

1 **Original Research Article**  
2 **Pharmacological Evaluation of Ovulation Inducing**  
3 **Potential of Siddha Herbomineral Formulation**  
4 ***Arputha Mathirai* on Estradiol Valerate Induced Poly**  
5 **Cystic Ovarian Syndrome Wistar Albino Rat Model**  
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12 **ABSTRACT**

13 **Aim:** The aim of this study was to examine the potential of Arputha Mathirai, a tablet-based Siddha  
14 herbomineral formulation, in promoting ovulation in Wistar Albino rats with polycystic ovary syndrome  
15 (PCOS) induced by Estradiol Valerate (EV).  
16

17 **Place of Study**

The study took place at C.L. Baid Metha College of Pharmacy located in Thorapakkam, Chennai - 600  
092, Tamil Nadu.

18 **Methodology**

19 Arputha Mathirai, a Siddha herbomineral formulation, was prepared in accordance with Good  
20 Manufacturing Practices (GMP) guidelines. Prior to conducting the study, approval was obtained from  
21 the Institutional Animal Ethics Committee (IAEC). The research adhered to ethical principles and  
22 guidelines established by the committee responsible for overseeing and regulating animal  
23 experimentation, ensuring proper control and supervision.

Female Wistar Albino rats were selected as the preferred rodent species for the study. Polycystic ovary  
syndrome (PCOS) was induced in the animals by administering a subcutaneous injection of 100µg  
Estradiol valerate (EV). The reproductive cycles of the rats were synchronized to ensure consistency.

The rats were divided into four groups, each containing six rats, as follows:

- Group I: Normal Control animals received 1ml/kg of Sodium Carboxymethyl Cellulose (CMC) solution.
- Group II: Rats were orally administered Arputha Mathirai at a dosage of 100mg/kg for 10 days.
- Group III: Rats were orally administered Arputha Mathirai at a dosage of 200mg/kg for 10 days.
- Group IV: Received Clomiphene at a dosage of 10mg/kg and served as the standard group.

At the end of the study, blood samples were collected from the rats through retro-orbital and cardiac  
puncture. The levels of luteinizing hormone (LH), follicle-stimulating hormone (FSH), estradiol, and  
progesterone were estimated using the ELISA method. Ovaries from the experimental rats were  
dissected out for histopathological studies.

28 **Results**

29 The results consistently showed that the higher dosage of the drug (group III) had a more pronounced  
30 effect in normalizing hormone levels compared to the lower dosage group. However, it is important to  
31 note that the lower dosage group also exhibited effectiveness. While the results in both groups did not  
32 closely match those of the standard group (group IV), the drug Arputha Mathirai demonstrated significant  
potential in the experimental groups, supporting its efficacy in inducing ovulation.

Furthermore, the histopathological analysis confirmed the drug's potential in inducing ovulation,  
particularly in the higher dosage group (group II) compared to the lower dosage group.

Overall, the findings suggest that Arputha Mathirai has promising ovulation-inducing activity, with the  
higher dosage showing more pronounced effects.

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35 **Conclusion**

36 It is hypothesized that the bioactive phytochemicals present in Arputha Mathirai may exert their  
37 effects at various stages to restore hormone levels and reverse the pathological condition associated  
38 with PCOS. These natural compounds have the potential to regulate hormone levels and promote  
39 ovulation. However, it is important to acknowledge that there is a scarcity of published evidence  
40 regarding the specific ability of these phytochemicals to address the underlying causes of the  
41 condition. Further research is needed to explore the mechanisms by which these compounds act and  
42 their potential in addressing the root causes of PCOS.

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44 *Keywords: Siddha, Arputha Mathirai, PCOS, Ovulation inducing Activity, Animal Study.*

45

UNDER PEER REVIEW

## 1. INTRODUCTION

Polycystic ovary syndrome (PCOS) is the most common endocrinologic condition in women, affecting from 8% to 13% of reproductive-aged women. Research has exposed the still-large gap between the available evidence and its translation to improved diagnostic timing and evidence-based treatments: The pathogenesis of PCOS is complex and multifactorial, including genetic, environmental, and transgenerational components. <sup>(1)</sup> Hyperandrogenism, anovulation, the presence of numerous ovarian cysts, abnormalities in the menstrual cycle, and fluctuating levels of gonadotropins are all symptoms of PCOS. <sup>(2)</sup>

