

INFLUENCE OF VARIOUS ORGANIC MANURES ON GROWTH PARAMETERS OF GINGER (*Zingiber officinale* Rosc.) UNDER COCONUT BASED CROPPING SYSTEM

ABSTRACT

Ginger is one of the most important spices which has been utilised for centuries as a food flavouring agent as well as a key ingredient in pharmaceutical products. Since organic food is considered to be healthy by consumers, the demand for organic products is increasing and organic farming is gaining popularity worldwide. As ginger is a shade-loving crop, it can be grown well under the coconut ecosystem. Therefore, a study was undertaken to evaluate the effect of organic amendments on the growth and yield of ginger (IISR Varada) under coconut-based cropping system. The results revealed that the maximum plant height (72.08 cm) was recorded in T₈ - poultry manure + microbial consortium followed by T₉ - goat manure + microbial consortium (71.86 cm), the number of leaves per plant was maximum (86.57) in the combined application of poultry manure and microbial consortium (T₈) followed by T₆ - FYM + microbial consortium (83.03), the largest leaf length (26.75 cm) was observed in T₈ - poultry manure + microbial consortium followed by T₉ - goat manure + microbial consortium (25.83 cm) and similarly, the largest leaf width was observed in T₈ - poultry manure + microbial consortium (3.92 cm) followed by T₉ - goat manure + microbial consortium (3.87 cm). The application of T₈ - poultry manure + microbial consortium produced more tillers (9.48) followed by T₆ - FYM + microbial consortium (9.23). Thus, the combined application of poultry manure and microbial consortium is considered the best and most effective organic amendment for ginger under coconut-based cropping system.

Key word: poultry manure, cropping system, Zingiberaceae, plant height

1. INTRODUCTION

Ginger (*Zingiber officinale* Rosc.) belongs to the family Zingiberaceae a perennial herbaceous monocotyledon plant that is commercially cultivated as an annual crop for its rhizomes. It has been cultivated in the country since ancient times and is a significant commercial spice crop [5]. It is a high-value tropical rhizomatous spice crop grown in tropical and subtropical climates. Despite being grown in numerous countries, it thrives best in humid, tropical climates. Ginger has a long history that goes back more than 5000 years. It is a South East Asian native, but over the years, it has spread across several regions of the world, including Africa [14].

The countries like UK, the USA, and Saudi Arabia import the majority of the harvest. Nigeria is the greatest amount of ginger growing country (56.23% of the world's total area), followed by Bangladesh, India, China, and Indonesia, which have approximately 23.6, 4.7, and 3.4 per cent of the total area under ginger, respectively [5].

Ginger is used for various kinds of purposes in India, including pickles, food additives, confections, and traditional remedies for stomach aches. It is a powerful antioxidant and has been used to treat all kinds of nausea, including those brought on by motion sickness, pregnancy, surgery, and nausea following chemotherapy [17]. Traditional

and modern medications both employ ginger as a common raw ingredient [19]. This is due to the volatile oleoresin and volatile oil found in ginger [1].

Regarding the rising global demand for organic products, ginger producers that cultivate organically should expect to receive significant profits on their investments. Organic has gained more attention due to enhanced quality, better market demand, and environmental protection [7]. The ginger rhizomes and leaves from organic sources have a much higher capacity for phytochemicals and antioxidants than those from non-organic sources [15]. PPFM bacteria have been shown to fix nitrogen and can also produce cytokinins, auxins, and inducible systemic resistance enzymes including pectinase and cellulase [9]. Prior research has shown that spraying soil and leaves simultaneously with PPFM bacteria could considerably improve ginger plant growth characteristics [24].

Ginger can be grown well in coconut's partial shade without harming the performance of the main crop. The ginger crop thrives in somewhat shaded conditions in the coconut garden, has reasonable market demand, is easy to process, and has a good shelf life [12]. Hence, this study was conducted to find out the effect of organic amendments on the growth and yield of ginger in the coconut ecosystem.

