Original Research Article

Mycoremediation of Heavy Metals from Electronic Waste Polluted Water Using Indigenous Fungi and Its Implications

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ABSTRACT

Background: Heavy metals (HM) pollutants are crucial environmental and public health problems due to their toxicity, which has implications on public health.The site of study has been reported to be densely polluted with heavy metals as a result of dumping used electronic wastes into the water body.

Aim: This study was carried out to determine the biosorption potentials of indigenous fungi isolates in reducing heavy metal present in electronic wastes polluted water body.

Study Design: This was a laboratory based study

Place and Duration of Study: The study was conducted at the Microbial Resources Research Laboratory, Department of Pure and Applied Biology, Ladoke Akintola University of Technology Ogbomoso, Nigeria from July 2021 to June 2022.

Methodology: Heavy metal polluted water sample was collected near the dumpsite of e-waste in Alaba International market, Lagos Nigeria**.**Fungi were isolated from the polluted water sample by carrying out serial dilution. Pure colonies were obtained and stored at 4 ^{o}C. Media formulations (MF) trials for the biosorption process was achieved using brewery waste and honeycomb extracts. Exactly 200 ml of MF were dispensed into a 500 ml Elmeryer flask containing 100 ml of the e-waste polluted water and fugal discs.Atomic absorption spectrophotometer was used to determine the heavy metal concentration in the water samples. The biological interactions of the fungi with the polluted water sample was monitored using Fourier transform infrared (FTIR) spectroscopy. The pH, electrical conductivity and other physicochemical parameters were also determined.

Results:*Trichoderma harzianum, T. viride*, *Fusarium oxysporum, Aspergillus flavus*and *A. niger*were isolated from the heavy metal polluted water samples. Heavy metal such as Pb (13.30 mg/L), Cd (16.50 mg/L), Cr (6.41 mg/L), Ni (3.81 mg/L), Zn (8.85 mg/L), Cu (8.33 mg/L), Fe (5.60 mg/L) had values which were higher than the acceptable limits. Biosorption efficiency of each of the fungus in reducing the metals present in the sample was in the increasing order of *A. niger<A. flavus<F. oxysporum<T. viride<T. harzianum*. FTIR showed that some peaks were shifted to lower wavenumber as a result of the interactions of the fungus with the heavy metal in the water sample. This study revealed that *T. harzianum* hadthe highest biosorption efficiency for the removal of heavy metal in the polluted water sample.

Conclusion: It is essential that heavy metals be removed from polluted water body to avoid its deleterious effects on public health. This can be achieved biologically using any of the fungi isolated in this work.

Keywords: E-waste polluted water, Heavy metal, Fungi, FTIR

1. INTRODUCTION

Environmental pollutioncould arise fromindustrialization and mining of natural resources leading to the contamination of soil, air, sediments and surface water with heavy metals [1]. Apart from these, human activities also lead to situations in which heavy metals are transformed into new compounds and may be spread worldwide [2]. Hence, there is a need for new techniques that focuses on the removal of the pollutants instead of the conventional methods of disposal that results in environmental pollution [3,4] Heavy metals can be divided into three based on their physiological effects and toxicity as:low toxicity (Iron, Molybdenum, Manganese), average toxicity (Zinc, Nickel, Copper, Vanadium, Cobalt, Tungsten, Chromium) and high toxicity (Arsenic, Silver, Antimony, Cadmium, Mercury, Lead, Uranium) [5].

Metal collections such as Lead, Cadmium, Chromium, Copper, Cobalt, Mercury and Nickel as well as Zinc generate dangerous effects on human health when amounts that cannot be processed are absorbed. However, heavy metals, such as cadmium were not known to be dangerous until the early 19th century [2].When found in the human body system, they result in severe health problems which hamper normal functions [6].Chronic exposure occurs which refers to contact with low levels of heavy metal over a long period [2]. The toxicity effects greatly depend on the concentrationof the contaminant present in the environment in the form(s) that can be assimilated by the organism [7].Exposure for a long duration and heavy metals accumulation often leads to dangerous health consequences on aquatic and human life, causing acute and chronic effects [8, 9].

