

Review Article

Strategies for Controlling *Campylobacter* in Poultry Production: A Comprehensive Review of Challenges and Potential Solutions

ABSTRACT

Campylobacteriosis, a major foodborne illness caused by *Campylobacter*, poses significant challenges in poultry production. This review examines strategies to mitigate *Campylobacter* prevalence and foodborne outbreaks in poultry. Antibiotic therapy is limited due to *Campylobacter*'s antibiotic resistance. Natural alternatives, including bacteriocins, phages, probiotics, essential oils, and plant-derived compounds, show promise in combating *Campylobacter* and improving meat safety. Biosecurity measures and hygiene practices are crucial in preventing *Campylobacter* introduction and colonization. Strict protocols and cleanliness reduce contamination. Nutritional interventions and vaccination strategies enhance disease resistance and immune responses in poultry. Nanotechnology, particularly ZnO nanoparticles, exhibits antimicrobial efficacy against *Campylobacter* and other bacteria. Electrostatic interaction with cell walls and the production of reactive oxygen species disrupt bacterial membranes and intracellular components. A comprehensive approach integrating natural alternatives, biosecurity, nutrition, and nanotechnology is necessary for effective *Campylobacter* control. Continued research and adherence to hygiene practices can reduce *Campylobacter* contamination, improve food safety, and protect public health.

Keywords: Campylobacter, Transmission, Poultry, Contamination, Campylobacteriosis

1. INTRODUCTION

The available data indicates that in recent years, Campylobacteriosis incidence has increased in both developed and developing countries [1], with individual cases and outbreaks of the infection common globally. Campylobacteriosis is caused by *Campylobacter spp.*, which are the commonest zoonotic pathogens globally. Most *Campylobacter* infections are caused by *Campylobacter jejuni*, and to a lesser extent, *C. coli*.

The name "*Campylobacter*" [kam"pə-lo-bak'tər] is derived from the ancient Greek meaning "curved rod", where "kampylos" means "curved" and "baktron" means "rod". However, instead of having a curved rod form, *Campylobacter*'s distinctive shape actually resembles a spiral or helical one. To adapt to challenging circumstances, *Campylobacter* may transform its structure into a filamentous or coccoid one [2]. *Campylobacter* was initially identified as a "Vibrio-like bacteria" after being isolated from a sheep abortion case [3]; however, after clearly differing in taxonomic profile from the Vibrio species, it was renamed "*Campylobacter*". *Campylobacter* species are highly diverse at the subspecies and strain levels [4]. Diversity involves variances in genetic and phenotypic features, as well as development requirements, which may explain their appearance in different hosts or ecological niches, such as various poultry and wild birds. Some *Campylobacter* species have a single polar flagellum or bipolar flagella (e.g., *C. concisus*, *C. coli*, *C. jejuni*, and *C. showae*), whereas others (e.g. *C. ureolyticus* and *C. hominis*) do not [5]. Also, within a location, there might be significant regional variations in *Campylobacter* cases. This is caused by various factors, including underreported cases, limited sensitivity of detection methods, population size and composition, disparities in public health standards, intervention strategies, surveillance systems, food safety norms, and the prevalence of *Campylobacter* in localised natural reservoirs.

Although *C. jejuni* and *C. coli* are the most prevalent, emerging *Campylobacter* species been identified to cause infections, including *C. concisus*, *C. curvus*, *C. fetus*, *C. gracilis*, *C. mucosalis*, *C. pinnipediorum*, *C. rectus*, *C. showae*, *C. sputorum*, *C. lari*, *C. ureolyticus*, *C. upsaliensis*, and *C. volucris* [6]. The virulence of most emerging *Campylobacter* species are often less severe than those of *C. jejuni* and *C. coli*. Additionally, according to a recent study, *C. concisus* and *C. foetus* infections were more frequent in older people (68.4 years old on average) than in young adults (28.6 years old) [7].

Traveling and person-to-person transfer, close contact with animals, and ingestion of contaminated food or water are a few risk factors that might result in *Campylobacter* infections. According to meta-analysis data, the most important risk factors for *Campylobacter* infections include domestic and international travel, intake of raw chicken, environmental exposure, and close contact with farm animals. Most *Campylobacter* outbreaks are attributed to poultry and untreated water. Poultry, particularly broiler chickens, makes up the majority of the transmission route of this pathogen and presents the primary source of *Campylobacter* transmission in humans. The environment and horizontal transmission between flock mates are the main modes of *Campylobacter* transmission to poultry. If a bird is *Campylobacter* infection positive, all poultry on the same farm will be infected within a week [8]. Poultry and poultry products are an important transmission source because chickens' gastrointestinal tracts contain *Campylobacter*. As a result, the procedures for slaughtering broilers and processing chicken meat constitute a significant threat. They frequently lead to cross-infection of broiler carcasses due to contamination by intestinal content [9]. Inflammatory reactions, septicemia, GBS, and other associated symptoms are brought on by *Campylobacter* adhering to and penetrating the intestinal epithelial cell after consumption of the infected broiler meat [10].

The importance of poultry and poultry products in the transmission of *Campylobacter* cannot be overemphasized. Understanding the transmission dynamics of *Campylobacter* species in poultry is important for preventing the infection, and ultimately, conquering Campylobacteriosis. This review consequently covers the transmission of *Campylobacter* in poultry and poultry products.