2. MATERIALS AND METHODS

A field experiment was conducted at a farmer's field, Kailasapatti near Periyakulam in Tamil Nadu. The field is located at an altitude of 356 m above MSL (mean sea level) at a geographical location of 10° 5' 50" N latitude and 77° 32' 2" E longitude. Composite soil samples were collected initially from 15 to 30 cm depth before sowing and analyzed for physical and chemical properties. The experiment was laid out in a randomized block design with three replications. The soil was brought to fine tilth by deep ploughing four times. Weeds, stubbles, roots etc. were removed. At the time of the last ploughing, 2t Neem cake/ha was applied. After levelling, the raised beds were formed to accommodate the treatments. Plots were divided at 5 m length to accommodate 10 treatments per replication.

The organic manures were applied at nitrogen equivalent doses such as FYM @ 30 kg/plot, Vermicompost @ 15 kg/plot, Poultry manure @ 5 kg/plot, and Goat manure @ 5 kg/plot. 50% of the phosphorous was applied as basal. 150 g P were applied in the control plot. Healthy and disease-free rhizomes weighing about 15-20 grams each were selected and treated with *Pseudomonas fluorescens* for 20 minutes, shade dried for 30 minutes and used for sowing on 03.02.2023. The seed rhizome bits with one or two well-sprouted buds were planted at a distance of 30 x 30 cm in raised beds. The seed rhizomes were planted by the dibbling method. Manual weeding was done at 30, 60, 90, 120 and 150 days after planting. The microbial consortium was sprayed @ 1% conc. at monthly intervals to the respective treatments.

The performance of treatments was assessed based on growth characteristics like plant height, number of leaves per plant, leaf length, leaf width, number of tillers per clump and leaf area index at monthly intervals.

Leaf Area Index was calculated using the following formula.

$$\text{LAI} = \frac{\text{Total leaf area of the plant (cm}^2\text{)}}{\text{The ground area occupied by the plant (cm}^2\text{)}}$$

Treatment details:

T₁ -FYM (30t/ha)

T₂ -Vermicompost (5t/ha)

T₃ -Poultry manure (3t/ha)

T₄ -Goat manure (2t/ha)

T₅ - Microbial consortium (1%)

T₆ -T₁ + Microbial consortium (1%)

T₇ -T₂+ Microbial consortium (1%)

T₈ -T₃ + Microbial consortium (1%)

T₉ -T₄ + Microbial consortium (1%)

T₁₀ -Control (Recommended Dose of fertilizers-75:50:25 kg/ha)

3. RESULTS AND DISCUSSION

3.1 Soil properties

The physical and chemical properties of soil are given in Table 1.

3.2 Effect of organic amendments on growth characters of ginger

3.2.1 Plant height

Poultry manure combined with foliar spraying of microbial consortium (T₈) recorded the highest plant height (72.08 cm) followed by T₉ - goat manure + microbial consortium (71.86 cm). However, the lowest values of plant height were recorded for control (Table 2). Any method used to modify plant height would have an impact on the rhizome because plant height is a key yield parameter for ginger yield. Similar outcomes are also reported by (Egbuchua *et al.*, 2013), (Lepcha *et al.*, 2019) in ginger and (Anupam Pariari *et al.*, 2019) in mango ginger (Shadia *et al.*, 2021) in strawberry. This might be attributed to a microbial consortium which enhances plant growth by a variety of mechanisms, including the production of plant growth regulators like cytokinin and auxins [13], which regulate plant development and several physiological processes [11], and the generation and production of enzymes like urease or laminocyclopropane-1-carboxylate deaminase (ACCD), which modulates plant growth [8]. This may also occurred as a result of poultry manure that increases nitrogen uptake, which as a component of protein and protoplasm, strongly encourages the vegetative growth of the plant. By favourably influencing root growth and development, the improved particle density, reduced bulk density, expanded pore space, texture, and soil availability of nutrients were able to contribute indirectly to the increased plant height [10].