Many allopathic and ayurvedic drugs are being prescribed by medical practitioners to address the secondary symptoms of PCOS. Although allopathic drugs like metformin, clomiphene citrate, tamoxifen, flutamide, oral contraceptive pills, myo-inositol etc., [\(Jitendra and Pravin, 2012\)](#) assist in overcoming infertility and improving PCOS symptoms, they pose several side effects like hepatotoxicity, hot flushes, nausea, vomiting, impotency, blurred vision and osteoporosis in PCOS patients [\(Domecq et al., 2013\)](#). Hence, phytomedicine from herbal plants are being constantly explored for treating PCOS. <sup>(3)</sup>

In recent times, there has been a growing trend in utilizing these medicinal preparations for addressing Poly Cystic Ovarian Syndrome (PCOS). Herbal remedies for PCOS have received attention as a form of lifestyle management in traditional medicine, in which the menstrual cycle and normal serum hormones levels can be recovered. <sup>(4)</sup> Different approaches to manage PCOS include the traditional system of drugs, herbal remedies, herbs, and phyto-ingredients that have proven their potential in treating PCOS. <sup>(5)</sup> In Siddha system of medicine, various herbal and herbo-mineral formulations have been used for the management of PCOS traditionally along with lifestyle changes and physical activity. <sup>(6)</sup> An attractive appeal of herbal medicine is the prospect to simultaneously target multiple pathophysiological mechanisms. <sup>(7)</sup>

The etiology of PCOS is unknown, likely reflecting multiple pathophysiological mechanisms, and an accepted animal model of the disease has not been established. A recent review concluded that multiple models may be needed, depending on whether the goal is to investigate ovarian morphology or a particular PCOS-related disorder. In both mechanistic and treatment studies, we and others have used a model in which PCO is induced by a single intramuscular (i.m.) injection of estradiol valerate (EV) in 8-week-old rats. The rats cease ovulating and develop characteristics of human PCOS, including large cystic follicles in the ovaries and altered concentrations of luteinizing hormone. <sup>(8)</sup>

New drug development process must continue through several stages in order to make a medicine that is safe, effective, and has approved all regulatory requirements. <sup>(9)</sup>

The process of developing a novel drug is time consuming and costly. To increase the chances of successfully completing a clinical trial leading to the approval of a new drug, the choice of appropriate preclinical models is of utmost importance. Identifying a safe, potent, and efficacious drug requires thorough preclinical testing, which evaluates aspects of pharmacodynamics, pharmacokinetics, and toxicology in in vitro and in vivo settings. Nevertheless, merely a small fraction of investigational new drugs tested in clinical trials after passing preclinical evaluation eventually lead to a marketed product. Hence, there is a need for optimising current standard preclinical approaches to better mimic the complexity of human disease mechanisms. <sup>(10)</sup>

The Siddha system of Medicine offers distinct combinations of medicines that provide solutions for managing PCOS. Among these formulations, one specific herbomineral formulation is *Arputha Mathirai* (AM), which is mentioned in the Siddha literature, *Koshayee Anuboga Vaithiya Bramma Ragasiyam*. This formulation includes 7 herbal and two mineral drugs. <sup>(11)</sup>

This preclinical evaluation was conducted to assess the ovulation-inducing activity of the mentioned formulation AM, in Wistar Albino rats with polycystic ovary syndrome (PCOS) induced by Estradiol Valerate (EV). Hence, the objective of this study was to evaluate the potential of the formulation to induce ovulation in Wistar Albino rats with EV-induced PCOS.

## 2. MATERIALS AND METHODS

106 **2.1 Study Drug**

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108 **2.1.1 Source and Authentication of Drugs**

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110 The herbal and mineral raw drugs were procured from **reputable raw drug store**, which were then  
111 identified and verified by the Botanist of Government Siddha Medical College, Chennai (Voucher  
112 number GSMC/MB- 566 – 571 & 614) and the Head of the Department of Gunapadam, Government  
113 Siddha Medical College, Chennai, Tamil Nadu, respectively, with the aim of ensuring authenticity.