There is a short life span of electronic products as a result of higher demands for newer and more efficient technologies leading to fast production by the manufacturing industry. This leads to the short life span of electronics making them becomes outdated at a swiftly escalating rate around the world [10, 11].As a result of this, they are being dumped into the environment by the users. These have the potential to release heavy metals into water bodies and the environment at large which needs to be removed. Heavy metal removal techniques involves expensive methodologies due to high energy and reagent requirements [12].Numerous fresh approaches to cleaning the surroundings are being developed and many are already in use as an option to physical methods which are relatively too expensive [13].

Improvement in scientific methods and industrial technology has led to the utilization of intrinsic traits of natural resources which include microorganisms, to swiftly bio-absorb harmful inorganic contaminants to safe levels in water and soils [13]. The safe levels are as determined by environmental health regulatory bodies to overcome environmental damages due to pollution [14].The site of study has been reported to be densely polluted with heavy metals as a result of dumping used electronic wastes into the water body. These releases their heavy metals into the water body.

This research aims to determine the bio-sorption potentials of five fungal species on toxic metals present in electronic waste (e-waste) polluted water using agro-waste formulated media.

2. MATERIALS AND METHODS

2.2 Description of the Study Site

The site of the study was Alaba International Market, Ojo, Lagos State with a Latitude of 6°23'N and a Longitude of 2°42'E (Figure 1). This market is known for the sales of new and used electronic devices. Around the market are residential quarters and illegal dumpsites which consist of abandoned and outdated electronics which are dismantled and discarded for crude recycling. Thus, leaving the rest of the fragments to be burnt (Plate 1) to reduce the waste volume, which tends to contaminate streams and well waters in the neighbourhood located near the e-waste open dumpsites. Wells and Streams which served the purpose of cooking, drinking and ablution as well as other domestic and commercial purposes by the inhabitants and workers in the area are situated very close to the e-waste open dumpsites. Contamination gets to the wells through percolation to the ground water

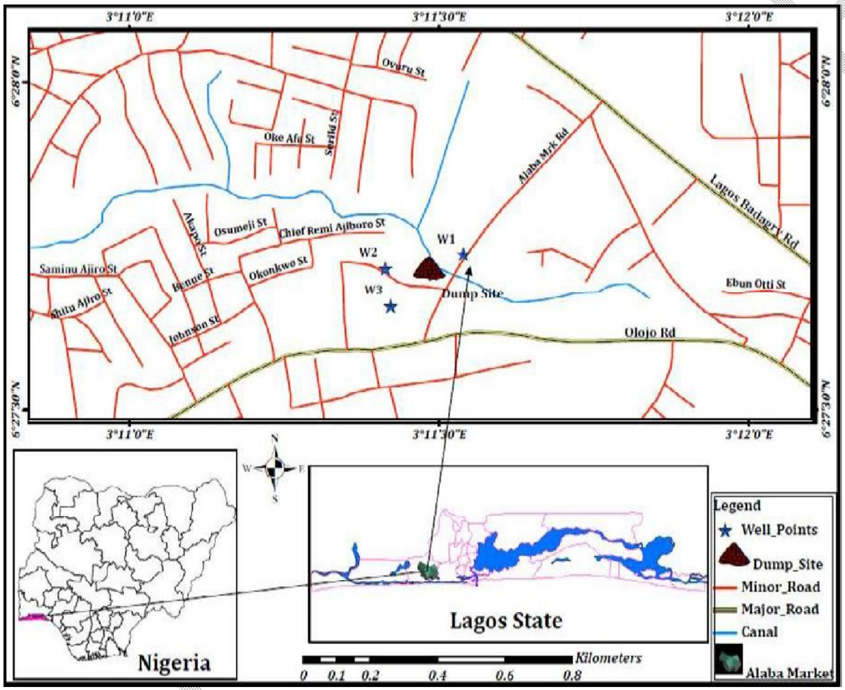

Fig. 1 Geographical View of Alaba International Electronics Market e-waste Dumpsite

Plate 1: Activities at the e-waste Recycling Sites Where Samples were collected

2.2 Collection of Electronic Wastewater

Samples were collected from electronic waste polluted water site (EWPWS) near the dumpsite of e-waste in Alaba International market using a clean and sterilized plastic container. The samples were stored in sterile bottles pre-washed with HNO_3 and rinsed thoroughly with deionized water. All samples were maintained at 4 °C in an ice box during transportation from the collection site to the laboratory [15]