3.2.2 Number of leaves

The highest number of leaves was recorded for T₈ - poultry manure + microbial consortium (86.57) followed by T₆ - FYM + microbial consortium (83.03) and the lowest number of leaves were recorded for T₁₀ - control (Table 3). Egbuchua *et al.* (2013) in ginger and Anupam Pariari *et al.* (2019) in mango ginger also reported that the application of poultry manure produced the highest number of leaves. Vadivukkarasi and Bhai 2020 also revealed that spraying microbial consortium can increase the number of leaves. This may be happened due to the methyl bacteria producing phytohormones like auxin, which stimulates plant elongation and cell division and poultry manure contains easily accessible nutrients and hormonal activity. More leaves are produced and as a result, increasing the available space for the photosynthetic process. The effectiveness of nutrient absorption by plants is correlated with an increase in the number of leaves on a ginger plant. The more leaves there are, the more effectively the leaves absorb light, resulting in better growth [20].

3.2.3 Leaf size

Table 4 shows that the combination of organic nutrient sources T₈ - poultry manure + microbial consortium produced the largest leaf length (26.75 cm) followed by T₉ - goat manure + microbial consortium (25.83 cm) which was at par with T₆ - FYM + microbial consortium (25.22cm). The lowest leaf length was observed in control condition T₁₀ (22.24 cm). The largest leaf width was observed in T₈ - poultry manure + microbial consortium (3.92 cm) followed by T₉ - goat manure + microbial consortium (3.87 cm). The lowest leaf width was recorded in the control (Fig 1). Lepcha *et al.*, (2019) in ginger and Anupam Pariari *et al.* (2019) in mango ginger also recorded the largest leaf size with the application of poultry manure. Similar findings were reported by Vadivukkarasi and Bhai in 2020 that spraying microbial consortium can increase the leaf length and leaf width in ginger. This might be due to the adequate release and supply of nutrients from the manures which ultimately resulted in triggering the production of plant growth hormones, viz. Indole-3-acetic acid and other hormones [10] or due to the generation of phytohormones, nodulation, nitrogen fixation, and nutrient acquisition from PPFM which helps to encourage plant growth [23].

3.2.3 Number of tillers

The numbers of tillers per plant significantly increased with the application of T₈ - poultry manure + microbial consortium (9.48) followed by T₆ - FYM + microbial consortium (9.23) and T₉ - goat manure + microbial consortium (9.04). There was no significant difference among those three treatments. The lowest number of tillers was recorded in the control condition (Fig 2). This may be attributed to the action of auxin which aids in the establishment of a massive root system, consequently, cytokinins signal the shoot system to create more tillers [18]. As revealed by Lepcha *et al.* (2019), Egbuchua *et al.* (2013) and Singh *et al.* (2009) in ginger, Balkrishnamurty *et al.* (2009) and Singh (2013) in turmeric, the number of tillers, which is also a significant yield-contributing trait in ginger, affects the yield and mother rhizome.

Similar findings were revealed by Aswathy *et al.* (2020) in rice as the application of microbial consortium increases the number of tillers.

3.3 Physiological characters

The leaf area of the plant increased with the number of leaves, which likewise increased sunlight absorption. A plant's capacity to tolerate shade is essentially shown by the increase in leaf area. The way leaves get light has an impact on how quickly a plant grows. The energy required to carry out photosynthesis and raise the number of photosynthesis increases as more light reaches the leaves [20]. Fig 3 shows that the leaf area index was higher in the combination of poultry manure and microbial consortium (8.09) followed by goat manure + microbial consortium (7.34). The lowest leaf area was calculated in control. Shadia *et al.* (2021) in strawberries and Aswathy *et al.* (2020) in rice reported that microbial consortium can increase the leaf area index. Egbuchua *et al.* (2013) in ginger and Anupam Pariari *et al.* (2019) in mango ginger also recorded the largest leaf area with the application of poultry manure.

4. CONCLUSION

According to the findings of the study, it is concluded that the growth and yield characteristics of ginger rhizome were increased with the combined application of two organic nutrient sources such as poultry manure and microbial consortium. Among the various treatment combinations, T₈ gave the best response to all the growth characters and yield attributes of ginger under coconut-based cropping system.

