

**Studies on the effect of biostimulants on quality of cut flower of Chrysanthemum
(*Dendranthemagrandiflora*) cv. DenjigarWhite.**

Abstract:

The experiment entitled- “Effect of biostimulants on quality of cut flower of Chrysanthemum (*Dendranthemagrandiflora*) cv. DenjigarWhite” was conducted during the Rabiseason of the year 2020-2021 at Ammapuram(V), Thorrur(M), Mahabubabaddistrict. Among the treatments, the treatment T₂(*Ascophyllumnodosum*@ 5 ml L⁻¹) recorded maximum flower diameter (7.48 cm), individual flower weight (6.55 g), stem length (72.10cm), vase life (21.60 days) and shelf life (5.84 days), gross returns (₹ 76,93,200), net returns (₹ 59,82,204) and best benefit cost ratio (3.50). However, control recorded minimum in all the parameters.

Key words: Bio stimulants, chrysanthemum, Denjigar White, *Ascophyllumnodosum*.

Introduction:

Flowers and flowering plants have been a fascinating part of our life. Chrysanthemum (*Dendranthemagrandiflora*) is a most beautiful and oldest flowering plant, commercially grown in different parts of the world. Chrysanthemum occupies a prominent place in ornamental horticulture and is one of the commercially exploited flower crop, belongs to the family 'Asteraceae' and referred as “Queen of the East”, “Autumn Queen”, and “Guldaudi” having diploid chromosome number 2n=18.

The word “Chrysanthemum” comes from two Greek words, *Chrysos* - golden and *anthos*-flower which means golden flower. It is native to the Northern hemisphere and is widely distributed in Europe and Asia. However, the origin of chrysanthemum is China (Carter, 1990).

After green revolution, the indiscriminate use of chemical fertilizers has led to negative impact on environment. To mitigate this, biostimulants have been emerged as a supplement to the mineral fertilizers and hold a promise to improve the yield as well as quality of the crop under protected conditions (Rawat and Vishal, 2002). Biostimulants are the materials other than the fertilizers that promote the plant growth when applied in minute quantities and are also referred as ‘metabolic enhancers’ (Zhang and Schmidt, 1997). Keeping in view, the need and importance of biostimulants, the

present investigation was conducted with an objective to study the effect of biostimulants on growth, yield and quality of cut flower of Chrysanthemum (*Dendranthemagrandidflora*) cv. Denjigar White.

Materials and methods:

The present investigation entitled “Effect of biostimulants on quality of cut flower of chrysanthemum (*Dendranthemagrandidflora*), cv. Denjigar White” was carried out during the *Rabi* season of the year 2020-2021 at the season of the year 2020-2021 at Ammapuram (V), Thorrur (M), Mahabubabad district. Healthy and rooted terminal cuttings were planted on the raised beds at a spacing of 15 cm x 15 cm under polyhouse. The design adopted was Randomized Block Design with nine treatments and replicated thrice. Treatments included T₁- *Ascophyllum nodosum* @ 2.5 ml L⁻¹, T₂-*Ascophyllum nodosum* @ 5 ml L⁻¹, T₃- Rhodophyte extract @ 0.2 g L⁻¹, T₄-Rhodophyte extract @ 0.4 g L⁻¹, T₅-Potassium humate @ 1.5 g L⁻¹, T₆-Potassium humate @ 3 g L⁻¹, T₇-Fulvic acid 10% + Seaweed 8% + Spirulina 6% @ 2 g L⁻¹, T₈- Fulvic acid 10% + Seaweed 8% + Spirulina 6% @ 4 g L⁻¹, T₉-Control (Water spray). These biostimulants were sprayed on the foliage at 3 intervals *i.e.* @ 30, 45 and 60 days after transplanting (DAT) and the observations recorded were flower diameter, individual flower weight, stem length, vase life, shelf life and economics were recorded and the data were statistically analysed.

Schedule of spray : 30, 45 and 60 days after transplanting (DAT).

Results and Discussion:

The results of the experiments have been presented in Tables 1-3.

The effect of biostimulants on quality of cut flower of chrysanthemum (*Dendranthema grandiflora*), cv. Denjigar White and the results of the experiment were presented in Table 1 to 3.

Flower diameter (cm)

With respect to the quality parameters in chrysanthemum, T₂ treatment (*Ascophyllum nodosum* @ 5 ml L⁻¹) recorded maximum flower diameter (7.48 cm). Whereas the minimum flower diameter was recorded in T₉ - Control (Water spray) (4.48 cm). The enlargement in size of the flower might be due to production of more food which was diverted to flowering area. There by due to presence of more food reserves in the flowering parts results in maximum diameter of flowers. Similar findings were reported by Hegdeet *al.* (2016) in chrysanthemum, Tartilet *al.* (2016) in pot marigold, Kahkashanet *al.* (2017) in tuberose, Hegdeet *al.* (2020) in orchids and Lingwalet *al.* (2017) in strawberry.

