

ISOLATION AND CHARACTERIZATION OF RICE BLAST AND BROWN SPOT DISEASE IN DIFFERENT REGIONS OF TAMIL NADU

ABSTRACT

Rice is the most important staple food crop cultivated in almost every district of Tamil Nadu. Among the rice fungal diseases, rice blast caused by *Pyricularia oryzae* and brown spot caused by *Bipolaris oryzae* are considered to be most potent threats causing major yield losses in Tamil Nadu. The present study aims in identification and characterization of rice blast and brown spot pathogen from different rice growing regions of Tamil Nadu. Three isolates of blast pathogen and five isolates of brown spot pathogen were collected and characterized based on colony morphology and conidial characters. *P. oryzae* and *B. oryzae* was identified based on cultural and morphological characters. Molecular characterization of blast and brown spot isolates were done with universal ITS primers. Pathogenicity test was carried out for all the isolates of blast and brown spot and the isolate showing maximum disease incidence was found as virulent one. The present work of morphological and molecular characterization of blast and brown spot pathogen will therefore be helpful to identify and manage the disease effectively.

Key words: *Rice, Pyricularia oryzae, Bipolaris oryzae, blast, brown spot, isolates, characterization*

1. INTRODUCTION

Rice (*Oryza sativa* L.) is the major staple food crop for more than half of the world's population belonging to the family Poaceae. China is the leading producer of rice followed by India with an area and production of 46.38 million hectares and 130.29 million tonnes respectively during the year 2021-22. Rice is grown in almost all districts of Tamil Nadu. The demand for rice is increasing due to continuous increase in population. There is a need to improve the production strategy of rice by reducing the losses caused by biotic and abiotic stresses.

Rice diseases are one of the biotic pressures that cause roughly 15.6% of losses annually [10]. Among various diseases of rice, the rice blast and brown spot are the most important fungal diseases causing significant yield loss. The pathogen *Pyricularia oryzae* (T: *Magnaporthe oryzae*) is the causal organism of rice blast. The pathogen infects almost all parts of the plant viz., leaf, collar, neck, panicle etc. The symptom appears as minute spots which later become enlarged spindle shaped lesions with grey centre and dark brown margin. The *Bipolaris oryzae* (Syn: *Helminthosporium oryzae*) causes brown spot of rice. This pathogen infects the leaves and the panicle, symptom appears as minute brown dots which later become oval shaped resembling sesame seed.

Various studies on morphological and molecular characterization of rice blast pathogen and brown spot pathogen were carried out. Many studies on morphological and molecular characterization of rice blast pathogen *Pyricularia oryzae* (Gowrisri *et al.*, 2019; Kumar *et al.*, 2019) and rice brown spot pathogen *Bipolaris oryzae* was performed (Jaiganesh & Kannan., 2019; Monisha *et al.*, 2019).

In this study, isolation of the pathogen causing blast and brown spot in rice were done by collecting infected leaf samples from different regions of Tamil Nadu. The isolates were identified and characterized based on morphological characteristics like colony characters, mycelial growth, conidial characters etc. The molecular characterization was done by PCR amplification of ITS regions for all the isolates.

2. MATERIALS AND METHODS

2.1. Isolation of blast and brown spot pathogen of rice

Blast and brown spot infected rice leaf samples were collected from different rice growing regions of Tamil Nadu viz., Coimbatore, Perambalur and Thanjavur districts. The infected leaf samples were cut into small pieces along with healthy portion and surface sterilized in 1% sodium hypochlorite for 1 minute followed by subsequent washing with sterile distilled water for three times to remove excess sodium hypochlorite. Then the sterilized leaf bits were blot dried in sterile filter paper and transferred into sterile petri-plate containing Potato Dextrose Agar (PDA) medium. The plates were incubated at $28 \pm 2^\circ\text{C}$ and provided with 12 hours of alternate light and darkness. After 3 days, the actively growing mycelium were sub-cultured and purified by single hyphal tip method. The purified cultures were maintained in PDA slants at 4°C for further study.

