

The Use of Dimethyl biguanide as a Potential Treatment for Alzheimer's Disease in Rats

The manuscript type: Original Research Article

ABSTRACT

Amis: Alzheimer's disease (AD) is one of the most rapidly growing diseases in recent times. Despite extensive research to find an appropriate medicine, there has been no effective drug until now.

Study Design: The present study was designated to investigate the therapeutic impact of antihyperglycemic **dimethyl biguanide** on Alzheimer's disease symptoms.

Methodology: Alzheimer's disease was induced in male rats by AlCl_3 and D-galactose at doses of 50 and 120 mg/kg daily for one month. Then, for the next four weeks, rats were given oral **dimethyl biguanide** (200 mg/kg daily).

Results: The obtained data indicated an increase in the arrival time of the AD rat group (G2) compared to the control group (G1). In addition, the AD rat group showed an elevation in glucose level, oxidative stress, liver, and kidney function. Importantly, **dimethyl biguanide** was able to **ameliorate** these unpleasant outcomes in G3. Interestingly, **dimethyl biguanide** was able to reduce GFAB immunoreactivity in the **dimethyl biguanide**-treated group (G3) compared to the AD group (G2).

Conclusion: In fact, dimethyl biguanide can delay the symptomatology of AD.

Keywords: **Dimethyl biguanide**; Alzheimer's disease; D-galactose; AlCl_3 .

1. INTRODUCTION

Alzheimer's disease (AD) is a neurological condition that worsens over time and is brought on by hereditary, epigenetic, and environmental factors [1, 2]. There are two different types of AD, including the more prevalent late-onset AD and an early-inception AD. **Severe memory loss and decreased cognitive function are hallmarks of AD [3]. Furthermore, while the increase of amyloid deposition and**

neurofibrillary plaques in the brain is considered the most acceptable hypothesis of AD formation, the actual mechanism for the emergence of AD symptoms remains relatively unknown [4]. There are currently no effective therapies for AD [5]. As a result, extensive research is required to study AD pathology and its relationship with other diseases in order to create a complete map of AD pathways that will aid in the development of appropriate medication.

Previous research has found a clear link between diabetes, particularly type 2, and Alzheimer's disease. Moreover, it was reported that insulin levels and the insulin pathway in the brain were impaired in AD patients [6, 7]. From this point forward, there is a growing trend surrounding the use of anti-diabetic drugs to treat Alzheimer's disease and mild neurodegenerative symptoms.

Dimethyl biguanide, a member of the biguanide family of antihyperglycemics, is currently the most used for treating type 2 diabetes mellitus (T2DM) [8]. Previous biomedical reports have even shown that dimethyl biguanide has several valuable impacts, including cardiovascular protection, anti-cancer activity, and anti-inflammatory properties [9]. Based on the anti-inflammatory effect of dimethyl biguanide, we and others suggested that it can protect against AD [10, 11]. The aim of this study was to investigate the impact of dimethyl biguanide against Alzheimer's disease.

2. MATERIAL AND METHODS

2.1. Chemicals

Dimethyl biguanide, D- galactose were gained from Sigma-Aldrich (St. Louis, MO, USA). $AlCl_3$ was obtained from Loba chemie, India. Calretinin antibody (No. IR627) and polyclonal GFAB (NO. Z0334) were provided by Agilent Dako, Denmark. All other chemicals obtained in highly pure grade.

2.2. Experimental section

2.2.1. Alzheimer's disease induction design

The current research was performed corresponding to the general guidelines of Faculty of Science, Tanta University Egypt (Approved ethical No. IACUC-SCI-TU-0259) for handling of laboratory animals. 200- 220 g on average, thirty male **Wistar rats** were bought from Faculty of Pharmacy, Cairo University, Egypt. Rats were maintained according to general care guidelines (free access to water and food *ad libitum*/ 12 h day cycle at 25 °C) [12]. Five groups were generated (n= 10/ each group). G1; is normal rat. G2; is AD rats that received 50 mg/Kg of AlCl₃ and 120 mg/ Kg of D- galactose daily for constitutive 4 weeks [13]. G3; Biguanide treated that firstly received AlCl₃ and D-galactose as the same as G4 for one month, then treated with 200 mg/Kg **dimethyl biguanide** for one month [14]. **Before the scarification, the memory impairment was checked by the classical labyrinth test [15].** Finally, the rats were slaughtered by decapitation and the skulls were opened with fine scissors and the brains were excised. Hippocampus were quickly removed and divided into two segments; one was fixed in 10% neutral buffer formalin, for histopathological examination and the remaining was washed and stored at -80 °C for preparation of tissue homogenates.