2.3 Isolation and Morphological Identification of Fungi

Serial dilution was performed on the water samples by introducing 1ml sample into 9ml of distilled water until diluents of 10^{-1} to $10^{-5}$was achieved in separate test tubes. 1ml of different concentrations of the diluents were introduced into separate plates in the pour plate method, then potato dextrose agar was added to the plates, and the plate was swirled gently and allowed to solidify. Plates were incubated at $30\pm2^{o}C$ for 72 hours. Different colonies of each of the organisms were sub-cultured into newly prepared Potato dextrose agar plates and incubated as mentioned earlier. Pure culture of each of the organisms was obtained and then stored at 4℃ [16].Morphological Identification of the Isolated fungi was performed using a bright field binocular Microscope. The cellular morphology and microscopic features of each of the isolates were observed and recorded [17,18]

2.5 Medium Formulation Trials (MFT)

Brewery waste and honeycomb extracts were used for the media formulation trials (MFT) for the biosorption processes. A series of trial tests (1:1:1, 1:1:1/2, 1:1:1¼, etc) were performed to know the optimum volume of brewery waste extract, honeycomb extract and polluted water required for the process respectively.

2.6 Heavy Metals Biosorption from e-waste Polluted Water

For the biosorption process, the best media trial concentrations were used according to a modified method [19, 20]. Biosorption of heavy metals was done in ten flasks containing 200 ml of the e-waste polluted water, and 100 ml each of brewery waste and honeycomb extracts. The flasks were first sterilized using an autoclave and allowed to cool down. Thereafter, five fungal discs for each of the isolates were introduced into the medium, while the control was without fungi discs.

The experiment was carried out for 28 days and incubated in a shaker incubator at 25 ^{o}C.A Same sample volume were taken from the experimental set-ups for analysis using Atomic absorption spectrophotometry (AAS) for each of the heavy metals (Pb, Cd, Cr, Ni, Zn, Cu, Fe) at seven days intervals throughout the incubation period using a modified method [19]. The percentage heavy metal (HM) removal was calculated using the equation below. Experiments were performed in triplicates while the average mean value was calculated as (X±SEM).

$$\% \, HM\ Removal = \frac{\text{HM (untreated water sample)} - \text{HM (treated water sample)}}{\text{HM (untreated water sample)}} \qquad \ldots\ldots\ldots\ 1.1$$

2.7 Physicochemical Analysis of e-waste Sample

The following physicochemical parameters such as pH, Electrical conductivity (EC), Total solid (TS), Dissolved oxygen (DO),Total suspended solids (TSS), Biological oxygen demand (BOD), Carbon oxygen demand (COD), Sulphate, Nitrate, Iron (Fe), Lead (Pb), Cadmium (Cd), Chromium (Cr), Copper (Cu), Zinc (Zn), Nickel (Ni), were determined for each of the experimental set-ups [21].This was done to know the pre-and post physicochemical properties of the water sample.

2.8 Heavy Metals Analysis of the e-waste Polluted Water

To determine the heavy metal concentration 100 ml of the samples were introduced into Pyrex beakers containing 10 ml of concentrated HNO_3. It was gently heated on a hot plate to reduce the volume and to discharge metals that are organic bounded, particulate or adsorbed on particulates. The beakers were allowed to cool. Thereafter another 5 ml of Conc. HNO_3 was added. It was heated again with the addition of Conc. HNO_3, as required till digestion, was completed.

The samples were finally evaporated to dryness; followed by the addition of 5 ml of HCl solution (1:1 v/v) and heated again. The addition of 5 ml of 5 M NaOH was done, then filtered. The filtrates were decanted into 100 ml volumetric flasks and diluted to the mark with distilled water. The digested samples were analyzed using a Buck Scientific Flame Atomic Absorption Spectrophotometer Thermo M5 Model [22, 23] in duplicate for the determination of heavy metals and the mean result was recorded. Fourier transform infrared (FTIR) was used to monitor the biological interactions of the fungi with the heavy metal polluted water sample.

