

*Original Research Article*

**RESPONSE OF RICE (*Oryza sativa* L.) VARIETIES TO SALICYLIC ACID,  
POTASSIUM SILICATE AND TAMARIND EXTRACT IN SALINE SOILS OF KANO  
RIVER IRRIGATION SCHEME**

**Abstract**

The experiment was carried out in the Eastern and Western sectors of Kano River Irrigation Scheme, Kano state latitude 11° 45' N and 12° 05' N and longitude 8° 45' E and 9° 05' E in the Sudan Savanna Agro-ecology of Nigeria, during the dry season of 2020 and 2021. The experiment was therefore conceived with the aim of introducing improved salt tolerant varieties to farmers in addition to enhancing their tolerance using synthetic and natural plant growth regulators. The treatments consisted of four rice varieties made up of 2 salt tolerant accessions {Arica 1 and Arica 2, 1 blast resistant variety (Gawal R1) and a variety (FARO 44) as a check} these were factorially combined with foliar application of salicylic acid (0.2g in 1L of water), potassium silicate (16 gL<sup>-1</sup> of K<sub>2</sub>SiO<sub>3</sub>), tamarind extract 8% (320ml of extract in 1 liter of water) and a zero control (0). The treatments were laid out in a Randomized Complete Block Design (RCBD) which consisted of 16 treatments replicated six times with a farmer constituting a replication. Results have shown that plant height, leaf area index (LAI), photosynthetically active radiation (PAR), chlorophyll content and the yield were all significantly affected by varietal differences except total number of tillers per plant in 2020. However, in 2021 LAI, PAR and chlorophyll content were not significantly affected by differences due to varieties but the remaining characters differed significantly. Exogenous factors had no effect on all the characters in 2020 but significantly influenced the chlorophyll content, total number of tillers and the yield in 2021. Arica 1 and Faro 44 gave the highest yield. Among the exogenous factors Tamarind extract had significantly heavier paddy yield than the control but was similar to other exogenous factors in terms of effectiveness. Tamarind extract could therefore be suggested to farmers when growing rice under saline or sodic condition in the irrigation scheme due to its lower cost and availability.

Keywords: Growth regulators, Irrigation, Rice, Salinity.

## **Introduction**

Nigeria is the Africa's leading consumer of rice, one of the largest producers of rice in the continent and simultaneously one of the largest rice importers in the world (FAO, 2019). Nigeria has a total of 5.4 million tonnes of paddy rice in 2017 (FAO, 2017). Rice is an important food security crop, it is an essential cash crop for it is mainly small-scale producers who commonly sell 80 per cent of total production and consume only 20 per cent. Rice generates more income for Nigerian farmers than any other cash crop in the country (FAO, 2019).

Soil salinity poses a major threat to rice crop productivity. Salinity is found to induce both biochemical and physiological changes causing growth inhibition and yield loss (Fukuda *et al.*, 2007; Kang *et al.*, 2007 Colom and Vazzana, 2003; Chang and Sung, 2003).

Soil salinity is a potential problem in some parts of Kano River Irrigation Project (KRIP). Saline soils exhibit structural problems such as crusting, hard setting and waterlogging associated with poor infiltration (Naidu and Rangasamy, 1993). Visual evidence of salinity, such as crusting and waterlogging have been found on many farmers' fields in Kano River Irrigation Scheme (KRIS). About 73% of farmers in these areas have problems associated with waterlogging and salinity/sodicity (Jibrin *et al.*, 2008).

Salinity causes several injuries in plants such as tissue burning, yield reduction, and finally plant death (Zekri and Parsons, 1992). Salinity leads to the abandoning of land and also reduces production. Farmers, in this case, are restricted to grow only salt tolerant crops. High salt concentrations severely affect rice plants' normal physiology, especially during early stages of growth, and as such, there is need to test some salt tolerant rice varieties.

Most rice farmers in Nigeria are smallholders, applying a low-input strategy to agriculture, with minimum input requirements and low output (USAID 2009; IFAD

2009). Rice productivity in Nigeria is among the lowest within neighbouring countries, with average yields of 1.51 tone/ha (FAO 2009).

Despite the importance of rice to the nation, many lands have been abandoned for rice cultivation which have been proven to be due to soil salinity, leading to importation of rice from India, Thailand, United States of America, etc. putting farmers out of work (Osagie, 2014). To ameliorate the effect of soil salinity, foliar application of growth promoters or regulators/ antitranspirants like; salicylic acid, potassium silicate and tamarind extract and the use of some salt tolerant varieties could be used.

Salicylic acid is an endogenous growth regulator (Sakhabutdinova *et al.*, 2003) and belongs to a group of phenol compounds. It participates in the regulation of physiological processes (Hayat *et al.*, 2010) and also provides protection against biotic and abiotic stress such as salinity (Kaya *et al.*, 2009). It also has a role in germination under stress condition, although its definite role and the underlying physiological mechanisms have not been fully elucidated (Asadi *et al.*, 2013, Alonso-Ramirez *et al.*, 2009, Rajjou *et al.*, 2006 and Borsani *et al.*, 2001).

Potassium silicate is a source of highly soluble potassium and silicon. It is used in agricultural production systems primarily as a silica amendment and has the added benefit of supplying small amounts of potassium (USDA, 2003), and also promotes greater tolerance to salinity (Epstein, 2001).

Tamarind tree plant is available almost everywhere in dry areas of Northern Nigeria, its leaf extract can be utilized as a growth promoter to reduce the risks involved in the use of synthetic growth promoters and also promote organic farming.

The abandoned land due to salinity could be put into use if tolerant varieties are screened. This will increase the level of productivity; reduce the amount of rice importation; increase farmers' income; help to attain food security and increase the economic growth, and development of the nation.

The information obtained will be useful to extension agents and policy makers in respect to rice production.

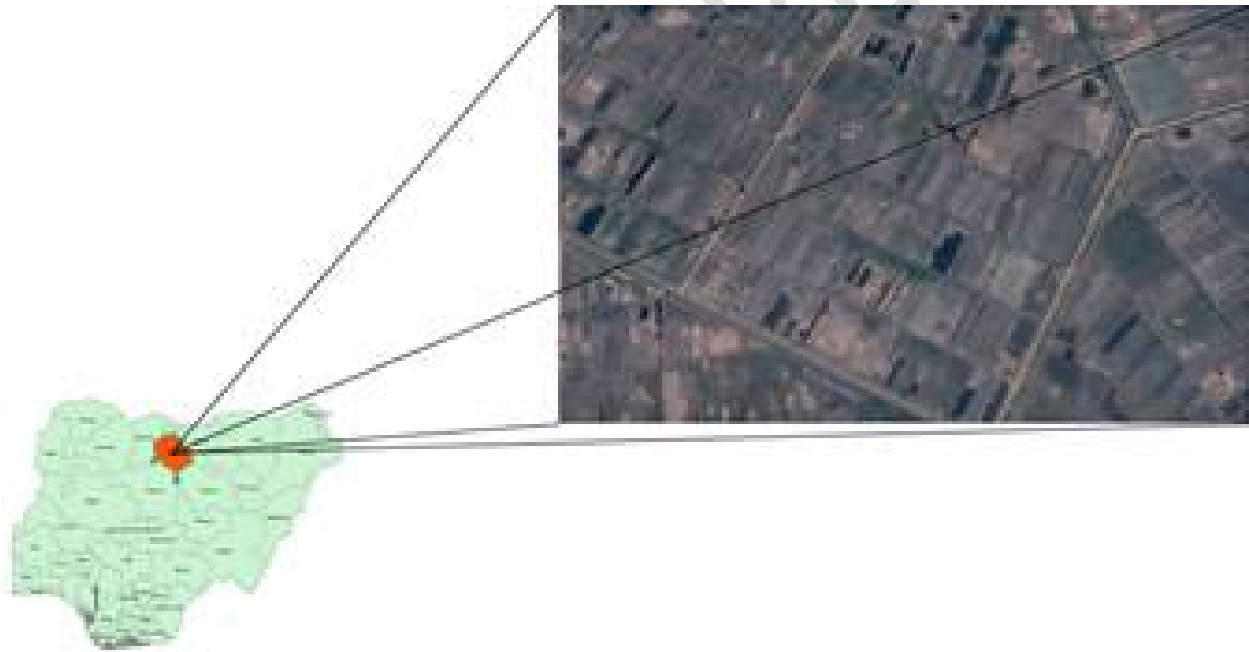
## Objectives

1. To evaluate the growth and yield of rice varieties as affected by Salicylic acid under the saline soils of KRIS.
2. To evaluate the growth and yield of rice varieties as affected by Potassium silicate under the saline soils of KRIS.
3. To evaluate the growth and yield of rice varieties as affected by Tamarind extract under the saline soils of KRIS

## Materials and Methods

### Experimental site

The experiment was carried out in the Eastern and Western sectors of Kano River Irrigation Scheme, Kano state, between latitudes  $11^{\circ} 45' N$  and  $12^{\circ} 05' N$  and longitude  $8^{\circ} 45' E$  and  $9^{\circ} 05' E$  in the Sudan Savanna Agro-ecology of Nigeria, during the dry season of 2020.