2.2.2. Biochemical assessment in serum

Under anesthesia, a blood sample was taken from the eye, and serum was obtained by centrifuging the clotted blood samples. Fasting blood glucose (FBG), **liver function [Alanine transaminase (ALT) and Aspartate transaminase (AST)]** and kidney function (urea and creatinine) were conducted according to the instruction of commercial **kits procured from (Bio diagnostic, Egypt).**

2.2.3. Evaluation oxidative and anti-oxidative parameter in hippocampus

Glutathione **(GSH)** and glutathione peroxidase **(GPx)** were assessed employing profitable kits from **(Bio diagnostic, Egypt)**. Further, **Malondialdehyde (MDA)**, lipid peroxidation indicator, was measured according to [16].

2.2.4. Histopathological investigation

Brains were excised and fixed in 10% formalin in phosphate buffered saline pH 7.4 for 24 h at 4°C. Fixed tissues were dehydrated through a graded series of ethanol and

embedded in paraffin according to standard procedures. Paraffin sections (5µm thick) were mounted on gelatin chromalum-coated glass slides and used for Haematoxylin and eosin stains as a routine method [17].

2.2.5. Glial fibrillary acidic protein (GFAP) immunohistochemistry

Expression of GFAB-ir (GFAB immunoreactivity) in brain sections (hippocampus) were detected using the avidin biotin peroxidase complex method. Briefly, sections were incubated with Polyclonal rabbit anti-GFAP immunoglobulin (Z0334, Dako) [18] at a dilution of 1:1000 for 16 h at room temperature.

2.3. Statistical analysis

GraphPad Prism v. 6 (GraphPad Software, San Diego, CA, USA) was used in the current study. Using one way ANOVA and multiple comparison the significances between groups were statistically presented. $P < 0.05$ was considered as significant.

3. RESULTS

3.1 Dimethyl biguanide ameliorate memory, glucose level, liver kidney function in AD induced rats

Table 1 indicated that there was a significant increase in the time taken during Labyrinth in the AD-induced rats (G2). This time was clearly reduced in biguanide-treated groups (G3). Furthermore, in G2, liver function, including ALT and AST, and kidney parameters, such as creatinine and urea, were elevated, but improved after Biguanide administration in G3.

Table 1: Impact of dimethyl biguanide on biochemical parameters in AD induced rats

parameters	Rat groups
------------	------------

	G1	G2	G3
Arrival time (sec)	43±4.51	150.4±6 [#]	92.3±3.2 ^{#,*}
FBG (mg/dl)	79.3±1.23	214.23±3.23 [#]	126.2±2.16 ^{#,*}
ALT (IU)	35.2±2.1	66.72±1.54 [#]	44.3±1.98 ^{#,*}
AST (IU)	44.3±1.2	98.2±2.3 [#]	65±1.78 ^{#,*}
Urea(mg/dl)	24.8±2.54	57.2±1.97 [#]	35.4±1.1 ^{#,*}
Creatinine (mg/dl)	0.8±0.02	2.7±0.1 [#]	1.6±0.13 ^{#,*}

Data was presented as mean± SD, P<0.05 was considered as significant. #,* are the significancy compared to control (G1) and AD group (G2) respectively.

3.2.Impact of dimethyl biguanide on the hippocampal oxidative stress

Administration of AlCl₃ and D-galactose induced oxidative stress in hippocampus with visible decline in antioxidant parameters in G2 compared to control (G1). Indeed, the treated group with dimethyl biguanide showed a magnificent drop in MDA and an elevation of GPx and GSH as antioxidant parameters as shown in Figure 1.

