

Surveillance of Mycotoxin contamination and Production of Aflatoxin by *A. flavus* in Contaminated Maize Seeds in Bihar

Abstract

Aflatoxin-producing fungi like *Aspergillus flavus* contaminate maize crops in the agricultural field at harvest, post-harvest, and during storage making them one of the most widespread and dangerous mycotoxins. It has been directly correlated to adverse health effects, such as liver cancer in many animal species as well as plant systems.

Maize samples were collected from seven districts of Bihar viz, Begusari, East Champaran, Khagaria, Bhagalpur, Saharsa, Madhepura, and Samastipur. The collected sample was examined for associated mycoflora and aflatoxin-producing prospective of *Aspergillus flavus*. It was determined that *A. flavus* was of presiding occurrence accompanied by *Penicillium* spp., *Fusarium*, *A. nigar*, *Rhizopus*, *Trichoderma*, *Mucor* and *A. ochraceus*. 119 strains of the total *A. flavus* isolates were toxigenic, producing aflatoxin B1, B2, and G1. The highest Aflatoxin is all probability due to complementary environmental conditions, undeveloped agricultural practices, poor storage circumstances of grains, and because of yearly flood problems in this region.

Key words: Maize seeds, Aflatoxin, Mycoflora, *Aspergillus*.

INTRODUCTION

Mycotoxin (Greek word Mykes = Fungus and Toxin = Toxin) is a toxic secondary metabolite produced by an organism of Kingdom Fungi (Richard JL, 2007; Zain, 2011; Cimen et. al., 2022) and is capable of causing disease and death in humans and animals (Bennett, J. W and Klick, M. 2003). They are stable, invisible, and toxic chemical compounds, found to be common in the farm environment surviving in several places as well as on many different types of feedstuffs.

The growing moulds and production of mycotoxin are most encouraged by environmental factors like temperature (cool/hot) and moisture (wet/dry) (Janse van Rensburg et al. 2015; Vaughan et al. 2014). However, mycotoxins may also be produced by moulds when other stress conditions occur to the host plant or the mould.

Aflatoxins are a type of mycotoxin produced by *Aspergillus* spp. such as *A. flavus* and *A. parasiticus* (Deiner et. al., 1987; Kurtzman et. al., 1987; Martins et. al., 2001; Reddy et. al., 2009). The umbrella term aflatoxin refers to four different compounds which are B1, B2, G1, and G2 (Yin YN et. al., 2008; Abbas et. al., 2010) where Aflatoxin B1, the most toxic, is a potent carcinogen and has been directly correlated to adverse health effects, such as Liver cancer in many animals as well as plant system (Martins et. al., 2001; Ragni K and Prasad G., 2022a & b). Aflatoxin is largely associated with commodities produced in the tropics and subtropics region in cotton, wheat, millet, spices, rice, sorghum, peanuts, sunflower, pistachios, and maize crops (Martins et. al., 2001; Yin YN et. al., 2008).

Maize is widely cultivated throughout the world and a greater weight of maize is produced each year than any other grain (International grains council, 2013). In 2021, total world production was 1.2 billion tonnes. Maize is

the most widely grown grain crop throughout America. Out of 38 administrative districts of Bihar only seven districts, viz. Begusarai, East Champaran, Khagaria, Bhagalpur, Saharsa, Madhepura, and Samastipur constitute about half of the total maize acreage. These districts are historically flood-prone areas during the rainy season and fall north of the Ganges, having several seasonal river tributaries.

These seven districts of Bihar have recorded in April 2021, the highest maize productivity in the world. Maize cultivation provides livelihood to approx. 1.3 million farmers in Bihar state. Maize has become a staple food in many parts of the world with the total production surpassing that of wheat or rice. In addition to being consumed directly by humans (often in the form of masal). Maize is used for corn ethanol, animal feed, and other products, such as corn starch and corn syrup (Foley, J. 2019). It is also used in making ethanol and other biofuels.

Therefore, an attempt has been made to surveillance of mycotoxigenic fungi and the production of aflatoxin by *A. flavus* in contaminated maize seeds of major growing districts of Bihar.

