

Isolation and Characterization of Plant Growth Promoting Rhizobacteria form *Raphanussativus*(Radish)

Abstract:

Rhizobacteria are present in rhizosphere region of plant root system, which enhance the plant growth by various way like biological nitrogen fixation, siderophore production, phosphate solubilization and phytohormone production. The soil samples were collected from rhizosphere region of *Raphanussativus*, after enrichment rhizobacteria was isolated by serial dilution method, diluted sample were spread on respective solid agar media plates. Isolated rhizobacteria was identify by biochemical and molecular characterization methods. The isolated PGPRs was *Bacillus subtilis* which was showed phosphate solubilization activity and they were enhanced 30% more green gram seed germination. The method of current research is screening, isolation and biochemical characterization of rhizobacteria form rhizosphere region of *Raphanussativus*. Phosphate solubilizing bacteria, solubilized the unavailable phosphate and provide to plant. The main purpose of this research paper is to widen the understanding of the role of phosphate solubilizing bacteria in crop production as biofertilizers.

Keywords: Rhizobacteria, Rhizosphere, Solubilization, Biochemical, Phosphate.

Introduction:

The groups of microorganism which is colonized in rhizospheric region of plant roots called as plant growth-promoting rhizobacteria (PGPR), they enhanced plant growth by nutrient immobilization. Some common examples of PGPR genera exhibiting plant growth promoting activity are *Pseudomonas*, *Azospirillum*, *Azotobacter*, *Bacillus*, *Burkholdaria*, *Enterobacter*, *Rhizobium*, *Erwinia*, *Mycobacterium*, *Mesorhizobium*, *Flavobacterium*, etc (Odoh, 2017, Birhanu, 2022). PGPRs have been shown to directly enhance plant growth by several mechanisms, including the fixation of atmospheric nitrogen transferred to the plant, the production of siderophores that chelate iron and make it available to the plant root, the solubilization of minerals like phosphorus, and the synthesis of phytohormones (Bhattacharyya et. al., 2012, Rawat et al., 2021). The potentiality of PGPR offers an attractive way to replace the use of chemical fertilizers, pesticides and other supplements to increase agriculture yield (Aloet

al., 2022, Gulnazet *al.*, 2017). The different growth-promoting characteristics of the rhizobacteria identified in the rhizosphere of *Raphanussativus* are discussed in the current work, as well as their morphological characterization, biochemical characterization and molecular characterization of the isolated strains.

Purpose of current research to isolate and identify the potential phosphate solubilizing rhizobacteria, which enhances the growth and yield of crops. Most soils possess considerable amounts of phosphate, phosphorus is a very reactive element and does not exist as elemental form in the soil, and large proportion is bound to soil particles. A group of rhizobacteria capable of hydrolyzing organic and inorganic insoluble phosphate compounds to soluble phosphate form that can easily be assimilated by plants. The major objective of current study:

- To screening and isolation of plant growth promoting rhizobacteria from rhizosphere of *Raphanussativus* (Radish) root.
- Biochemical and molecular characterization of PGPR.
- Effect of rhizobacterial isolates on seed germination.

Review of literature:

Plant growth promoting rhizobacteria consist of a group of rhizobacteria that colonize in rhizosphere which can increase the root growth and influence the plant physiology. These rhizobacteria have a crucial role in sustaining soil fertility and plant health (Glick, 2012, Alori, 2017). Plant growth promoting rhizobacteria increase plant growth through either direct mechanism by providing readily available nutrient for plants such as nitrogen, phosphorus and plant hormones or through indirect mechanism by synthesizing hydrogen cyanide (HCN), antibiotics, and siderophores (Mahanteshet *al.*, 2015). The mechanism of solubilization of insoluble phosphate, rhizobacteria having ability to secrete organic acids and phosphatase enzymes which solubilized the insoluble phosphate into soluble form, which decreases the dependency of high cost phosphatic fertilizers in agriculture practice. Hashem et al. 2019, reported the activity of *B. subtilis* in the rhizosphere, *Bacillus* species are a significant type of rhizobacteria which can form spores that can survive in the soil for long period of time under adverse conditions and also it is a root colonizer increase crop productivity under conditions of biotic and abiotic stress. Agustiyani et al. 2022, they characterized the PGPR isolated from rhizospheric soils of various plant and checked its effect on growth of radish

