

## Original Research Article

# IN VITRO ANTIFUNGAL ACTIVITY PROPERTY OF CASSIA FISTULA L. EXTRACTS AGAINST THE FUNGAL PATHOGEN MACROPHOMINAPHASEOLINA (TASSI.) GOID.

### ABSTRACT

**Aims:** To evaluate the ~~anti-fungal~~ ~~anti-fungal~~ ~~properties~~ ~~activity~~ of *Cassia fistula* ~~alcoholic~~ and aqueous extracts against *Macrophominaphaseolina*.

**Methods:** Ethanolic, methanolic, and aqueous extracts were prepared; ~~and~~ qualitative and quantitative ~~analysis estimation~~ of phytoconstituents was done. The antioxidant and antifungal activity of the extracts were determined.

**Results:** It was observed that the methanolic extract had the highest (15.65%) extraction yield, whereas ethanolic extract had the lowest (12.45%) ~~extraction yield~~. While screening for secondary metabolites, cardiac glycosides were found to be lacking in all three ~~solvent extract~~ ~~solvent~~ ~~extracts~~, while sterols and terpenoids ~~were~~ ~~are~~ absent in the aqueous extract. ~~The methanolic~~ ~~Methanolic~~ extract contained ~~the~~ highest amounts of phenolic compounds (13.38±0.060 mg GAE/g dw.), flavonoid (10.58±0.074 mg QUE/g dw.), tannin (11.43±0.052 mg TAE/g dw.), and alkaloid (16.18±0.062 mg AE/g dw.). ~~The highest~~ ~~Highest~~ antioxidant activity was observed in the methanolic extract at 300 µg/ml (% inhibition, 63.02%). Antifungal activity against *M. phaseolina* was also highest (68.07%) for the methanolic fraction.

**Conclusion:** *C. fistula* extracts may be further ~~be~~ ~~explored~~ ~~explored~~ to formulate antifungal agents.

**Keywords:** Antioxidant activity, antifungal activity, *Cassia fistula*, *Macrophominaphaseolina*

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## 1. INTRODUCTION

Chickpea (*Cicer arietinum* L.), commonly known as Bengal gram, is a significant annual plant of the pea family (Fabaceae) that is widely ~~produced known~~ for its nutritional seed value in many countries. Chickpeas are ~~a commercially significant widely produced plant~~ in India, Africa, Central, and South America [1]. It provides a substantial ~~amount number~~ of vital proteins and vitamins that are beneficial to human health [2]. Furthermore, they ~~are significant as were also used as source of raw materials material~~ for food processing and value-added industrial products [3].

There is a high demand ~~for~~ chickpea seeds for ~~their~~ nutritional value, mainly in the vegetarian diet in developing nations such as India, necessitating increased attention to its sustainable production [4].

About 172 pathogens are responsible for diseases in ~~chickpea chickpeas~~. The most destructive soil-borne disease is Dry Root Rot, which is ~~spread on infected~~ by *Macrophomina phaseolina* (Tassi) Goid (*Rhizoctonia bataticola* Taub.). The disease's severity varies according to the temperature and moisture content of the chickpea plant and causes significant losses ~~on the plant~~ from blooming to podding [5].

Although synthetic fungicides have long been used to control phytopathogenic fungi, their excessive use can be detrimental to human beings, ~~the the~~ environment, and non-target species, negatively impacting biodiversity. Phytochemicals generated from plants exhibit antifungal capabilities [6]. Today, the global endeavor in contemporary agriculture is to limit the use of toxic chemicals such as ~~fungicides fungicide, weedicides weedicide, pesticides pesticide~~, and so on, by adopting new biological and ecological ways. Utilizing the chemical interactions between plants is one of these strategies [7]. Allelopathy and competition are two examples of the impacts that individual plants in a shared ecosystem have on their nearby ~~neighbors neighbours~~. Competition entails the active absorption of limited resources by one organism, resulting in a decrease in supply and hence growth inhibition of other organisms; nevertheless, allelopathy occurs when one species stops developing due to chemicals emitted by another species [8,9]. However, while defining allelopathy, some researchers also ~~consider the stimulatory effects of growth take the stimulatory effects of growth into consideration~~ [10].