Occurrence of mycotoxin in maize crop

Aflatoxin-producing fungi like *A. flavus* contaminate maize crops in the agricultural field at harvest and during storage making them one of the most widespread and dangerous mycotoxins. In Kenya (2004), about 125 people died, and nearly approx. 200 others required medical treatment after eating aflatoxin-contaminated maize (Lewis L. et. al., 2005). The death was associated with home-grown maize that had not been treated with fungicides or properly dried before storage. At that time, due to food shortages, farmers may have been harvesting maize earlier than normal to prevent thefts from their agricultural fields, so the grain had not fully matured and was more susceptible to infection with *A. flavus*.

MATERIALS AND METHODS

Sample collection

Maize seed samples were collected from a farmer's field in seven districts viz. Begusarai, East champaran, Khagaria, Bhagalpur, Saharsa, Madhepura and Samastipur of Bihar. Maize seed samples were kept in sterile polythene bags for further experiments. The moisture content of each sample was recorded with the help of an OSAW moisture meter and pH readings were taken by using a digital pH meter (IJBAF, 2013).

Isolation and identification of Mycoflora

100 kernels of all seven- sampling site (districts) was surface sterilized in 2% NaOCl and plated on moist blotting paper in sterile Petri dishes (ISTA, 1966). Plates were incubated at room temperature for 7 days followed by macro and microscopic identification (Mc clenny, N. 2005; Adame- Garcia, J. et. al., 2015). Fungal colonies were maintained on PDA media for further use and identification.

Aflatoxin analysis of toxigenic isolates

Aspergillus flavus isolates were allowed to grow on SMKY liquid media (Diener and Davis, 1966) for one week at $30 \pm 2^\circ\text{C}$ and aflatoxin was extracted (Thomas et. al., 1975). It was extracted with chloroform, and dried in a water bath and concentrated extracts were kept in screw-tight bottles for qualitative and quantitative analysis.

Qualitative and Quantitative analysis

Qualitative analysis of aflatoxin was done using TLC (Thin layer chromatography), toluene- isoamyl alcohol-methanol (90: 32: 2 v/v) solvent system (Reddy et. al., 1970), and for chemical conformation trifluoroacetic acid (Stack and Pohland, 1975) or 25% sulphuric acid spray was used.

Quantitative estimation of aflatoxin was done spectrophotometrically (Nabney and Nesbitt, 1965).

RESULTS

Table -1 and fig. 1, indicates the association of mycoflora in seven districts of Bihar in flood-prone areas during the rainy season (Agroclimatic zone in Fig- 2) and observed during the study, moisture content, and pH of the sample. *A. flavus*, *Penicillium* spp., *Fusarium*, *A. nigar*, and *Rhizopus* were present in all the seven districts in large amounts whereas *Trichoderma*, *Mucor*, and *A. ochraceus* were present in a rare amount of all districts but absent in Khagaria and Saharsa districts, respectively (Fig- 3). pH ranged between 5.8 to 6.7 and moisture content was recorded at 10.8, 10.5, 10.0, 9.6, 10.1, 10.4, and 10.7 for Begusarai, East Champaran, Khagaria, Bhagalpur, Saharsa, Madhepura, and Samastipur, respectively.

Table-2 represents that *A. flavus* sample of 7 districts having 385 isolates of *A. flavus*. 119 strains were found to be toxigenic with which 86 positives to AFT- B1, 24 positives to AFT-B1B2, and only 9 positives for AFT-B1B2G1. The amount of AFT-B1 was, however very low at 0.2- 1.3, 0.0- 1.0, and 0.0- 1.1 $\mu\text{g/ml}$, respectively. Whenever, the maximum aflatoxin was recorded from Begusarai samples at 0.2- 1.8 $\mu\text{g/ml}$ followed by East Champaran, Khagaria, Saharsa, Bhagalpur, Samastipur, and Madhepura districts, respectively.

Maize samples collected from seven districts in flood-prone areas during the rainy season were highly contaminated with *A. flavus* and other genera of fungi like, *Penicillium*, *Fusarium*, *A. nigar*, *Rhizopus*, *Trichoderma*, *Mucor*, and *A. ochraceus*. Mycotoxin-producing fungi like *Aspergillus* and *Fusarium* spp. were of predominant occurrence (Fig. 4 and 5). The moisture contents were also observed in all the samples which influence the aflatoxin production.