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115 **2.1.2 Ingredients of *Arputha Mathirai* (AM) <sup>(11)</sup>**

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117 <i>Cuminum cyminum</i> L. (Cumin)	-70 g
118 <i>Piper nigrum</i> L. (Black Pepper)	-70g
119 <i>Zingiber officinale</i> Roscoe. (Dried Ginger)	-70g
120 <i>Piper longum</i> L. (Long Pepper)	-70g
121 <i>Allium sativum</i> L. (Garlic)	-70g
122 <i>Ferula asafoetida</i> L. (Asafetida)	-70g
123 Purified rock salt	-70g
124 Purified Sulphur	-70g
125 <i>Citrus limon</i> Linn. (Lemon juice)	- Sufficient Quantity

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127 **2.1.3 Preparation of AM**

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129 **2.1.3.1 Purification of Raw Drugs**

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131 All the herbal and mineral raw drugs were purified individually according to the purification methods  
132 mentioned in the classical Siddha texts, “*Sikitcha Ratna Deepam Ennum Vaidhiya Nool*” and  
133 “*Gunapadam Thathu Jeeva Vaguppu*” respectively, as mentioned below. <sup>(12)(13)</sup>

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135 **a) Cumin**

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137 Unwanted soil particles and dusts were removed, winnowed and sun dried.

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139 **b) Black Pepper**

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141 Soaked in sour buttermilk for 3 hours (1 *saamam*) and sun dried.

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143 **c) Dried Ginger**

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145 One part of dried ginger was bleached with 2 parts of lime stone (*kal sunnambu*) for 3 hours  
146 (1 *saamam*), washed, dried and the outer skin was peeled.

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148 **d) Long Pepper**

149

150 Soaked in the leaf juice of **Plumbago indica** (*Kodiveli*) for 24 minutes (1 *naazhiga*) and then  
151 sun dried.

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153 **e) Garlic**

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155 Outer dry papery skin and the tip were removed and washed.

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157 **f) Asafetida**

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159 Roasted in coal fire and then powdered.

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161 **g) Rock Salt**

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163 Soaked in vinegar (*kaadi*) for 3 days and then sun dried.

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165 **h) Sulphur**

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Crude Sulphur (*Gandhagam*) was melted in an iron ladle containing little butter and the melted content was poured into a container containing juice of banana stem. This was repeated for 10 times.

## i) Lemon Juice

Lemons were washed, pat dry, juice was extracted, and the seeds were removed.

### 2.1.3.2 Preparation of Sample

The herbomineral siddha formulation *Arputha Mathirai* (AM) was prepared as per Siddha text.<sup>(11)</sup>

- ❖ All the purified ingredients except garlic and lemon juice were taken in the equal quantities (70 g), grounded individually, manually in an iron mortar with pestle, into a very fine powder, and sieved using a sieving cloth individually.
- ❖ Then all the powdered single drugs were mixed together.
- ❖ Garlic (70 g) was grounded and made into a very fine paste in a stone mortar, and mixed and grounded with the above powder.
- ❖ The above mixture was grounded together in a stone mortar for about 12 hours with adequate lemon juice and then rolled into pills of 500 mg (*sundai alavu*)<sup>(14)</sup> size.
- ❖ The rolled pills were dried in shade and stored in a clean, dry, air tight container.

## 2.2 Ethical Approval and Experimental Animals

### 2.2.1 Ethical Approval

Following approval from the Institutional Animal Ethics Committee (IAEC) at C.L. Baid Metha College of Pharmacy in Thorapakkam, Chennai 600 092, Tamil Nadu, preclinical evaluations were initiated. All experiments were carried out in full adherence to ethical principles and guidelines established by the committee responsible for overseeing and regulating animal experimentation, ensuring proper control and supervision. (IAEC NO: 05/31/PO/Re/S/01/CPCSEA/dated 06/04/2022).

### 2.2.2 Selection of Animals Species

Sexually matured Female Wistar albino rats (150-200 gm) were obtained from Mass Biotech, Chennai. All the animals were kept under standard environmental condition (22±3°C). The animals had free access to water and standard pellet diet (Sai Meera Foods, Bangalore).

The preferred rodent species was female Wistar Albino rat, although other rodent species may be used. Healthy young adult animals were commonly used. All females were nulliparous and non-pregnant. Each animal, at the commencement of its inducing, were between 6 to 8 weeks old. The weight of the animals were 150-200gm, and the weight of the animals fell in an interval within±20 % of the mean weight of any previously dosed animals.