Table 1. Properties of Experimental Soil

Sl. No.	Properties	Content
1.	Textural class	Sandy clay loam
2.	Soil pH	8.21
3.	EC (dSm ⁻¹)	0.31
4.	Available nitrogen (kg/ha)	163.09

5.	Available phosphorus (kg/ha)	27.18
6.	Available potassium (kg/ha)	289.11
7.	Organic carbon (%)	0.24

Table 2. Effect of various organic amendments on plant height (cm) of ginger at different growth stages

Treatments	30 DAP	60 DAP	90 DAP	120 DAP	150 DAP
T₁	26.82	29.71	60.14	61.28	64.89
T₂	26.01	28.99	59.52	60.41	62.6
T₃	27.95	30.34	60.88	61.87	65.49
T₄	24.54	29.16	58.23	59.62	61.16
T₅	23.36	27.98	55.27	58.45	59.23
T₆	25.42	31.23	61.63	66.34	69.54
T₇	25.76	30.64	61.08	64.79	68.77
T₈	27.64	33.97	64.36	69.52	72.08
T₉	26.52	31.72	62.47	67.95	71.86
T₁₀	24.63	29.82	52.25	54.18	56.41
Mean	25.86	30.35	59.58	62.44	65.20
SE(d)	0.67	0.78	1.54	1.61	1.67
CD (0.05)	1.41	1.64	3.24	3.38	3.52

Table 3. Effect of organic amendments on number of leaves per plant of ginger at different growth stages

Treatments	30 DAP	60 DAP	90 DAP	120 DAP	150 DAP
T₁	4.82	18.74	47.84	66.65	70.85
T₂	4.57	17.65	45.61	61.59	65.72
T₃	5.96	19.52	48.42	67.94	74.92
T₄	6.08	19.24	47.12	64.12	68.32
T₅	4.31	15.87	44.68	58.24	62.76
T₆	5.68	20.07	51.58	72.53	83.03
T₇	3.99	18.98	48.91	71.15	78.69
T₈	6.24	20.43	52.72	74.39	86.57
T₉	5.13	19.86	50.63	71.76	81.26
T₁₀	3.32	13.92	41.87	55.98	60.84
Mean	5.01	18.42	47.938	66.43	73.29
SE(d)	0.13	0.47	1.23	1.70	1.87
CD (0.05)	0.27	1.00	2.59	3.58	3.93

Table 4. Effect of organic amendments on leaf length (cm) of ginger at different growth stages

Treatments	30 DAP	60 DAP	90 DAP	120 DAP	150 DAP
T₁	9.54	13.85	18.92	22.15	23.46
T₂	9.02	13.31	16.71	20.42	23.25
T₃	10.46	14.29	19.22	22.98	24.37
T₄	10.28	14.01	18.41	21.68	23.91
T₅	8.72	12.64	15.75	20.03	22.89
T₆	10.09	15.47	20.15	23.86	25.22
T₇	9.39	14.78	19.63	22.74	24.64
T₈	10.63	16.46	20.89	24.37	26.75
T₉	9.81	15.96	20.57	23.39	25.83
T₁₀	8.34	11.55	15.19	19.84	22.24
Mean	9.628	14.232	18.544	22.146	24.25
SE(d)	0.24	0.36	0.47	0.56	0.62
CD (0.05)	0.52	0.76	0.99	1.19	1.31