2.2. Morphological characterization

Morphological characterization of all the isolates of *Pyricularia oryzae* and *Bipolaris oryzae* were carried out in PDA medium and incubated at room temperature for 7 days. After incubation, colony growth, colony characters, conidium shape, colour, and septations were observed for all the isolates. For the sporulation of *P. oryzae* and *B. oryzae*, a 9mm mycelial disc were placed at centre of a sterile glass slide under aseptic condition kept on sterile petri-plate with moist sterile cotton. The petri-plates were sealed and incubated at $25 \pm 2^\circ\text{C}$ for 3 days with alternate 12 hours light and dark period. After 3 days of incubation, the spores were collected using sterile distilled water and observed under compound microscope. The stress condition was given to the pathogen for inducing sporulation by growing in plain agar medium.

2.3. Pathogenicity test

To prove the pathogenicity of the pathogen, variety CO39 susceptible to both blast and brown spot were raised in pots under glasshouse condition. 21 days old seedlings were inoculated with spore suspension of 5×10^5 conidia/ml of each isolate of *Pyricularia oryzae* and *Bipolaris oryzae*. The inoculated plants were given frequent watering and covered with polythene bags to maintain humid condition favourable for disease development. For each pathogen 5 pots were maintained. A healthy plant without inoculation of pathogen served as control. After symptom expression, the pathogen was re-isolated from blast and brown spot infected leaves. The re-isolated culture was confirmed with original culture and confirmed based on cultural and morphological characters.

2.4. Molecular characterization

The total genomic DNA from all the isolates of *Pyricularia oryzae* and *Bipolaris oryzae* was extracted by the CTAB method [1]. The isolates were grown in Potato Dextrose broth for 15 days. The mycelial mats were collected through filter paper, dried at room temperature for 24 hours and then used for DNA extraction. The genomic DNA extracted was electrophoresed on 0.8% agarose gel and confirmed the presence of genomic DNA by documentation under image analyser.

2.4.1. PCR amplification of ITS region

The universal primers ITS-1 (5'- TCCGTAGGTGAACCTGCGG -3') and ITS- 4 (5'- TCCTCCGCTTATTGATATGC -5') were used for identification of all the isolates of *P. oryzae* and *B. oryzae*. The PCR amplification reaction was carried out for each isolate with a final volume of 10 μ l, which consists of 5 μ l Master mix, 1 μ l of forward primer (ITS-1), 1 μ l of reverse primer (ITS-4), 2 μ l of sterile distilled water and 1 μ l of DNA. The PCR amplification was carried out under the following condition in Thermocycler: initial denaturation at 95°C for 2 minutes, followed by 40 cycles of denaturation at 95°C for 1 minute, annealing at 58°C for 1 minute, extension at 72°C for 1 minute and final extension at 72°C for 5 minutes, and then hold at 10°C . Then the PCR products were electrophoresed in 1% agarose gel with ethidium bromide for 45 minutes in 1X TAE buffer and then visualized under UV light in gel documentation unit.

2.4.2. Sequencing of PCR products

A PCR reaction was performed for a total volume of 40 μ l, using Emerald Amp[®] GT PCR master mix, forward primer (ITS-1), reverse primer (ITS-4) for genomic DNA of virulent *P. oryzae* isolate (PO CBE1) and *B. oryzae* isolate (BO CBE1). The PCR amplification was carried out in thermocycler. The PCR products were resolved in 1% agarose gel electrophoresis. The PCR products was then sequenced. The obtained nucleotide sequence was used as input sequence in nucleotide blast analysis program at NCBI database. The output data obtained were analysed and the organism

showing major score was considered as the closely related species to the test fungus (*P. oryzae* and *B. oryzae*).