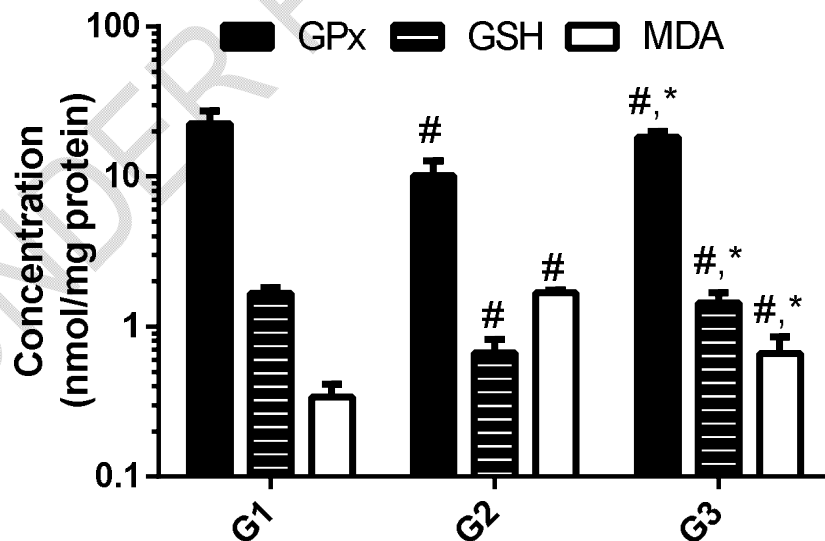


Figure 1. The antioxidant/ oxidative stress parameters in treated rats. Data was presented as mean± SD, P<0.05 was considered as significant. #,* are the

significance compared to control (G1) and AD group (G2) respectively. GPx: Glutathione peroxidase, GSH: Glutathione, MDA: Malondialdehyde

3.3. Histopathology assessment

Histopathology examination of hippocampus of treated rats demonstrated a typical structure with tightly packed layers of pyramidal cells in control groups (Figure 2A). Further, AD-induced rats (G2) revealed oedema, disseminate vacuolar deterioration, vacuolated neurocytes and degenerated, reduction and alteration pyramidal cells (Figure 2B). On the other hand, dimethyl biguanide treatment rats (G3) showed little tissue damage with a few neuronal injuries without extensive vacuolar atrophy (Figure 2C).

3.4. Immunohistochemistry of GFAB

GFAB immunoreactivity in AD rats (G2) showed moderately positive immunoreactivity compared to control (G1), which significantly dropped in the dimethyl biguanide post-treatment group (G3) as shown in Figure 3.