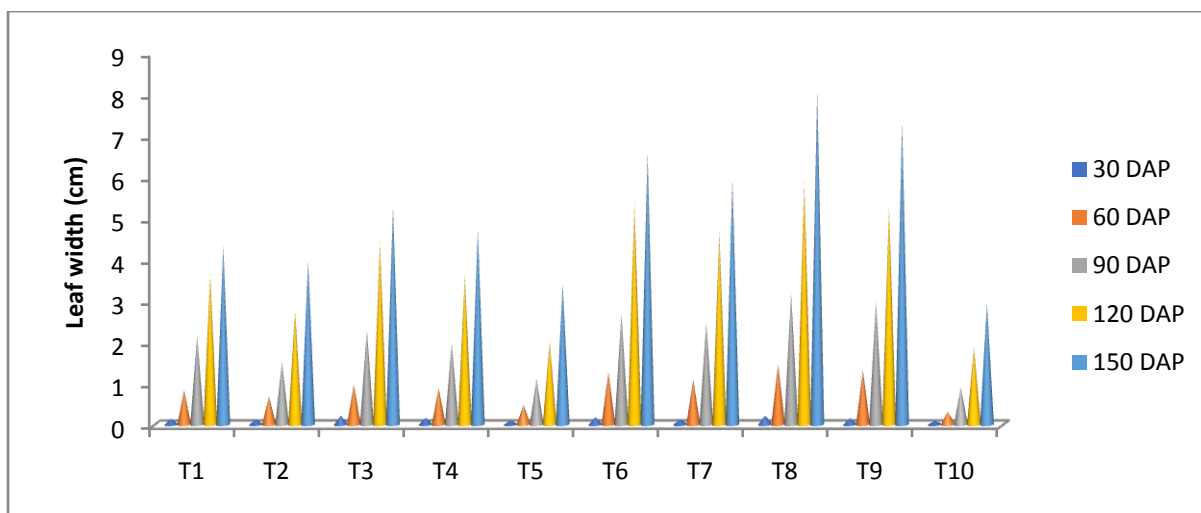


Fig. 1. Effect of organic amendments on leaf width of ginger at different growth stages

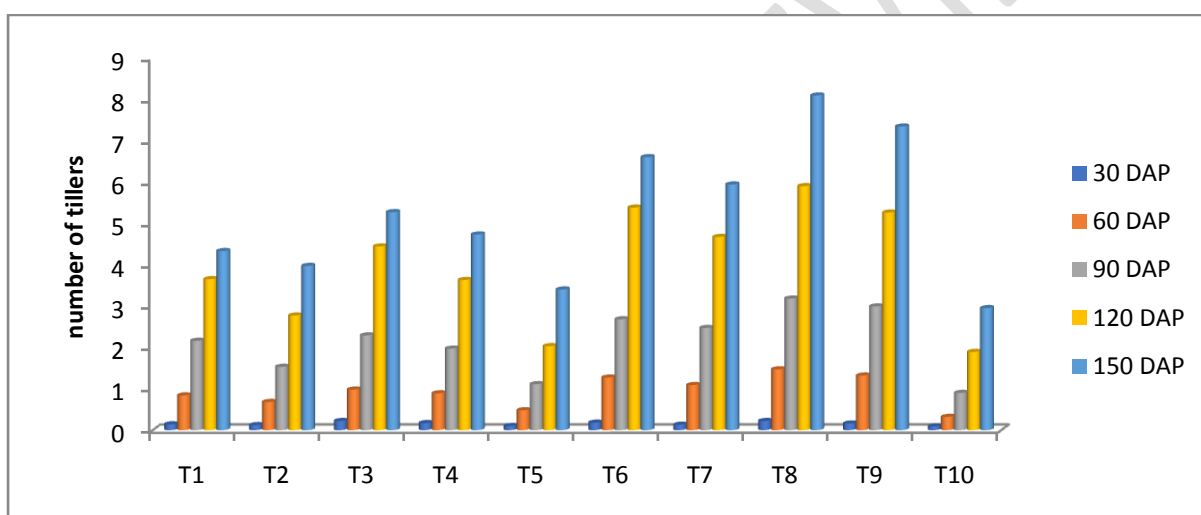


Fig. 2. Effect of organic amendments on number of tillers of ginger at different growth stages