The Indian laburnum, sometimes known as the "golden shower tree," or *Cassia fistula* L. (Fabaceae), is a tree with therapeutic benefits [11]. Secondary metabolites including tannins, terpenoids, alkaloids, flavonoids, glycosides, and others are abundant in plant tissues, ~~and which~~ have been shown to exhibit antibacterial [12,13,14]; anti-inflammatory [15]; antioxidant [16]; wound healing [17]; antifungal [18] and anticancer activity [19], characteristics when tested in vitro. *C. fistula* includes important bio-natural components that are highly beneficial for providing crucial medicinal benefits.

In the present work, ~~the~~ phytochemical, and antioxidant activity of *Cassia fistula* L., were determined, and antifungal activity ~~was~~ checked against *Macrophomina phaseolina*.

## 2. Material and Methods

### 2.1 Sample collection

Fresh *Cassia fistula* leaves were randomly selected from the trees growing ~~in~~ campus of Banasthali Vidyapith in Tonk, Rajasthan, with ~~no obvious physical or microbiological problems~~. The leaves were cleaned with distilled water before being ~~dried~~ for two to three

**Comment [pp2]:** I would strongly suggest for you to site the methods, if you have obtain or referred to any source.

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**Comment [pp4]:** Air dried or any other methods?

days in the shade. The dried samples were then processed into a fine powder using a grinder and kept tight in a container until extraction was needed.

## 2.2 Extraction of Leaf Samples

20g of leaf sample was dissolved in 100 ml of solvent (ethanol, methanol, and water) for 24–42 hours at room temperature. Then, the extracts were filtered through sterile muslin cloth. The filtrates from each solvent were evaporated in a desiccator. The dried extracts were packaged in airtight containers, labeled, and kept in a refrigerator (2–4°C) for further use.

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Extraction yield (%) was calculated as follows:

Extraction yield (%) =  $W2/W1 * 100$ , where W2=weight of the extract after evaporating solvent, W1=initial dry weight of the sample

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## 2.3 Qualitative and Quantitative Phytochemical Analysis

Phytochemical evaluation for saponins, flavonoids, cardiac glycosides, terpenoids, steroids, tannins, phenol, anthraquinone, alkaloids, and tannins was done using standard methods [20,21]. The total phenolic [22,23], flavonoid [24,25], tannin [24,26], and alkaloid [20] content of various fractions of extracts (methanol, ethanol, and aqueous) were determined using standard procedures.

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## 2.4 Determination of Antimicrobial Activity

*Macrophomina phaseolina* (Tassi.) Goid, culture strains (NFCCI No. 4832) were procured from NFCCI (National Fungal Culture Collection of India), Agharkar Research Centre, Pune, India, and maintained at the Department of Bioscience and Biotechnology, Banasthali Vidyapeeth, Rajasthan. The antifungal assay was performed on a Potato dextrose broth medium [27]. The growth of fungal mycelia at 28–30°C was determined by observing the dry weight on the 7th day.

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## 2.5 Determination of Total Antioxidant Activity by DPPH assay:

To determine the total antioxidant activity, DPPH (1, 1-dihydroxy-2-picrylhydrazyl, Sigma Aldrich) free radical scavenging assay was conducted; 1.0 ml of DPPH solution (0.135 mM DPPH in methanol) was added to (100–300 µg/ml) plant extract [28]. The reaction mixture was kept in the dark for 30 minutes and its absorbance at 517 nm was measured with a spectrophotometer. Ascorbic acid was utilized as a reference.

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## 2.6 Statistical Analysis:

The experiments were repeated three times, and the findings were provided as Mean ± S.E. All tests were designed using a random block design and ANOVA at  $p < 0.05$ . Appropriate post-hoc tests were performed, including the "Duncan Multi-Distance Test" (DMRT) and the "Least Significant Difference" (LSD).