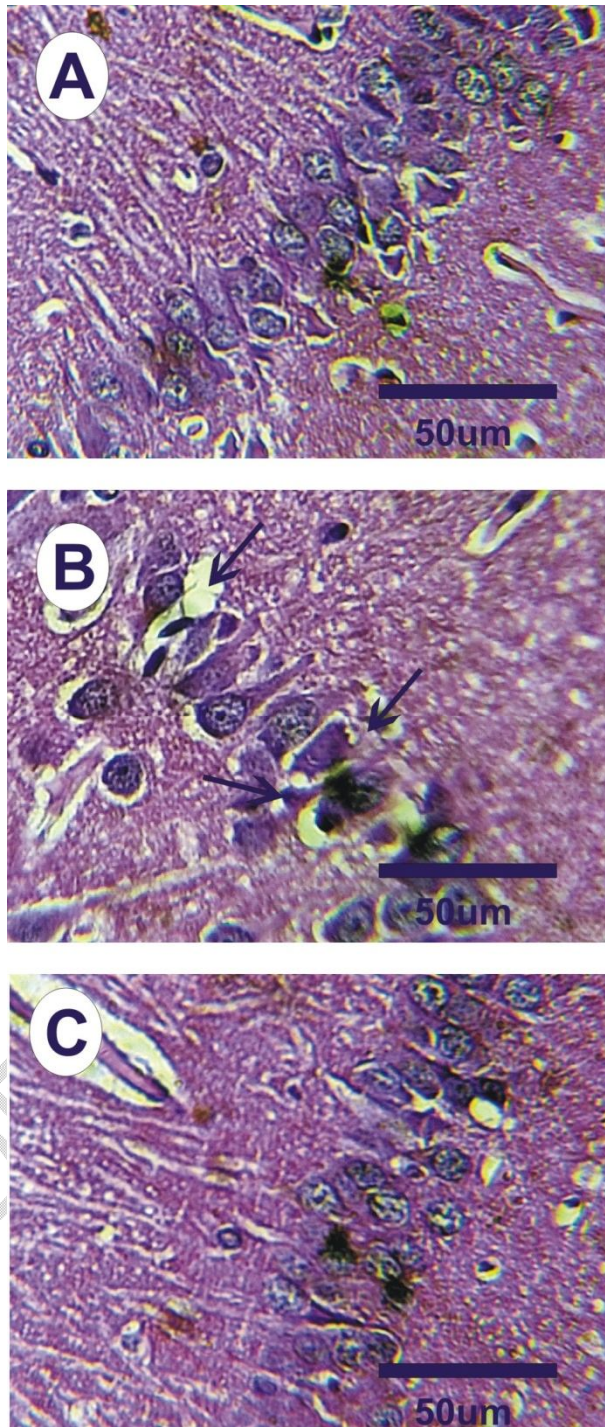


Figure 2: Histopathological Changes in hippocampus structure in different groups. A: Control, B: Alzheimer's disease (AD) revealed oedema, disseminate vacuolar deterioration, vacuolated neurocytes (arrows) and degenerated, reduction and alteration pyramidal cells, C: **Dimethyl biguanide** post-treatment group.

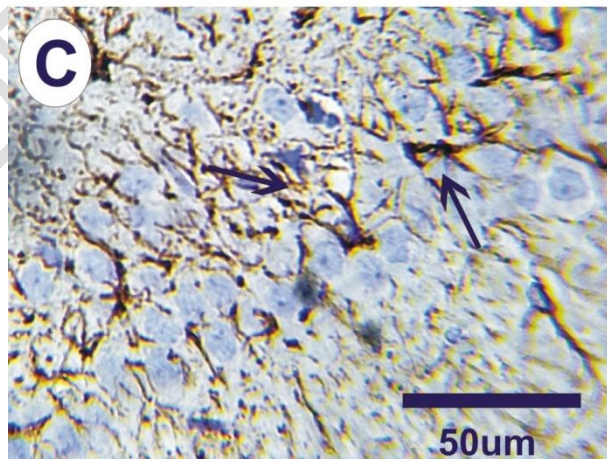
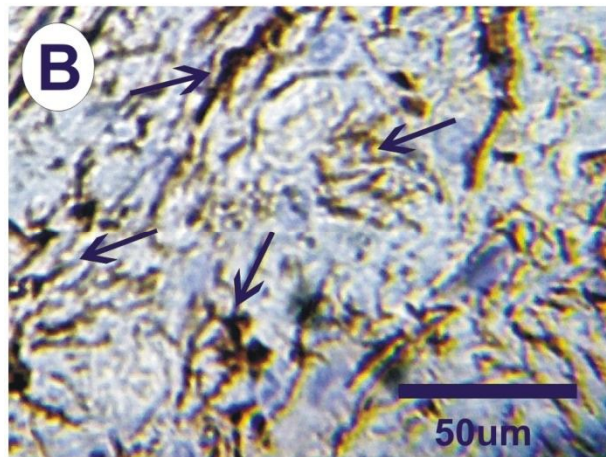
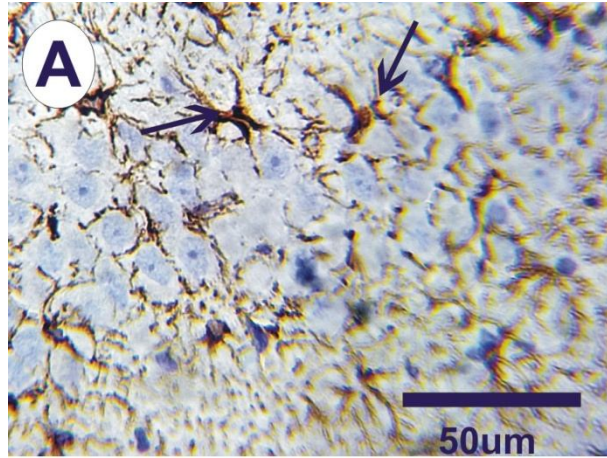


