

Original Research Article

Flat Tummy Water Attenuates Lipid Profile and Serum Glucose of High-Fat Diet-Induced Obese Female Wistar Rats

Abstract

Background

Flat tummy water (FTW) is herbal tonic preparation made from cucumber (*Cucumis sativus*), lemon (*Citrus limon*), ginger (*Zingerber officinale*) and mint leaf (*Mentha piperita*) and is believed to be effective in the reduction of central/abdominal fat. While the potency and efficacy of FTW are hardly known, the present study presents a scientific investigation of its effect on high-fat-induced obese Wistar rat models.

Method

FTW was constituted using measured quantities of cut and infused cucumber, lemon and mint leaf while the individual plants were extracted and mixed to get an extract mixture (EM). Twenty-five (25) female Wistar rats were used for the study. They were divided into five (5) groups of five (5) animals each. Groups 1 & 2 served as the normal and negative controls and received standard rat chow/distilled water and high-fat diet/distilled water respectively. Groups 3, 4 & 5 served as the experimental groups. Group 3 received FTW *ad libitum* while groups 4 & 5 received 25 and 50mg/kg of extract mixtures (EM) respectively alongside the formulated high-fat diet. Lipid profile and serum glucose were determined using standard colourimetric methods. The body weight, naso-anal length, abdominal and thoracic circumferences (AC/TC) of the animals were measured before and after the experiment.

Result

There was a significantly reduced total cholesterol, triglycerides, low-density lipoproteins, very low-density lipoproteins and serum glucose and significantly increased high-density proteins among the obese animals treated with FTW and EM compared to the untreated obese negative control ($p < 0.05$). The study also observed reduced body weight, AC/TC ratio and lee obesity index among treated obese animals when compared with the untreated negative control.

Conclusion

The present study shows that flat tummy water (FTW) as well as extracts mixtures (EM) from the component of FTW indeed caused a mild reduction in central abdominal obesity. FTW and EM showed potent hypolipidaemic, hypoglycaemic and anti-obesity activity. This study concludes that these observed effects could be due to the synergistic action of cucumber (*Cucumis sativus*), lemon (*Citrus limon*), ginger (*Zingerber officinale*) and mint leaf (*Mentha piperita*).

Key Words: flat tummy water, lipid profile, serum glucose, AC/TC ratio, lee index

INTRODUCTION

The incidence of obesity has received global attention as a health burden and preceding factor in cases of terminal illnesses like diabetes, cancer, cardiovascular diseases and some neurological disorders [1, 2]. Over twenty (20) years ago, the World Health Organization (WHO) declared obesity a global epidemic with more than 39% of the adult population affected [3, 4]. Obesity results from the interplay between nature (genetic makeup) and nurture (high-energy diets and sedentary lifestyles) [5, 6]. In the face of the increased economic burden associated with the management of obesity, there has been an increase in the use of natural agents derived from plants, herbs and vegetables as potential solutions for achieving weight loss [7-9]. These preparations are used alone or in combination with other plants, herbs and vegetables. Some of the plants screened for their anti-obesity potential include *Amorphophallus koniac*, *Camellia Sinensis*, *Nigella sativa*, *Hibiscus, sabdariffa*, *Ginkgo biloba*, *Phaseolus vulgaris* and *Dioscorea bulbifera* [8, 10, 11]. The mechanism of action of some of these natural preparations includes improving lipid metabolism, appetite suppression and anti-lipase activity [7, 8, 11-13].

There are many other plant preparations which are yet to undergo scientific evaluation for their efficacy as anti-obesity agents. One such is the widely acclaimed “flat tummy water” (FTW). The origin of the FTW water is largely unknown and has continued to gain popularity in society, especially among the many health, wellness and fitness centres and online blogs. As the name suggests, it is said to be an indigenous herbal tonic preparation used specifically in the reduction of central/abdominal fat. Though a few variations on the composition of FTW water exist, it is typically composed of cucumber (*Cucumis sativus*), lemon (*Citrus limon*), ginger (*Zingiber officinale*) and mint leaf (*Mentha piperita*). They are cut and soaked in water for about eight (hours) after which the aqueous mixture is regularly taken to attain a slimmer body shape, especially in the abdominal region [14, 15].