2. CAMPYLOBACTER INFECTION: UNDERSTANDING THE ROLE OF RESERVOIRS AND SOURCES

Since the inception of poultry rearing, poultry has long been recognized as the primary source of *Campylobacter* species transmission to humans through food [11]. *Campylobacter* naturally colonize the cecum of birds and are transmitted among avian populations through the fecal-oral route [11,12]. Additionally, although present in smaller quantities, *Campylobacter* can also be found in the liver and small intestine of birds [11,12].

3. ASSESSING THE PREVALENCE AND RISK FACTORS OF CAMPYLOBACTER CONTAMINATION IN POULTRY

In poultry production systems, including those for broilers, layers, turkeys, and ducks, *Campylobacter* is quite common. *Campylobacter* is most typically absent in commercial broilers less than 2-3 weeks old, while experimental inoculation of newly born chicks with *Campylobacter* can effectively establish colonization. The causes of this lag period are unknown, although it might be due to a combination of factors such as maternal antibodies, antibiotic feed additives, intestinal development, and intestinal microbial ecology.

In the chicken microbiome, *C. jejuni* is one of the most predominant commensal bacteria. Other *Campylobacter* species like *C. concisus*, *C. lari*, and *C. upsaliensis* are also common [13]. These bacteria often spread horizontally to flocks from several environmental sources. *Campylobacter* species have been shown to be prevalent in the environment surrounding chicken farms, including soil, water sources, dust, surfaces, and air. *Campylobacter* can be transmitted from the environment to poultry farms via animal feed and/or drinking water. Farmers and farm visitors who carry *Campylobacter* can spread it to poultry farms. Several investigations identified *Campylobacter* from wild bird feces near poultry houses, indicating that wild birds helped spread these bacteria into chicken houses. Other species, including as flies, insects, amoebae, yeasts, and molds, have been discovered to be key drivers of *Campylobacter* horizontal transmission into poultry buildings [13]. The presence of amoebae, yeasts, and molds in *Campylobacter* cells allows them to live longer. In poultry facilities, a smaller mealworm beetle and its larvae (*Alphitobius diaperinus*) have been found as major transmitters of *C. jejuni*. They might spread *C. jejuni* not only within batches, but also throughout flocks in subsequent rearing cycles [14]. Furthermore, microbial eukaryotes may operate as a *Campylobacter* reservoir in the environment. Numerous *C. jejuni* strains, for example, may penetrate, multiply, and survive inside an amoeba host (*Acanthamoeba polyphaga*) [14,15]. Because eukaryotes are often found in both drinking water systems and microbial biofilms on farms [14-16], infected eukaryotes may contribute to *C. jejuni* transfer to poultry materials.

There has long been debate about whether *Campylobacter* may be transferred vertically from one generation of poultry to the next. A study of 60,000 progeny parent breeders found no indication of vertical transmission of *Campylobacter* to chickens [17]. However, research has shown that egg passage can result in the transmission of fecal bacteria like *Campylobacter*, which can then infect the shell, shell membrane, and albumen of newly laid and viable eggs [17,18]. This can result in *Campylobacter* ingestion when chicks hatch from their eggs, as well as *Campylobacter* colonization and dissemination in chicken houses. In contrast, *Salmonella* is well-known for vertical transmission because they contaminate the egg within the reproductive canal before the shell is produced or pierce the eggshell and enter the yolk of the post-lay egg. Furthermore, *Salmonella* is the most common cause of foodborne outbreaks related with chicken eggs, but *Campylobacter* egg-associated outbreaks are relatively rare [13]. *Campylobacter* was rarely recovered from the

internal egg contents, according to a systematic review of 4,316 references [19], which was further verified by multiple on-farm investigations.

4. CAMPYLOBACTER COLONIZATION IN CHICKENS

Since *Campylobacter* infection in chicken does not result in a significant inflammatory response or intestinal tissue damage, the slow and moderate *Campylobacter*-specific antibody response is not surprising. The interaction between *Campylobacter* and the chicken immune system that initiates the immune response is still mostly unclear. Developing immunization-based strategies to prevent *Campylobacter* infections in poultry would be substantially aided by understanding the intricate interplay between *Campylobacter* and the chicken immune systems. Young chickens' spleen, liver, and blood were also shown to have *Campylobacter* in certain investigations, which raises the possibility that the pathogen may infiltrate intestinal epithelial cells and spread throughout the body.

In chickens, *Campylobacter* colonization is mainly caused via horizontal transmission from the environment, such as drinking water or animal feed. After one week, *Campylobacter* multiplies and colonizes the intestinal tracts (crop, small intestine, and ceca) of the majority of chickens which contract them. *C. jejuni* levels inside these niches might be as high as 10^9 cells/gram of intestinal tract until slaughter, with no symptoms or evident adverse consequences. According to one study, *C. jejuni* is more than simply a commensal bacterium in broiler chickens; it can also induce chronic inflammation, gut tissue damage, and diarrhea. In contrast, independent of chicken growth rate or breed, four combined and eight individual chicken genotypes revealed no difference or negative effect on *C. jejuni* colonization and proliferation [13].