Figure 3: Immunohistochemical Changes in hippocampus structure stained with GFAB in different groups. A: Mild positive reactions (arrows) in Control, B:

Moderate positive reactions in Alzheimer's disease (AD) (arrows), C: Mild positive reactions (arrows) in **dimethyl biguanide** post-treatment group.

4. DISCUSSION

The current aim of this study is to investigate the curing role of the antidiabetic **dimethyl biguanide** against Alzheimer's disease. The data collected revealed that the arrival time of AD rats was 3.48 times longer than that of control rats. This high-fold change was dropped after dimethyl biguanide uptake. Moreover, AD rats showed higher glucose levels than the control rats, which were further ameliorated by dimethyl biguanide treatment. Our data agreed with previously published reports, which indicated an elevation in arrival time with hyperglycemia [19-21]. Furthermore, the alteration in liver and kidney functions in AD syndrome was well-known due to impairment in lipid metabolism with elevated inflammatory markers [22, 23]. Our findings were completely consistent with this phenomenon.

Later research found a buildup of free radicals to be involved in the etiology of AD or moderate cognitive impairment [24]. Furthermore, an elevation of the oxidative stress marker MDA, along with a considerably declining antioxidant system (GPx and GSH), were clearly presented in our data. Previously published reports revealed similar findings [25, 26].

On the level of histopathology changes, our results indicated a harmful alteration with distortion in the hippocampus region of AD rats that was greatly ameliorated with **dimethyl biguanide** medication. Previous reports suggested that the brain is shielded from harm by **dimethyl biguanide**. The findings of our investigation are consistent with these studies [27-30], and we found that the rats' hippocampus significantly reduced the amount of neuronal tissue damage.

In our work, AD model rats showed substantial hippocampus astrogliosis (heightened GFAP intensity with a reduction in calretinin). In these rats, a dose of 200 mg/kg dimethyl biguanide resulted in significant improvement. Our findings are completely in line with those of *Pilienko et al. (2020)* who indicated that **dimethyl biguanide** improved GFAB levels in AD rats [31].

Conclusion

Antihyperglycemic **dimethyl biguanide** was able to improve neural damage caused by Alzheimer's disease and other cognitive disorders. As a result, Alzheimer's disease pathogenesis is more complicated and related to other metabolic disorders, such as diabetes mellitus type 2. **More research is needed to determine whether they have the same origin or if they have different pathways.**

DISCLAIMER

The products used for this research are commonly and predominantly used products in our area of research and country. There is absolutely no conflict of interest

between the authors and producers of the products because we do not intend to use these products as an avenue for any litigation but for the advancement of knowledge. Also, the research was not funded by the producing company rather it was funded by personal efforts of the authors.

CONSENT

Not applicable.

ETHICAL APPROVAL

Animal Ethic committee approval has been collected and preserved by the author(s).