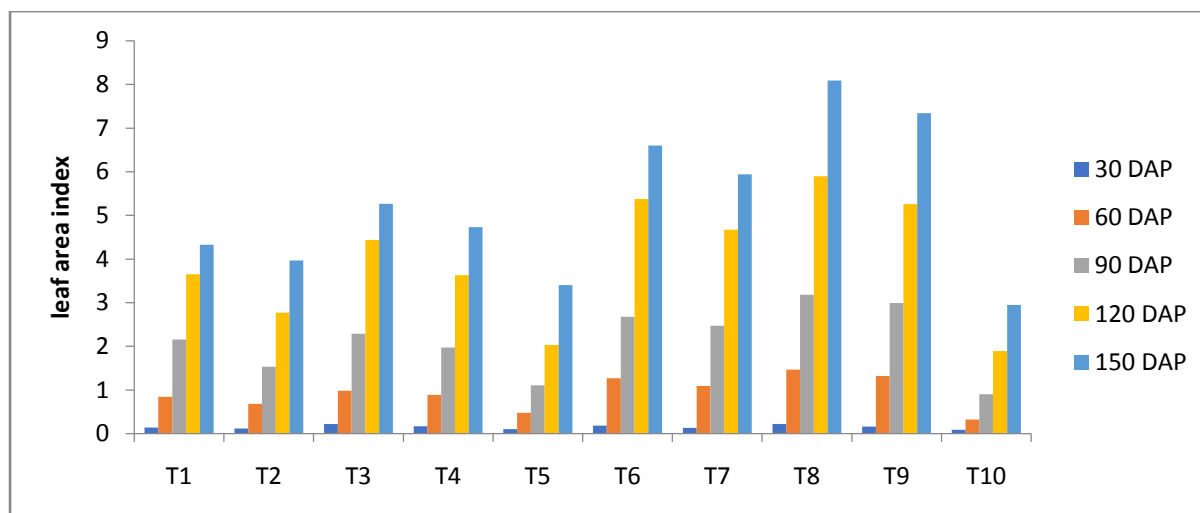


Fig. 3. Effect of organic amendments on leaf area index of ginger at different growth stages

REFERENCES

1. Aidin, N. Sahiri, and I. Madauna, "The Effect of Rhizome Types and The Composition of Planting Media on The Growth of Red Ginger (*Zingiber Officinale* Rosc.) (Pengaruh Jenis Rimpang dan Komposisi Media Tanam Terhadap Pertumbuhan Bibit Jahe Merah)," *J. Agrotekbis*, vol. 4, no. 4, pp. 394–402, 2016.
2. Aswathy, J.C., Pillai, P.S., John, J. and Meenakumari, K., 2020. Effect of pink pigmented facultative methylotrophs (PPFM) on growth and growth attributes of rice (*Oryza sativa* L.). *Int J Chem Stud*, 8(4), pp.3209-13.
3. Balakrishnamurty G, Kamal K R and Prabu T. 2009. Standardization of organic manures and bioregulators for organic production of turmeric (*Curcuma longa* L.), pp 123–30. *Production Technology for Organic Spices*.(Eds.)M Tamil Selvan, Homey Cheriyan, K. Manoj, Kumar and BabulalMeena).Director, Directorate of Arecanut and Spices Development, Calicut, Kerala, India.
4. Egbuchua, C.N. and Enujeke, E.C., 2013. Growth and yield responses of ginger (*Zingiber officinale*) to three sources of organic manures in a typical rainforest zone, Nigeria. *Journal of Horticulture and Forestry*, 5(7), pp.109-114.
5. Gohain, N., Atibudhi, H.N. and Kumar, A., 2020. A study on economics of ginger cultivation in Assam—a case study of Tinsukia district. *Plant Archives*, 20(2), pp.2522-2526.
6. Ismail, S.A. and Fafy, A.M., 2021. Influence of Pink Pigmented Facultative methylotrophic bacteria (PPFM) as a foliar spraying on the growth and productivity of strawberry (*Fragaria xananassa* Duch.). *bioRxiv*, pp.2021-12.
7. Jyotsna, N., Ghosh, M., Ghosh, D.C., Meitei, W.I. and Timsina, J., 2013. Effect of biofertilizer on growth, productivity, quality and economics of rainfed organic ginger