### 2.2.3 Housing and Feeding Conditions

The temperature in the experimental animal room was maintained between 22°C ± 3°C. Although the relative humidity should be at least 30% and preferably not exceed 70% other than during room cleaning the aim was 50-60%. Lighting was artificial, the sequence being 12 hours light, 12 hours dark. For feeding, conventional laboratory diets were used with an unlimited supply of drinking water. Animals were group-caged by dose, but the number of animals per cage did not interfere with clear observations of each animal.

### 2.2.4 Preparation of Animals

The animals were randomly selected and marked to permit individual identification, and kept in their cages for at least 7 days prior to inducing PCOS to allow for acclimatization to the laboratory conditions.

226 **2.2.5 Induction of PCOS**

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228 PCOS was induced in the animals by subcutaneous injection of 100µg Estradiol valerate (EV).

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230 **2.2.6 Examination of Oestrous Cycle**

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232 Before starting drug treatment, the reproductive cycles of the rats were synchronized by the following  
233 method. 100µg estradiol valerate dissolved in 2 ml olive oil was injected subcutaneously. All rats after  
234 a 24 hr period, received intramuscular injections of 50 µg progesterone dissolved in olive oil. After few  
235 hours, vaginal smears were obtained by vaginal lavage to monitor ovulation and oestrous cycle.  
236 Vaginal smears were prepared by washing vaginal opening with 0.9% w/v of sodium chloride with a  
237 glass dropper and placed in a clean glass slide and viewed under light microscope at 40X  
238 magnification. Examination of vaginal smears showed that all the animals were in the estrous stage.

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240 **2.3 Evaluation of Ovulation Inducing Activity**

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242 All the animals were weighed daily after drug administration for 10 days. The suitable sensitive rats  
243 were divided into four groups of six each as follows.

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245 **2.3.1 Experimental design**

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247 ❖ Group I : Normal Control animals received 1ml/kg of Sodium Carboxymethyl Cellulose (CMC)  
248 solution.

249 ❖ Group II : Rats were administered Arputha Mathirai 100mg/kg for 10days,

250 ❖ Group III : Rats were administered Arputha Mathirai 200mg/kg for 10 days

251 ❖ Group IV : Received Clomiphene 10mg/kg and served as standard.

252 All the drugs were given orally.

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254 **2.3.2 Sample Collection**

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256 At the end of the study, prior to sacrificing the animals, an overnight fasting period was implemented  
257 while ensuring they had access to water. The animals were then sacrificed using an excessive  
258 amount of anesthesia. Blood samples were collected through retro-orbital and cardiac puncture  
259 methods. Hormone estimation was subsequently performed using the Cobas e411 immunoassay  
260 analyzer.

261 After that 2ml of blood was collected by retro orbital puncture. Blood samples were centrifuged for 15  
262 minutes at 4000 rpm and the separated serum samples were frozen at -20°C and kept for later  
263 estimation of LH, FSH, Estradiol and progesterone by ELISA method.

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265 **2.3.3 Hormonal Assay**

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267 **2.3.3.1 Estimation of Serum Luteinizing Hormone (LH)**

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269 The method employed was Microwell Enzyme Linked Immunosorbent Assay (ELISA) using analytical  
270 grade reagents. 0.05 ml of the serum was pipetted into the assigned wells. 0.001 ml of LH-Enzyme  
271 reagent was added to all the wells. The microplate was swirled for 20-30 seconds and covered, this  
272 mixture was allowed to incubate for 60 minutes at room temperature. After which, the contents were  
273 discarded by decantation. 350 µl of wash buffer was added and decanted for 3 times. 100 µl of  
274 working substrate solution was added to all the wells and was allowed to incubate for fifteen minutes.  
275 50 µl of stop solution was added to all the wells and gently mixed for 20 seconds. The optical density  
276 was read at 450 nm in a microplate reader within 30 mins. The mean absorbance values for each set  
277 of standards, controls, and test samples were calculated and a standard curve was constructed by  
278 plotting a mean absorbance obtained from each of the reference standard against its concentration  
279 from the standard curve.