**Fig 1. Experimental site**

### Treatments and experimental design

The treatments consisted of four rice varieties made up of 2 salt tolerant accessions (Arica 1 and Arica 2), 1 blast resistant variety (Gawal R1) and a variety (FARO 44) as a check; these were factorially combined with foliar application of salicylic acid (0.2g in 1L of water), potassium silicate ( $16 \text{ gL}^{-1}$  of  $\text{K}_2\text{SiO}_3$ ), tamarind extract 8% (320ml of extract in 1 liter of water) and a zero

control (0). The treatments were laid out in a Randomized Complete Block Design (RCBD) which consisted of 16 treatments replicated ten times.

### **Plot size**

The gross plot size was 3m x 3m (9m<sup>2</sup>), consisting of 225 stands of rice, while the net plot was 3m x 1m (3m<sup>2</sup>) consisting of 75 stands of rice. A gap of 0.5m was left between plots.

### **Seed source and description of varieties**

The rice varieties were obtained from Africa Rice, these are: Arica 1, Arica 2, Gawal-R1 and Faro 44. The Arica 1 and Arica 2 are both accessions.

- i. Faro 44: this variety was released in 1992 with an old variety name as SIPI 692033. It is planted in irrigated swamp/shallow swamp ecologies. It is a long grain type with 100-115 days of grain growth duration. It has a yield potential of 4.0-6.0 t ha<sup>-1</sup> and also resistant to blast.
- ii. Gawal R1: this variety was released in 2017 with an old variety name as Chaotan. It is planted in irrigated swamp/shallow swamp ecologies. It is a long grain type with 90-100 days of grain growth duration. It has a yield potential of 10.4 t ha<sup>-1</sup> and also resistant to blast.
- iii. Arica 1 WAB 2094-WAC 2-TGR 2-B: It is grown in rainfed lowland areas and high yielding over NERICA L19 and BW 348-1. Suitable for Mali and Burkina Faso countries.
- iv. Arica 2 WAB-2056-2-FKR 2-5-TGR 1-B: It is grown in rainfed lowland areas and high yielding over BW348-1, NERICA L19 and WITA 12. Suitable for Nigeria and Mali countries.

ARICA stands for Advance Rice Varieties for Africa. Both Arica 1 and Arica 2 have particular advantages of being iron toxicity tolerant, cold tolerant, salt tolerant and good grain quality (high milling recovery, low chalky and short cooking time). Research showed that Arica 1 and Arica 2 respectively showed 20-44% and 50-111% higher yield than NERICA-L 19 which is in wide use.

### **Preparation of exogenous factors**

#### **Tamarind leaf extract**

About 5kg of fresh leaves and shoots were crushed with 2 liters of water using mortar and pestle, which were then filtered out as a stock solution after 2 days; the liquid extract of 320ml (8%) was then diluted with 1 liter of water.

#### **Salicylic acid**

Salicylic acid comes in solid crystals. 0.2 grams of salicylic acid was crushed to a fine powder, and then 0.5 liter of water was poured to help dissolve it for a day. And on application the other 0.5 liter of water was applied to balance the 1 liter of water.

## Potassium Silicate

Potassium silicate comes in liquid form. 16g of it was weighed which was then diluted with 1 litre of water.

### **Cultural practices**

#### **Nursery practices**

The soil was flooded, puddled and slightly raised where a significant amount of rice was sown.

#### **Land preparation**

The land was cleared, harrowed and made into basins of 3m x 3m, which were then demarcated by 0.5m between plots and 1m between replications so as to provide easy movement.

#### **Transplanting**

The seedlings raised in the nursery were transplanted 5 weeks after sowing and spaced 20cm x 20cm inter and intra row spacing.

#### **Fertilizer application**

Fertilizer was applied in two split doses, basal application of NPK 15:15:15 was used to supply 60 kg each of N, P<sub>2</sub>O<sub>5</sub>, and K<sub>2</sub>O at 3 weeks after transplanting while the balance of 60 kg nitrogen was supplied using urea (46% N) as top dressing at 6 weeks after transplanting.

#### **Weed control**

The experimental plots were kept weed free using the manual method. This was done at 3<sup>rd</sup> and 6<sup>th</sup> weeks after transplanting.

#### **Pest and Disease control**

During the course of the study, stem borer was identified. The chemical used was Lambda-cyhalothrin insecticide at the rate of 1.5g a.i. /ha in 100L of water. It acts as a contact and systemic poison.

#### **Harvesting**

Rice was harvested at physiological maturity stage when the grains were hard and turned yellow or brown in color. The harvesting was done by cutting the rice at the base and threshed by smashing the panicles on a drum and later winnowed and weighed. The plot yield was converted in to yield per hectare by multiplying paddy from the plot by a factor of 3333.33 obtained by dividing 10, 000 m<sup>2</sup> by 3 m<sup>2</sup>.

### **Data collection**

#### **Plant height**

The heights of three sampled plants were measured using meter rule from the ground level to the tip of the flag leaf at 9<sup>th</sup> weeks after transplanting. Mean values for each plot were recorded.

### **Leaf Area Index**

The leaf area index of three sampled plants were obtained from the sampled plants by measuring, using AccuPAR model LP-80 PAR/LAI Ceptometer (Decagon Device, Inc. Pullman, USA) at 8<sup>th</sup> weeks after transplanting.

### **Photosynthetically Active Radiation**

Intercepted photosynthetically active radiation was measured using AccuPAR model LP-80 PAR/LAI Ceptometer (Decagon Device, Inc. Pullman, USA) at 8<sup>th</sup> weeks after transplanting.

### **Total number of Tillers Plant<sup>-1</sup>**

The total number of tillers were counted and recorded from three sampled plants at 9<sup>th</sup> weeks after transplanting. Mean values for each plot were recorded.

### **Total Chlorophyll Content**

The chlorophyll content was measured at 8<sup>th</sup> week after transplanting using SPAD (SPAD meter – 502, Konica - Minotta).

### **Meteorological data**

Meteorological data were collected from Centre for Dryland Agriculture, Geographic Information System (GIS) Laboratory, Bayero University, Kano.

### **Soil sampling and analysis**

Soils of the experimental sites were randomly collected using auger at a depth of 0-20cm before land preparation. The samples were bulked and sampled, then subjected to routine analysis according to standard procedure (Black 1965).

### **Data analysis**

Data collected were subjected to analysis of variance (ANOVA) as described by Snedecor and Cochran (1967) using Genstat 17<sup>th</sup> edition (VSNI, 2015). The treatment means were separated using Student Newman-Keuls test (SNK) at 5% probability level.

## **Results**

### **Physical and chemical properties of soil of the experimental sites**

Results of the soil analysis from the experimental sites (Table 1) shows that the soils did not differ much in textural class and other chemical and physical properties. Soil at Garun mallam was sandy loam with particle distribution of sand (648.0 g kg<sup>-1</sup>), silt (218.0 g kg<sup>-1</sup>) and clay (134.0 g kg<sup>-1</sup>). For

chemical composition of the soil, nitrogen, phosphorus and organic carbon contents were low. The pH of the area places the soil in a moderately alkaline state. For exchangeable cations; Ca, Mg, and K were medium while Na was low.

Soil at Kura was loam with particle distribution of sand (522.3 g kg<sup>-1</sup>), silt (334.6 g kg<sup>-1</sup>) and clay (143.1 g kg<sup>-1</sup>). The nitrogen, phosphorus and organic carbon contents of the soil were low. The pH of the area places the soil in a neutral state. For exchangeable cations; Ca was low, Mg and K were medium while Na was low. The salt contents of both areas were above normal range but were not high enough to define the soils as saline, sodic or saline-sodic, but literature showed that plants may be starved of nutrients (Sonon *et al.*, 2015).

Table 1: Physical and chemical properties of soil at Garun mallam and Kura during 2020 dry season.