3. RESULTS

3.1. Isolation of *P. oryzae* and *B. oryzae*

Blast infected leaf samples showing characteristic spindle shaped lesions and brown spot infected leaf samples showing oval shaped lesions were collected from different regions of Tamil Nadu. The pathogen *P. oryzae* and *B. oryzae* were isolated from blast and brown spot infected leaf samples respectively. A total of three isolates of *Pyricularia oryzae* and five isolates of *Bipolaris oryzae* were obtained from different regions of Tamil Nadu (Table 1)

Table 1. Isolates of *Pyricularia oryzae* and *Bipolaris oryzae*

S. NO.	ISOLATES	LOCATION	DISTRICT
Isolates of <i>Pyricularia oryzae</i>			
1.	PO CBE1	Paddy Breeding Station, TNAU	Coimbatore
2.	PO CBE2	Wetland, TNAU	Coimbatore
3.	PO ECK1	AC & RI, Eachangkottai	Thanjavur
Isolates of <i>Bipolaris oryzae</i>			
1.	BO BSR1	Research station	Bhavani Sagar
2.	BO CBE1	Paddy Breeding Station, TNAU	Coimbatore
3.	BO CBE2	Wetland, TNAU	Coimbatore
4.	BO PBR1	Farmer's field	Perambalur
5.	BO ECK1	AC & RI, Eachangkottai	Thanjavur

3.2. Morphological characterization of *P. oryzae* and *B. oryzae*

The colony morphology, growth pattern, radial mycelial growth, conidial characters of all the isolates of *P. oryzae* and *B. oryzae* were studied on PDA medium and presented in Table 2 and Table 3 respectively. The colony characters varied for different isolates of *P. oryzae* (Fig 1) and *B. oryzae* (Fig 2). The diameter of the mycelial growth was measured at 7th day after incubation for all the isolates. Among the three isolates of *Pyricularia oryzae*, the PO CBE1 isolate grew well on PDA medium with mycelial growth of 6.72 cm. The isolate BO CBE1 showed maximum mycelial growth of 8.92 cm compared to other isolates of *Bipolaris oryzae*.

All the isolates produced spores on PDA medium. The spores were observed under phase contrast microscope and size, shape and number of septations were studied. The conidia of *B. oryzae* isolates varied from oval to cylindrical in shape, slightly curved with a bulge in the middle and tapering towards the ends (Fig 3). The size of conidia and number of septations also varied among different isolates of *B. oryzae*. The conidia of *P. oryzae* isolates were typically pyriform with a narrow apex, round base and a short hilum, 2 – 3 septate (Fig 4).

Table 2. Morphological characteristics of *P. oryzae*

S.No.	Isolate	Radial mycelial growth (in cm)*	Colony characters
1.	PO CBE1	6.72 (2.59)	Greyish brown, flat mycelium with concentric growth
2.	PO CBE2	6.08 (2.47)	Light grey to white, flat mycelium with concentric growth
3.	PO ECK1	6.24 (2.50)	Greyish white, slightly raised mycelium with concentric growth
SE(d)		0.03	
CD (0.05)		0.065	

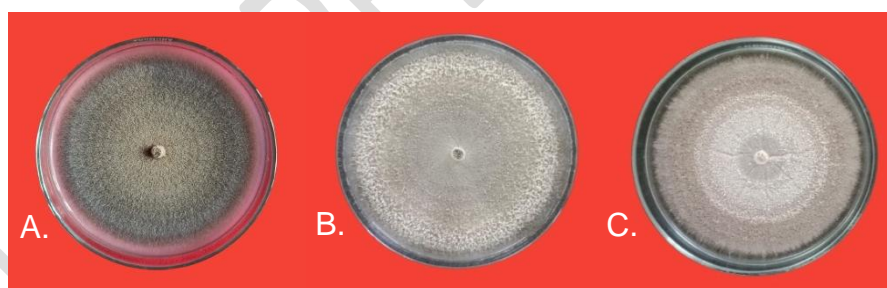
* - Mean of five replications

Table 3. Morphological characteristics of *B. oryzae*

S.No.	Isolate	Radial mycelial growth (in cm)*	Colony characters
1.	BO BSR1	6.68 (2.59)	Mixture of grey and white, fluffy mycelium
2.	BO CBE1	8.92 (2.99)	Greyish black, fluffy mycelium with grey cottony mycelium in centre
3.	BO CBE2	7.68 (2.77)	Greyish, fluffy mycelium
4.	BO PBR1	8.64 (2.94)	Greyish brown, fluffy mycelium
5.	BO ECK1	7.52 (2.74)	Greyish brown, fluffy mycelium with white dots
SE(d)		0.022	
CD (0.05)		0.045	