6. REFERENCE

1. Apter, J.T., K. Shastri&K. Pizano, *Update on disease-modifying/preventive therapies in Alzheimer's disease*. Current Geriatrics Reports. 2015. **4**(4): p. 312-317.
2. Uwishema, O., A. Mahmoud, J. Sun, I.F.S. Correia, N. Bejjani, M. Alwan, A. Nicholas, A. Oluyemisi&B. Dost, *Is Alzheimer's disease an infectious neurological disease? A review of the literature*. Brain and Behavior. 2022. **12**(8): p. e2728.
3. Lecanu, L.&V. Papadopoulos, *Modeling Alzheimer's disease with non-transgenic rat models*. Alzheimer's Research & Therapy. 2013. **5**(3): p. 17.
4. Ali, S.K.&R.H. Ali, *Effects of antidiabetic agents on Alzheimer's disease biomarkers in experimentally induced hyperglycemic rat model by streptozocin*. PLoS One. 2022. **17**(7): p. e0271138.
5. Rygiel, K., *Novel strategies for Alzheimer's disease treatment: An overview of anti-amyloid beta monoclonal antibodies*. Indian J Pharmacol. 2016. **48**(6): p. 629-636.
6. Gasparini, L., W.J. Netzer, P. Greengard&H. Xu, *Does insulin dysfunction play a role in Alzheimer's disease?* Trends in pharmacological sciences. 2002. **23**(6): p. 288-293.
7. Knezovic, A., S. Budisa, A. Babic Perhoc, J. Homolak&J. Osmanovic Barilar, *From Determining Brain Insulin Resistance in a Sporadic Alzheimer's Disease Model to Exploring the Region-Dependent Effect of Intranasal Insulin*. Molecular Neurobiology. 2023.
8. Ferrannini, E., *The Target of Metformin in Type 2 Diabetes*. New England Journal of Medicine. 2014. **371**(16): p. 1547-1548.
9. Malínská, H., O. Oliyarnyk, V. Škop, J. Šilhavý, V. Landa, V. Zídek, P. Mlejnek, M. Šimáková, H. Strnad, L. Kazdová, et al., *Effects of Metformin on Tissue Oxidative and Dicarbonyl Stress in Transgenic Spontaneously Hypertensive Rats Expressing Human C-Reactive Protein*. PLOS ONE. 2016. **11**(3): p. e0150924.
10. Sonnen, J.A., E.B. Larson, K. Brickell, P.K. Crane, R. Woltjer, T.J. Montine&S. Craft, *Different patterns of cerebral injury in dementia with or without diabetes*. Archives of neurology. 2009. **66**(3): p. 315-322.
11. Beerli, M., J. Schmeidler, J. Silverman, S. Gandy, M. Wysocki, C. Hannigan, D. Purohit, G. Lesser, H. Grossman&V. Haroutunian, *Insulin in combination with other diabetes medication is associated with less Alzheimer neuropathology*. Neurology. 2008. **71**(10): p. 750-757.