Properties	Garun mallam	Kura
<u>Physical %</u>		
Sand	64.80	52.23
Silt	21.80	33.46
Clay	13.40	14.31
Textural Class	Sandy loam	Loam
<u>Chemical</u>		
pH in water	8.16	6.84
EC(dS/m)	0.21	0.22
Organic Carbon (g kg <sup>-1</sup> )	0.47	0.45
Total Nitrogen (g kg <sup>-1</sup> )	0.06	0.04
Available Phosphorus (mg kg <sup>-1</sup> )	5.52	4.29
<u>Exchangeable Bases (cmol<sup>+</sup> kg<sup>-1</sup>)</u>		

Ca <sup>2+</sup>	2.30	1.67
Mg <sup>2+</sup>	0.79	0.79
K <sup>+</sup>	0.17	0.16
ECEC	3.39	2.75
ESP (%)	2.51	2.93

---

Analyzed at Center for Dryland Agriculture laboratory.

UNDER PEER REVIEW

## **Effects of exogenous factors on plant height, LAI, PAR, Chlorophyll content, total number of tillers and yield of rice varieties**

Table 2 shows plant height, leaf area index (LAI), photosynthetically active radiation (PAR), chlorophyll content, total number of tillers and yield of rice varieties as affected by potassium silicate, salicylic acid, tamarind extract and the control in 2020 and 2021 dry season at Kano River Irrigation Project (KRIP). Gawal R1 had significantly taller plants than Arica 1, Arica 2 and Faro 44 which were at par. The exogenous factors had no effect on plant height in 2020. However, in 2021, Arica 1 was significantly taller than Arica 2 and Faro 44 which were statistically similar but significantly taller than Gawal R1. The exogenous factors had no effect on plant height in the same year.

The differences in LAI among the varieties were significant in 2020 only, where Arica 1, Arica 2 and Faro 44 were at par but significantly higher than Gawal R1. The exogenous factors had no significant effect on LAI in both years. Gawal R 1 had significantly more PAR than all the other varieties which were observed to trap the same PAR but the effect of exogenous factors was not significant in 2020. The data for 2021 was not available because we could not get the equipment to measure it at the time. Chlorophyll contents differed significantly among varieties in 2020 only. Arica 1 had the highest chlorophyll content but was at par with Arica 2 and significantly higher than Faro 44 which had significantly higher chlorophyll than Gawal R1. The effect of the exogenous factors was also significant in 2021 only where Potassium silicate and Salicylic acid had statistically similar chlorophyll content but significantly higher than Tamarind extract and the control which were also at par.

The total number of tillers was not affected by varietal differences in 2020, but in 2021, Arica 2 and Faro 44 were statistically similar but had significantly more number of tillers than Gawal R1 which also had significantly more number of tillers  $t\ ha^{-1}$  than Arica 1. The effect of the exogenous factors was not significant in 2020, however, in 2021, Tamarind extract, Salicylic acid and Potassium silicate were at par but Tamarind extract and Salicylic acid gave more number of tillers than the control.

The paddy yield was significantly influenced by differences in varieties of rice in both years. In 2020, Arica 1 gave the highest yield but was at par with Arica 2 and Faro 44, however, Arica 1 and Faro 44 had significantly heavier grains than Gawal R 1. In 2021, Faro 44 had significantly heavier grains than Arica 2 which was significantly heavier than Arica 1 which was also significantly heavier than Gawal R1. The effect of exogenous factors was not significant in 2020, however, in 2021, Tamarind extract had significantly heavier paddy yield than the control but was at par with all the other exogenous factors.

**Table 2: Effects of exogenous factors on plant height, LAI, PAR, Chlorophyll content, total number of tillers and yield of rice varieties in 2020 and 2021 at KRIP**

Treatments	Plant height (m)		LAI		PAR (nm)		Chlorophyll content		Total number of tillers		Yield (kg ha <sup>-1</sup> )	
	2020	2021	2020	2021	2020	2021	2020	2021	2020	2021	2020	2021
<b>Rice Var. RV</b>												
Arica 1	0.82 <sub>b</sub>	0.81 <sub>a</sub>	3.5 <sub>a</sub>	2.0	162.6 <sub>b</sub>		43.4 <sub>a</sub>	21.3	18.4	20.5 <sub>c</sub>	7249.1 <sub>a</sub>	7942 <sub>c</sub>
Arica 2	0.82 <sub>b</sub>	0.77 <sub>b</sub>	3.3 <sub>a</sub>	2.0	171.8 <sub>b</sub>	NA	42.6 <sub>ab</sub>	19.9	19.1	26.5 <sub>a</sub>	6603.5 <sub>ab</sub>	9701 <sub>b</sub>
Faro 44						NA						
	0.82 <sup>b</sup>	0.73 <sub>b</sub>	3.4 <sub>a</sub>	2.3	156.8 <sub>b</sub>		42.0 <sub>b</sub>	18.9	20.2	26.5 <sub>a</sub>	7161.5 <sub>a</sub>	9743 <sub>a</sub>
Gawal R 1	0.90 <sub>a</sub>	0.71 <sub>c</sub>	2.2 <sub>b</sub>	2.0	366.3 <sub>a</sub>	NA	39.1 <sub>c</sub>	22.7	19.0	23.4 <sub>b</sub>	4290.7 <sub>b</sub>	7861 <sub>d</sub>
SE ±	0.015	0.014	0.12	0.15	12.41	NA	0.45	1.43	1.07	0.91	284.0	2007
<b>Exogenous Factors (EF)</b>												
Control	0.82	0.75	2.9	2.0	211.0	NA	42.2	37.2 <sub>b</sub>	19.2	22.5 <sub>b</sub>	5976.9	7701 <sub>b</sub>
Potassium silicate	0.86	0.73	3.2	2.1	209.3	NA	42.1	43.5 <sub>a</sub>	20.8	23.4 <sub>ab</sub>	6521.4	8567 <sub>ab</sub>
Salicylic acid	0.84	0.77	3.2	1.9	221.5	NA	41.8	44.8 <sub>a</sub>	18.8	25.4 <sub>a</sub>	6618.1	8903 <sub>ab</sub>
Tamarind Extract	0.84	0.77	3.1	2.3	215.7	NA	41.1	38.6 <sub>b</sub>	18.0	25.9 <sub>a</sub>	6188.6	10,076 <sub>a</sub>
SE ±	0.015	0.014	0.12	0.15	12.4	NA	0.45	1.43	1.07	0.91	284.0	2007
<b>Interaction</b>												
RV X EF	NS	NS	NS	NS	NS	NA	NS	NS	NS	NS	NS	NS

Means followed by the same letter(s) in column and row are not significantly different at 5% probability level using Student Newman-Keul Test

Table 3: Meteorological data covering the experimental period at Kura LGA Kano in 2020 and 2021.

Month	Temperature °C				Rainfall (mm)		Relative Humidity (%)		Wind speed ms <sup>-1</sup>	
	Minimum		Maximum		2020	2021	2020	2021	2020	2021
	2020	2021	2020	2021						
<b>March</b>	19.75	19.9	37.27	36.02	0.00	0.37	22.26	26.85	2.64	2.60
<b>April</b>	23.13	21.63	37.12	39.10	120.55	1.11	47.66	25.88	2.71	2.53
<b>May</b>	23.68	23.66	36.18	39.36	185.99	12.71	58.98	41.42	2.28	2.29
<b>June</b>	23.28	25.58	34.67	39.99	158.50	197.40	64.87	39.11	2.10	2.36

Table 4: Meteorological data covering the experimental period at Garun mallam LGA Kano in 2020 and 2021.

Month	Temperature °C				Rainfall (mm)		Relative Humidity (%)		Wind speed ms <sup>-1</sup>	
	Minimum		Maximum		2020	2021	2020	2021	2020	2021
	2020	2021	2020	2021						
<b>March</b>	19.85	19.14	36.28	34.88	0.00	3.72	27.54	35.95	3.47	3.39
<b>April</b>	22.68	21.37	34.56	37.39	203.77	6.0	57.97	35.94	3.49	3.21
<b>May</b>	23.01	23.17	33.48	36.89	91.89	65.41	68.24	51.87	2.74	2.85
<b>June</b>	22.59	24.87	30.03	37.17	212.65	376.2	72.10	46.69	2.60	3.08

Source: Centre for Dryland Agriculture, BUK, GIS Laboratory.

## **DISCUSSION**

From the results of the rice experiment the yield was observed to be high (Table 2) this may be as a result of the seed quality, since the seed was obtained directly from Africa Rice and from the varietal description it was reported that Gawal R1 could give a yield up to the level ( $10 \text{ t ha}^{-1}$ ) we got from in this experiment. Other reasons may be due to higher moisture content as a result of inadequate drying period of the paddy because the rice was harvested during rainy season (Table 3 and 4). Good management of the trial which was jointly managed by the researchers and farmers might have contributed to the high yield that was obtained more over this was similarly reported by other research teams under TRIMING project namely Africa Rice and the one led by Institute for Agricultural Research, Ahmadu Bello University, Zaria who worked on Sustainable Rice Intensification at Bakolori Irrigation Scheme Nigeria.