Several factors influence *Campylobacter* colonization in chickens. These include the chicken strain, the *Campylobacter* strain, the amount of viable *Campylobacter* cells, and the season [20]. Under in-vivo experimental conditions, the colonization potential of some *Campylobacter* strains could be increased by 1,000-folds or 10,000-folds, making it difficult to predict the ability of *Campylobacter* wild strains to colonize chicken flocks in real commercial farms [13]. Summer has a greater rate of colonization than any other season of the year. The amount of colonization and strain type are also seasonally dependent. Aside from high temperatures and humidity, summer requires greater ventilation in poultry buildings, exposing the birds to more *Campylobacter* from the outside environment than any other time of year [21].

Furthermore, geographical regions, flock size, and production system type (i.e., organic or conventional) can all impact *Campylobacter* colonization in chicken flocks [20]. One research found that up to 100% of organic and free-range flocks tested positive for *Campylobacter* [22]. This is most likely due to the birds' exposure to the outdoors environment and the longer time it takes them to reach slaughter size compared to indoor raised flocks. In situations where *Campylobacter* colonization was detected at the species level, *C. jejuni* was the dominant group, colonizing around 90% of *Campylobacter*-positive birds. *C. coli* and *C. lari* colonized the remaining ones roughly evenly. Several European investigations concluded that the indoor-grown flocks were largely colonized by one or two *C. jejuni* strains. Several *C. jejuni* strains colonized the indoor-grown flocks in other studies conducted in North America and Australia. This might be attributed to various degrees of biosecurity requirements in different nations, as *C. jejuni* colonization can be caused by exposure to several sources containing different strains or a single source containing many strains (e.g., feed or water).

The ability of *Campylobacter* strains to adapt to their environments and their responses thereto is another crucial element in chicken colonization. For instance, *C. jejuni* has a remarkable capacity to change quickly throughout storage, culture, and condition transfer, according to Gaynor and colleagues [23,24]. Before live chickens enter poultry processing plants, certain processes, such as feed withdrawal and transportation, have an impact on the prevalence of *Campylobacter* in the animals. Farmers frequently remove animal feeds from chicken houses 3 to 18 hours before slaughter, a procedure known as "feed withdrawal" [24]. In order to limit cross-contamination during the processing of chicken, this method aims to empty the gastrointestinal tract and lower the amount of fecal matter in the body. Byrd and colleagues demonstrated that the removal of feed might raise the frequency of *Campylobacter* in broiler chicken crops at the time of slaughter [24]. Whyte and colleagues revealed that poultry overcrowding and stress during transportation significantly increased *Campylobacter* shedding in broiler feces and contributed to cross-contamination of their carcasses during processing [24,25].

5. CONTROL OF CAMPYLOBACTERIOSIS IN POULTRY AND POULTRY PRODUCTS

Most poultry with *Campylobacteriosis* recover without treatment. Although antibiotics are commonly used to kill the bacteria, they cannot completely prevent *Campylobacteriosis*, and the growing antibiotic resistance of *Campylobacter* poses a global public health concern for both humans and animals. Moreover, antibiotic use in chicken farms can disrupt

the gut flora. Therefore, it is crucial to quickly identify natural alternatives that can limit or prevent *Campylobacter* colonization in poultry farms, thus helping to contain outbreaks of foodborne diseases [26].

Recent efforts to manage *Campylobacter* have focused on preventing and reducing its colonization in broiler chicks, as reducing the numbers of *Campylobacter* in broiler carcasses would lead to decreased meat contamination and, consequently, a decline in human Campylobacteriosis. Three main strategies can be identified as fundamental approaches to prevent and control *Campylobacter* prevalence in poultry flocks: (1) implementing biosecurity and hygiene measures to prevent the introduction of *Campylobacter* to poultry farms; (2) employing nutritional interventions to improve disease resistance and immune responses; and (3) utilizing vaccination interventions to enhance antibody responses and vaccination rates.

Implementing a well-designed biosecurity program at the farm level is a key strategy for combating pathogens like *Campylobacter* in poultry farms [27]. Over the past two decades, there has been an increasing focus on enhancing and advancing biosecurity standards. Previous research has indicated that chicken farms with excellent biosecurity and cleanliness standards tend to have lower levels of *Campylobacter* colonization. Research conducted over the past 20 years has also explored the use of natural alternatives to prevent and manage *Campylobacter* colonization in poultry farms. These studies have shown that pre-harvest nutritional supplementation with plant-derived compounds, prebiotics, probiotics, bacteriocins, and bacteriophages has significant antibacterial activity against *Campylobacter* and can improve meat safety following processing in slaughterhouses. Similarly, the development of poultry vaccines against *Campylobacter* can help reduce human Campylobacteriosis cases and the prevalence of the bacteria in poultry flocks.

Various control measures have been implemented at different stages of chicken production, including primary production, slaughterhouses, and food processing, with varying degrees of effectiveness.