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281 **2.3.3.2 Estimation of Serum Follicle Stimulating Hormone (FSH)**

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283 The method employed was Microwell immunoassay (ELISA) using analytical grade reagents. 0.05 ml  
284 of the serum was pipetted into the assigned wells. 0.001 ml of FSH-Enzyme reagent was added to all  
285 the wells. The microplate was swirled for 20-30 seconds and covered, this mixture was allowed to

286 incubate for 60 minutes at room temperature. After which, the contents were discarded by  
287 decantation. 350 µl of wash buffer was added and decanted for 3 times. 100 µl of working substrate  
288 solution was added to all the wells and was allowed to incubate for fifteen minutes. 50 µl of stop  
289 solution was added to all the wells and gently mixed for 20 seconds. The optical density was read at  
290 450 nm in a microplate reader within 30 mins. The mean absorbance values for each set of  
291 standards, controls, and test samples were calculated and a standard curve was constructed by  
292 plotting a mean absorbance obtained from each of the reference standard against its concentration  
293 from the standard curve.

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#### 295 **2.3.3.3 Determination of Serum Progesterone Levels**

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297 The method employed was Microwell immunoassay (ELISA) using analytical grade reagents. 0.025  
298 ml of the serum was pipetted into the assigned wells. 0.05 ml of progesterone Enzyme reagent was  
299 added to all the wells. The microplate was swirled for 20 seconds to mix, 0.05 ml progesterone biotin  
300 reagent was added to all the wells, the mixture was swirled for 20 seconds to mix and covered, this  
301 mixture was allowed to incubate for 60 minutes at room temperature. After which, the contents were  
302 discarded by decantation. 350 µl of wash buffer was added and decanted for 3 times. 100 µl of  
303 working substrate solution was added to all the wells and was allowed to incubate for twenty minutes.  
304 50 µl of stop solution was added to all the wells and gently mixed for 20 seconds. The optical density  
305 was read at 450 nm in a microplate reader within 30 mins. The mean absorbance values for each set  
306 of standards, controls, and test samples were calculated and a standard curve was constructed by  
307 plotting a mean absorbance obtained from each of the reference standard against its concentration  
308 from the standard curve.

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#### 310 **2.3.3.4 Determination of Serum Estradiol Levels**

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312 The method employed was Microwell immunoassay (ELISA) using analytical grade reagents. 0.025  
313 ml of the serum reference was pipetted into the assigned wells. 0.05 ml of Estradiol Biotin reagent  
314 was added to all the wells. The microplate was swirled for 20 seconds to mix, the mixture was  
315 incubated at room temperature for 30 mins, 0.05 ml Estradiol enzyme reagent was added to all the  
316 wells, the mixture was swirled for 20 seconds to mix and covered, this mixture was allowed to  
317 incubate for 90 minutes at room temperature. After which, the contents were discarded by  
318 decantation. 350 µl of wash buffer was added and decanted for 3 times. 100 µl of working substrate  
319 solution was added to all the wells and was allowed to incubate for twenty minutes. 50 µl of stop  
320 solution was added to all the wells and gently mixed for 20 seconds. The optical density was read at  
321 450 nm in a microplate reader within 30 mins. The mean absorbance values for each set of  
322 standards, controls, and test samples were calculated and a standard curve was constructed by  
323 plotting a mean absorbance obtained from each of the reference standard against its concentration  
324 from the standard curve.

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#### 326 **2.3.4 Histopathology**

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328 Ovaries from the experimental rats were dissected out and fixed in 10% buffered neutral formal saline  
329 and processed. After fixation, tissues were embedded in paraffin. Fixed tissues were cut at 10 µm and  
330 stained with haematoxylin and eosin. The sections were examined under light microscope for  
331 histological changes.

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### 333 **3. RESULTS**

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335 All the results were tabulated and depicted in figures as hereunder.

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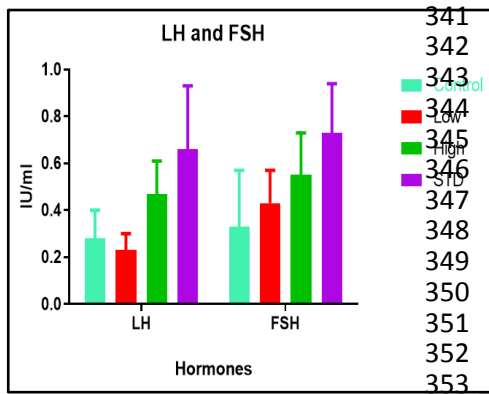
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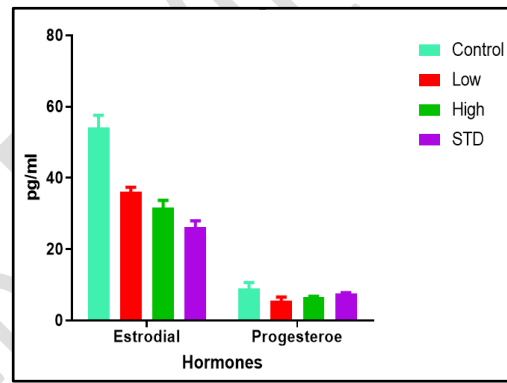
**Table 1 Effect of *Arputha Mathirai* on Serum Concentration of Reproductive Hormones of Female Wister Albino Rat**