2.9 Statistical Analysis

Data collected were arranged using Excel spreadsheet 2010 and results were presented as percentages. Statistical Package for Social Sciences (SPSS) software for Windows (V. 21.0) was used for the data analysis. Significant differences between the means were determined and P values < 0.05 were regarded as significant.

3. RESULTS

The result of the physicochemical parameters of the effluent shows that it has a pH of 4.5. The metals present include Lead (13.30 mg/L), Cadmium (16.50 mg/L), Chromium (6.41 mg/L), Nickel (3.81mg/L), Zinc (8.85 mg/L), Copper (8.33 mg/L), Manganese (3.65 mg/L), Sodium (150 mg/L), Potassium (18.55 mg/L), Calcium (210 mg/L), Magnesium (30 mg/L), Iron (5.60 mg/L). Moreover, the result of showed that it contains TS (550 mg/L), TSS (25.9mg/L), DO (2.5 mg/L), BOD (18.33 mg/L), COD (33.15 mg/L), Phosphate (11.33 mg/L), Sulphate (580 mg/L), Nitrate (65.30 mg/L) and EC (134.35 μs/cm) as seen in Table 1

Table 1. Physico-chemical analysis of e-waste polluted water

Parameters	Value (mg/L)
pH	6.8
Pb	13.3
Cd	16.5
Cr	6.41
Ni	3.81
Zn	8.85
Cu	8.33
Mn	3.65
Na	4.65
K	18.55
Ca	6.85
Mg	7.2
Fe	5.6
Total solid	55
Total suspended solids	25.9
Turbidity	very turbid
Dissolved Oxygen	6.8
Biochemical Oxygen demand	18.33
Chemical Oxygen demand	33.15

Phosphate	11.35
Sulphate	13.7
Nitrate	2.32
Electrical conductivity (µs/cm)	134.35

3.1 Isolation, Identification and Characterization of Isolates

A total of five (5) fungal species were isolated and identified to be: *Trichoderma harzianum*, *T. viride*, *Aspergillus niger*, *A. flavus* and *Fusarium oxysporum*. The cellular morphological description is presented in Table 2 below.

Table 2. Morphological Description of the Isolated Fungi used for the Biosorption Studies

Fungal Identity	Description
Trichoderma spp. AG1	Colony at 20°C attained a diameter of 4.5-7.5 cm in 5 days. Initially more or less hyaline, later becoming whitish green in tufted conidia areas in blue-green shades. Reverse colourless. Conidiophores pyramidally branched.
Trichoderma spp. AG2	Colony at 20°C attaining a diameter of more than 9 cm in 5 days, more or less hyaline, whitish-green becoming olive-green in tufted conidia areas. Reverse colourless. Conidiophores pyramidally branched. Chlamydospores were present in mycelia of older colonies, intercalary, sometimes terminal, mostly globose, hyaline and smooth-walled.
Aspergillus spp. AG1	Colonies at 25°C attained a diameter of 4-5 cm within 7 days. Consist of a compact white or yellow basal felt having a dense layer of dark brown to black conidiophores. Vesicles globose to sub-globose.
Aspergillus spp. AG2	Colonies at 25°C attained a diameter of 3-5 cm within 7 days. Consist of a dense felt of yellow-green conidiophores. Colonies on MEA are thinner but sporulating densely.
Fusarium spp.	It is white often with a peach tinge. Aerial mycelium floccose, whitish or peach which often changes to brownish (14-21 days). Sporodochia is absent. Conidia 3-5 cm within 7 days. Septate fusiform straight or somewhat curved, wedged shaped, apical cell beaked.

3.2 Biosorption Studies of Heavy Metals Present in the e-waste Samples

3.2.1 Percentage Removal of Lead

The lowest biosorption value of lead by *A. niger*was recorded on day 7 (18.33 %), while the highest biosorption value was recorded on day 28 (70.34 %). For lead, the lowest biosorption by *A. flavus*was observed on day 7 (16.37 %), while the highest biosorption was recorded on day 28 (65.79 %). Also, the lowest biosorption of lead by *T. viride* was on day 7 (24.78 %), while the highest biosorption was found on day 28 (71.16 %). *T. harzianum* gave the least biosorption of lead on day 7 (23.21 %), while the highest biosorption was recorded on day 28 (84.35 %). *F. oxysporum* recorded the least biosorption of lead on day 7 (20.69 %), while the highest biosorption was recorded on day 28 (71.25 %).