6. ON-FARM INTERVENTIONS TO CONTROL CAMPYLOBACTER COLONIZATION IN POULTRY

In recent years, early-stage on-farm management of *Campylobacter* in broiler chickens has received increased attention due to the successful colonization of chickens by *Campylobacter* from the earliest stages of their lives, as well as its persistence throughout the poultry processing line. Various control options, such as phage treatment, bacteriocins, probiotics, fatty acids, and essential oils, have been studied for their potential in managing *Campylobacter* (Table 1). Bacteriocins, which are antimicrobial peptides produced by commensal bacteria in the chicken gut microbiota, have shown promise in inactivating *Campylobacter* both in vitro and in vivo. For instance, the bacteriocin L-1077 was found to significantly reduce *C. jejuni* numbers (>4 log CFU/g) in cecal content [28]. Another study investigated the development of *C. jejuni* resistance through oral administration of three different bacteriocins from *Lactobacillus salivarius* (OR-7) and *Enterococcus faecium* (E-760 and E50-52) in broiler chickens [29]. The CmeABC multidrug efflux pump in *C. jejuni* was found to play a critical role in both innate and acquired resistance to bacteriocins. To effectively inactivate *C. jejuni* and prevent the emergence of antibiotic resistance, the use of bacteriocins and efflux pump inhibitors may be considered. The effectiveness of *Campylobacter* phages in reducing the number of *Campylobacter* or preventing their colonization in chicken broilers has also been investigated in several studies. In one study, using an artificial infection model and a highly colonizing strain of *C. jejuni*, numerous phages isolated from the same environment as the bacterial host showed significant reductions [36]. Some phages, when applied at a high Multiplicity of Infection, led to up to 3 log reductions within the first 24 hours, while others resulted in approximately a 1 log decrease for up to 30 days (MOI).

Table 1. The advantages and disadvantages of various *Campylobacter* preventive and control measures in chicken production (Adapted from [13])

Strategies	Advantage	Disadvantage
Bacteriocins	Reduced <i>C. jejuni</i> by more than >4 log CFU/g in in-vitro conditions. [28]	Antimicrobial resistance is being developed via the CmeABC multidrug efflux pump. [29]
Bacteriophages	<i>C. jejuni</i> levels in commercial broiler flocks were reduced by up to 5 log CFU/g. [30]	Over time, gastrointestinal dilution and resistance development [31]
Probiotics	A portion of the microbiota of chicken gut [1]	<i>C. jejuni</i> decrease was limited after 15 days of oral administration [32]
Short chain fatty acids	Capability to penetrate gut epithelial cells [33]	The reduction of <i>C. jejuni</i> under in-vitro settings is limited [33]
Vaccination	Prophylactic and promising [34]	Expensive, demanding, and very specific [1]

7. CAMPYLOBACTER CONTAMINATION IN POULTRY PROCESSING FACILITIES

Chicken meat is considered a sustainable primary source of dietary protein due to its significantly lower feed conversion ratio (FCR) compared to beef. With an FCR of around 40%, chicken is highly efficient in converting feed into edible weight. However, poultry production, which involves animal farming and food processing, is also a common source of foodborne outbreaks, with live chickens and poultry meat serving as important reservoirs for *Campylobacter* and other foodborne pathogens [1]. Implementing effective on-farm and in vivo *Campylobacter* control measures is challenging due to the complexity and diversity of the industry (Table 2).

The processing of poultry is a labor-intensive operation that requires well-trained employees. Any lapse in sanitation or hygiene standards can result in multiple cases of foodborne diseases. *Campylobacter*, present in the cecal content at levels up to 10⁹ cells/g, can enter a processing facility through potentially infected birds [13]. Birds from different farms, with variations in age, size, geographical origin, production methods, and biosecurity practices, are typically processed together, increasing the risk of *Campylobacter* contamination. The processing procedures, including receiving, hanging, and packing, involve various stages where *Campylobacter* can start to proliferate or contaminate chicken carcasses (Table 2). Processes such as scorching, defeathering, evisceration, nick removal, and inside-out washing can all contribute to the cross-contamination of *Campylobacter* spp.

Evisceration, in particular, is a critical step for cross-contamination. The densely populated *Campylobacter* present in the gastrointestinal tract of poultry birds can spread extensively, especially in cases of gut leakage. Numerous in-plant studies have shown a significant increase in the prevalence of *Campylobacter*-positive carcasses after the evisceration process [36]. During evisceration, when birds are typically hung upside-down by their feet, *Campylobacter* leakage from the stomach can contaminate the lower half of the carcasses (breast and neck) more than the upper half (thighs and drumstick). *Campylobacter* is frequently detected in the hanging necks of the carcasses [13].

To prevent bacterial growth after evisceration, rapid cooling of poultry carcasses is crucial. Many processing facilities use a combination of chilling and antimicrobial treatment by washing the carcasses with cold chlorinated water to conserve energy and inhibit bacterial development [36]. Additionally, after evisceration, poultry carcasses are often cleaned by dipping or spraying them in chlorinated water to remove contaminants, blood, tissues, and fragments. However, under

commercial processing conditions, dipping can lead to cross-contamination of carcasses, especially when a large number of birds are processed simultaneously [37].