### **Growth and Yield Response of Rice Varieties**

The differences among the varieties were significant in 2020 and 2021. Gawal R1 had significantly taller plants than Arica 1, Arica 2 and Faro 44 which were at par. However, in 2021, Arica 1 was significantly taller than Arica 2 and Faro 44 which were statistically similar but significantly taller than Gawal R1. This could be as a result of inherent genetic ability of the varieties. There were significant differences in leaf area index and photosynthetically active radiation due to varietal differences. At Garun mallam Arica 1 recorded the highest LAI in 2020, even though it was statistically similar to Faro 44 and Arica 2. This was expected because resistant varieties grow vigorously, healthy, very active and also able to withstand external factors. Gawal-R1 was statistically higher in PAR in 2020. It is a well-known fact that taller varieties are more likely to trap more solar radiation than shorter varieties which is closely related to its inherent ability. Pal *et al.* (2004) reported a reduction in rate of photosynthesis of a salinity tolerant rice cultivar to be associated with decreased Rubisco activity due to the accumulation of ions in the chloroplast.

Numbers of tillers  $\text{plant}^{-1}$  was significant in 2021 with Faro 44 and Arica 1 producing more tillers which could be attributed to their genetic make-up. With regards to chlorophyll content, it is considered as an index to measure leaf injury under salt stress (James *et al.*, 2002). A significant effect was observed statistically at both locations, even though Arica 1 recorded the highest chlorophyll content.

The paddy yield was significantly influenced by differences in varieties of rice in both years. In 2020, Arica 1 gave the highest yield but was at par with Arica 2 and Faro 44, however, Arica 1 and Faro

44 had significantly heavier grains than Gawal R 1. In 2021, Faro 44 had significantly heavier grains than Arica 2 which was significantly heavier than Arica 1 which was also significantly heavier than Gawal R1. The high yield observed in Arica 1 might be due to its high LAI and high chlorophyll content that might have facilitated better light interception leading to higher photosynthesis, more dry matter production and grain filling hence higher yield. However, in 2021 Faro 44 might have had higher yield due to its high LAI and better tillering ability that ensured better light interception and more grain number and weight for higher yield. Faro 44 being the equivalence of SIPI a local variety used by farmers may be genetically more pure than the farmers SIPI which may have suffered genetic erosion and adulteration over the years. Farmers should therefore be encouraged to purchase and use Faro 44 from reputable sources instead of the seeds they saved.

### **Effect of Plant Growth Regulators on Growth and Yield Characters of Rice**

Application of plant growth regulators did not significantly affect the growth parameters of rice at both locations. This could be as a result of the treatments being applied once and at later vegetative growth stage prior to booting stage. The treatments should be applied at growth, booting and panicle initiation stages, but due to the COVID-19 pandemic in 2020, the treatments were applied only once. This agrees with the findings of Frasetya *et al.* (2019) who reported that the role of Si element in vegetative phase do not affect rice growth especially plant height, and in contrast to the findings of Singh *et al.* (2015) who observed that foliar spray of SA at 20 ppm on hybrid rice had a significant effect on plant height. A study by Khoshbakht and Asgharei (2014) also observed a significant increase in plant height and leaf area when 1.00mM was applied to citrus crop. Bekheta and Talaat (2009) observed that the spray of SA at 15 mg/L significantly increased plant height in mung bean. However, in 2021 total number of tillers were also significantly increased by application of SA and Tamarind extract when compared to the control this is in agreement with Hussein *et al.* (2007) who reported that foliar spray of SA to the foliage of wheat crop enhanced productivity due to an improvement in all growth characteristics. Sairam and Tyagi (2004) reported that there was a decrease in growth at higher sodium concentrations due to a decrease in the uptake of  $K^+$  and  $Ca^{2+}$ , this confirmed what we observed in our experiment where  $Ca^+$  and  $K^+$  concentrations were very low. Sardoei *et al.* (2014) reported that the exogenous spray of SA and  $ZnSO_4$  levels had significant effect on number of tillers. Increase in shoot growth in rice by exogenous spray of SA was reported by Meguro and Sato (2014). Ahmad *et al.* (2013) reported that Maximum number of productive tillers was found in 0.05% silicon aqueous solution application while minimum in 0.025% silicon aqueous solution application. A similar report by Li *et al.* (2002), Rodrigues *et al.* (2003) and Mobasser *et al.* (2008) stated that applied silicon enhanced the number of productive tillers and total

number of tillers in a quadrant. Ahmad *et al.* (2013) also observed a significant increase in spikes per panicle when 1.00% silicon aqueous solution was applied to rice plant.

Coronado *et al.* (1998) reported that aqueous solutions of SA as a spray to shoots of soybean significantly increased the growth of shoots and roots in either greenhouse or field conditions. An increase in growth parameters of salt affected plants in response to SA might be related to the protective role of SA on membranes that might increase the tolerance of plants to salt stress (Aftab *et al.* 2010). It is well known that the promoting effect of SA on leaf area is attributed to its important roles on activating cell division and the biosynthesis of organic foods (Khoshbakht and Asgharei 2014). In addition, Raskin (1992) mentioned that enhancing effect of SA on the availability and movement of nutrients could result in stimulating different nutrients in the leaves. Foliar application of SA increased the leaf area of sugarcane (Zhou *et al.* 1999). The exogenous application of SA helps in preventing the lowering of indole-3-acetic acid (IAA) and cytokinin levels in salinity stressed wheat plants resulting in the betterment of cell division in root apical meristem, thereby increasing growth and productivity of plants (Shakirova *et al.* 2003). Exogenous application of SA results in accumulation of abscisic acid (ABA) which contributes to pre-adaptation of seedlings to salinity stress as ABA induces the synthesis of a wide range of anti-stress proteins, thereby providing protection to plants (Shakirova *et al.* 2003).

No significant difference was observed in relation to electrolyte leakage, proline content and chlorophyll content except at Kura where potassium silicate had the highest chlorophyll content. This result is in line with the findings of Esmaeili (2016) who reported that silicon increased chlorophyll content, relative water content and visual quality under non saline water and low concentrations of salinity in both short and long term exposures of Kentucky bluegrass. Dabo *et al.* (2018) also reported an increase in chlorophyll content when aqueous leaf extract of tamarind was applied to maize plant which was a result of growth promotion effects of chemicals like phenolics, terpenoids, carbohydrate and amino acids present in tamarind leaves which increases growth at low concentrations. Parvez *et al.* (2003) reported that plant extract of some tree crops can influence crop growth. A contradictory report concerning electrolyte leakage goes to a strawberry plant being treated with SA which showed far less electrolyte leakage than control plants following salt stress (Yildirim *et al.*, 2008). Data also exist which showed that SA causes increases in the activities of anti-oxidant enzymes which, in turn, protect plants against the generation of Reactive Oxygen Species (ROS) and membrane injury, or may result in the synthesis of other substances which have a protective effect on plants growing under salt stress (Xu *et al.*, 2008). Proline on the other hand is one of the important components of the adaptation of plants to salinity; pretreatment with SA will contribute to the accumulation of this amino acid under stress through maintaining an enhanced level of ABA in seedlings (Kuznetsov and Shevyakova 1999).

Silicon is an important micronutrient for healthy and competitive growth of all cereals including rice (Brunings *et al.*, 2009). Role of silicon in plant health and growth has been investigated in silicon accumulating crops and it seemed to be effective (Jinab *et al.*, 2008). Research evidences proved that adequate uptake of silicon (Si) can increase the tolerance of agronomic crops especially rice to both biotic and abiotic stresses (Ma and Takahashi 2002). Silicon uptake by plants reduces the susceptibility to chewing insects such as stem borer; it might be by rendering plant tissue less digestible or by greatly damaging the mandibles of feeding insects (Massey and Hartley 2006). Silicon deficiency in plants makes them more susceptible to insect feeding, fungal diseases, germs attack and abiotic stresses that adversely affect crop yield and quality. Low silicon uptake has been proved to increase the susceptibility of rice to diseases such as rice blast, leaf blight of rice, brown spot, stem rot and grain discoloration (Rodrigues *et al.*, 2001; Massey and Hartley., 2006).

Many scientists working on role of silicon in plant growth have concluded that reduced amount of silicon in plant develops necrosis, disturbance in leaf photosynthetic efficiency, growth retardation and reduction of grain yield in cereals (Shashidhar *et al.*, 2008). Although silicon has not been considered important for vegetative growth but it aids the plant in healthy development under stresses in different grasses especially in rice.