- (*Zingiber officinale* Rosc.) bhaisey cv. in north-eastern region of India. *J. Agro. Scie. and Tech*, 3, pp.83-98.
8. Kwak MJ, Jeong H, Madhaiyan M, Lee Y, Sa TM, Oh TK, et al. Genome information of *Methylobacterium oryzae*, a plant-probiotic methylotroph in the phyllosphere. *PLoS One*. 2014;9(9).
 9. Lee, H.S., Madhaiyan, M., Kim, C.W., Choi, S.J., Chung, K.Y. and Sa, T.M., 2006. Physiological enhancement of early growth of rice seedlings (*Oryza sativa* L.) by production of phytohormone of N₂-fixing methylotrophic isolates. *Biology and fertility of soils*, 42, pp.402-408.
 10. Lepcha, B.O.N.I.F.A.C.E., Avasthe, R.A.V.I.K.A.N.T., Singh, R.A.G.H.A.V.E.N.D.R.A., Singh, N.J. and Phukan, P.A.L.L.A.B.I., 2019. Effect of organic nutrient sources on productivity, profitability and quality of ginger (*Zingiber officinale*) in acid soils of Eastern Himalayas. *Indian J. Agric. Sci*, 89, pp.1103-1107.
 11. Madhaiyan M, Poonguzhali S, Sundaram SP, Sa T. A new insight into foliar applied methanol influencing phylloplane methylotrophic dynamics and growth promotion of cotton (*Gossypium hirsutum* L.) and sugarcane (*Saccharum officinarum* L.). *Environ Exp Bot*. 2006;57(1-2):168-76.
 12. Maheswarappa, H.P., Nanjappa, H.V. and Hegde, M.R., 2007. Influence of agronomic practices on growth, productivity and quality of galangal (*Kaempferia galanga* L.) grown as intercrop in coconut garden.
 13. Nadali I, Paknejad F, Moradi F, Vazan S, Tookalo M, Al-Ahmadi MJ, et al. Effects of methanol on sugar beet (*Beta vulgaris*). *Aust J Crop Sci*. 2010;4(6):398-401.
 14. Nair, K.P. and Nair, K.P., 2019. The agronomy and economy of ginger. *Turmeric (Curcuma longa L.) and Ginger (Zingiber officinale Rosc.)-World's Invaluable Medicinal Spices: The Agronomy and Economy of Turmeric and Ginger*, pp.245-315.
 15. Ozkur, M., Benlier, N., Takan, I., Vasileiou, C., Georgakilas, A.G., Pavlopoulou, A., Cetin, Z. and Saygili, E.I., 2022. Ginger for healthy ageing: A systematic review on current evidence of its antioxidant, anti-inflammatory, and anticancer properties. *Oxidative medicine and cellular longevity*, 2022.
 16. Pariari, A., Karthik, C.S. and Bhattacharya, S., 2019. Effect of organic manures on growth, yield and quality of mango ginger (*Curcuma amada* Roxb.) in the Gangetic Alluvial Plains of West Bengal. *Int J Curr Microbiol App Sci*, 8(11), pp.1030-1034.
 17. Prasad, S. and Tyagi, A. K. (2015). Ginger and its constituents: Role in prevention and treatment of gastrointestinal cancer. *Gastroenterology Research and Practice*. <https://doi.org/10.1155/2015/142979>
 18. Raghavendra, J. and Santhosh, G.P., 2019. Effect of efficient strains of pink pigmented facultative methylotrophs on plant growth parameters of direct seeded rice. *Int. J. Curr. Microbiol. Appl. Sci*, 8(7), pp.1473-1487.

19. S. Soeparjono, "Pengaruh Komposisi Media Organik Terhadap Pertumbuhan, Hasil dan Kualitas Rimpang Tiga Varietas Jahe (*Zingiber officinale* Rosc.)," Prosiding Seminar Ilmiah Perhorti 2013, pp. 613– 618, 2013.
20. Samanhudi, A.Y., Pujiasmanto, B. and Rahayu, M., 2014. Effect of organic manure and arbuscular mycorrhizal fungi on growth and yield of young ginger (*Zingiber officinale* Rosc.). *IOSR Journal of Agriculture and veterinary Science*, 7(5), pp.1–5.
21. Singh S P, Chaudhary R and Mishra A K. 2009. Effect of different combination of organic manures on growth and yield of ginger (*Zingiber officinale* Rose). *Journal of Eco-friendly Agriculture* 4: 22–4.
22. Singh S P. 2013. Response of different combination of organic manures for production of turmeric (*Curcuma longa* L.). *Journal of Eco-friendly Agriculture* 8: 35–8.
23. Soundarajan, S., Marimuthu, R., Arunkumar, K., Dhanaphal, K., Venkatesan, T. and Sivakumar, G., 2022. Pink pigmented facultative methylotrophic bacteria (PPFMs) as microbial farmers in small cardamom plantation. *Pharma Innovafion J*, 11(4), pp.607–610.
24. Vadivukkarasi, P. and Bhai, R.S., 2020. Phyllosphere-associated Methylobacterium: a potential biostimulant for ginger (*Zingiber officinale* Rosc.) cultivation. *Archives of microbiology*, 202, pp.369-375.