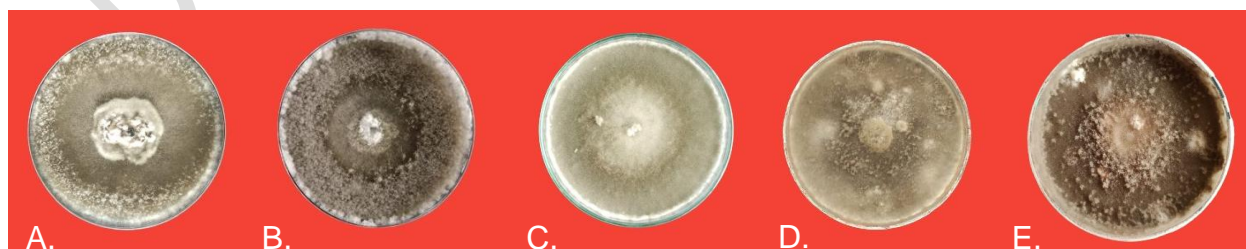
* - Mean of five replications

Fig 1. Colony characters of *P. oryzae* isolates



A. PO CBE1; B. PO CBE2; C. PO ECK1

Fig 2. Colony characters of *B. oryzae* isolates



A. BO BSR1; B. BO CBE1; C. BO CBE2; D. BO PBR1; E. BO ECK1

Fig 3. Conidia of *P. oryzae*



Fig 4. Conidia of *B. oryzae*



3.3. Pathogenicity test

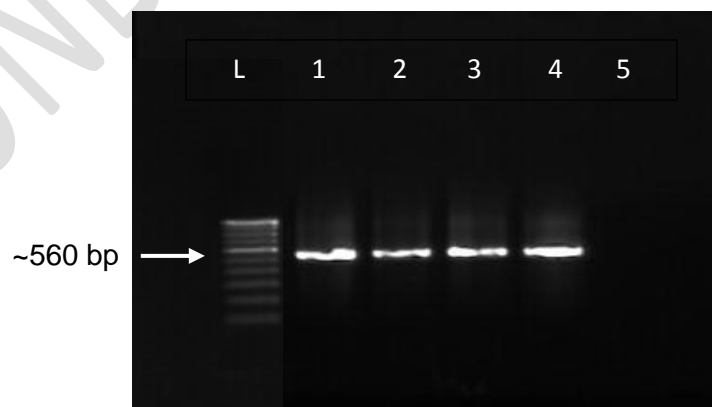
The pathogenicity test was performed using variety CO39 susceptible to brown spot and blast. The symptom of brown spot started to appear 3rd day after inoculation. The pathogen initially produced minute, brown spots, which gradually enlarged to oval shaped spots. The blast symptom initiated after 4 days of spraying. The symptom initially appeared as minute spots which later turned into characteristic spindle shaped, brown lesions. The pathogens were re-isolated from the leaf showing typical symptoms. The re-isolated pathogen was confirmed as *P. oryzae* and *B. oryzae* based on colony characters and conidial morphology. The colony characters and conidial morphology of the re-isolated pathogen was observed under compound microscope and confirmed as *P. oryzae* and *B. oryzae*.

From the pathogenicity test, the isolate PO CBE1 and BO CBE1 taken from Paddy Breeding station, Coimbatore were found to be virulent showing highest percent disease incidence when compared to other isolates.