(*Raphanussativus*L.), total 15 PGPR isolates were isolated from various plant roots and in vitro screening was done for different plant growth promotion activities. PGPR on Radish growth in the green house showed that all isolates had effects on increasing growth and tuber formation compared to control.

Materials and methods:

Collection of rhizospheric soil samples

Rhizospheric soil sample was collected from Radish plant agriculture field Prayagraj, Uttar Pradesh, India. The roots and adherent topsoil were put in sterile sample collecting bags and delivered to the laboratory for isolation.

Screening and isolation of PGPR

Screening of PGPR by serial dilution method, diluted suspension were spread on Pikovskaya agar media, Jensen media, Azospirillum media (BTB media), King's B media. Spread plant were incubated at 32°C for 72 hrs. Gown colony were screening on the basis of colour change and clear hydrolytic zone around the colony.

Morphological and biochemical characterization of isolates

Colony morphology were observed by Gram staining technique under 40X objective of the microscope (Cappuccino and Sherman, 1996). The biochemical identification like Indole Test, Methyl Red Test, Voges-Proskauer (VP) Test, Citrate Utilization Test, Gelatin Hydrolysis Test, Starch Hydrolysis Test, Hydrogen Sulphide Test, NO₂ Reducing Test, Nitrogen Fixation Test, IAA Production, Siderophore Production, Ammonium Production and Phosphate Solubilization was done as per the procedure given by (Cappuccino and Sherman, 1996) and (Aneja, 2003).

Molecular characterization of the isolated strains

Molecular characterization of the isolated strains was based on 16S r-DNA Sequence methods (Altschulet *al.*, 1997).

Application of the isolated Rhizobacteria along with phosphate rich organic manure

200 ml of full grown rhizobacterial culture mixed with 800 gm phosphate rich organic manure, and which was incubate at room temperature to maintain the moisture 25% for 30 days, now that were used in pot soil before sowing of green gram seed.

Result and Discussion:

The aim of current study to isolate the PGPR bacteria from soil samples and isolated culture identification was based on biochemical characterization.

Screening and isolation of PGPR

Screening and isolation of PGPR bacteria based on various function viz. halo zone around the bacterial colony and colour change around the colony. Total 4 bacterial isolates (Isolates - I, II III and IV) were selected. Isolates were grown on only Pikovskaya agar media and they showed halo zone around the bacterial colony. No any isolates was grown and shown positive character on Jensen media, BTB media and King's B media.

Morphological and biochemical characterization of isolates

The morphology of the selected isolates were tested by gram's staining method, isolates I, II, III and IV was Gram positive and rod shaped bacteria. According to table 1 and Figure 1, isolates was positive for Indole production, Acids produced, Citrate utilization, Gelatin hydrolysis, Starch hydrolysis, Hydrogen sulphide, NO₂ reduction, Ammonium production and Phosphate solubilisation. While, isolates was negative for Voges-Proskauer (VP) test, Nitrogen fixation test, Siderophore production and in case of IAA production all isolates are negative only isolates IV was positive.