Table -1: Mycoflora associated with Maize Seeds collected from seven Districts of Bihar

| Sl. No. | Fungus | Maize Seed Samples from Seven District | | | | | | | | | | | | | | | | | | | | |
|---------|-------------------------|--|------------------|-----|------------------------|------------------|-----|----------------------|------------------|-----|----------------------|------------------|-----|----------------------|------------------|-----|----------------------|------------------|-----|----------------------|------------------|-----|
| | | Begusari (N=100) | | | East Champaran (N=100) | | | Khagaria (N=100) | | | Bhagalpur (N=100) | | | Saharsa (N=100) | | | Madhepura (N=100) | | | Samstipur (N=100) | | |
| | | No. of infested seed | Moisture content | PH | No. of infested seed | Moisture content | PH | No. of infested seed | Moisture content | PH | No. of infested seed | Moisture content | PH | No. of infested seed | Moisture content | PH | No. of infested seed | Moisture content | PH | No. of infested seed | Moisture content | PH |
| 1 | <i>A. flavus</i> | 75 | 10.8 | 6.7 | 66 | 10.5 | 6.3 | 63 | 10.0 | 6.0 | 57 | 9.6 | 5.8 | 46 | 10.1 | 5.9 | 41 | 10.4 | 6.2 | 37 | 10.7 | 6.5 |
| 2 | <i>Penicillium Spp.</i> | 68 | | | 60 | | | 57 | | | 49 | | | 42 | | | 38 | | | 33 | | |
| 3 | <i>Fusarium Spp.</i> | 62 | | | 58 | | | 43 | | | 34 | | | 30 | | | 28 | | | 25 | | |
| 4 | <i>A. nigar</i> | 74 | | | 65 | | | 62 | | | 56 | | | 45 | | | 40 | | | 36 | | |
| 5 | <i>Rhizopus Spp.</i> | 35 | | | 32 | | | 24 | | | 21 | | | 19 | | | 16 | | | 13 | | |
| 6 | <i>Trichoderma</i> | 9 | | | 5 | | | 0 | | | 3 | | | 6 | | | 9 | | | 4 | | |
| 7 | <i>Mucor</i> | 4 | | | 9 | | | 1 | | | 7 | | | 0 | | | 1 | | | 5 | | |
| 8 | <i>A. ochraceus</i> | 5 | | | 8 | | | 0 | | | 4 | | | 7 | | | 2 | | | 1 | | |

Table- 2: *Aspergillus flavus* isolates from Maize Seed samples

| Sl. No. | Districts of Bihar | No. of <i>A. flavus</i> strains isolates | No. of toxigenic isolates of <i>A. flavus</i> | Positive isolates | | | Range of aflatoxin B ₁ concentration µg/ml (ppm) |
|---------|--------------------|--|---|-------------------|-------------------------------|--|---|
| | | | | B ₁ | B ₁ B ₂ | B ₁ B ₂ G ₁ | |
| 1 | Begusari | 75 | 25 | 15 | 7 | 3 | 0.2- 1.8 |
| 2 | East Champaran | 66 | 19 | 13 | 5 | 1 | 0.1- 1.6 |
| 3 | Khagaria | 63 | 18 | 12 | 4 | 2 | 0.2- 1.4 |
| 4 | Bhagalpur | 57 | 13 | 11 | 1 | 1 | 0.2- 1.3 |
| 5 | Saharsa | 46 | 17 | 14 | 2 | 1 | 0.0- 1.2 |
| 6 | Madhepura | 41 | 12 | 10 | 2 | 0 | 0.0- 1.0 |
| 7 | Samstipur | 37 | 15 | 11 | 3 | 1 | 0.0- 1.1 |
| | Total | 385 | 119 | 86 | 24 | 9 | - |