Cucumber (*Cucumis sativus*) is the most widely cultivated plant from the family of *Cucurbitaceae*. It is a cylindrical creeping fruit which is used in salads and also for medicinal purposes. The fruit is used in the folk remedy treatment of bronchitis, hepatitis, asthma and dyspepsia [16-19]. It has also been shown to possess hypolipidaemic, wound healing, anti-cancer, anti-diabetic and hypolipidaemic properties [19-23]. Lemon (*Citrus limon*) is a flowering plant from the family of *Rutaceae*. Lemon juice is consumed as a homemade drink as lemonade and cocktails. It is also used as a cleaning agent and as a preservative for baking. Folk medicinal applications include uses as an appetizer, treatment for a boil, throat and tonsillar abscesses, anthelmintic and remedy against drunkenness [24, 25]. Lemon has also been shown to possess antibacterial, hypolipidaemic, hepatoprotective and antioxidant properties [26-29]. Ginger (*Zingiber officinale*) is a herbaceous flowering plant commonly known by its root or rhizome. It belongs to the family *Zingiberaceae* and is mostly used as a spice and in folk medicine for the treatment of cough, rheumatism, nausea, asthma and loss of appetite [30, 31]. Mint (*Mentha piperita*) is a strongly scented glabrous perennial herb thought to be a natural hybrid between watermint (*Mentha aquatica*) and spearmint (*Mentha spicata*), all from the family of *Lamiaceae* [32, 33]. Their scented nature makes them applicable as a flavouring and cosmetic agent with varying uses in traditional medicine such uses in the treatment of headaches, fever, influenza and

red eyes [33, 34]. Some of the pharmacological effects of the mint leaf include antibacterial, anti-nociceptive, antioxidant, hepato-protective and anti-carcinogenic activity [35-39].

With increasing testimonies highlighting the efficacy of FTW as an effective herbal tonic in reducing truncal adiposity, the present study, therefore, is aimed at evaluating the effect of flat tummy water on lipid profile, body weight and body mass index using female Wistar models. This study is an attempt to provide a scientific basis for the use of FTW.

MATERIALS AND METHODS

Plant Materials

Freshly harvested cucumber, lemon, ginger and mint leaf were sourced from the Fruit Garden Market, Port-Harcourt, Nigeria. The plants were identified and authenticated by a botanist at the Plant Science and Biotechnology Department of the University of Port Harcourt.

Preparation of Flat Tummy Water

The FTW was prepared according to the already existing formulation using 550g of cucumber, 130g of lemon, 10g of ginger and 15g of mint leaves [14, 15]. The plants were cut into and soaked in 6L of water for eight (8) hours. The mixture was refrigerated at 5°C pending administration.

Preparation of Plant Extracts and Phytochemical Screening

Measured quantities of cucumber, lemon, ginger and mint leaves were constituted according to flat tummy formulation, sliced and air-dried for two (2) weeks. Individual grounded plant materials (1kg) were subjected to ethanolic extraction using standard methods. [40].

Phytochemical screening of each plant was performed by methods described by Trease and Evans [41]. The extracts were mixed and re-constituted to simulate the standard FTW composition.

Research Animals

Fifteen (15) female mice (20-30g) and twenty-five (25) female Wistar rats (155 – 214g) were used for the study. They were sourced from the animal house of the Department of Human Physiology, Faculty of the Basic Medical Sciences University of Port Harcourt. They were allowed two (2) weeks of acclimatization under the natural light/dark cycle and access to standard rat chow and water *ad libitum*.

Determination of LD₅₀ of Plant Extract Mixture (EM)

An acute toxicity test of the plant EM was carried out using fifteen (15) female mice according to the method described by Karber [42].

Induction of Obesity

A high-fat diet was formulated using a standard rat chow (80%) and rendered cow fat. The animals were allowed to feed on the formulated high-fat diet for seven (7) weeks. The length and weight of the animals were measured and Lee obesity index was calculated as follows:

$$\text{Lee Obesity Index} = \frac{\sqrt[3]{\text{weight}(g)}}{\text{naso} - \text{anal length (cm)}}$$

Abdominal and thoracic circumferences were also measured to obtain an abdominal/thoracic ratio (AC/TC ratio) as follows:

$$\text{AC/TC ratio} = \frac{\text{Abdominal Circumference (cm)}}{\text{Thoracic Circumference (cm)}}$$

After seven (7) weeks on a high-fat diet, animals with up to 35% weight increase or a Lee obesity of >0.30 were considered obese and selected for the study [43, 44].

Experimental Design

Thirty (25) female Wistar rats were used for the experiment. They were assigned to five (5) groups of five (5) animals each and treated as follows

Groups	Name	Treatment
1	Control	Normal rat chow + Distilled water
2	Negative Control	High-fat diet + Distilled water
3	FTW	High-fat diet + FTW (<i>ad libitum</i>)
4	Low Dose EM	High-fat diet + 25mg/kg EM
5	High Dose EM	High-fat diet + 50mg/kg EM

The oral administration of the FTW, plant extract mixture (EM) lasted for fourteen (14) days. Animals in group 3 (High-fat diet + FTW) were allowed to drink from the formulated FTW *ad libitum*.

Determination of food Intake

The quantity of rat chow left after twenty-four (24) hours of feeding were collected and measured three (3) times a week to determine food intake as follows

$$\text{Food intake} = \frac{\text{Amount of chow given} - \text{Amount of chow left after 24 hours}}{\text{Number of animals in a cage}}$$

Blood Collection and Analysis

The animals were anaesthetised by cervical dislocation following overnight fasting. The blood sample was collected by cardiac puncture and analysed for triglycerides (TG), total cholesterol (TC) and high-density lipoproteins (HDL) were determined using standard spectrophotometric methods while fasting blood glucose was measured using a standard glucometer (Accu-Chek,

UK). Low-density lipoproteins (LDL), and very-low-density lipoproteins (VLDL) are estimated using the standard formula [45, 46].