3.4. Molecular characterization of *P. oryzae* and *B. oryzae*

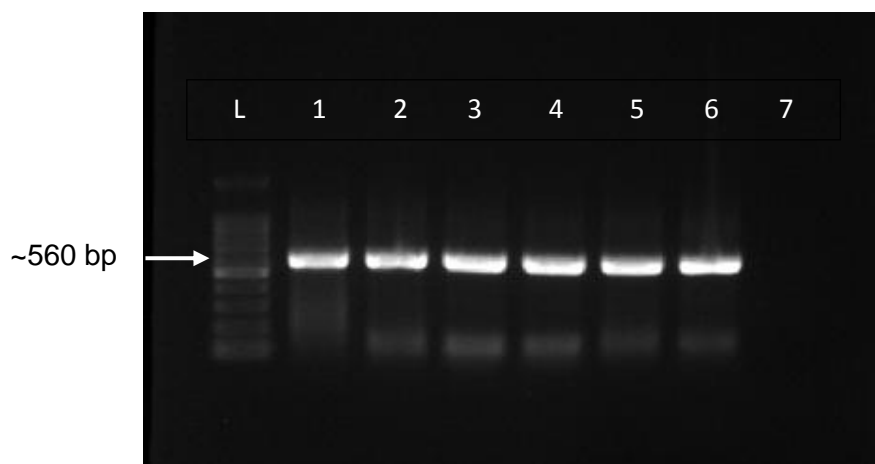
PCR amplification of ITS-1 and ITS-4 regions using universal primers showed an amplicon of size 560bp for all the isolates of *P. oryzae* (Fig 5) and *B. oryzae* (Fig 6). The unpurified PCR product of the virulent isolate of *P. oryzae* and *B. oryzae* were sequenced. The newly obtained sequences of PO CBE1 and BO CBE1 were then deposited in GenBank (NCBI database) and accession number were obtained.

Fig 5. PCR amplification of ITS region of *P. oryzae*



L – Ladder, Lane 1 – PO CBE1, Lane 2 – PO CBE2, Lane 3 – PO ECK1, Lane 4 – Positive control (PC), Lane 5 – Negative control (NC)

Fig 6. PCR amplification of ITS region of *B. oryzae*



L – Ladder, Lane 1 – BO BSR1, Lane 2 – BO CBE1, Lane 3 – BO CBE2, Lane 4 – BO PBR1, Lane 5 – BO ECK1, Lane 6 – Positive control (PC), Lane 7 – Negative control (NC)

4. DISCUSSION

4.1. Isolation of *P. oryzae* and *B. oryzae*

The pathogen *P. oryzae* causing rice blast and *B. oryzae* causing brown spot of rice were isolated from infected leaves showing typical symptoms using Potato dextrose agar medium and sub-cultured using single hyphal tip method. Different media like Oat meal agar medium, Rice straw extract agar medium, etc can be used for culturing of the pathogen. The pathogen was identified as *P. oryzae* and *B. oryzae* based on descriptions of colony characters and conidial characters given by Ou (1985). Six isolates of *P. oryzae* was isolated from different regions of Tamil Nadu. [3]. Five isolates of *B. oryzae* were collected from different villages in Cuddalore district, Tamil Nadu and characterized on PDA medium (Jaiganesh & Kannan., 2019).

4.2. Morphological characterization of *P. oryzae* and *B. oryzae*

In the present study, all the isolates of *P. oryzae* and *B. oryzae* showed variations in cultural and morphological characteristics on PDA medium. The mycelium of *P. oryzae* varied from flat to slight raised, greyish white to greyish brown, showing concentric growth pattern. The results were in accordance with the prior work of Gowrisri *et al* (2019) on the morphological and molecular characterization of *Magnaporthe oryzae*, causing rice blast. They have grown 6 isolates of *Magnaporthe oryzae* on PDA medium and studied the variation in colony characters and conidial size. The mycelial growth showed variation in colour from brownish grey to pure white and both flat and aerial. The spore size also varied between the isolates. Kumar *et al* (2019) grouped 71 isolates collected from three states of eastern India into five groups based on colony characters. Sahu *et al* (2018) identified and characterized the rice blast pathogen from different rice growing regions of Odisha and categorized 20 isolates into three groups.

The colony characters of *B. oryzae* varied from greyish brown to greyish black in colour, slightly raised or aerial, fluffy mycelium showing white dots. The conidial size, shape and number of septations also varied among the different isolates. The similar results were obtained by Jaiganesh and Kannan (2019). They studied the cultural characters and pathogenicity of *Helminthosporium oryzae* causing brown spot of rice. The colony morphology of the isolates showed light brown to black, septate, aerial or submerged, branched mycelium. The conidia were light brown to brown in colour, slightly curved with bulge in the middle and tapering toward the end. Monisha *et al* (2019) collected five isolates of *B. oryzae* from different locations of Tamil Nadu and characterized them based on morphological and cultural characters. Meghana *et al* (2019) studied the cultural, morphological and molecular variability

of *B. oryzae* causing brown spot of rice in Northern Karnataka. Among different media test, Potato dextrose agar medium showed better growth of *B. oryzae*.