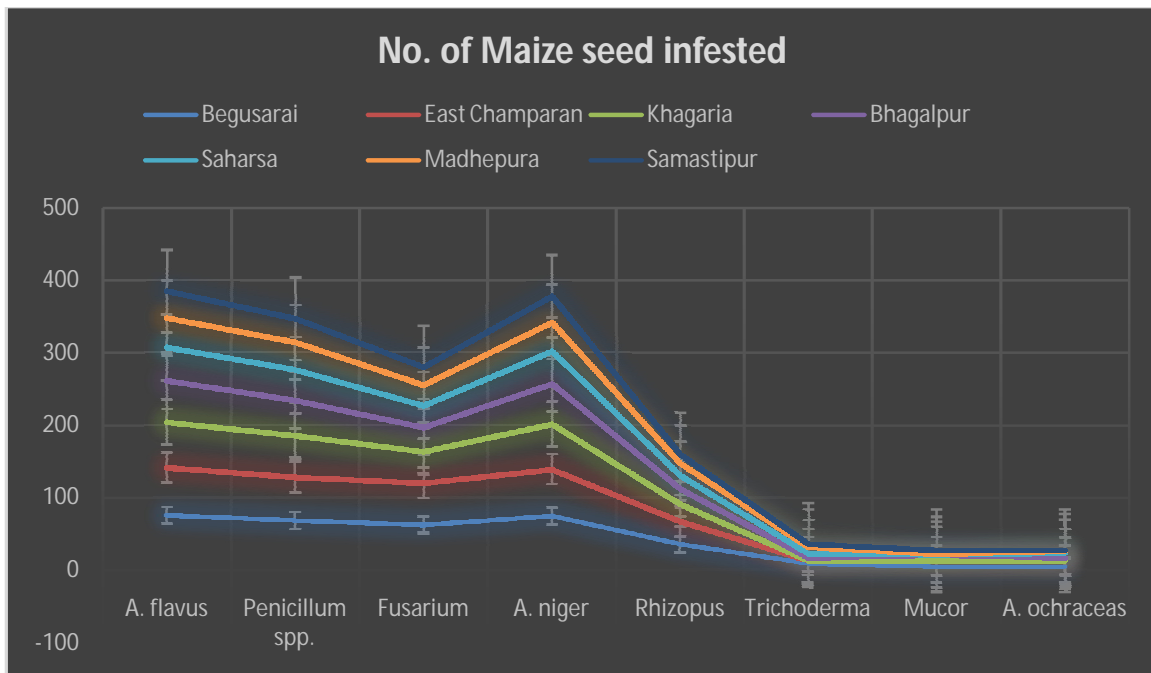


Fig- 1: Line showing No. of Maize Seed infested in Seven Districts of Bihar

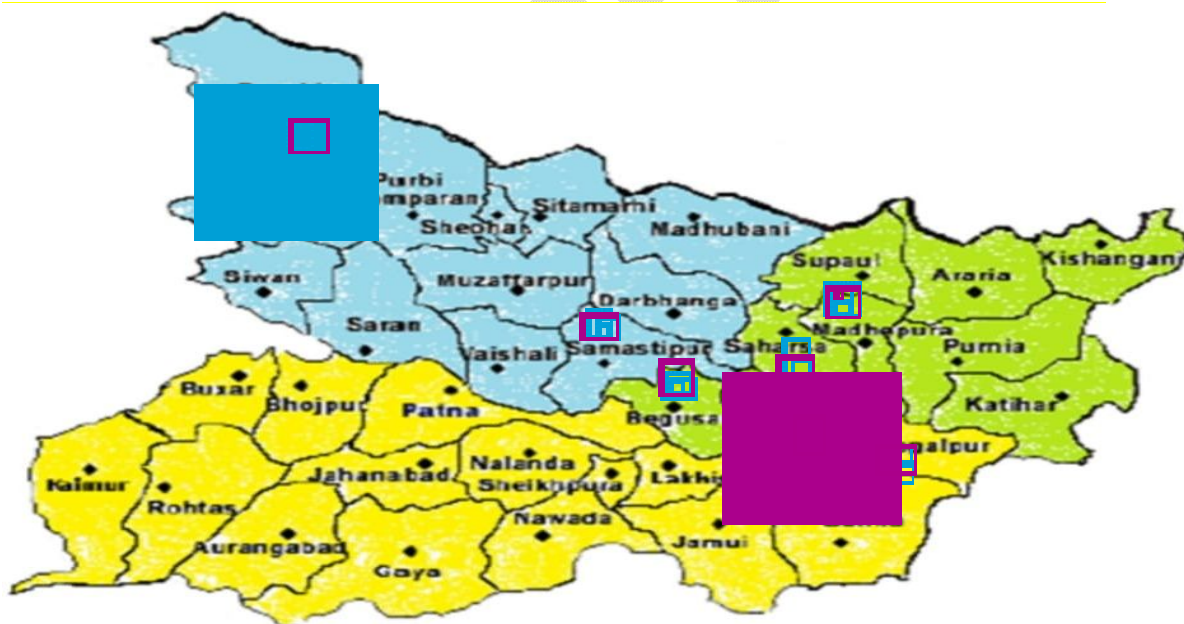