Statistical Analysis

The statistical analysis was done using IBM Statistical Product and Service Solutions (SPSS) version 25. The only-way ANOVA was used to determine the difference among the groups followed by Fisher's Least Significant Difference (LSD). A paired t-test was used to determine the difference in initial and final body weight, AC/TC ratio and Lee body index of the animals. A p-value of less than 0.05 was considered statistically significant.

UNDER PEER REVIEW

RESULTS

Table 1: Qualitative Phytochemical Screening of ethanolic extracts of cucumber (*Cucumis sativus*), lemon (*Citrus limon*), ginger (*Zingerber officinale*) and mint leaf (*Mentha piperita*).

Phytochemical	Test	Cucumber	Lemon	Ginger	Mint leaf
Alkaloids	Meyers	+	-	++	++
	Drangendorffs	+	++	++	++
	Hagers	+	+	++	++
Tannins	Ferric chloride	+	++	++	+
	Frothing	-	+	++	++
Saponin	Emulsion	-	+	-	++
	Shinola	+	+	++	++
Flavonoids	NaOH	+	+	++	++
	Molisch	+	+	++	++
Carbohydrate	Fehlings solution	+	+	++	++
	Free anthraquinone	-	+	-	-
Anthraquinones	Combined	+	+	+	-
	Keller killiani	+	+	++	+
	Liebernzano	-	-	++	+
Cardiac glycoside	Salkwoski	+	-	++	-
	Keddeksp	+	-	++	-
	Cyanogenic glycoside	-	-	-	-
Fixed oil		-	++	++	++
Phenols		++	++	++	++

= Absent, + = Present, ++ = Largely Present

Qualitative Phytochemical Screening of ethanolic extracts of Cucumber, Lemon, Ginger and Mint leaf (Table 1) show the presence of alkaloids, tannins, flavonoids, carbohydrate and phenols among all the extracts with ginger and mint leaf having higher quantities of these phytochemicals. It was observed that saponin was not detected in cucumber extract while mint leaf extract did not contain any detectable anthraquinones. Cardiac glycosides were not found in any of the extracts

Table 2: Effect of flat tummy water (FTW) and extract mixture (EM) on lipid profile of high-fat diet obese Wistar rats

Groups	TC (mmol/l)	TG (mmol/l)	HDL (mmol/l)	LDL (mmol/l)	VLDL (mmol/l)	Glucose (mmol/l)
Control	1.86±0.51	0.96±0.80	0.52±0.42	0.90±0.41	0.44±0.04	6.12 ± 0.47
Negative control	2.94±0.86	1.57±0.10	0.39±0.18	1.52±0.86	0.71±0.05	9.58 ± 0.34
Positive control	2.14±0.51	1.28±0.45	0.45±0.37	1.11±0.40	0.58±0.02	6.76 ± 0.43
FTW	2.31 ^a ±0.14	1.16 ^a ±0.23	0.59 ^{ab} ±0.59	1.36 ^b ±0.13	0.53 ^a ±0.01	6.04 ^a ± 0.56
25mg/kg EM	2.24 ^a ±0.14	1.27 ^a ±0.46	0.51 ^a ±0.37	1.35±0.46	0.58 ^a ±0.04	7.16 ^a ± 0.42
50mg/kg EM	2.21 ^a ±0.15	1.21 ^a ±0.92	0.66 ^{ab} ±0.30	1.28 ^a ±0.36	0.55 ^a ±0.41	7.88 ^a ± 0.34

mean±standard error of mean. significantly different compared to negative^a and positive^b control

Table 2 shows the effects of FTW and the ethanolic mixture of cucumber, lemon, ginger and mint leaf. Mean values of serum total cholesterol (TC), triglycerides, low-density lipoproteins (LDL), very low-density lipoproteins (VLDL) and glucose were found to be significantly lower among the obese animals treated with FTW (Group 4) and the EMs (Groups 5&6) compared to the negative control ($p<0.05$). In the same way, mean values of serum high-density lipoproteins (HDL) among experimental groups (Groups 4, 5 & 6) were found to be significantly higher compared to the negative control (group 2) ($p<0.05$).