Application of growth regulators did not influence paddy yield in 2020. This was expected because some of the plants were eaten by stray animals at Kura in few instances, thereby retarding the growth of the plant. It could also be because of the environmental stress being experienced by the plant. This contradicts the results of Mobasser *et al.* (2008) and Malidareh *et al.* (2011), however, Mauad *et al.* (2003), reported that silicon application does not affect paddy yield. Effects of silicon on yield are related to the deposition of the element under the leaf epidermis which results in a physical mechanism of defense, reduces lodging, increases photosynthesis capacity and decreases transpiration losses (Korndörfer *et al.*, 2004). However, in 2021 the paddy yield was significantly increased by application of Tamarind extract when compared to the control but was at par with all the other exogenous factors. The result was in agreement with the findings of Dabo *et al.* (2018) who observed an increase in grain yield and harvest index of maize when aqueous leaf extract of tamarind was applied, which could be attributed to the presence of pectin in the extract; an important cell wall polysaccharide that allows primary cell wall extension and plant growth (El-Siddig *et al.*, 2006). Singh *et al.* (2015) also observed in contrast to this that seed yield increased significantly when (40 ppm) of SA treatment was applied over control. Mohammad and Tarpley (2011) reported that SA in treated plants increased grain yield by 13 % over control in rice. The concentration of the exogenous factors used in our research may not have reached the levels that could cause appreciable increase in most of the characters measured or the

environment had different climatic and soil variables than that of the authors who reported appreciable increase in these characters.

## CONCLUSION

The paddy yield was significantly influenced by differences in varieties of rice and Arica 1 and Faro 44 gave the highest yield. Among the exogenous factors Tamarind extract had significantly heavier paddy yield than the control but similar to other exogenous factors in terms of effectiveness. Tamarind extract could be suggested to farmers when growing rice under saline or sodic condition in KRIS. Arica 1 and Faro 44 rice varieties are suggested to be grown by farmers under saline or sodic condition in the same location.

## References

1. Abdel-Monaim, M.F., Abo-Elyousr, K.A.M., and Morsy, K.M. (2011). Effectiveness of Plant Extracts on Suppression of Damping-off and Wilt Diseases of Lupine (*Lupinus termis* Forsik). *Crop Protection*, 30(2): 185-191.
2. Aftab, T., Khan M.M.A., and Idrees M. (2010). Salicylic Acid Acts as Potent Enhancer of Growth, Photosynthesis and Artemisinin Production in *Artemisia annua* L. *Crop Science. Biotechnology*, 13: 183-188.
3. Agarie, S., Hanaoka, N., and Ueno, O. (1998). Effects of Silicon on Tolerance to Water Deficit and Heat Stress in Rice Plants (*Oryza sativa* L.), Monitored by Electrolyte Leakage. *Plant Production. Science*, 1: 96–103.
4. Ahmad, A., Afzal, M., Ahmad, A., and Tahir, M. (2013). Effect of Foliar Application of Silicon on Yield and Quality of Rice (*Oryza Sativa* L), *Cercetari Agronomice in Moldova* 46:21-28.
5. Ahmad, R., Zaheer, S.H., and Ismail, S. (1992). Role of Silicon in Salt Tolerance of Wheat (*Triticum aestivum* L.). *Plant Sci.*, 85: 43-50.
6. Aldesuquy, H. S., Mankarios, A. T., and Awad, H.A. (1998). Effect of some Antitranspirants on Growth, Metabolism and Productivity of Saline Treated Wheat Plants. Induction of Stomatal Closure, Inhibition of Transpiration and Improvement of Leaf Turgidity. *Acta Bot. Hungarica*, 41: 1-10.
7. All Chap (compile) - Farmer. (n.d.). Retrieved from <http://www.farmer.gov.in/imagedefault/pestanddiseasescrops/rice.pdf>.

8. Alonso-Ramirez, A., Rodriguez, D., Reyes, D., Jimenez, J.A., Nicolas, G., Lopez-Climent, M., Gomez-Cadenas, A., and Nicolas, C. (2009). Evidence for a Role of Gibberellins in Salicylic Acid-modulated Early Plant Responses to Abiotic Stress in Arabidopsis Seeds. *Plant Physiol*, 150, (30), 1335-1344.
9. Analysis Of Incentives and Disincentives For Rice In Nigeria. (n.d.). Retrieved from <http://www.fao.org/3/a-at581e.pdf>.
10. Andrade Pinto, J.M., Souza, E.A., and Oliveira, D.F. (2010). Use of Plant Extracts in the Control of Common Bean Anthracnose. *Crop Protection*, 29(8): 838-842.
11. Arberg, B. (1981). Plant growth regulators, Monosubstituted benzoic acid. *Swed. Agric. Res.* 11: 93-105.
12. Arfan, M., Athar, H.R., and Ashraf, M. (2007). Does Exogenous Application of Salicylic Acid through the Rooting Medium Modulate Growth and Photosynthetic Capacity in two Differently Adapted Spring Wheat Cultivars under Salt Stress. *Plant Physiol.*, 6(4): 685-694.
13. Asadi, M., Heidari, M. A., Kazemi, M., and Filinejad, A. R. (2013). Salicylic Acid Induced Changes in some Physiological Parameters in Chickpea (*Cicer arietinum* L.) under salt stress. *J Agric Technol*, 9, (2), 311-316.
14. Asadujjaman, M., Mishuk, A.U., Hossain, M.A., and Karmakar, U.K. (2004). Medicinal potential of *Passiflora foetida* L. Plant Extracts: Biological and Pharmacological Activities. *Journal of Integrative Medicine*, 12 (2): 121-126.
15. [Ashraf](#), M., [Akram](#), N. A., [Arteca](#), R. N., and [Foolad](#) M. R. (2010). The Physiological, Biochemical and Molecular Roles of Brassinosteroids and Salicylic Acid in Plant Processes and Salt Tolerance. *Critical reviews in Plant sciences*, 29: 162-190.
16. Bajaj, S., and Mohanty, A. (2005). Recent Advances in Rice Biotechnology towards Genetically Superior Transgenic Rice. *Plant Biotechnol J*, 3, 275-307.

17. Bandurska, H., and Stroinski. (2005). The Effect of Salicylic Acid on Barley Response to Water Deficit. *Acta Physiol. Plant.*, 27: 379-386.
18. Barkosky, R. R., and Einhellig, F. A. (1993). Effects of Salicylic Acid on Plant–Water Relationships. *J. Chem. Ecol.* 19: 237-247.
19. Bates, L. S. (1973). Rapid Determination of Free Proline for Water Stress Studies. *Plant Soil*, 39, 205-207.
20. Bayou Farms and Industries Limited, Rice Value Chain Development Plan, Kaduna Rice Industry Supply Chain Development Programme, submitted to MSME Nigeria, April 2009.
21. Bekheta, M. K. and Imam, M. T. (2009). Physiological response of mung bean “Vignaradiata” plants to some bioregulators. *Applied Botany and Food Quality*, 83: 76-84.
22. Black, C. A. (1965). *Methods of Soil Analysis II, Chemical and Microbiological Properties*, Madison Wisconsin. American Society of Agronomy, 341-350.
23. Borsani, O., Valpuesta, V., and Botella, M. A. (2001). Evidence for a Role of Salicylic Acid in the Oxidative Damage Generated by NaCl and Osmotic Stress in Arabidopsis Seedlings. *Plant Physiol*, 126 (3), 1024-1030.
24. Brunings, A.M., Datnoff, L.E., Ma, J.F., Mitani, N., Nagamura, Y., Rathinosabapathi, B., and Kirst, M. (2009). Differential Gene Expression of Rice in Response to Silicon and Rice Blast Fungus *Magnaporthe oryzae*. *Ann. Appl. Biol.*, 155:161-170.
25. Castillo, E., Phuc, T. G., Abdelbaghi, M. A., and Kazuyuki, I. (2007). Response to Salinity in Rice: Comparative Effects of Osmotic and Ionic Stress. *Plant Prod. Sci.*, 10 (2), 159-170.
26. Chang, C. S., and Sung, J. M. (2004). Nutrient Uptake and Yield Responses of Peanuts and Rice to Lime and Fused Magnesium Phosphate in Acid Soil. *Field Crops Res*, 89, 319-325.