Individual flower weight (g)

Among all the treatments, T₂ treatment (*Ascophyllum nodosum* @ 5 ml L⁻¹) recorded highest individual flower weight (6.55 g). While T₉ - Control (Water spray) recorded lowest individual flower weight (5.09 g). The increase in the individual flower weight might be due to translocation of food reserves from vegetative parts to reproductive parts. Due to the increase of food and carbohydrate reserves in flower portion, the size of the flower increases thereby, the weight of the flower increases. Similar findings were reported by Hegde *et al.* (2016) in chrysanthemum, Kahkashan *et al.* (2017) in tuberose, Selvakumari and Venkatesan (2017) in tomato, Hegde *et al.* (2020) in orchids and Lingwalet *et al.* (2017) in strawberry.

Stem length (cm)

Irrespective of the treatments, T₂ treatment (*Ascophyllum nodosum* @ 5 ml L⁻¹) recorded maximum stem length (72.10 cm). Whereas the minimum stem length was recorded in T₉ - Control (Water spray) (62.87 cm). The highest stem length was observed in sea weed extract sprayed plants as they are the precursors of auxin, cytokinin and micronutrients. The increase in the stem length might be due to the fact that, gibberellins promoted the efficacy of plants in terms of photosynthetic activity, uptake of nutrients and their translocation, better partitioning of assimilates into reproductive parts. Similar findings were reported by Aziz *et al.* (2012) in *Amaranthus tricolor* plants, Hegde *et al.* (2016) in chrysanthemum, Tartilet *et al.* (2016) in pot marigold, Kahkashan *et al.* (2017) in tuberose, Majeed Khadim Al-Hamzawi (2019) in Chinese carnation and Gazania splendens and Hegde *et al.* (2020) in orchids.

Vase life (days)

The data recorded on the vase life depicts that treatment T₂ (*Ascophyllum nodosum* @ 5 ml L⁻¹) recorded highest vase life (21.60 days). Whereas T₉ - Control (Water spray) recorded lowest vase life (14.82 days). The increase in the vase life of chrysanthemum may be due to the entry of sea weed extract into the plant, might have mediated the respiration by acting as a hydrogen acceptor, thus altering the carbohydrate metabolism of plants promoting the accumulation of sugar (Cacco and Dell Agnola, 1984) and also sea weed extract contain cytokinin and auxin that might have increased the antioxidant levels and resistance to senescence leading to enhanced longevity of stem. Similar findings were reported by Hegde *et al.* (2016) in chrysanthemum, Tartilet *et al.* (2016) in pot marigold, Kahkashan *et al.* (2017) in tuberose and Hegde *et al.* (2020) in orchids.

Shelf life (days)

Among all the treatments, T₂ treatment (*Ascophyllumnodosum* @ 5 ml L⁻¹) recorded maximum shelf life (5.84 days). While the minimum shelf life was recorded in T₉ - Control (Water spray) (3.52 days). The increase of shelf life in *Ascophyllumnodosum* treated plants over control may be due to that sea weed extract biostimulant induced photosynthesis that might have led to recombination of nutrients in flower that is used for remaining long days. Similar results were reported by Hegde *et al.* (2016) in chrysanthemum, Povolny (1976) in tomato.

Economic analysis

Among the biostimulant treatments, T₂ treatment (*Ascophyllumnodosum* @ 5 ml L⁻¹) recorded highest gross returns (₹ 76,93,200), net returns (₹ 59,82,204) and benefit cost ratio (3.50). Whereas T₉ - Control (Water spray) recorded lowest gross returns (₹ 50,13,200), net returns (₹ 33,92,200) and benefit cost ratio (2.09). It is evident from the data that, maximum gross return was recorded in T₂ treatment (*Ascophyllumnodosum*@ 5 ml L⁻¹) which might be due to higher number of flower stalks hectare⁻¹ as compared to others.

Table 1. Effect of biostimulants on flower diameter and individual flower weight of chrysanthemum cv. Denjigar White.