Table 3: Effect of flat tummy water (FTW) and extract mixture (EM) on Body weight, AC/TC ratio and Lee index of high-fat diet obese Wistar rats

Groups	Body Weight (kg)		AC/TC ratio		Lee Index	
	Initial	Final	Initial	Final	Initial	Final
Control	202.00±9.71	216.00±10.07	1.12±0.01	1.11±0.01	0.295±0.004	0.298±0.004
Negative control	155.00±4.17	168.00±6.23	1.09±0.01	1.08±0.02	0.305±0.003	0.308±0.002
Positive control	183.40 ±3.06	174.80±4.02	1.10±0.02	1.06±0.02	0.308±0.004	0.297±0.002
FTW	194.40±2.69	190.80 ^a ±3.56	1.13±0.01	1.07 ^a ±0.01	0.301±0.001	0.302±0.003
25mg/kg EM	201.20±3.38	199.00±3.69	1.14±0.11	1.08 ^a ±0.01	0.303±0.001	0.296±0.001
50mg/kg EM	214.20±8.08	212.00±6.16	1.13±0.31	1.03 ^a ±0.01	0.304±0.001	0.293 ^a ±0.002

mean±standard error of mean. significantly different compared to the initial (P<0.05)

The effect of FTW and EM on body weight, AC/TC ratio and Lee index of high-fat diet obese Wistar rats are shown in table 3 above. The results indicate that there was a significant reduction in the final body weight, AC/TC ratio of obese animals treated with FTW and EM compared to their initial (P<0.05). Only obese rats treated with 50mg/kg of EM had a significant reduction in their final Lee index compared to the initial (P<0.05).

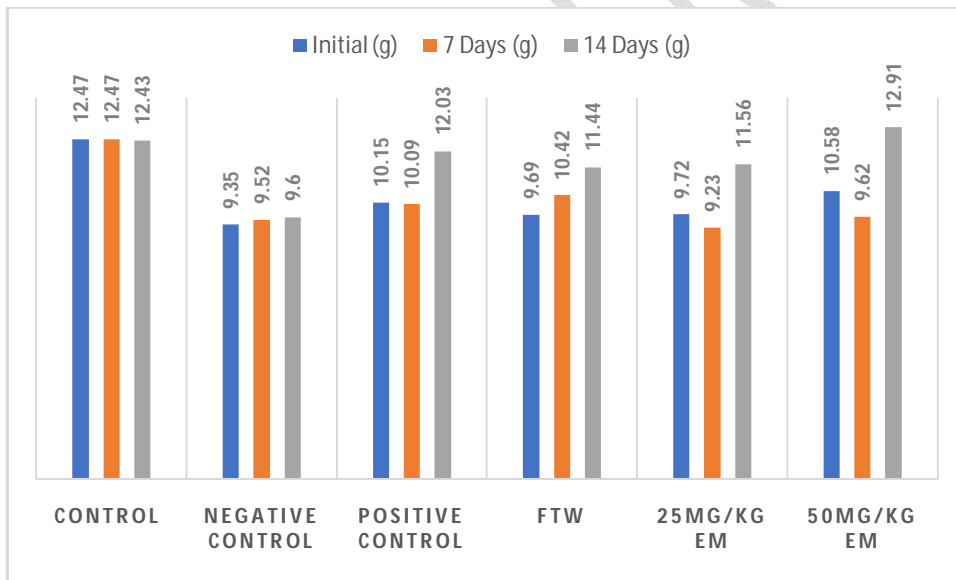


Figure 1: Effect of flat tummy water (FTW) and extract mixture (EM) on food consumption of high-fat diet obese Wistar rats

The effect of FTM and EM on the food consumption of high-fat diet obese Wistar rats is shown in Figure 1 above. The results indicate that there was a slight reduction in food consumption among animals treated with FTW and EMs after the first week of administration (p<0.05).

However, the food consumption significantly increased after the second week of administration compared to the initial food consumption ($p < 0.05$)

DISCUSSION

Flat tummy water (FTW) is herbal tonic preparation made from cucumber (*Cucumis sativus*), lemon (*Citrus limon*), ginger (*Zingiber officinale*) and mint leaf (*Mentha piperita*) and is believed to be effective in the reduction of central/abdominal fat. While the potency and efficacy of FTW are hardly known, the present study presents a scientific investigation of its effect on high-fat-induced obese Wistar rat models.

Effect on Lipid Profile

The result from the study shows a significantly reduced serum TC, LDL and VLDL among obese animals treated with FTW and extract mixture (EM) of cucumber, citrus lemon and ginger compared to the untreated animals (negative control) while serum HDL increased among FTW and EM treated animals compared to the negative control ($p < 0.05$). This is attributable to the similarity of the individual phytochemical constituents of FTW such as flavonoids, tannins, alkaloids and saponins (Table 1). Flavonoids are known to impact lipid metabolism by reducing cholesterol synthesis or increasing LDL receptor expression [47, 48]. Also, saponins have been shown to suppress cholesterol absorption, increase lipid peroxidation and depress the rates of intestinal hepatic and intestinal cholesterol synthesis [48, 49]. *Cucumis sativus* has been demonstrated as a potent antihyperlipidemic agent in both human and animal studies [23, 50]. This is heavily linked to the linoleic acid and resin content of their seeds which have been severally demonstrated to lower LDL and TG [51, 52]. Similarly, *Citrus limon* is effective in attenuating lipid profiles due to its antioxidant nature and its ability to modulate lipid peroxidation [53-55]. Furthermore, *Mentha piperita* also possesses antihyperlipidemic activities as has been previously demonstrated [49, 56, 57]. This has been attributed to its constituent oils: menthofuran, menthol, menthyl acetate, neomenthol, menthone and isomenthone which are known to exhibit antioxidant and lipid peroxidation activity [56]. Hence, the improved lipid profile observed in this study suggests a synergistic antihyperlipidemic activity of the constituents of FTW and EM.