4.3. Pathogenicity test

The pathogenicity of *P. oryzae* and *B. oryzae* were proved by artificial inoculation of spore suspension on the susceptible rice variety CO39 and humid condition was provided. The symptom development was observed 7 days after inoculation and the virulent isolate was obtained based on maximum percent disease incidence. The similar method was followed for proving the pathogenicity of *Magnaporthe oryzae* (Gowrisri *et al.*, 2019; Kumar *et al.*, 2019; Panda *et al.*, 2017) and *Bipolaris oryzae* on rice (Jaiganesh & Kannan., 2019; Monisha *et al.*, 2019).

4.4. Molecular characterization of *P. oryzae* and *B. oryzae*

The PCR amplification of ITS regions were done using universal primers ITS-1 and ITS-4 for molecular identification of *P. oryzae* and *B. oryzae*. An amplicon size of ~560bp were obtained from all the isolates of *P. oryzae* and *B. oryzae*. The similar method was used for molecular confirmation of isolates of *P. oryzae* (Gowrisri *et al.*, 2019; Mohammadpourlima *et al.*, 2017) and *B. oryzae* (Monisha *et al.*, 2019; Meghana *et al.*, 2019). The confirmation of the pathogen at species level can be done using gene specific primers. Molecular characterization of *Magnaporthe oryzae* was done by PCR amplification of Pot2 transposon region using specific primers (Gowrisri *et al.*, 2019).

5. CONCLUSION

Three isolates of *Pyricularia oryzae* causing rice blast and five isolates of *Bipolaris oryzae* causing brown spot of rice were collected from different locations of Tamil Nadu. The isolates were characterized based on morphological and molecular characteristics. The isolates were confirmed as *P. oryzae* and *B. oryzae* based on morphological and cultural characters. All the isolates showed variations in colony growth, growth pattern, mycelial characters, conidia size, shape etc. The cultural variability can also be studied by growing the pathogen in different media.

From the pathogenicity test for both the pathogen, the virulent isolate was found and can be used for further screening. Molecular confirmation of *P. oryzae* and *B. oryzae* was done using PCR amplification of ITS regions using primers ITS-1 and ITS-4. Further confirmation of the pathogen at species level can be done using gene specific primers.

REFERENCES

1. Doyle JJ, Doyle JL. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochemical bulletin*. 1987.
2. El-Shafey RA, Elamawi RM, El-Wahsh SM, AbdKhalek AF, Badr EA, El-Refae YZ. Morphopathological and molecular variation studies among different pathotypes of *Helminthosporium fungus* causing brown spot disease of rice. *Egypt. J. Phytopathol.* 2011;39(1):135-53.
3. Gowrisri N, Kamalakannan A, Malathi VG, Rajendran L, Rajesh S. Morphological and molecular characterization of *Magnaporthe oryzae* B. Couch, inciting agent of rice blast disease. *Madras Agricultural Journal.* 2019 Jun 2;106.
4. Jagadeesh D, Devaki NS. Morphological and biochemical variation of rice blast fungus *Magnaporthe oryzae* in Karnataka, India. *International Journal of Agricultural Technology.* 2020;16(4):799-818.
5. Jaiganesh V, Kannan C. Studies on the cultural characters and pathogenicity studies of brown leaf spot of rice caused by *Helminthosporium oryzae* (Syn: *Bipolaris oryzae*). *Plant Archives.* 2019;19:585-7.
6. Kulkarni K, Peshwe S. Screening, isolation and molecular identification of rice pathogen *Magnaporthe oryzae*. *Int. J. Adv. Res.* 2019;7:428-33.
7. Kumar YM, Aravindan S, Prabhukarthikeyan SR, Keerthana U, Raghu S, Archana B, Pankajini S, Motilal B, Kumari KM, PC R, Pramesh D. Characterization and molecular phylogeny of *Magnaporthe oryzae* causing rice blast disease in eastern India. *Research Journal of Biotechnology Vol.* 2019 Jun;14:6.