27. Chen, W., Yao, X., and Cai, K. (2011). Silicon Alleviates Drought Stress of Rice Plants by Improving Plant Water Status, Photosynthesis and Mineral Nutrient Absorption. *Chen Biological trace element research*, 142: 67–76.
28. Colom, M. R., and Vazzana, C. (2003). Photosynthesis and PS II Functionally of Draught Resistant and Drought Sensitive Creeping Love Grass Plants. *Environ Exp Bot*, 49, 135-144.
29. Coronado, M.A.G., Lopez C.T., and Saavedra A.L. (1998). Effects of Salicylic Acid on the Growth of Roots and Shoots in Soybean. *Plant Physiol. Biochem*, 8: 563-565.
30. Dabo, Z.G., Garba, I.I., and Adnan, A.A. (2018). Response of Early Maturing Maize (*Zea mays* L.) to Foliar Application of Tamarind (*Tamarindus indica* L.) Leaf Extracts under Low Nitrogen in the Dryland Ecology. *Savannah Journal of Agriculture*, 13(1): 46-58 6.
31. Diallo, B.O., Joly, H.I., McKey, D., Hosaert-McKey, M., and Chevallier, M.H. (2007). Genetic Diversity of *Tamarindus indica* Populations. *African Journal of Biotechnology*. 6 (7).
32. Econometric Analysis of The Effect Of Rice Production And ... (n.d.). Retrieved from <http://directresearchpublisher.org/wp-content/uploads/2015/12/Wudil-et-al.pdf>.
33. El-khawaga, H.A. (2018). Effect of Silica on Physiological and Ultrastructure Characters in Barley (*Hordeum vulgare* L.) Plant under Salt Stress. *Al-Azhar Bulletin of Science*, 29(9): 1-17.
34. El-Siddig, E., Gunasena, H.P.M., Prasad, B.A., Pushpa kumara D.K.N.G., Ramana, K.V.R., Vijayanand, P., and Williams, J.T. (2006). Tamarind (*Tamarindus indica* L.). Southampton Centre for underutilized crops, England. ISBN 0854328599.

35. Epstein, E. (1994). The Anomaly of Silicon in Plant Biology. Proceedings of the National Academy of Sciences 91: 11-17.
36. Epstein, E. (2001). Silicon in Plants: Facts vs. Concepts. Silicon in Agriculture. Amsterdam, Holand: Elsevier, 1-15.
37. Esmaeili, S., and Salehi, H. (2016). Kentucky Bluegrass (*Poa pratensis* L.) Silicon-treated Turfgrass Tolerance to Short- and Long-term Salinity Condition. Adv. Hort. Sci., 30(2): 87-94.
38. Falah, A. (2010). The Effects of Salinity at Different Growth Stage on Rice. Proceeding of 11<sup>th</sup> national congress open agronomy.
39. FAO (2004). FAO Land and Plant Nutrition Management Service.
40. FAO, (2017). FAOSTAT Database, 2017. FAO, Paris [www.fao.org/economic/RRM](http://www.fao.org/economic/RRM).
41. FAO, (2019). Nigeria at a Glance | Fao In Nigeria | Food And ... (n.d.). Retrieved from <http://www.fao.org/nigeria/fao-in-nigeria/nigeria-at-a-glance/en/>
42. Fariduddin, Q., Hayat, S., and Ahmad, A. (2003). Salicylic Acid Influences Net Photosynthetic Rate, Carboxylation Efficiency, Nitrate Reductase Activity and Seed Yield in Brassica juncea. Photosynthetica, 41: 281-284.
43. Frasetya, B., Harisman, K., Sudrajat, D., and Subandi, M. (2019). Utilization of Rice Husk Silicate Extract to Improve the Productivity of Paddy Ciherang cultivar. Bulgarian Journal of Agricultural Science, 25(3): 499-505.
44. Fukuda, T., Saltu, A., Wasaki, J., Shinana, T., and Osaki, M. (2007). Metabolic Alterations Proposed by Proteome in Rice Roots Grown under a Low P and High Al Concentrations under Low pH. Plant Science, 172, 1157-1165.

45. Ghosh, B., Ali, M. N., and Saikat, G. (2016). Response of Rice under Salinity Stress: A Review Update. *J Res Rice* 4, 167. doi:10.4172/2375-4338.1000167.
46. Harkamal, W., Clyde, W., Linghe, Z., Abdelbagi, M. I., and Pascal, C. (2007). Genome Wide Transcriptional Analysis of Salinity Stressed Japonica and Indica Rice Genotypes during Panicle Initiation Stage. *Plant Mol Biol*, 63, 609-623.
47. Hattori, T., Sonobe, K., Inanaga, S. (2008). Effects of Silicon on Photosynthesis of Young Cucumber Seedlings under Osmotic Stress. *Plant Nutr* 31:1046–1058.
48. Hayat, Q., Hayat, S., Irfan, M., and Ahmad, A. (2010). Effect of Exogenous Salicylic Acid under Changing Environment: A review *Environ Exp Bot*, 68 (1), 14-25.
49. Horváth, E., Szalai, G., Janda, T. (2007). Induction of Abiotic Stress Tolerance by Salicylic Acid Signaling. *Plant Growth Regul*, 26:290–300.
50. Hussein, M. M., Balbaa, L. K., and Gaballah, M. S. (2007). Salicylic acid and salinity effects on growth of maize plants, *Res. J. Agric. Biol. Sci.* 3(4): 321-328.
51. IFAD, (2009). Country Programme Evaluation, Federal Republic of Nigeria.
52. Improvement of Salt Tolerance in Rice (*Oryza sativa* L.) By .(n.d.). Retrieved from [http://www.cropl.com/hoque\\_10\\_1\\_2016\\_50\\_56.pdf](http://www.cropl.com/hoque_10_1_2016_50_56.pdf).
53. Ismail, B.S., and Chong, T.V. (2002). Effects of Aqueous Extracts and Decomposition of *Mikania micrantha* H.B.K. Debris on Selected Agronomic crops. *Weed Biology and Management*, 2(1): 31- 38.
54. James, R.A., Rivelli, A.R., Munns, R., and Von-Caemmerer, S. (2002). Factors Affecting CO<sub>2</sub> Assimilation, Leaf Injury and Growth in Salt Stressed Durum Wheat. *Funct. Plant Bio.*, 29: 1393-1403.
55. Jayakannan, M., Bose, J., Babourina, O., Shabala, S., Massart, A., Poschenrieder, C., and Rengel, Z. (2015). NPR1-dependent Salicylic Acid Signalling Pathway is Pivotal for Enhanced Salt and Oxidative Stress Tolerance in Arabidopsis. *Exp Bot.* doi:[10.1093/jxb/eru528](https://doi.org/10.1093/jxb/eru528).

56. Jayakannan, M., Bose, J., Babourina, O., Shabala, S., Massart, A., Poschenrieder, C., and Rengel, Z. (2015). NPR1-dependent Salicylic Acid Signalling Pathway is Pivotal for Enhanced Salt and Oxidative Stress Tolerance in Arabidopsis. *Exp Bot.* doi:[10.1093/jxb/eru528](https://doi.org/10.1093/jxb/eru528).
57. Jbilou, R., Amri, H., Bouayad, N., Ghailani, N., Ennabili, A., and Sayah, F. (2008). Insecticidal Effects of Extracts of Seven Plant Species on Larval Development,  $\alpha$ -amylase Activity and Offspring Production of *Tribolium castaneum* (Herbst) (Insecta: Coleoptera: Tenebrionidae). *Bioresource Technology*, 99(5): 959-964.
58. Jibrin, J. M, Abubakar, S. Z., and Suleiman A. (2008). Soil Fertility Status of the Kano River Irrigation Project Area in the Sudan Savanna of Nigeria. *J Applied Sciences* 8: 692-696.
59. Jinab, H., Solond, M., and Varietel, M. (2008). *Functional Food Product Development*. Smith & Charter, Book. 354.
60. Joseph, B., Jini, D., and Sujatha, S. (2010). Biological and Physiological Perspectives of Specificity in Abiotic Salt Stress Response from Various Rice Plants. *Asian Journal of Agricultural Sciences*, 2(3): 99-105.
61. Kang, D. J., Futakuchi, K., Dumnoeng, S., and Ishii, R. (2007). High Yielding Performance of New Rice, IR 53650 in Mildly Improved Acid Sulfate Soil Conditions. *Plant Prod Sci* 10, 64-67.
62. Karathansis, A., Barton, C., and Chalfant, G. (2002). Influence of Acidic Atmospheric Deposition on Soil Solution Composition in the Daniel Boone National Forest, Kentucky, USA. *Environment Geology*, 41: 672-682.
63. Kavosi, M. (1995). *The Best Model for Rice Yield Prediction in Salinity Condition*. Dissertation of MSc. Tabriz University.
64. Kaya, C., Tuna, A., and Yokas, I. (2009). *Salinity and Water Stress* Springer, Netherlands. 45-50.