Table 2: Examples of *Campylobacter* prevalence and load across the chicken processing chain. (Adapted by [13])

Stage	Source	<i>Campylobacter</i> prevalence (%) and/or average load	Reference
Plant	Pre-scald	77%, > 6 log CFU/g of feather or skin ($n = 40$)	[38]
	Defeathering	3.9 log CFU/ml of carcasses rinse ($n = 24$)	[39]
	Evisceration	96–100%, 2.7 log CFU/carcass ($n = 48$)	[40]
	Pre-chill	98%, 4.75 log CFU/ml of carcasses rinse ($n = 450$)	[41]
	Post-chill	84.7%, 3.03 log CFU/ml of carcasses rinse ($n = 450$)	[41]
	Pre-wash	87%, 4.78 log CFU/ml of carcasses rinse ($n = 30$ [4 processing plants])	[42]
	Post-wash	80%, 4.30 log CFU/ml of carcasses rinse ($n = 30$ [4 processing plants])	[42]
Retail	90%, > 4 log CFU/carcass ($n = 552$)	[43]	

8. TREATMENT OF POULTRY PLANTS USING ANTIMICROBIALS

Numerous studies have investigated the efficacy of various permitted antimicrobials, including chlorine, chlorite, acidified sodium, cetylpyridinium, trisodium phosphate, chlorine dioxide, and peroxyacetic acid, in reducing the levels of *Campylobacter* in chicken meat by up to 5 logs. However, despite the effectiveness of these antimicrobials, the temperature of the water, or the washing mechanism employed, in-plant poultry washers have demonstrated limited capability in inactivating *Campylobacter* in chicken meat.

This limited effectiveness can be attributed to several factors. Firstly, chicken meat contains large molecules such as proteins and lipids, which may hinder the action of antimicrobials and prevent their full penetration into the meat. Additionally, changes induced by processing, the sensitivity of chicken skin to heat, oxidation, and discoloration, as well as the initial microbial load of carcasses, can contribute to the survival of *Campylobacter* despite washing treatments. The number of processed carcasses per minute, as well as the interaction or masking of antimicrobials, such as chlorine, by organic materials present in the processing water, can further impact their efficacy. Moreover, the quality of the water used during processing and the ability of *Campylobacter* to survive under certain conditions are also influential factors.

It is important to highlight that unlike the pasteurization stage in the processing of milk, there is no single effective critical control point for killing *Campylobacter* during the processing of raw chicken [2]. This emphasizes the need for comprehensive approaches and multiple interventions throughout the poultry processing chain to address *Campylobacter* contamination effectively.

9.0 ALTERNATIVE CONTROL STRATEGIES FOR CAMPYLOBACTER CONTAMINATION IN AGRI-FOODS

The use of herbs and essential oils in medicine initially stemmed from their beneficial properties such as antibacterial, anti-inflammatory, and antioxidant effects. Over time, these compounds found their way into the agricultural and food industry during the nineteenth century, primarily for their aromatic qualities and flavor profiles. These antimicrobials are classified as secondary metabolites and serve as essential components in plant defense systems, aiding in the protection against microbial diseases [38].

In addition to their antimicrobial properties, essential oils can also act as growth promoters in agricultural animals, similar to antibiotics [39]. A histology study demonstrated that feeding various plant extracts to chicken broilers resulted in a thickened mucus layer in their glandular stomach and jejunum [40]. This modification in the gastrointestinal tract was associated with a significant alteration in the gut microbiome, which has the potential to promote the growth and development of birds.

Several plants have been scientifically proven to possess anti-*Campylobacter* properties. For instance, cinnamon, which encompasses approximately 250 different species within the *Cinnamomum* genus, exhibits antimicrobial activity against *Campylobacter*. Another plant with anti-*Campylobacter* potential is turmeric (*Curcuma longa*), which contains curcumin as its major active compound. These examples highlight the diverse range of plants that hold promise in combating *Campylobacter* contamination.

10. NANOPARTICLES OF METAL OXIDE

In recent times, there has been a significant interest in exploring new applications of nanotechnology and nanomaterials. One area of focus is the utilization of metal oxide nanoparticles (such as Al_2O_3 , TiO_2 , and ZnO NPs) to deactivate various pathogens responsible for foodborne illnesses in different agri-food systems [41]. Studies have demonstrated that ZnO nanoparticles exhibit greater effectiveness against both Gram-negative and Gram-positive bacteria compared to other metal oxides like CuO and Fe_2O_3 [42]. Moreover, ZnO NPs have shown superior anti-*C. jejuni* properties compared to other Gram-negative bacteria like *E. coli* O157:H7 and *S. enterica* [43].

The direct electrostatic interaction between positively charged ZnO nanoparticles and the negatively charged bacterial cell wall leads to the destabilization and disruption of the bacterial outer cell membrane. Furthermore, ZnO's semiconductive nature allows the generation of reactive oxygen species, which can attach to and attack various cytoplasmic and extracellular targets [44]. These mechanisms contribute to the antimicrobial action of ZnO nanoparticles against *Campylobacter* and other pathogens.

8. CONCLUSION

Addressing the challenges of *Campylobacter* in poultry production and the associated foodborne outbreaks requires a comprehensive and multi-faceted approach. Strategies such as natural alternatives, biosecurity measures, nutritional interventions, and vaccinations have been investigated to minimize *Campylobacter* contamination and ensure food safety. Bacteriocins, phages, and antimicrobials have shown promise in reducing *Campylobacter* levels in chicken meat, although their effectiveness can be influenced by complex on-farm and in vivo controls, processing-related cross-contamination, and the presence of organic materials.

Emerging technologies, particularly nanotechnology, offer potential solutions. Metal oxide nanoparticles, especially ZnO nanoparticles, exhibit antimicrobial properties against *Campylobacter* and other bacteria. Through electrostatic interaction with bacterial cell walls and the generation of reactive oxygen species, ZnO nanoparticles disrupt bacterial membranes and attack intracellular targets. Further exploration and development of nanomaterial-based approaches hold promise for *Campylobacter* control in the future.