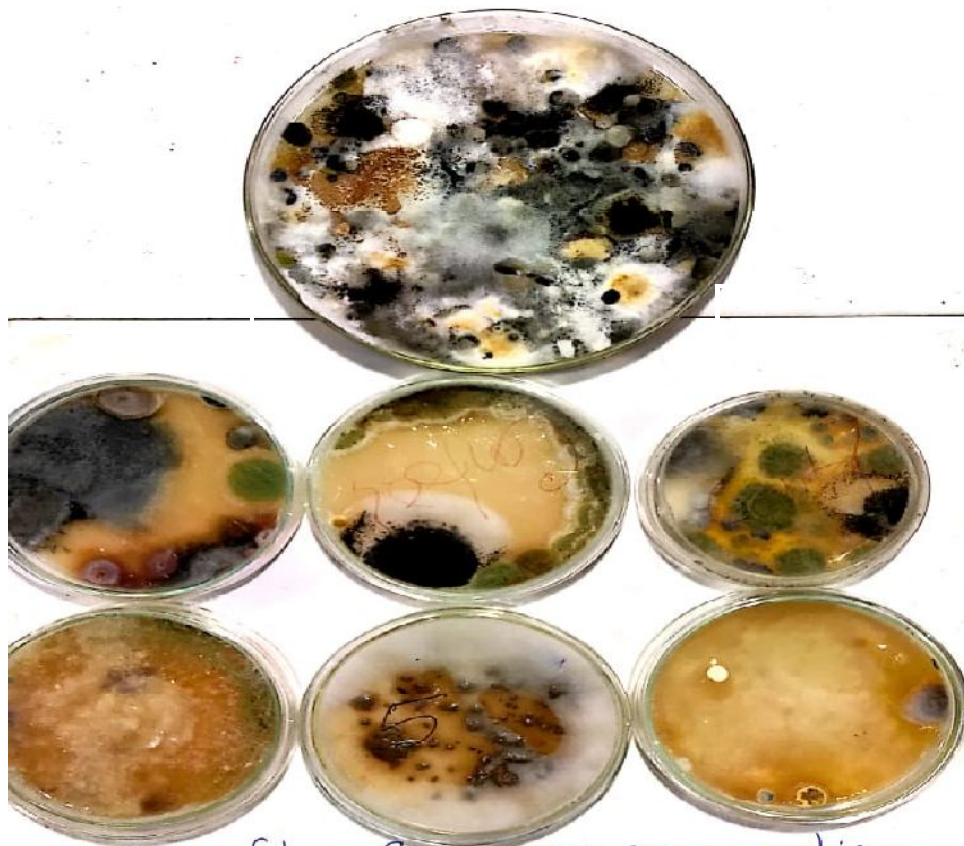


Fig 2: Pink dot indicates Agroclimatic Zone of Seven Districts of Bihar



Mycoflora Grown on PDA-Media
on Various Districts - Bihar.