Effect on Serum Glucose

Similarly, data (Table 2) from the present study indicate that serum glucose was significantly reduced among obese animals treated with FTW and EM compared to the untreated animals (negative control) ($P < 0.05$). The similar phytochemical constituents of FTW and EM suggest that they could have a glucose-lowering effect. *Cucumis sativus* extracts are effective in attenuating blood sugar levels among diabetic animals [58-60]. They are thought to increase glycolysis and peripheral insulin sensitivity [60, 61]. In the same way, *Citrus limon* has also been shown to be a potent antihyperglycaemic agent against diabetic animals [55, 62]. This is thought to be by improving glucose tolerance, reducing insulin resistance and restoring oxidant balance [62, 63]. Also, *Citrus limon* has been demonstrated to decrease blood glucose levels among diabetic animals [64, 65] by improving insulin secretion, and reducing lipid peroxidation and oxidative stress [56, 66]. These glucose-attenuating actions of FTW and EM can be due to the

improved lipid metabolism as observed in this study. Randle's glucose-fatty acid cycle postulates that an increase in plasma triglycerides leads to an increase in free fatty acid (FFA), hence their oxidation tends to weaken glucose metabolism and insulin action, leading to hyperglycaemia. Therefore, the attenuation of FFA will lead to a reduction in plasma glucose [67]. Therefore, the improved serum glucose observed in this study suggests a synergistic antihyperglycaemic activity of the constituents of FTW and EM.

Effect on Body weight, AC/TC ratio, Lee Index and Food Intake

Data from the present study show that FTW significantly reduced the final body weight and AC/TC ratio of the obese animals while 50mg/kg of the EM significantly reduced the final Lee index of the obese animals (Table 3). The AC/TC ratio and the Lee Index serve as anthropometric obesity measurements for laboratory animals [44, 68]. A reduction in the AC/TC ratio suggests a decrease in central abdominal fat while a decrease in the Lee index a reduction in adipose tissue among the obese animals treated with FTW and EM. This goes further to show the effect of reduced plasma lipids as observed in this study (Table 2). There was a slight reduction in food intake for the obese animals treated with FTW and EM in the first week of the administration followed by a significant rise in food consumption within the last week of the experiment. This slight reduction in food intake appears to follow the second stage observed among high fat-induced obesity which is characterized by a reduction in food intake due to increased leptin production with retained central leptin sensitivity. The third stage involves increased food intake with a reduction in central leptin sensitivity [69].

CONCLUSION

The present study shows that flat tummy water (FTW) as well as mixtures of the extracts (EM) from the component of FTW indeed caused a mild reduction in central abdominal obesity. FTW and EM showed potent hypolipidaemic, hypoglycaemic and anti-obesity activity. This study concludes that these observed effects could be due to the synergistic action of cucumber (*Cucumis sativus*), lemon (*Citrus limon*), ginger (*Zingerber officinale*) and mint leaf (*Mentha piperita*).

Ethical Approval

The animals were treated according to the highest ethical standard as the experimental design and protocol were approved by the Centre of Research Management and Development (CRMD) of the University of Port Harcourt. (UPH/CEREMAD/REC/04)

REFERENCES

1. Wilson PWF, D'Agostino RB, Parise H, Sullivan L, Meigs JB. Metabolic syndrome as a precursor of cardiovascular disease and type 2 diabetes mellitus. *Circulation*. 2005;112(20):3066-72.