To achieve safer poultry production and reduce the risk of *Campylobacteriosis* outbreaks, a continuous focus on research, technological advancements, and strict adherence to hygiene and biosecurity practices is crucial. Implementing preventive measures throughout the entire production process, from farm to fork, is essential. This includes optimizing the application of antimicrobial agents, improving processing techniques, and ensuring proper handling and sanitation practices.

By integrating these measures, we can effectively combat *Campylobacter* and minimize the occurrence of foodborne illnesses. This comprehensive approach safeguards both human and animal health and contributes to the sustainability and safety of the poultry industry. Continued efforts in research and the adoption of innovative strategies will pave the way for a future where *Campylobacter*-related risks are mitigated, ensuring the well-being of consumers and the industry as a whole.

REFERENCES

1. Kaakoush, N. O., Castaño-Rodríguez, N., Mitchell, H. M., & Man, S. M. (2015). Global Epidemiology of *Campylobacter* Infection. *Clinical Microbiology Reviews*, 28(3), 687–720. <https://doi.org/10.1128/CMR.00006-15>
2. Tresse, O., Alvarez-Ordóñez, A., & Connerton, I. F. (2017). Editorial: About the Foodborne Pathogen *Campylobacter*. *Frontiers in Microbiology*, 8, 1908. <https://doi.org/10.3389/fmicb.2017.01908>
3. McFadyean, S.J., Stockman, S.S., (1913). Report of the Departmental Committee Appointed by the Board of Agriculture and Fisheries to Inquire into Epizootic Abortion. Appendix to Part III, Abortion in Sheep. HM Stationery Office.
4. Vidal, A. B., Colles, F. M., Rodgers, J. D., McCarthy, N. D., Davies, R. H., Maiden, M. C. J., & Clifton-Hadley, F. A. (2016). Genetic Diversity of *Campylobacter jejuni* and *Campylobacter coli* Isolates from Conventional Broiler Flocks and the Impacts of Sampling Strategy and Laboratory Method. *Applied and Environmental Microbiology*, 82(8), 2347–2355. <https://doi.org/10.1128/AEM.03693-15>
5. Man, S. M. (2011). The clinical importance of emerging *Campylobacter* species. *Nature Reviews Gastroenterology & Hepatology*, 8(12), 669–685. <https://doi.org/10.1038/nrgastro.2011.191>
6. Costa, D., & Iraola, G. (2019). Pathogenomics of Emerging *Campylobacter* Species. *Clinical Microbiology Reviews*, 32(4), e00072-18. <https://doi.org/10.1128/CMR.00072-18>
7. Bessède, E., Lehours, P., Labadi, L., Bakiri, S., & Mégraud, F. (2014). Comparison of Characteristics of Patients Infected by *Campylobacter jejuni*, *Campylobacter coli*, and *Campylobacter fetus*. *Journal of Clinical Microbiology*, 52(1), 328–330. <https://doi.org/10.1128/JCM.03029-13>