8. Manjunatha B, Krishnappa M. Morphological characterization of *Pyricularia oryzae* causing blast disease in rice (*Oryza sativa* L.) from different zones of Karnataka. *Journal of Pharmacognosy and Phytochemistry*. 2019;8(3):3749-53.
9. Mohammadpourlima M, Yassoralipour A, Tong PE, Ahmad ZA, Yun WM. Morphological and molecular characterizations of rice blast fungus, *Magnaporthe oryzae*. *Pakistan Journal of Agricultural Sciences*. 2017 Dec 1;54(4).
10. Mondal D, Ghosh A, Roy D, Kumar A, Shamurailatpam D, Bera S, Ghosh R, Bandopadhyay P, Majumder A. Yield loss assessment of rice (*Oryza Sativa* L.) due to different biotic stresses under system of rice intensification (SRI). *Journal of Entomology and Zoology Studies*. 2017;5(4):1974-80.
11. Monisha S, Praveen NM, Ramanathan A. Isolation, characterization and management of brown spot disease of rice. *Journal of Pharmacognosy and Phytochemistry*. 2019;8(3):4539-45.
12. Nayak MS, Hiremath SV. Cultural, morphological and molecular characterization of *Bipolaris oryzae* causing brown leaf spot of rice in Northern Karnataka. *Journal of Pharmacognosy and Phytochemistry*. 2019;8(2):1235-9.
13. Ou SH. Rice diseases. IRRI; 1985.
14. Panda G, Sahu C, Yadav MK, Aravindan S, Umakanta N, Raghu S, Prabhukarthikeyan SR, Lenka S, Tiwari JK, Kar S, Jena M. Morphological and molecular characterization of *Magnaporthe oryzae* from Chhattisgarh. *ORYZA-An International Journal on Rice*. 2017;54(3):330-6.
15. Priyanka P, Kumar YM, Prabhukarthikeyan SR, Raghu S, Keerthana U, Umakanta N, Pramesh D. Morpho-molecular characterization of Indian population of *Magnaporthe oryzae*, an incitant of blast disease in rice. *Research Journal of Biotechnology* Vol. 2021 Jan;16:1.
16. Sahu C, Yadav MK, Panda G, Aravindan S, Umakanta N, Raghu S, Prabhukarthikeyan SR, Keerthana U, Adak T, Sharma V, Mohanty MR. Morphological and molecular characterization of *Magnaporthe oryzae* causing rice blast disease in Odisha. *ORYZA-An International Journal on Rice*. 2018;55(3):467-72.
17. Shaw S, Prabhukarthikeyan SR, Keerthana U, Aravindan S, Yadav MK, Raghu S, Baite MS, Naveenkumar R, Parida S, Panda G, Rath PC. Morphological and molecular characterization of *Magnaporthe grisea* and bio-efficacy of *Bacillus* strains against *M. grisea*. *Int. J. Curr. Microbiol. App. Sci*. 2019;8(6):1900-8.
18. Sobanbabu G, Sabarinathan KG, Parthiban VK, Ramamoorthy V. Isolation, screening and identification of virulent isolates of *Bipolaris oryzae* causing rice brown spot and *Sarocladium oryzae* causing sheath rot disease. *International Journal of Current Microbiology and Applied Sciences*. 2018;7(09):930-9.
19. Sonah H, Deshmukh RK, Parida SK, Chand S, Kotasthane A. Morphological and genetic variation among different isolates of *Magnaporthe grisea* collected from Chhattisgarh. *Indian Phytopathology*. 2009;62(4):469.
20. Srivastava D, Shamim MD, Kumar D, Pandey P, Khan NA, Singh KN. Morphological and molecular characterization of *Pyricularia oryzae* causing blast disease in rice (*Oryza sativa*) from North India. *International Journal of Scientific and Research Publications*. 2014;4(7):2250-3153.