## 3. RESULTS:

### 3.1 Determination of Extraction Yield:

The % yield for the different solvent extracts of *C. fistula* leaf is shown in Table 1. When yields are compared, the Methanolic extract has the most significant.

**Table 1.** Percentage yield of various solvent extracts of leaf of *C. fistula*

Solvent	Yield percent (%)
Methanol	15.65 <sup>c</sup>
Ethanol	12.15 <sup>a</sup>
Aqueous	13.55 <sup>b</sup>

### 3.2 Qualitative Phytochemical Analysis

The phytochemical analysis of three solvent extracts of *C. fistula* is listed in Table 2. All three different solvent extracts gave a variety of compounds.

**Table 2.** Phytochemical screening of various solvent extracts of *C. fistula*

Phytochemical compound	Screening test	Solvents		
		Ethanol	Methanol	Aqueous
Alkaloids	Wagner's test	+	+	+
	Mayer's test	+	+	+
Cardiac glycosides	Keller kellani test	-	-	-
Flavonoids	Alkaline test	+	+	+
Phenolics	Ferric chloride test	+	+	+
Saponins	Foam test	+	+	+
Sterols	Liebermann-Burchard test	+	+	-
Tannins	Ferric chloride test	+	+	+
Terpenoids	Salkowaski's test	+	+	+

(+) indicates presence (-) indicates absence

### 3.3 Quantitative analysis:

The results for the quantitative estimation of leaf extract of *C. fistula* are tabulated in Table 3. A maximum amount of phenolic, flavonoid, tannin, and alkaloid content was observed in the methanolic extract while the least was found in the aqueous extract.

**Table 3.** Quantitative analysis of *C. fistula* leaf extracts

Solvents	Total Phenolic Content (mg GAE/g) DWdw.	Total Flavonoid Content (mg QUE/g) DWdw.	Total Tannin Content (mg TAE/g) DWdw.	Total Alkaloid Content (mg AE/g) DWdw.
Methanol	13.38±0.060 <sup>c</sup>	10.58±0.074 <sup>c</sup>	16.18±0.062 <sup>c</sup>	11.43±0.052 <sup>c</sup>

Ethanol	10.58±0.063 <sup>b</sup>	8.68±0.106 <sup>b</sup>	8.91±0.065 <sup>b</sup>	9.85±0.050 <sup>b</sup>
Aqueous	5.88±0.153 <sup>a</sup>	6.56±0.059 <sup>a</sup>	7.5±0.157 <sup>a</sup>	7.91±0.063 <sup>a</sup>

(Results are mean ± standard deviation ;*p*=0.05 following ANOVA and LSD)

### 3.4 Antifungal activity:

The percentage of mycelial growth inhibition of *M. phaseolin* was studied using plant extracts in various solvents. The antifungal effects of *C. fistula* plant extract were tested at five different concentrations (100, 150, 200, 250, and 300 µg/ml) and represented in Table 4. At 300 µg/ml, methanol extract effectively reduced the emergence of selected [pathogens/pathogen](#), i.e., 68.07% while aqueous extract appears less significant (60.71%). Results were compared with standard Bavistin are presented.

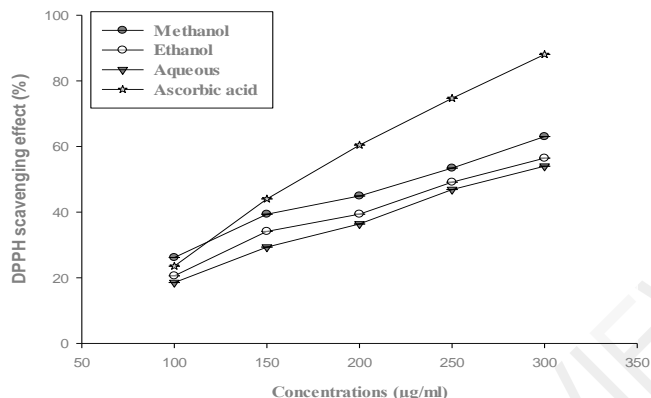
**Table 4.** Antifungal activity of *C. fistula* leaf extracts with different [solvents/solvent](#) at various concentrations.