S. No	Group	Treatment and dose	LH (IU/ml)	FSH (IU/ml)	Estrodial (pg/ml)	Progesterone (pg/ml)
1.	Normal	2 ml/kg 2% CMC	0.28±0.12	0.33±0.24	54.10±3.5	9.03±1.65
2.	Low	100 mg /kg	0.23±0.07	0.43±0.14	36.11±1.3	5.7±1.22
3.	High	200 mg /kg	0.47±0.04	0.55±0.18	31.64±2.1	6.5±0.32
4.	STD	Clomiphene 10 mg/kg	0.66±0.27	0.73±0.21	26.16±18	7.5±0.32

**Fig 1 Effect of *Arputha Mathirai* on Serum LH and FSH**

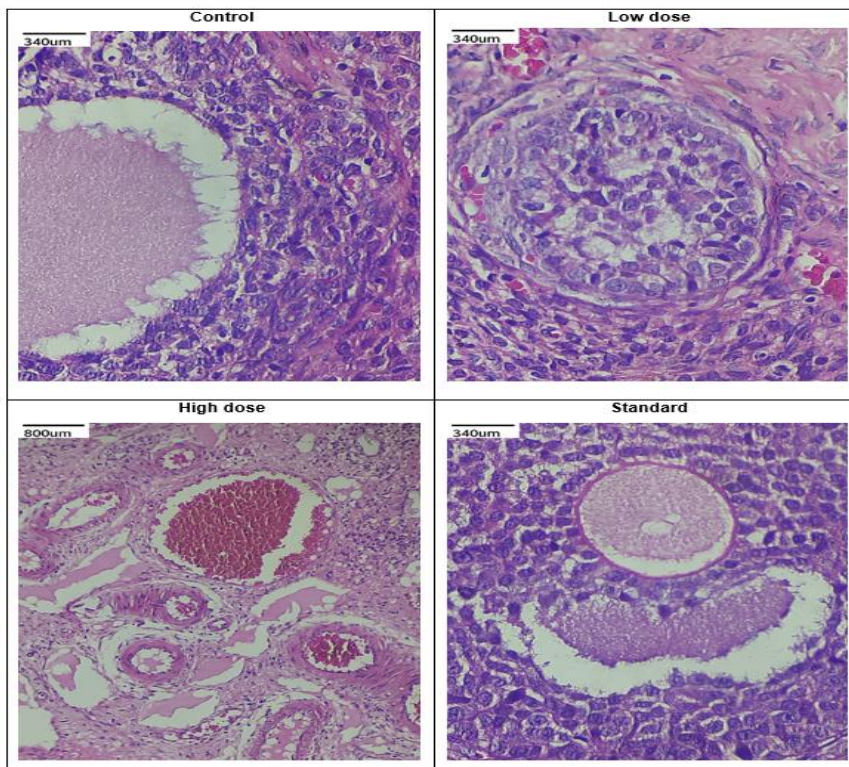


**Fig 2 Effect of *Arputha Mathirai* on Serum Estradiol and Progesterone**



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**Fig 3 Histopathology of Ovary in all Experimental Groups**



368 **4. DISCUSSION**

369 Weight of ovary sample belonging to group I rats showed progressive increase when compared to  
370 group IV. Treatment with AM at both the dose level, in group II and group III, showed decrease in the  
371 weight of the ovaries.

372 The serum LH levels in group II and group III were measured to be 0.23+0.07 IU/ml and 0.47+0.04  
373 IU/ml respectively. In comparison, the control group I had a level of 0.28+0.12 IU/ml, while the  
374 standard group IV showed a level of 0.66+0.27 IU/ml. The results clearly demonstrate a significant  
375 difference in the serum LH level in the high-dose group (group III), suggesting that the drug is  
376 effective at a high dosage of 200 mg/kg, in normalizing the serum LH levels.