65. Kenny, O., Smyth, T.J., Walsh, D., Kelleher, C.T., Hewage, C.M., and Brunton, N.P. (2014). Investigating the Potential of Under-utilised Plants from the Asteraceae Family as a Source of Natural Antimicrobial and Antioxidant Extracts. *Food Chemistry*, 161(0): 79-86.
66. Khaled, A.A.A, Mazrou, Y.S.A., and Hafez, Y.M. (2020). Silicon Foliar Application Mitigates Salt Stress in Sweet Pepper Plants by Enhancing Water Status, Photosynthesis, Antioxidant Enzyme Activity and Fruit Yield. *Plants*, 9(6): 733.
67. Khoshbakht, D., and Asgharei, M. R. (2015). Influence of Foliar-Applied Salicylic Acid on Growth, Gas-Exchange Characteristics, and Chlorophyll Fluorescence in Citrus under Saline Conditions. *Photosynthetica*. 53 (3), 410-418.
68. Kijne, J. W. (2006). Biotic Stress and Water Scarcity. *Iranian Journal of Soil and Waters Sciences Spring*; 20 (1), 73-83. and Resolving Conflicts from Plant Level to the Global Level . *Field Crops Research*. 97, 3–18.
69. Kogel, K. H., Beckhove, U., Dreschers, J., Munch, S., and Romme, Y. (1994). Acquired Resistance in Barley. *Plant Physiology*, 106: 1269-1277.
70. Kohler, A., Schwindling, S., and Conrath, U. (2002). Benzothiadiazole Induced Priming for Potentiated Responses to Pathogen Infection, Wounding, and Infiltration of Water into Leaves Require the NPR1/NIM1 Gene in Arabidopsis. *Plant Physiology*, 128: 1046-1056.
71. Korndorfer, G. H., Pereira, H. S., and Nolla, A. (2004). Silicon Analysis in Soil, Plant and Fertilizers. Uberlândia, Brazil, GPSi/ICIAG/UFU.
72. Kumar, P., Dube, S. D., and Chauhan, V. S. (1999). Effect of Salicylic Acid on Growth, Development and Some Biochemical Aspects of Soybean (*Glycine max* L. Merrill). *Ind. J. Plant Physiology*, 4: 327-330.
73. Kuznetsov V.L.V., and Shevyakova N.I.(1999). Proline under Stress Conditions: Biological Role, Metabolism, and Regulation. *Russ. J. Plant Physiol.*, 46: 321-336.

74. Lamb, C., and Dixon, R. A. (1997). The Oxidative Burst in Plant Disease Resistance. *Annual Review. Plant Physiology Plant Molecular Biology*, 48: 251-275.
75. Larque-Saavedra, A., and Martin-Mex, R. (2007). Effects of Salicylic Acid on the Bioproductivity of Plants (Chapter 2). In S. Hayat and Dr. A. Ahmad, En: *Salicylic Acid: A plant hormone*. The Netherlands, Dordrecht, 15–23.
76. Lee, S.K., Sohn, E.Y., Hamayun, M., Yoon, J.Y., and Lee, I.J. (2010). Effect of Silicon on Growth and Salinity Stress of Soybean Plant Grown under Hydroponic System. *Agroforest. Syst.*, 80: 333–340. Doi:10.1007/s10457-010-9299-6.
77. Li, J., Zhang, Y.L., Liu, M.D., Yu, N., Huang, Y., and Yang, L.J. (2002). Study on Silicon-supply Capacity and Efficiency of Siliceous Fertilizer in Paddy Soils in Liaoning Province. *Chinese Journal of Soil Science*, 2: 61-71.
78. Liang, Y.C., Cheng, Q., Liu, Q., Zhang, W.H., and Ding, R.X. (2003). Exogenous Silicon (Si) Increases Antioxidant Enzyme Activities and Reduces Lipid Peroxidation in Roots of Salt-stressed Barley (*Hordeum vulgare* L.). *Plant Physiol.*, 160: 1157-1164.
79. Liang, Y., Sun, W., Si, J., and Römheld, V. (2005). Effects of Foliar and Root Applied Silicon on the Enhancement of Induced Resistance to Powdery Mildew in *Cucumis sativus*. *Plant Pathology* 54: 678-685.
80. Liang, Y.C., Sun, W.C., Zhu, Y.G., and Christie, P. (2007). Mechanisms of Silicon-mediated Alleviation of Abiotic Stresses in Higher Plants. *A Review of Environmental Pollution*. 147: 422-428.
81. Liu, W., Zhang, Y., Yuan, X., Xuan, Y., Gao, Y., and Yan, Y. (2016). Exogenous Salicylic Acid Improves Salinity Tolerance of *Nitraria tangutorum*. *Russian Journal of Plant Physiology*, 63, 132–142.
82. Ma, J.F. (2003). Functions of Silicon in Higher Plants. In: *Silicon biomineralization*, Springer, 127-147.

83. Ma, J.F., and Takahashi, E. (2002). Soil Fertilizer and Plant Silicon Research in Japan. Elsevier Science, 73-106.
84. Makandar, R., Essig, J. S., Schapaugh, M. A., Trick, H. N., and Shah, J. (2006). Genetically Engineered Resistance to Fusarium Head Blight in Wheat by Expression of Arabidopsis NPR1. *Mol. Plant-Microbe Interact.* 19: 123-129.
85. Malidareh, A.G., (2011). Silicon Application and Nitrogen on Yield and Yield Components in Rice (*Oryza sativa*) in two Irrigation Systems. *World Academy of Sci. Engr. and Tech*, 78:88-95.
86. Massey, F.P., and Hartley, S.E. (2006). Experimental Demonstration of the Antiherbivore Effects of Silica in Grasses: Impacts on Foliage Digestibility and Vole Growth Rates. *Proc. R. Soc. B*, 273:2299-2304.
87. Mauad, M., Crusciol, C.A.C., Grassi Filho, H., and Corrêa, J.C. (2003). Nitrogen and Silicon Fertilization of Upland Rice. *Scientia Agricola* 60: 761-765.
88. Mbega, E.R., Mortensen, C.N., Mabagala, R.B., and Wulff, E.G. (2012). The Effect of plant Extracts as Seed Treatments to Control Bacterial Leaf Spot of Tomato in Tanzania. *Journal of general plant pathology*, 78: 277-286.
89. Meguro, A., and Sato, Y. (2014). Salicylic Acid Antagonizes Abscisic Acid Inhibition of Shoot Growth and Cell Cycle Progression in Rice. *Sci Rep* 4(1): 1–11.
90. Mishra, A., and Choudhuri, M.A. (1999). Effects of Salicylic Acid on Heavy Metal-induced Membrane Degradation Mediated by Lipoxygenase in Rice. *Biol. Plant*, 42: 409-415.
91. Mishra, M., Wungramoha, S., Kumar, G., Singla-Pareek, S.L., and Pareek, A. (2020). How do Rice Seedlings of Landrace Pokkali Survive in Saline Fields after Transplantation? *Physiology, Biochemistry and Photosynthesis*. *Photosynthesis Research*, <https://doi.org/10.1007/s1120-020-00771-6>.

92. Mobasser, H.R., Malidarh, G.A., and Sedghi, H. (2008). Effects of Silicon Application to Nitrogen Rate and Splitting on Agronomic Characteristics of Rice (*Oryza sativa*). Silicon in Agriculture: 4<sup>th</sup> International Conference 26-31 October, South Africa: 76.
93. [Mohammed](#), A. R. and [Tarpley](#), L. (2011). Effects of Night Temperature, Spikelet Position and Salicylic Acid on Yield and Yield-Related Parameters of Rice (*Oryza sativa* L.) Plants. Journal of agronomy and crop science, <https://doi.org/10.1111/j.1439-037X.2010.00439.x>.
94. [Morton, Julia F.](#) (1987). [Fruits of Warm Climates](#). Wipf and Stock Publishers, 115–121. ISBN 978-0-9653360-7-9.
95. Munns, R., and Termaat, A. (1986). Whole-Plant Responses to Salinity. Aust. J. plant physiol., 13, 143 -160.
96. Naidu, R., and Rangasamy, P. (1993). Ion Interactions and Constraints to Plant Nutrition in Australian Sodic Soils. Aust. J. Soil Res., 31, 801-819.
97. Nepomuceno, A. L., Oosterhuis, D. M., and Stewart J. M. (1998). Physiological Response of Cotton Leaves to Water Deficit Induced by Polyethylene Glycol. Environ. Exp. Bot., 40, 29-41.
98. Osagie, C. (2014). 2015 Rice Importation Ban: Disregard US report, FG urged. Available from <http://www.thisdaylive.com/articles/2015-rice-importation-ban-disregard-us-report-fg-urged/168731/> [Accessed 17/05/14].
99. [Pal](#), M., [Singh](#), D.K., [Rao](#), L.S., and [Singh](#), K.P. (2004). Photosynthetic Characteristics and Activity of Antioxidant Enzymes in Salinity Tolerant and Sensitive Rice Cultivars. Indian J. Plant Physiol, 9(4): 407-412.