2. Reilly JJ, Methven E, McDowell ZC, Hacking B, Alexander D, Stewart L, et al. Health consequences of obesity. *Archives of disease in childhood*. 2003;88(9):748-52.
3. Jackson SE, Llewellyn CH, Smith L. The obesity epidemic–Nature via nurture: A narrative review of high-income countries. *SAGE open medicine*. 2020;8:2050312120918265.
4. Organization WH. Obesity: preventing and managing the global epidemic. 2000.
5. Billington CJ, Epstein LH, Goodwin NJ, Hill JO, Pi-Sunyer FX, Rolls BJ, et al. Overweight, Obesity, And Health Risk. *Archives of Internal Medicine*. 2000;160(7):898-904.
6. Kopelman PG. Obesity As A Medical Problem. *Nature*. 2000;404(6778):635-43.
7. Kazemipoor M, Radzi CWJWM, Cordell GA, Yaze I. Potential Of Traditional Medicinal Plants For Treating Obesity: A Review. *arXiv*. 2012;1208.1923:1-8.
8. Moro CO, Basile G. Obesity And Medicinal Plants. *Fitoterapia*. 2000;71:S73-S82.
9. Kazemipoor M, Cordell GA, Sarker MMR, Radzi CwJBWM, Hajifaraji M, En Kiat P. Alternative treatments for weight loss: Safety/risks and effectiveness of anti-obesity medicinal plants. *International Journal of Food Properties*. 2015;18(9):1942-63.
10. Hasani-Ranjbar S, Jouyandeh Z, Abdollahi M. A systematic review of anti-obesity medicinal plants-an update. *Journal of Diabetes & Metabolic Disorders*. 2013;12(1):1-10.
11. Ikete PW, Chinko BC. Inhibition of Serum Lipase as a Mechanism of Action for Anti-Obesity Potentials of *Dioscorea bulbifera* Extracts on Wistar Rats. *Asian Journal of Health Sciences*. 2022;8(1):ID29-ID.
12. Marrelli M, Statti G, Conforti F. A Review Of Biologically Active Natural Products From Mediterranean Wild Edible Plants: Benefits In The Treatment Of Obesity And Its Related Disorders. *Molecules*. 2020;25(3):649.
13. Verma RK, Paraidathathu T. Herbal Medicines Used In The Traditional Indian Medicinal System As A Therapeutic Treatment Option For Overweight And Obesity Management: A Review. *Int J Pharm Pharm Sci*. 2014;6(2):40-7.
14. Ali S. Home Made Detox Water / Flat Belly Water. *Spoon, Fork and Food*; 2016.
15. Jaiswal S. How To Prepare Flat Tummy Water And How It Works. *Health Tips, Weight Loss Tips: Ayurveda*; 2020.
16. Sahu T, Sahu J. *Cucumis sativus* (cucumber): A review on its pharmacological activity. *Journal of Applied Pharmaceutical Research*. 2015;3(1):4-9.
17. Sharma S, Dwivedi J, Paliwal S. Evaluation of antacid and carminative properties of *Cucumis sativus* under simulated conditions. *Scholars Research Library Der Pharmacia Lettre*. 2012;4(1):234-9.
18. Mallik J, Das P, Das S. Pharmacological activity of *Cucumis sativus* L.–a complete overview. *Asian Journal of Pharmaceutical Research and Development*. 2013;1(1):1-6.
19. Karthiyayini T, Kumar R, Kumar KS, Sahu RK, Roy A. Evaluation of antidiabetic and hypolipidemic effect of *Cucumis sativus* fruit in streptozotocin-induced-diabetic rats. *Biomedical and Pharmacology Journal*. 2015;2(2):351-5.
20. Patil MVK, Kandhare AD, Bhise SD. Pharmacological evaluation of ameliorative effect of aqueous extract of *Cucumis sativus* L. fruit formulation on wound healing in Wistar rats. *Chronicles of young scientists*. 2011;2(4).
21. Patil MVK, Kandhare AD, Bhise SD. Effect of aqueous extract of *Cucumis sativus* Linn. fruit in ulcerative colitis in laboratory animals. *Asian Pacific Journal of Tropical Biomedicine*. 2012;2(2):S962-S9.

22. Agarwal M, Kumar A, Gupta R, Upadhyaya S. Extraction of polyphenol, flavonoid from *Emblica officinalis*, *Citrus limon*, *Cucumis sativus* and evaluation of their antioxidant activity. *Oriental Journal of Chemistry*. 2012;28(2):993-8.
23. Soltani R, Hashemi M, Farazmand A, Asghari G, Heshmat Ghahdarijani K, Kharazmkia A, et al. Evaluation of the Effects of *Cucumis sativus* Seed Extract on Serum Lipids in Adult Hyperlipidemic Patients: A Randomized Double-Blind Placebo-Controlled Clinical Trial. *Journal of food science*. 2017;82(1):214-8.
24. Arias BA, Ramón-Laca L. Pharmacological properties of citrus and their ancient and medieval uses in the Mediterranean region. *Journal of Ethnopharmacology*. 2005;97(1):89-95.
25. Mohanapriya M, Ramaswamy L, Rajendran R. Health and medicinal properties of lemon (*Citrus limonum*). *International Journal of Ayurvedic and Herbal Medicine*. 2013;3(1):1095-100.
26. Saeb S, Amin M, Gooybari RS, Aghel N. Evaluation of antibacterial activities of *Citrus limon*, *Citrus reticulata*, and *Citrus grandis* against pathogenic bacteria. *International Journal of Enteric Pathogens*. 2016;4(4):3-37103.
27. Bhavsar SK, Joshi P, Shah MB, Santani DD. Investigation into Hepatoprotective Activity of *Citrus limon*. *Pharmaceutical Biology*. 2007;45(4):303-11.
28. Otang WM, Afolayan AJ. Antimicrobial and antioxidant efficacy of *Citrus limon* L. peel extracts used for skin diseases by Xhosa tribe of Amathole District, Eastern Cape, South Africa. *South African journal of botany*. 2016;102:46-9.
29. Oboh G, Bello FO, Ademosun AO, Akinyemi AJ, Adewuni TM. Antioxidant, hypolipidemic, and anti-angiotensin-1-converting enzyme properties of lemon (*Citrus limon*) and lime (*Citrus aurantifolia*) juices. *Comparative Clinical Pathology*. 2015;24(6):1395-406.
30. Banerjee S, Mullick HI, Banerjee J, Ghosh A. *Zingiber officinale*: 'a natural gold'. *Int J Pharmaceutical Bio-Sci*. 2011;2:283-94.
31. Kumar G, Karthik L, Rao KB. A review on pharmacological and phytochemical properties of *Zingiber officinale* Roscoe (*Zingiberaceae*). *Journal of Pharmacy Research*. 2011;4(9):2963-6.
32. Shah PP, Mello PMD. A review of medicinal uses and pharmacological effects of *Mentha piperita*. *Natural Product Radiance*. 2004;3(4):214-21.
33. Mahendran G, Rahman LU. Ethnomedicinal, phytochemical and pharmacological updates on Peppermint (*Mentha piperita* L.)—A review. *Phytotherapy Research*. 2020;34(9):2088-139.
34. Li H, Lu C, Liu Y, Wang Z, Zhang J. *Dietary Chinese Herbs*. Springer Verlayr Wien. 2015:488.
35. Pramila DM, Xavier R, Marimuthu K, Kathiresan S, Khoo M, Senthilkumar M, et al. Phytochemical analysis and antimicrobial potential of methanolic leaf extract of peppermint (*Mentha piperita*: Lamiaceae). *J Med Plants Res*. 2012;6(2):331-5.
36. Taher YA. Antinociceptive activity of *Mentha piperita* leaf aqueous extract in mice. *Libyan Journal of Medicine*. 2012;7(1):16205.
37. Singh R, Shushni MAM, Belkheir A. Antibacterial and antioxidant activities of *Mentha piperita* L. *Arabian Journal of Chemistry*. 2015;8(3):322-8.