377 Group II and III exhibited serum FSH levels of 0.43+0.14 IU/ml and 0.55+0.18 IU/ml, respectively.  
378 These levels were significantly higher compared to the control group, which had a level of 0.33+0.24  
379 IU/ml. The standard group showed a serum FSH level of 0.73+0.21 IU/ml. These findings further  
380 confirm the efficacy of the drug in significantly normalizing the serum FSH levels.

381 When analyzing the serum estradiol levels, it was observed that group II and group III had levels of  
382 36.11±1.3 pg/ml and 31.64±2.1 pg/ml, respectively. In comparison, the normal control group had a  
383 level of 54.10±3.5 pg/ml, while the standard group exhibited a level of 26.16±18 pg/ml. These results  
384 further validate the effectiveness of the drug in both groups, suggesting a potential impact in the high-  
385 dose group as well.

386 The serum progesterone levels in group II and III were found to be 5.7+1.22 pg/ml and 6.5+0.32  
387 pg/ml, respectively. In comparison, group I and IV had levels of 9.03+1.65 pg/ml and 7.5+0.32 pg/ml,  
388 respectively. The significant difference observed in both the low and high dose groups confirms the  
389 efficacy of the drug in reducing serum progesterone levels.

390 The histopathology examination of samples from group I demonstrated a noteworthy increase in the  
391 number of follicles at various stages and corpus luteum. Treatment with AM at doses of 100 mg/kg  
392 and 200 mg/kg in group II & III respectively, reduced the follicle count compared to the control group  
393 to a significant level, and the histology of the corpus luteum was restored to a similar state as that of  
394 the normal control rats. The histopathology report of group IV showed the normal appearance of  
395 graafian and antral follicles, as well as the presence of corpora lutea (CL), atretic follicles (AF), and  
396 interstitial tissue (IT) that appeared normal.

397 The aforementioned findings consistently indicated that the high dose of the drug (group III)  
398 demonstrated a greater potential in normalizing hormone levels compared to the low dose group.  
399 However, it is important to acknowledge that the low dose group also exhibited efficacy. Despite the  
400 fact that the results in both groups did not closely resemble those of the standard group IV, the drug  
401 AM demonstrated significant potential in the test groups, providing support for its efficacy in inducing  
402 ovulation.

403 Additionally the histopathological report also proved the potential ovulation inducing activity of the  
404 drug in the experimental groups, particularly with specific to high dose group II when compared to the  
405 low dose group.

406 The ingredients of the drug already possess anti diabetic, anti-hyperglycemic, hypoglycemic,  
407 emmenagogue effects individually. The combined effect of these drugs could be the reason behind  
408 this efficacy. The phytochemicals and the phytochemicals of the drug could be the reason for this  
409 efficacy.<sup>[15]</sup>

410 **5. CONCLUSION**

411 Initial analysis of the phytochemical content of Arputha Mathirai in tablet form revealed the presence  
412 of bioactive compounds that could be active components and have the potential to regulate hormone  
413 levels and stimulate ovulation. However, there is a lack of published evidence regarding the ability of  
414 these phytochemicals to repair the underlying causes of the condition. Our hypothesis suggests that  
415 these natural compounds may function at different stages to restore hormone levels and revert the

416 pathological condition back to normal. Nonetheless, it is crucial to identify the specific phytochemical  
417 component the mechanism that is responsible for reversing the pathogenesis of polycystic ovary  
418 syndrome (PCOS). Identifying the specific phytochemical component and its underlying mechanism is  
419 crucial in understanding how it contributes to the reversal of the pathogenesis of polycystic ovary  
420 syndrome (PCOS).

421 The study highlights the efficacy of "Ayurved" which is an ancient tradition, used in some  
422 parts of India. This ancient concept should be carefully evaluated in the light of modern  
423 medical science and can be utilized partially if found suitable.

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<sup>15</sup> 2023/JPRI/102854 : Exploration of Physicochemical, Phytochemical, and HPTLC Analysis of Arputha Mathirai: A Siddha Herbo Mineral Formulation for Poly Cystic Ovarian Syndrome (PCOS)

UNDER PEER REVIEW