Fig 3: Association of Mycoflora in PDA media of Seven Districts of Bihar

No. of toxigenic isolates of *A. flavus*

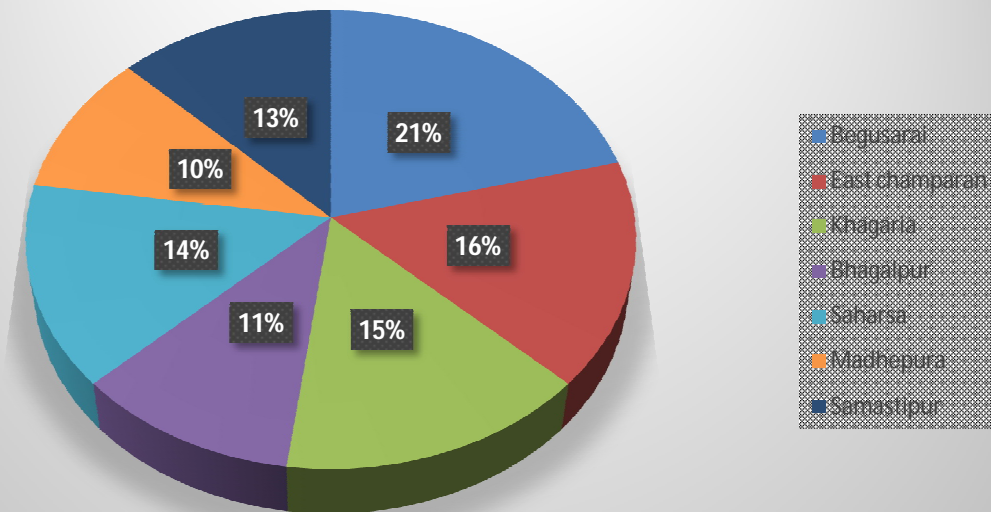


Fig- 4: Pie chart showing No. of toxigenic isolates of *A. flavus* (%) in Seven District of Bihar



Fig- 5: Association of Mycoflora in maize comb during flood situation

DISCUSSION

Recently, about 125 people died, and nearly approx. 200 others required medical treatment after eating aflatoxin-contaminated maize (Lewis L. et. al., 2005). In Kenya (2004), the death was associated with homegrown maize that had not been treated with fungicides or properly dried before storage. At that time, due to food shortages, farmers may have been harvesting maize earlier than normal to prevent thefts from their agricultural field, so the grain had not fully matured and was more susceptible to infection with *A. flavus* and aflatoxin-producing potentiality of toxigenic strains of *A. flavus* were higher, due to moisture content in rainy season in a flooded area as well as poor storage conditions that provide an opportunity for fungal growth like Aflatoxin to easily invade the maize seeds.

Bihar has experienced serious aflatoxicosis outbreaks associated with maize which has claimed lives as well as maize yield losses. Our findings from the various sites (districts) revealed that potentially mycotoxigenic fungal isolates were found on maize samples. Maize samples from each district were more infested by a specific fungal genus. Maize grains from Begusarai were heavily contaminated by *Aspergillus* spp. while those collected from Khagaria and Saharsa were not contaminated with *Trichoderma* and *Mucor*.

CONCLUSION

Maize samples from the seven districts tested were infested by different mycotoxigenic fungi. The existence of mould on the maize samples shows the possibility of the occurrence of more than one mycotoxin but dominant by Aflatoxin B1. The maize has the potential to enhance the income of 1.3 million maize growers in Bihar State, thereby significantly reducing the poverty of the poorest states in India.

So, it was important to determine the distribution and incidence of fungi that exist in maize from different districts.