8. Gahamanyi, N., Mboera, L. E. G., Matee, M. I., Mutangana, D., & Komba, E. V. G. (2020). Prevalence, Risk Factors, and Antimicrobial Resistance Profiles of Thermophilic *Campylobacter* Species in Humans and Animals in Sub-Saharan Africa: A Systematic Review. *International Journal of Microbiology*, 2020, 1–12. <https://doi.org/10.1155/2020/2092478>.
9. Abd El-Hack, M. E., El-Saadony, M. T., Salem, H. M., El-Tahan, A. M., Soliman, M. M., Youssef, G. B. A., Taha, A. E., Soliman, S. M., Ahmed, A. E., El-kott, A. F., Al Syaad, K. M., & Swelum, A. A. (2022). Alternatives to antibiotics for organic poultry production: Types, modes of action and impacts on bird's health and production. *Poultry Science*, 101(4), 101696. <https://doi.org/10.1016/j.psj.2022.101696>.
10. Johnson, T. J., Shank, J. M., & Johnson, J. G. (2017). Current and Potential Treatments for Reducing *Campylobacter* Colonization in Animal Hosts and Disease in Humans. *Frontiers in Microbiology*, 8. <https://doi.org/10.3389/fmicb.2017.00487>
11. Skirrow, M. B. (1977). *Campylobacter* enteritis: A “new” disease. *BMJ*, 2(6078), 9–11. <https://doi.org/10.1136/bmj.2.6078.9>
12. Chaloner, G., Wigley, P., Humphrey, S., Kemmett, K., Lacharme-Lora, L., Humphrey, T., & Williams, N. (2014). Dynamics of Dual Infection with *Campylobacter jejuni* Strains in Chickens Reveals Distinct Strain-to-Strain Variation in Infection Ecology. *Applied and Environmental Microbiology*, 80(20), 6366–6372. <https://doi.org/10.1128/AEM.01901-14>
13. Hakeem, M. J., & Lu, X. (2021). Survival and Control of *Campylobacter* in Poultry Production Environment. *Frontiers in Cellular and Infection Microbiology*, 10, 615049. <https://doi.org/10.3389/fcimb.2020.615049>
14. Hazeleger, W. C., Bolder, N. M., Beumer, R. R., & Jacobs-Reitsma, W. F. (2008). Darkling Beetles (*Alphitobius diaperinus*) and Their Larvae as Potential Vectors for the Transfer of *Campylobacter jejuni* and *Salmonella enterica* Serovar Paratyphi B Variant Java between Successive Broiler Flocks. *Applied and Environmental Microbiology*, 74(22), 6887–6891. <https://doi.org/10.1128/AEM.00451-08>
15. Axelsson-Olsson, D., Waldenström, J., Broman, T., Olsen, B., & Holmberg, M. (2005). Protozoan *Acanthamoeba polyphaga* as a Potential Reservoir for *Campylobacter jejuni*. *Applied and Environmental Microbiology*, 71(2), 987–992. <https://doi.org/10.1128/AEM.71.2.987-992.2005>
16. Snelling, W. J., McKenna, J. P., Hack, C. J., Moore, J. E., & Dooley, J. S. G. (2006). An examination of the diversity of a novel *Campylobacter* reservoir. *Archives of Microbiology*, 186(1), 31–40. <https://doi.org/10.1007/s00203-006-0119-3>
17. Callicott, K. A., Friðriksdóttir, V., Reiersen, J., Lowman, R., Bisailon, J.-R., Gunnarsson, E., Berndtson, E., Hiett, K. L., Needleman, D. S., & Stern, N. J. (2006). Lack of Evidence for Vertical Transmission of *Campylobacter* spp. In Chickens. *Applied and Environmental Microbiology*, 72(9), 5794–5798. <https://doi.org/10.1128/AEM.02991-05>
18. Cox, N. A., Richardson, L. J., Maurer, J. J., Berrang, M. E., Fedorka-Cray, P. J., Buhr, R. J., Byrd, J. A., Lee, M. D., Hofacre, C. L., O’Kane, P. M., Lammerding, A. M., Clark, A. G., Thayer, S. G., & Doyle, M. P. (2012). Evidence for Horizontal and Vertical Transmission in *Campylobacter* Passage from Hen to Her Progeny. *Journal of Food Protection*, 75(10), 1896–1902. <https://doi.org/10.4315/0362-028.JFP-11-322>
19. Newell, D. G., Elvers, K. T., Dopfer, D., Hansson, I., Jones, P., James, S., Gittins, J., Stern, N. J., Davies, R., Connerton, I., Pearson, D., Salvat, G., & Allen, V. M. (2011). Biosecurity-Based Interventions and Strategies To Reduce *Campylobacter* spp. On Poultry Farms. *Applied and Environmental Microbiology*, 77(24), 8605–8614. <https://doi.org/10.1128/AEM.01090-10>
20. Newell, D. G., & Fearnley, C. (2003). Sources of *Campylobacter* colonization in broiler chickens. *Applied and environmental microbiology*, 69(8), 4343–4351. <https://doi.org/10.1128/AEM.69.8.4343-4351.2003>
21. Hudson, J. A., Nicol, C., Wright, J., Whyte, R., & Hasell, S. K. (1999). Seasonal variation of *Campylobacter* types from human cases, veterinary cases, raw chicken, milk and water. *Journal of applied microbiology*, 87(1), 115–124. <https://doi.org/10.1046/j.1365-2672.1999.00806.x>
22. Heuer, O. E., Pedersen, K., Andersen, J. S., & Madsen, M. (2001). Prevalence and antimicrobial susceptibility of thermophilic *Campylobacter* in organic and conventional broiler flocks. *Letters in applied microbiology*, 33(4), 269–274. <https://doi.org/10.1046/j.1472-765x.2001.00994.x>
23. Gaynor, E. C., Wells, D. H., MacKichan, J. K., & Falkow, S. (2005). The *Campylobacter jejuni* stringent response controls specific stress survival and virulence-associated phenotypes. *Molecular microbiology*, 56(1), 8–27. <https://doi.org/10.1111/j.1365-2958.2005.04525.x>
24. Byrd, J. A., Corrier, D. E., Hume, M. E., Bailey, R. H., Stanker, L. H., & Hargis, B. M. (1998). Incidence of *Campylobacter* in crops of preharvest market-age broiler chickens. *Poultry Science*, 77(9), 1303–1305. <https://doi.org/10.1093/ps/77.9.1303>
25. Whyte, P., Collins, J. D., McGill, K., Monahan, C., & O’Mahony, H. (2001). The Effect of Transportation Stress on Excretion Rates of *Campylobacters* in Market-age Broilers. *Poultry Science*, 80(6), 817–820. <https://doi.org/10.1093/ps/80.6.817>