Table 1: Biochemical test of the isolates:

Biochemical Test	Isolate-I	Isolate-II	Isolate-III	Isolate-IV
Phosphate solubilisation	+Ve	+Ve	+Ve	+Ve
Indole production	+Ve	+Ve	+Ve	+Ve
Acid produced	+Ve	+Ve	+Ve	+Ve
Citrate utilization	+Ve	+Ve	+Ve	+Ve
Gelatin hydrolysis	+Ve	+Ve	+Ve	+Ve
Starch hydrolysis	+Ve	+Ve	+Ve	+Ve
Hydrogen sulphide	+Ve	+Ve	+Ve	+Ve
NO ₂ reduction	+Ve	+Ve	+Ve	+Ve
Ammonium production	+Ve	+Ve	+Ve	+Ve
Voges-Proskauer (VP) test	+Ve	+Ve	+Ve	+Ve
Nitrogen fixation test,	+Ve	+Ve	+Ve	+Ve
Siderophore production	+Ve	+Ve	+Ve	+Ve
IAA production	-Ve	-Ve	-Ve	+Ve

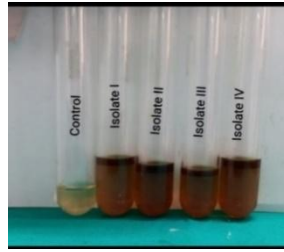


Figure1A:Indoletest of isolates; **Figure1B:**Methyl test of isolates; **Figure1C:**VP test of isolates

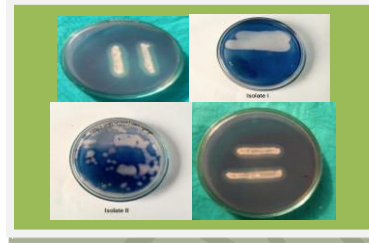
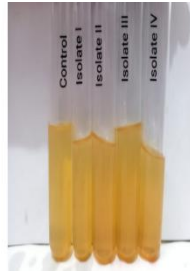
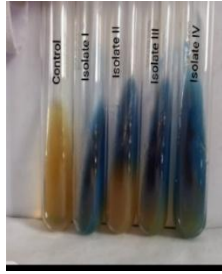


Figure1D:Citrate test;**Figure1E:**Gelatine hydrolysis test; **Figure1F:** Starch hydrolysis test

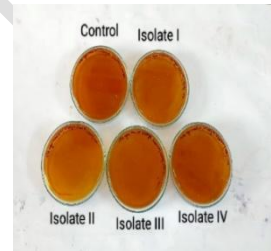
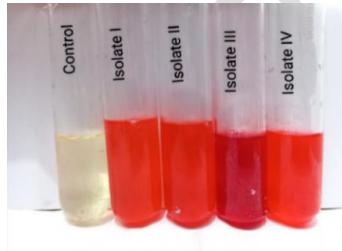
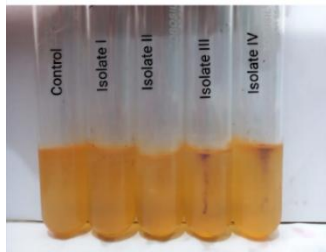


Figure1G:Showing H₂S test; **Figure1H:** showing NO₂test; **Figure1I:**Showing Nfb test

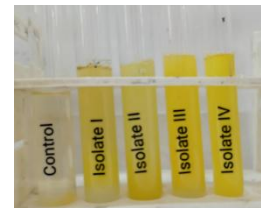
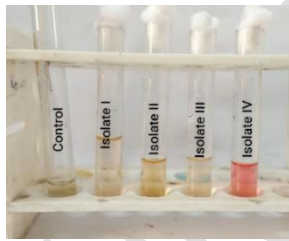


Figure1J: Showing IAA test **Figure1K:** Siderophore test**Figure1L:** Ammonium test

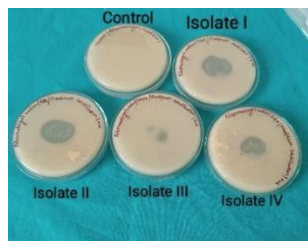


Figure1M:Showing PSB test of isolate

Molecular characterization of the isolated strains

During molecular characterization of the isolated strains genomic DNA was extracted and 16S-rDNA fragment was amplified by PCR methods. Amplified PCR products were sequenced and sequences were aligned and examined with their closest neighbors. The microbe was found to be *Bacillus subtilis* as the *Bacillus subtilis* strain AS1 16S ribosomal RNA gene, partial sequence was found to have the highest percent identity.