12. Kilkenny, C., W. Browne, I. Cuthill, M. Emerson & D. Altman, *Improving Bioscience Research Reporting: The ARRIVE Guidelines for Reporting Animal Research*. *PLoS Biol.* 8 (6): e1000412. 2010.
13. Mallikarjuna, N., K. Praveen & K. Yellamma, *Role of Lactobacillus plantarum MTCC1325 in membrane-bound transport ATPases system in Alzheimer's disease-induced rat brain*. *BioImpacts: BI.* 2016. 6(4): p. 203.
14. Zhang, S., H. Xu, X. Yu, Y. Wu & D. Sui, *Metformin ameliorates diabetic nephropathy in a rat model of low-dose streptozotocin-induced diabetes*. *Experimental and therapeutic medicine.* 2017. 14(1): p. 383-390.
15. Gasmi, S., *Classic Labyrinth Test for Neurobehavioral Evaluation in Wistar Rats*. *Bio Protoc.* 2018. 8(18): p. e3007.
16. Radwan, A.M., E.F. Aboelfetoh, T. Kimura, T.M. Mohamed & M.M. El-Keiy, *Fenugreek-mediated synthesis of zinc oxide nanoparticles and evaluation of its in vitro and in vivo antitumor potency*. *Biomedical Research and Therapy.* 2021. 8(8): p. 4483-4496.
17. Tousson, E., D.M. Beltagy, M.S.A. El-Gerbed, M.A. Gazia & M.A. Akela, *The ameliorating role of folic acid in rat hippocampus after propylthiouracil-induced hypothyroidism*. *Biomedicine & Aging Pathology.* 2012. 2(3): p. 104-110.
18. Beltagy, D.M., N.F. Nawar, T.M. Mohamed, E. Tousson & M.M. El-Keey, *Beneficial consequences of probiotic on mitochondrial hippocampus in Alzheimer's disease*. *J Complement Integr Med.* 2021. 18(4): p. 761-767.
19. Beltagy, D.M., N.F. Nawar, T.M. Mohamed, E. Tousson & M.M. El-Keey, *Beneficial consequences of probiotic on mitochondrial hippocampus in Alzheimer's disease*. *Journal of Complementary and Integrative Medicine.* 2021. 18(4): p. 761-767.
20. Burns, C.M., K. Chen, A.W. Kaszniak, W. Lee, G.E. Alexander, D. Bandy, A.S. Fleisher, R.J. Caselli & E.M. Reiman, *Higher serum glucose levels are associated with cerebral hypometabolism in Alzheimer regions*. *Neurology.* 2013. 80(17): p. 1557-1564.
21. Mohamed, T.M., M.A.M. Youssef, A.A. Bakry & M.M. El-Keiy, *Alzheimer's disease improved through the activity of mitochondrial chain complexes and their gene expression in rats by boswellic acid*. *Metabolic Brain Disease.* 2021. 36(2): p. 255-264.
22. Gillani, S.W., N. Ghayedi, P. Roosta, P. Seddigh & O. Nasiri, *Effect of Metformin on Lipid Profiles of Type 2 Diabetes Mellitus: A Meta-analysis of Randomized Controlled Trials*. *J Pharm Bioallied Sci.* 2021. 13(1): p. 76-82.
23. Ormazabal, V., S. Nair, O. Elfeky, C. Aguayo, C. Salomon & F.A. Zuñiga, *Association between insulin resistance and the development of cardiovascular disease*. *Cardiovasc Diabetol.* 2018. 17(1): p. 122.
24. Torres, L.L., N.B. Quaglio, G.T. de Souza, R.T. Garcia, L.M.M. Dati, W.L. Moreira, A.P. de Melo Loureiro, J.N. de Souza-Talarico, J. Smid & C.S. Porto, *Peripheral oxidative stress biomarkers in mild cognitive impairment and Alzheimer's disease*. *Journal of Alzheimer's Disease.* 2011. 26(1): p. 59-68.
25. Greilberger, J., C. Koidl, M. Greilberger, M. Lamprecht, K. Schroecksnadel, F. Leblhuber, D. Fuchs & K. Oettl, *Malondialdehyde, carbonyl proteins and albumin-disulphide as useful oxidative markers in mild cognitive impairment and Alzheimer's disease*. *Free radical research.* 2008. 42(7): p. 633-638.

26. Gustaw-Rothenberg, K., K. Kowalczyk & M. Stryjecka-Zimmer, *Lipids' peroxidation markers in Alzheimer's disease and vascular dementia*. *Geriatrics & gerontology international*. 2010. **10**(2): p. 161-166.
27. Patil, S.P., P. Jain, P. Ghumatkar, R. Tambe & S. Sathaye, *Neuroprotective effect of metformin in MPTP-induced Parkinson's disease in mice*. *Neuroscience*. 2014. **277**: p. 747-754.
28. Tang, G., H. Yang, J. Chen, M. Shi, L. Ge, X. Ge & G. Zhu, *Metformin ameliorates sepsis-induced brain injury by inhibiting apoptosis, oxidative stress and neuroinflammation via the PI3K/Akt signaling pathway*. *Oncotarget*. 2017. **8**(58): p. 97977.
29. Akinola, O., M. Gabriel, A.-A. Suleiman & F. Olorunsogbon, *Treatment of alloxan-induced diabetic rats with metformin or glitazones is associated with amelioration of hyperglycaemia and neuroprotection*. *The Open Diabetes Journal*. 2012. **5**(1): p. 8-12.
30. Sangi, S.M.A. & N.A. Al Jalaud, *Prevention and treatment of brain damage in streptozotocin induced diabetic rats with Metformin, Nigella sativa, Zingiber officinale, and Punica granatum*. *Biomedical Research and Therapy*. 2019. **6**(7): p. 3274-3285.
31. Pilipenko, V., K. Narbutė, J. Pupure, I.K. Langrate, R. Muceniece & V. Kluša, *Neuroprotective potential of antihyperglycemic drug metformin in streptozocin-induced rat model of sporadic Alzheimer's disease*. *European Journal of Pharmacology*. 2020. **881**: p. 173290.