Concentration (µg/ml)	Percent Inhibition(%)			
	Methanol	Ethanol	Aqueous	Bavistin
100	27.23±0.014 <sup>a</sup>	25.49±0.014 <sup>a</sup>	21.78±0.024 <sup>a</sup>	25.08±0.038 <sup>a</sup>
150	35.53±0.014 <sup>b</sup>	30.81±0.014 <sup>b</sup>	28.49±0.024 <sup>b</sup>	55.60±0.038 <sup>b</sup>
200	47.84±0.024 <sup>c</sup>	43.54±0.043 <sup>c</sup>	38.49±0.037 <sup>c</sup>	71.06±0.014 <sup>c</sup>
250	59.63±0.14 <sup>d</sup>	55.75±0.029 <sup>d</sup>	50.38±0.014 <sup>d</sup>	86.70±0.029 <sup>d</sup>
300	68.07±0.024 <sup>e</sup>	65.31±0.025 <sup>e</sup>	60.71±0.024 <sup>e</sup>	94.67±0.025 <sup>e</sup>

(Results are mean ± standard deviation ;*p*=0.05 following ANOVA and LSD)

### 3.5 Total antioxidant activity:

The efficacy of three distinct solvent extracts to scavenge DPPH free radicals was tested and compared to the standard, ascorbic acid. The methanolic extract of *C. fistula* was observed to be more active than the aqueous and ethanolic extracts (Figure 1). However, the extracts' DPPH radical scavenging capacities were lower than those of ascorbic acid (82.29%) at 300 µg/ml. This result clearly shows that the extracts have [proton-donating/proton donating](#) potential and might be used as free radical inhibitors or scavengers, possibly functioning as primary anti-oxidants.



**Figure 1.** Total antioxidant activity of *C. fistula* leaf extracts in different solvents and Ascorbic acid as standard.

#### 4. DISCUSSION:

In the present study, the presence of high TPC values in the methanolic extract of *C. fistula* were observed. Plant phenolic compounds are powerful antioxidants [29,30] as well as antimicrobials [31,32]. Various studies have demonstrated flavonoids in plant extracts to have antioxidant and antifungal effects [33,34]. The presence of tannins in all extracts may explain their strong bioactivities, as tannins are known to have significant antioxidant and antimicrobial activities [30]. Alkaloids also have antimicrobial and antioxidant properties [35].

These findings give scientific evidence to support traditional therapeutic usage and highlight the possibility of developing an antimicrobial and antioxidant agent from the *C. fistula* plant. Sharma [36] revealed that methanolic extracts of *C. fistula* flowers, leaves, stem bark, and pulp exhibit strong antioxidant activities. Rajput et al., [37] evaluated the antifungal effects of *C. fistula* leaf extracts produced in acetone, methanol, and diethyl ether against *C. albicans* and found substantial results. Variable activities of plant components such as flowers, leaves, pods, seeds, and stem bark have been recorded in various solvents [38,39,40]. Previously, methanol extracts of *C. fistula* showed strong antibacterial and antifungal activity on microorganisms [41]. According to in vitro data, this medicinal plant looks to be fascinating and promising as a potential source for emerging antimicrobial and antioxidant pharmaceuticals.

#### 5. CONCLUSION:

To design realistic management methods, it is necessary to understand the compatibility of bio-control agents with other components of the production system. According to current research, certain botanical plant extracts are a source of cost-effective and non-hazardous fungicides, towards as well as they have no human and environmental health hazard or implications thus, so *C. fistula* could be a good antifungal efficacy, which may be used for formulating new, safer, and eco-friendly fungicides.

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