Application of the isolated Rhizobacteria along with phosphate rich organic manure

Rhizobacteria-enriched phosphate-rich organic manure was used in pot soil before sowing of seeds. Rhizobacteria-treated pot showed 30% more green gram seed germination.



Figure 2: Observation of percentage seed germination in presence of rhizobacteria

Conclusion

Conclusion of the current work, isolated bacteria from the rhizospheric region of *Raphanus sativus* (Radish) root was *Bacillus subtilis*. Isolated rhizobacteria was a plant growth promoting rhizobacteria and showed phosphate solubilizing activity which was shown to increase green gram seed germination up to 30% more.

References:

Agustiyani D., Purwaningsih S., Dewi T.K., Nditasari A., Nugroho A.A., Sutisna, E., Mulyani N., Antonius S. (2022), Characterization of PGPR isolated from rhizospheric soils of various plants and its effect on growth of radish (*Raphanus sativus* L.), IOP Conference Series: Earth and Environmental Science, 976 :012037.

Aloo B. N., Tripathi V., Makumba B. A., Mbega E.R. (2022), Plant growth-promoting rhizobacterial biofertilizers for crop production: The past, present, and future, *Front. Plant Sci.* 13:1002448.

Alori E.T., Glick R.R., Babalola O.O. (2017), Microbial phosphorus solubilization and its potential for use in sustainable agriculture, *Frontiers in Microbiology*, 8:971

Altschul S.F., Madden T.L., Schaffer A.A., Zhang J., Zhang Z., Miller W., Lipman D.J. (1997), Gapped BLAST and PSI-BLAST: a new generation of protein database.

Aneja K.R. (2003), *Experiments in microbiology plant pathology and biotechnology* (4th Edition). New Age International Publishers, New Delhi, India.

Bhattacharyya P.N and Jha D.K. (2012), Plant growth promoting rhizobacteria (PGPR)-emergence in agriculture, *World Journal Microbiol Biotechnol*, 28 (4):1327-1350.

Birhanu B. (2022), Phosphate solubilizing Rhizobacteria and Their Growth Promoting Ability from Sorghum Rhizosphere soil. *Int. J. Adv. Res. Biol. Sci.*, 9(6): 69-85.

Cappuccino J.G. and Sherman N. (1996), *Microbiology: A Laboratory Manual*. Benjamin/Cummings, Menlo Park. pp. 129-186.

Glick B. (2012), Plant growth-promoting bacteria: mechanisms and applications, *Scientifica*, 20(12):20-35.

Gulnaz Y., Fathima P.S, Denesh G.R, Kulmitra A.K., Shivraj Kumar H.S., Sathisha C., Ajagol P., Nagesh C.R. (2017), Effect of plant growth promoting Rhizobacteria (PGPR) and PSB on growth and yield of irrigated maize under varying levels of phosphorus, *International Journal of Chemical Studies* 2017; 5(5): 1008-1010.

Hashem A., Tabassum B. and Allah E.F.A. (2019), *Bacillus subtilis*: A plant-growth promoting rhizobacterium that also impacts biotic stress, *Saudi J. Biol. Sci.*, 26(6): 1291–1297.

Mahantesh S. P., Patil C. S., Himanshu V. (2015), Isolation and characterization of potent phosphate solubilizing bacteria. *ISOIJ. Microbiol. Biotechnol. Food Sci.* 1: 23-28.

Odoh C. K. (2017), Plant Growth Promoting Rhizobacteria (PGPR): A Bioprotectant bioinoculant for Sustainable Agrobiolgy. A Review. *International Journal of Advanced Research in Biological Sciences*, 4(5): 123-142.

Rawat P., Das S., Shankhdhar D., Shankhdhar S.C. (2021), Phosphate-solubilizing microorganisms: mechanism and their role in phosphate solubilization and uptake, *Journal of Soil Science and Plant Nutrition*, 21(1): 49–68.

UNDER PEER REVIEW