Treatments / Biostimulants (T)	Flower diameter (cm)	Individual flower weight (g)
T ₁ - <i>Ascophyllumnodosum</i> @ 2.5 ml L ⁻¹	6.46 ^a	6.08 ^a
T ₂ - <i>Ascophyllumnodosum</i> @ 5 ml L ⁻¹	7.48 ^a	6.55 ^a
T ₃ - Rhodophyte extract @ 0.2 g L ⁻¹	4.83 ^b	5.14 ^c
T ₄ - Rhodophyte extract @ 0.4 g L ⁻¹	5.02 ^b	5.19 ^c
T ₅ - Potassium humate @ 1.5 g L ⁻¹	5.12 ^b	5.23 ^c
T ₆ - Potassium humate @ 3 g L ⁻¹	5.64 ^b	5.74 ^b
T ₇ - Fulvic acid 10% + Seaweed 8% + Spirulina 6% @ 2 g L ⁻¹	5.82 ^b	5.87 ^b
T ₈ - Fulvic acid 10% + Seaweed 8% + Spirulina 6% @ 4 g L ⁻¹	6.63 ^a	6.39 ^a
T ₉ - Control (Water spray)	4.48 ^b	5.09 ^c

S.E. m±	0.53	0.20
CD@ 5%	1.62	0.59

Table 2. Effect of biostimulants on stem length, vase life and shelf life of chrysanthemum cv. Denjigar White.

Treatments / Biostimulants (T)	Stem length (cm)	Vase life (days)	Shelf life (days)
T₁- <i>Ascophyllum nodosum</i> @ 2.5 ml L⁻¹	69.23 ^a	19.59 ^b	4.89 ^b
T₂- <i>Ascophyllum nodosum</i> @ 5 ml L⁻¹	72.10 ^a	21.60 ^a	5.84 ^a
T₃- Rhodophyte extract @ 0.2 g L⁻¹	64.66 ^c	15.56 ^f	3.88 ^c
T₄- Rhodophyte extract @ 0.4 g L⁻¹	65.49 ^c	16.48 ^e	4.16 ^c
T₅- Potassium humate @ 1.5 g L⁻¹	66.82 ^b	17.56 ^d	4.40 ^b
T₆- Potassium humate @ 3 g L⁻¹	67.90 ^b	18.53 ^c	4.56 ^b
T₇- Fulvic acid 10% + Seaweed 8% + Spirulina 6% @ 2 g L⁻¹	68.51 ^b	19.27 ^b	4.75 ^b
T₈- Fulvic acid 10% + Seaweed 8% Spirulina 6% @ 4 g L⁻¹	70.39 ^a	20.84 ^a	5.31 ^a
T₉- Control (Water spray)	62.87 ^c	14.82 ^f	3.52 ^c
S.E. m±	1.06	0.39	0.26
CD@ 5%	3.19	1.17	0.78

Table 3. Effect of biostimulants on economics of chrysanthemum cv. Denjigar White.

Treatments / Biostimulants (T)	Concentration	Number of flower stalks ha⁻¹ (Yield)	Total cost of cultivation ha⁻¹ (₹)	Gross returns (₹)	Net returns (₹)	B:C ratio
T₁- <i>Ascophyllum nodosum</i>	2.5 ml L ⁻¹	17,98,500	16,65,993	71,94,160	55,28,167	3.32
T₂- <i>Ascophyllum nodosum</i>	5 ml L ⁻¹	19,23,300	17,10,996	76,93,200	59,82,204	3.50
T₃- Rhodophyte extract	0.2 g L ⁻¹	13,75,950	16,52,635	55,03,800	38,51,165	2.33
T₄- Rhodophyte extract	0.4 g L ⁻¹	14,84,750	16,84,270	59,39,000	42,54,730	2.53

T₅ -Potassium humate	1.5 g L ⁻¹	15,52,600	16,59,500	62,10,400	45,50,900	2.74
T₆ -Potassium humate	3 g L ⁻¹	16,37,200	16,98,000	65,48,800	48,50,800	2.86
T₇ -Fulvic acid 10% + Seaweed 8% + Spirulina 6%	2 g L ⁻¹	17,35,000	16,66,329	69,40,000	52,73,671	3.16
T₈ -Fulvic acid 10% + Seaweed 8% + Spirulina 6%	4 g L ⁻¹	18,73,350	17,11,658	74,93,400	57,81,742	3.38
T₉ -Control (Water spray)	-	12,53,300	16,21,000	50,13,200	33,92,200	2.09

Conclusion

On the basis of the results obtained in the present investigation it is concluded that *Ascophyllum nodosum* @ 5 ml L⁻¹ proved for improving the quality of cut chrysanthemum, the present study also confirmed that the use of bio-stimulant is an eco-friendly technique to enhance crop production. Thus, it may be recommended that the chrysanthemum plants can be sprayed to get maximum *Ascophyllum nodosum* @ 5 ml L⁻¹ vase life and flower yield which may ensure us to get a maximum net returns.

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