REFERENCES

1. Abbas HK, Reddy KRN, Salleh B, Saad B, Abel CA and Shier WT (2010). An overview of mycotoxin contamination in foods and its implications for human health. *Toxin Reviews* 29: 3-26.
2. Adame- Garcia, J. Rodriguez- Guerra, R. Iglesias- Andreu, LG. Ramos- Prado, JM. Luna- Rodriguez, M. (2015). Molecular identification and Pathogenic variation of *Fusarium* species isolated from vanilla planifolia in Papantla Mexico. *Botanical Sciences* 93 (3): 669-678.
3. Bennett, J.W; Klich, M (2003). Mycotoxins. *Clinical microbiology Reviews*. 16 (3). 497-516.
4. Cimen, Duygu, Bereli, Nilay, Denizli, Adil (2022). "Patulin Imprinted Nanoparticles Decorated surface Plasmon Resonance chips for Patulin Detection". *Photonic Sensors*. 12 (2): 117-129.
5. Diener, U.L. and Davis, N. D. (1966). Aflatoxin production by isolates of *Aspergillus flavus*. *Phytopathology*, 56, 1390-1393.
6. Deiner UL, Cole RJ, Sanders TH, Payne GA, Lee LS and Klich MA (1987). Epidemiology of aflatoxin formation by *A. flavus*. *Annual Review of Phytopathology* 25: 240-270.
7. Foley Jonathon (2019). "Its Time to Rethink Americas corn sys. *Scientific American*. Retrieved. Feb 15, 2019.
8. International grains council (International organization 2013). International grains council market Report 28 Nov 2013.
9. IJBAF (2013). Mycoflora and aflatoxin contamination of some foodstuffs. **ORANUSI SU AND OLAREWAJU SA*. 1 (1): 9-18.
10. Janse van Rensburg, B., McLaren, N.W., Flett, B.C. and Schoeman, A. (2015). Fumonisin producing *Fusarium* spp. and fumonisin contamination in commercial south African maize. *European Journal of plant Pathology*. 141: 491-504.
11. Kurtzman CP, Horn BW and Hesseltine C (1987). *Aspergillus nomius*, a new aflatoxin- producing species related to *Aspergillus flavus* and *Aspergillus tamarii*. *Antonie van leauwenhoek* 53: 147-158.
12. Lewis, L. Onsongo, M. Njapau, H. *et. al.*, (2005). "Aflatoxin contamination of commercial maize products during an outbreak of acute aflatoxicosis in eastern and central Kenya. *Environ. Health Perspect*. 113 (12): 1763-67.
13. Liu. Yue, Yamdeu, Joseph Hubert Golani, Gong, Yun Yun, orfila, Caroline (2020). "A review of postharvest approaches to reduce fungal and mycotoxin contamination of foods". *Comprehensive Reviews in Food Science and Food safety*. 19 (4): 1521-1560.
14. Martins, ML. Martins, HM, Bernardo, F. (2001). "Aflatoxins in spices marketed in Portugal". *Food Addit. Contam*. 18 (4): 315-19.
15. Mc Clenny N. (2005). Laboratory detection and identification of *Aspergillus* traditional approach. *Medical Mycology* 43 (Supplement- D): S125- S128.
16. Miller JD. (2008). Mycotoxins in small grains and maize: old problems, new challenges. *Food Additives and Contaminants Part A* 25: 219-230.
17. Nabney, J. and Nesbitt, B. F. (1965). A spectrophotometric method for determining the aflatoxins Analysis. 90: 155-160.
18. Ragni K and Prasad G (2022a). Efficacy of bitter plant extracts (*Adhatoda vasica*) on prevention of aflatoxin B₁ production and reverse the physiology of maize seeds (*zea mays L.*). *International Journal of Scientific Development and Research* 7: 662-668.

19. Ragni K. and Prasad G. (2022b). Efficacy of Bitter plant extracts (*Azadirachta indica*) on Seed Germination, Chlorophyll and Carotenoid Synthesis in Mycotoxin especially Aflatoxin B₁ treated Maize Seeds (*Zea Mays* L.) *Research Journal of Agricultural Sciences An International Journal* 13:1783-1786.
20. Reddy, T.V. Viswanathan, L. and Venkita subramania, J.A. (1970). Thin layer chromatography of aflatoxins. *Analytical Biochemistry*, 38: 568-571.
21. Reddy, K.R.N., Reddy, C.S. and Muralidharan, K. (2009). Detection of *Aspergillus* spp. and aflatoxin B₁ in rice in India. *Food Microbiology* 26: 27-31.
22. Richard, JL (2007). "Some major mycotoxins and their mycotoxicosis- an overview". *Int. J. Food microbiol.* 119 (1-2): 3-10.
23. Stack, M.E. and Pohland, A.E. (1975). Collaborative study of a method for chemical conformation of identity of aflatoxin. *J. Assoc. off. Anal. Chem*, 58:110-113.
24. Thomas, F. Eppley, RM, Trucksess, MW. (1975). Rapid screening method for aflatoxins and zearalenone in corn. *Journal of the Association of official Analytical chemists.* 58 (1): 114-116.
25. Vaughan, M.M., Huffaker, A., Schmelz, E. A., Dafoe, N. J., Christensen, S., Sims, J., Martins, V.F., Swerbilow, J., Romero, M. and Alborn, H.T. (2014). Effects of elevated (CO₂) on maize defence against mycotoxigenic *Fusarium verticillioides*. *Plant, Cell and Environment.* 37: 2691-2706.
26. Yin, YN. Yan, LY. Jiang, JH. Ma, ZH (2008). "Biological control of aflatoxin contamination of crops". *J. Zhejiang univ. sci. B.* 9 (10): 787-92.
27. Zain, M.E. (2011). Impact of mycotoxin on human and animals. *Journal of Saudi Chemical Society.* 15: 129-144.