38. Sharma A, Sharma MK, Kumar M. Protective effect of *Mentha piperita* against arsenic-induced toxicity in liver of Swiss Albino mice. *Basic & clinical pharmacology & toxicology*. 2007;100(4):249-57.
39. Jain D, Pathak N, Khan S, Raghuram GV, Bhargava A, Samarth R, et al. Evaluation of cytotoxicity and anticarcinogenic potential of *Mentha* leaf extracts. *International Journal of Toxicology*. 2011;30(2):225-36.
40. Odebiyi OO, Sofowora EA. Phytochemical screening of Nigerian medicinal plants II. *Journal of Natural Products (Lloydia)*. 1977;41(3):234-46.
41. Evans WC. *Trease and Evans' Pharmacognosy*. 16th ed. New York: Elsevier Health Sciences; 2009.
42. Karber G. Contribution To The Collective Treatment Of Pharmacological Series Experiments. *Archives of Experimental Pathology and Pharmacology*. 1931;162:480-3.
43. Chinko BC, Dapper DV, Adienbo OM. Evaluation of Antihyperlipidaemic Activities of Hydromethanolic Extracts of *Dioscorea bulbifera*. *Journal of Advances in Medical and Pharmaceutical Sciences*. 2020;22(1):16-25.
44. Malafaia AB, Nassif PAN, Ribas CAPM, Ariede BL, Sue KN, Cruz MA. Obesity induction with high fat sucrose in rats. ABCD. *Arquivos Brasileiros de Cirurgia Digestiva (São Paulo)*. 2013;26:17-21.
45. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clinical Chemistry*. 1972;18(6):499-502.
46. Crook M. *Clinical Chemistry & Metabolic Medicine*. London: Edward Arnold publishers Ltd.; 2006.
47. Zeka K, Ruparelia K, Arroo RR, Budriesi R, Micucci M. Flavonoids And Their Metabolites: Prevention In Cardiovascular Diseases And Diabetes. *Diseases*. 2017;5(3):1-18.
48. You C-L, Su P-Q, Zhou X-X. Study On Effect And Mechanism Of *Scutellaria Baicalensis* Stem-Leaf Total Flavonoid In Regulating Lipid Metabolism. *China Journal of Chinese Materia Medica*. 2008;33(9):1064-6.
49. Barbalho SM, Damasceno DC, Spada APM, Silva VSd, Martuchi KA, Oshiiwa M, et al. Metabolic profile of offspring from diabetic Wistar rats treated with *Mentha piperita* (peppermint). *Evidence-Based Complementary and Alternative Medicine*. 2011;2011.
50. Aremu MO, Ajine PL, Omosibi MO, Baba NM, Onwuka JC, Audu SS, et al. Lipid Profiles and Health Promoting Uses of Carrot (*Daucus carota* L.) and Cucumber (*Cucumis sativus* L.). *International Journal of Sciences*. 2021;10(7):22-9.
51. Mensink RP, Katan MB. Effect of dietary fatty acids on serum lipids and lipoproteins. A meta-analysis of 27 trials. *Arteriosclerosis and thrombosis: a journal of vascular biology*. 1992;12(8):911-9.
52. Fu J, Zhang XW, Liu K, Li QS, Zhang LR, Yang XH, et al. Hypolipidemic activity in Sprague–Dawley rats and constituents of a novel natural vegetable oil from *Cornus wilsoniana* fruits. *Journal of food science*. 2012;77(8):H160-H9.
53. Norouzi F, Doulah A, Rafieirad M. Effects of Four Week Consumption of Lemon (*Citrus limon* L.) Essential Oil with Swimming Training on Lipid Profile and Lipid Peroxidation in Adult Male Mice. *Iranian Journal of Nutrition Sciences & Food Technology*. 2020;14(4):1-8.