26. Tabashsum, Z., Peng, M., Kahan, E., Rahaman, S. O., & Biswas, D. (2019). Effect of conjugated linoleic acid overproducing *Lactobacillus* with berry pomace phenolic extracts on *Campylobacter jejuni* pathogenesis. *Food & Function*, 10(1), 296–303. <https://doi.org/10.1039/C8FO01863D>
27. Sibanda, N., McKenna, A., Richmond, A., Ricke, S. C., Callaway, T., Stratakos, A. Ch., Gundogdu, O., & Corcionivoschi, N. (2018). A Review of the Effect of Management Practices on *Campylobacter* Prevalence in Poultry Farms. *Frontiers in Microbiology*, 9, 2002. <https://doi.org/10.3389/fmicb.2018.02002>
28. Svetoch, E. A., & Stern, N. J. (2010). Bacteriocins to control *Campylobacter* spp. In poultry—A review. *Poultry Science*, 89(8), 1763–1768. <https://doi.org/10.3382/ps.2010-00659>
29. Mavri, A., & Smole Možina, S. (2013). Development of antimicrobial resistance in *Campylobacter jejuni* and *Campylobacter coli* adapted to biocides. *International Journal of Food Microbiology*, 160(3), 304–312. <https://doi.org/10.1016/j.ijfoodmicro.2012.11.006>
30. Mavri, A., & Smole Možina, S. (2013). Development of antimicrobial resistance in *Campylobacter jejuni* and *Campylobacter coli* adapted to biocides. *International Journal of Food Microbiology*, 160(3), 304–312. <https://doi.org/10.1016/j.ijfoodmicro.2012.11.006>
31. Fischer, S., Kittler, S., Klein, G., & Glünder, G. (2013). Impact of a Single Phage and a Phage Cocktail Application in Broilers on Reduction of *Campylobacter jejuni* and Development of Resistance. *PLoS ONE*, 8(10), e78543. <https://doi.org/10.1371/journal.pone.0078543>
32. Santini, C., Baffoni, L., Gaggia, F., Granata, M., Gasbarri, R., Di Gioia, D., & Biavati, B. (2010). Characterization of probiotic strains: an application as feed additives in poultry against *Campylobacter jejuni*. *International journal of food microbiology*, 141 Suppl 1, S98–S108. <https://doi.org/10.1016/j.ijfoodmicro.2010.03.039>
33. Davidson, P. M., Sofos, J. N., & Branen, A. L. (Eds.). (2005). *Antimicrobials in Food* (0 ed.). CRC Press. <https://doi.org/10.1201/9781420028737>
34. McMillin K. W. (2008). Where is MAP Going? A review and future potential of modified atmosphere packaging for meat. *Meat science*, 80(1), 43–65. <https://doi.org/10.1016/j.meatsci.2008.05.028>
35. Wagenaar, J. A., Bergen, M. A. P. V., Mueller, M. A., Wassenaar, T. M., & Carlton, R. M. (2005). Phage therapy reduces *Campylobacter jejuni* colonization in broilers. *Veterinary Microbiology*, 109(3–4), 275–283. <https://doi.org/10.1016/j.vetmic.2005.06.002>
36. Keener, K. M., Bashor, M. P., Curtis, P. A., Sheldon, B. W., & Kathariou, S. (2004). Comprehensive Review of *Campylobacter* and Poultry Processing. *Comprehensive reviews in food science and food safety*, 3(2), 105–116. <https://doi.org/10.1111/j.1541-4337.2004.tb00060.x>
37. Demirok, E., Veluz, G., Stuyvenberg, W. V., Castañeda, M. P., Byrd, A., & Alvarado, C. Z. (2013). Quality and safety of broiler meat in various chilling systems. *Poultry science*, 92(4), 1117–1126. <https://doi.org/10.3382/ps.2012-02493>
38. Tajkarimi, M. M., Ibrahim, S. A., & Cliver, D. O. (2010). Antimicrobial herb and spice compounds in food. *Food Control*, 21(9), 1199–1218. <https://doi.org/10.1016/j.foodcont.2010.02.003>
39. Ahmadifar, E., Falahatkar, B., & Akrami, R. (2011). Effects of dietary thymol-carvacrol on growth performance, hematological parameters and tissue composition of juvenile rainbow trout, *Oncorhynchus mykiss*: Effects of thymol-carvacrol on rainbow trout. *Journal of Applied Ichthyology*, 27(4), 1057–1060. <https://doi.org/10.1111/j.1439-0426.2011.01763.x>
40. Jamroz, D., Wiertelcki, T., Houszka, M., & Kamel, C. (2006). Influence of diet type on the inclusion of plant origin active substances on morphological and histochemical characteristics of the stomach and jejunum walls in chicken. *Journal of animal physiology and animal nutrition*, 90(5-6), 255–268. <https://doi.org/10.1111/j.1439-0396.2005.00603.x>
41. Panea, B., Ripoll, G., González, J., Fernández-Cuello, Á., & Albertí, P. (2014). Effect of nanocomposite packaging containing different proportions of ZnO and Ag on chicken breast meat quality. *Journal of Food Engineering*, 123, 104–112. <https://doi.org/10.1016/j.jfoodeng.2013.09.029>
42. Azam, A., Ahmed, A. S., Oves, M., Khan, M. S., Habib, S. S., & Memic, A. (2012). Antimicrobial activity of metal oxide nanoparticles against Gram-positive and Gram-negative bacteria: a comparative study. *International journal of nanomedicine*, 7, 6003–6009. <https://doi.org/10.2147/IJN.S35347>
43. Xie, Y., He, Y., Irwin, P. L., Jin, T., & Shi, X. (2011). Antibacterial activity and mechanism of action of zinc oxide nanoparticles against *Campylobacter jejuni*. *Applied and environmental microbiology*, 77(7), 2325–2331. <https://doi.org/10.1128/AEM.02149-10>
44. Sirelkhatim, A., Mahmud, S., Seeni, A., Kaus, N. H. M., Ann, L. C., Bakhori, S. K. M., Hasan, H., & Mohamad, D. (2015). Review on Zinc Oxide Nanoparticles: Antibacterial Activity and Toxicity Mechanism. *Nano-micro letters*, 7(3), 219–242. <https://doi.org/10.1007/s40820-015-0040-x>