.* 2019; 257:012025.
- 239 **17.** Praveen Kumar Keshav, Chandrashekhar Banoth, Srinivas Naik, Kethavath, Bhima
240 Bhukya. Lignocellulosic ethanol production from cotton stalk: an overview on
241 pretreatment, saccharification and fermentation methods for improved bioconversion
242 process. *Biomass Conversion and Biorefinery.* 2021; 1-18.
- 243 **18.** Vazquez M, Oliva M, Tellez-Luis SJ, Ramírez JA. Hydrolysis of sorghum straw using
244 phosphoric acid: evaluation of furfural production. *Bioresour Technol.* 2007; 98:3053–
245 3060.
- 246 **19.** Wendhausen R, Fregonesi A, Moran PJS, Joekes I, Rodrigues JA, Tonella E, Althoff K.
247 Continuous fermentation of sugar cane syrup using immobilized yeast cells. *J Biosci*
248 *Bioeng.* 2001; 9(1): 48-52.

249
250