100. Parvez, S. S., Parvez, M. M., Nishihara, E., Gemma, H., and Fujii. Y. (2003). Tamarindus indica L. leaf is a source of allelopathic substance, Plant Growth Regulators., 40(2): 107–15.
101. Rady, M.M., B. Varma C, and Howladar, S.M. (2013). Common bean (Phaseolus vulgaris L.) Seedlings Overcome NaCl Stress as a Result of Presoaking in Moringa oleifera Leaf Extract. Scientia Horticulturae, 162: 63-70.
102. Rajasekaran, L. R., and Blake, T. J. (1999). New Plant Growth Regulators Protect Photosynthesis and Enhance Growth under Drought of Jack Pine Seedlings. J. Plant Growth Regul. 18: 175-181.
103. Rajjou, L., Belghazi, M., Huguet, R., Robin, C., Moreau, A., Job, C., and Job, D. (2006). Proteomic Investigation of the Effect of Salicylic Acid on Arabidopsis Seed Germination and Establishment of Early Defense Mechanisms. Plant Physiol, 141 (3), 910-923.
104. Rice - Sustainable Community Action. (n.d.). Retrieved from <http://sca21.wikia.com/wiki/Rice>
105. Rodrigues, F.Á., Datnoff, L.E. Korndörfer, G.H. Seebold, K.W., and Rush, M.C. (2001). Effect of Silicon and Host Resistance on Sheath Blight Development in Rice. Plant Dis, 85: 827-832.
106. Rodrigues, F.Á., Vale, F.X., Datnoff, L.E., Prabhu, A.S., and Korndörfer, G.H. (2003). Effect of Rice Growth stages and Silicon on Sheath Blight Development. Phytopathology, 93: 256-261.
107. Romero-Aranda, M.R., Jurado, O., and Cuartero, J. (2006). Silicon Alleviates the Deleterious Salt Effect on Tomato Plant Growth by Improving Plant Water Status. Plant Physiol., 163, 847–855. doi:10.1016/j.jplph.2005.05.010.
108. Saadalla, M. M., Shanahan, J. F., and Quick, J. S. (1990). Heat Tolerance in Winter Wheat: I. Hardening and Genetic Effects on Membrane Thermostability. Crop Sci., 30, 1243-1247.

109. Sairam, R. K., Rao, K. V., and Srivastava, G. C. (2002). Differential Response of Wheat Genotypes to Long Term Salinity Stress in Relation to Oxidative Stress, Antioxidant Activity, and Osmolyte Concentration. *Plant Sci.*, 163, 1037-1046.
110. Sairam, R.K., and Tyagi, A. (2004). Physiological and Molecular Biology of Salinity Stress Tolerance in Plants. *Current Sci.*, 86: 407–421.
111. Sakhabutdinova, A. R., Fatkhutdinova, D. R., Bezrukova, M. V., and Shakirova, F. M. (2003). Salicylic Acid Prevents the Damaging Action of Stress Factors on Wheat Plants *Bulg J Plant Physiol*, 29, 314-319.
112. Sardoei, A. L., Mojgan, S., Monir, R. Y., and Somayeh, G. (2014). Growth Response of Petunia hybrid to Zinc Sulphate and Salicylic Acid. *Advanced Biological and Biomedical Research*. 2(3): 622-627.
113. Sawhney, S., Sawhney, N., Kumar, S., and Nanda, K. K. (1979). Enzyme and Electrophoretic Pattern of Isoenzymes of Amylase, Catalase, and Peroxidase in Photo- and Gibberellin Induced Plants of *Impatiens balsamina* L. *New Phytologist*. 82: 41-47.
114. Shah, J., and Klessig, D. F. (1999). Salicylic Acid: Signal Perception and Transduction. In: Hooykaas, P.P.J., Hall, M.A., and Libbenga, K.R. (Eds.), *Biochemistry and Molecular Biology of Plant Hormones*. Elsevier, Amsterdam, Netherlands. 513-541.
115. Shah, M.A., Bosco, S.J.D., and Mir, S.A. (2014). Plant Extracts as Natural Antioxidants in Meat and Meat Products. *Meat Science*, 98(1): 21-33.
116. Shakirova, F.M., Sakhabutdinova A.R., and Bezrukova M.V. (2003). Changes in the Hormonal Status of Wheat Seedlings Induced by Salicylic Acid and Salinity. *Plant Sci.*, 164: 317-322.
117. Shashidhar H.E., N. Chandrashekhar, C. Narayanaswamy, A.C. Mehendra, N. B. Prakash, 2008 - Calcium silicate as silicon source and its interaction with nitrogen in

aerobic rice. Silicon in Agriculture: 4th International Conference 26-31 October, South Africa: 93.

118. Shirasu, K., Nakajima, H., Rajasekhar, V.K., Dixon, R.A., and Lamb, C. (1997). Salicylic Acid Potentiates an Agonist-dependent Gain Control that Amplifies Pathogen Signals in the Activation of Defense Mechanisms. *Plant Cell*, 9:261–270.
119. Singh, V.J., Srihima, G., Vikas, K. R., Chakraborti, S. K., and Amitava, B. (2015). Effect of Foliar Spray of Salicylic Acid on Sheath Infecting Pathogen and Yield Attributes in Hybrid Rice. *An International Quarterly Journal of Environmental Sciences* 9(1&2): 507-512.
120. Sonon, L.S., Saha U., and Kissel, D.E. (2015). *Agricultural and Environmental Services Laboratories, Soil Salinity: Testing, Data Interpretation and Recommendations*, Circular 1019.
121. Srivastava, M. K., and Dwivedi, U. N. (2000). Delayed Ripening of Banana Fruit by Salicylic Acid. *Plant Sci* 158: 87-96.
122. Surekha, M., Kiran, S., Ram Reddy, S., and Reddy, S.M. (2018). *Moulds and Mycotoxins of Paddy: Incidence, Impact and Management*. Scientific Publishers, <http://www.scientificpub.com>.
123. Talukder, M.A. I., Rahaman, M., Roy, B., and Saha, K.C. (2015). Effects of Herbal Plant Extracts on Germination and Seedling Growth of some Vegetables. *International Journal of Science and Nature*, 6 (3): 421-425.
124. Tascioglu, C., Yalcin, M., Sen, S., and Akcay, C. (2013). Antifungal Properties of Some Plant Extracts used as Wood Preservatives. *International Biodeterioration and Biodegradation*, 85(0): 23-28.
125. USAID, (William Grant, Dan Charette and Michael Field), (2009). *Global Food Security Response West Africa Rice Value Chain Analysis, Global Food Security Response Nigeria Rice Study*.

126. USDA. (2003) Potassium Silicate Tap1- Agricultural Marketing Service. (n.d). Retrieved from [https://www.ams.usda.gov/sites/default/files/media/Pot%20sil%20technica %20advis](https://www.ams.usda.gov/sites/default/files/media/Pot%20sil%20technica%20advis)
127. Van, V. O. (2017). Tamarind, *Tamarindus indica*. Retrieved June 4.
128. Waseem, M., Athar, H.U.R., and Ashraf, M. (2006). Effect of Salicylic Acid Applied through Rooting Medium on Drought Tolerance of Wheat. *Pak. J. Bot.*, 38(4): 1127-1136.
129. Xu, Q., Xu X., Zhao Y. (2008). Salicylic Acid, Hydrogen Peroxide and Calcium-induced Saline Tolerance Associated with Endogenous Hydrogen Peroxide Homeostasis in Naked Oat Seedlings. *Plant Growth Regul.*, 54: 249-259.
130. Yahia, E. M., and Salih, N. K. (2011). Tamarind (*Tamarindus indica* L.) Research Gate, DOI: 10.1533/9780857092618.442
131. Yildirim, E., Turan M., and Guvence I. (2008). Effect of Foliar Salicylic Acid Application on Growth, Chlorophyll, and Mineral Content of Cucumber Grown Under Salt Stress. *J.Plant Nut.*, 31: 593-612.
132. Zekri, M., and Parsons L.P.(1992). Salinity Tolerance in Citrus Rootstock: Effect of Salt on Root and Leaf Mineral Concentrations. *Plant Soil*, 147: 171-181.
133. Zhou, X.M., Mackeuzie A.F., and Madramootoo C.A. (1999). Effects of Some Injected Plant Growth Regulators with or without Sucrose on Grain Production Biomass and Photosynthetic Activity of Field-Grown Corn Plants. *Agro. Crop Sci.*, 183: 103-110.
134. Zhu, Z., Wei, G., Li, J., Qian, Q., and Yu, J. (2004). Silicon Alleviates Salt Stress and Increases Antioxidant Enzymes Activity in Leaves of Salt-stressed Cucumber (*Cucumis sativus* L.). *Plant Sci.*, 167: 527–533. doi:10.1016/.