54. Olukanni OD, Akande OT, Alagbe YO, Adeyemi OS, Olukann AT, Daramola GG. Lemon juice elevated level of reduced glutathione and improved lipid profile in Wistar rats. *American-Eurasian J. Agric. & Environ. Sci.* 2013;13(9):1246-51.
55. Raed AH, Khattab AS, Foad EA, Alkhateeb HH. Effects of Citrus limon leaf extract on blood glucose and lipid profile in alloxan monohydrate-induced diabetic rats. *Gazzetta Medica Italiana-Archivio per le Scienze Mediche.* 2021;180(11):730-7.
56. Badal RM, Badal D, Badal P, Khare A, Shrivastava J, Kumar V. Pharmacological action of *Mentha piperita* on lipid profile in fructose-fed rats. *Iranian journal of pharmaceutical research: IJPR.* 2011;10(4):843.
57. Barbalho SM, Machado FMVF, Oshiiwa M, Abreu M, Guiger EL, Tomazela P, et al. Investigation of the effects of peppermint (*Mentha piperita*) on the biochemical and anthropometric profile of university students. *Food Science and Technology.* 2011;31(3):584-8.
58. Sharmin R, Khan MRI, Akhtar MA, Alim A, Islam MA, Anisuzzaman ASM, et al. Hypoglycemic and hypolipidemic effects of cucumber, white pumpkin and ridge gourd in alloxan induced diabetic rats. *Journal of Scientific Research.* 2013;5(1):161-70.
59. Insanu M, Rizaldy D, Silviani V, Fidrianny I. Chemical compounds and pharmacological activities of cucumis genus. *Biointerface Res Appl Chem.* 2022;12(1):1324-34.
60. Minaiyan M, Zolfaghari B, Kamal A. Effect of hydroalcoholic and buthanolic extract of *Cucumis sativus* seeds on blood glucose level of normal and streptozotocin-induced diabetic rats. *Iranian journal of basic medical sciences.* 2011;14(5):436.
61. Cicero AF, Sahebkar A, Fogacci F, Bove M, Giovannini M, Borghi C. Effects of phytosomal curcumin on anthropometric parameters, insulin resistance, cortisolemia and non-alcoholic fatty liver disease indices: a double-blind, placebo-controlled clinical trial. *European journal of nutrition.* 2020;59(2):477-83.
62. Lv J, Cao L, Li M, Zhang R, Bai F, Wei P. Effect of hydroalcohol extract of lemon (*Citrus limon*) peel on a rat model of type 2 diabetes. *Tropical Journal of Pharmaceutical Research.* 2018;17(7):1367-72.
63. Sorrenti V, Consoli V, Grosso S, Raffaele M, Amenta M, Ballistreri G, et al. Bioactive compounds from lemon (*Citrus limon*) extract overcome TNF- α -induced insulin resistance in cultured adipocytes. *Molecules.* 2021;26(15):4411.
64. Bayani M, Ahmadi-Hamedani M, Javan AJ. Study of hypoglycemic, hypocholesterolemic and antioxidant activities of Iranian *Mentha spicata* leaves aqueous extract in diabetic rats. *Iranian Journal of Pharmaceutical Research: IJPR.* 2017;16(Suppl):75.
65. Wani SA, Naik H, Wagay JA, Ganie NA, Mulla MZ, Dar B. Mentha: A review on its bioactive compounds and potential health benefits. *Quality Assurance and Safety of Crops & Foods.* 2022;14(4):154-68.
66. Barbalho SM, Damasceno DC, Spada APM, Silva VSd, Martuchi KA, Oshiiwa M, et al. Metabolic profile of offspring from diabetic Wistar rats treated with *Mentha piperita* (peppermint). *Evidence-Based Complementary and Alternative Medicine.* 2011;2011:1-6.
67. Randle PJ. Regulatory interactions between lipids and carbohydrates: the glucose fatty acid cycle after 35 years. *Diabetes/metabolism reviews.* 1998;14(4):263-83.

68. Novelli ELB, Diniz YS, Galhardi CM, Ebaid GMX, Rodrigues HG, Mani F, et al. Anthropometrical parameters and markers of obesity in rats. *Laboratory animals*. 2007;41(1):111-9.
69. Lin S, Thomas TC, Storlien LH, Huang XF. Development of high fat diet-induced obesity and leptin resistance in C57Bl/6J mice. *International journal of obesity*. 2000;24(5):639-46.

UNDER PEER REVIEW