Moreover, *A. flavus* had the lowest biosorption on day 7 (16.37 %) when compared with other organisms while *T. viride* gave the highest biosorption on day 7 (24.78 %). On day 14, *A. flavus* had the lowest biosorption (21.56 %), while *T. viride* gave the highest biosorption (40.47 %). *A. flavus* recorded the least biosorption on day 21 (34.23 %), while the highest biosorption was recorded on day 21 by *T. harzianum* (56.12 %). *A. flavus* recorded the least biosorption on day 28 (65.79 %), while the highest biosorption was recorded on day 28 by *T. harzianum* (84.35 %) as found in Table 3.

Generally, lead biosorption increased as the time extended, where the highest biosorption percentages were observed at the 28th day of the experiment. *T. viridae* showed the highest biosorption capacities on the 7th and 14th days of treatment, which were 24.78 and 40.47%, respectively. On the other hand, *T. harzianum* recorded the highest biosorption of this metal on the 21st and 28th days, where the values were 56.12 and 84.35%, respectively.

Table 3. Percentage removal of lead (Pb) by different fungal isolates

Organisms/Day	% Biosorption at different days			
	D-7	D-14	D-21	D-28
A. niger	18.33	28.32	43.61	70.34
A. flavus	16.37	21.56	34.23	65.79
T. viride	24.78	40.47	50.67	71.16
T. harzianum	23.21	34.35	56.12	84.35
F. Oxysporum	20.69	26.47	42.36	71.25

3.2.2 Percentage Removal of Cadmium

In Table 4, *A. niger* gave the minimum biosorption value on day 7 (17.02 %), and the maximum value was given on day 28 (66.74 %). *A. flavus* gave the minimum value on day 7 (20.33 %), and the maximum value was given on day 28 (63.88 %). *T. viride* gave the minimum value on day 7 (26.45 %), and the maximum value was given on day 28 (73.18 %). Also, *T. harzianum* gave the minimum value on day 7 (31.10 %), and the maximum value was given on day 28 (80.76 %). *F. oxysporum* gave the minimum value on day 7 (16.95 %), and the maximum value was given on day 28 (65.66 %). However, on day 7, *A. niger* (17.02 %) had the least value while the highest was given by *T. harzianum* (31.10 %). On day 14,*A. flavus* (22.21 %) had the least value while the highest was given by *T. harzianum* (40.78 %). On day 21, *A. niger* (28.88 %) had the least value while the highest was given by *T.*

harzianum (59.80 %). On day 28, *A. flavus* (63.88 %) had the least value while the highest was given by *T. harzianum* at 63. 88 and 80.76 % respectively.

Table 4. Percentage removal of Cadmium (Cd) by different fungal isolates

Organisms/Day	% Biosorption at different days			
	D-7	D-14	D-21	D-28
A. niger	17.02	22.55	28.88	66.74
A. flavus	20.33	22.21	34.54	63.88
T. viride	26.45	36.46	46.91	73.18
T. harzianum	31.10	40.78	59.80	80.76
F. Oxysporum	16.95	23.65	29.64	65.66

3.2.3 Percentage Removal of Chromium

Table 5 showed that *A. niger* recorded the minimum biosorption (22.92 %) on day 7 (D-7) while the maximum value was recorded on day 28 *A. flavus* recorded the minimum value on day 7 (21.22 %) while the maximum value was recorded on day 28 (61.48 %). *T. viride* recorded the minimum value on day 7 (32.91 %) while the maximum value was recorded on day 28 (94.32 %). *T. harzianum* recorded the minimum value on day 7 (45.68 %) while the maximum value was recorded on day 28 (96.30 %). *F. oxysporum* recorded the minimum value on day 7 (31.12 %) while the maximum value was recorded on day 28 (78.77 %). Furthermore, on day 7, *A. flavus* (21.22 %) had the least value while the highest was given by *T. harzianum* (45.68 %). On day 14, *A. niger* (23.74 %) had the least value while the highest was given by *T. harzianum* (49.41 %). On day 21, *A. niger* (36.00 %) had the least value while the highest was given by *T. harzianum* (72.00 %). On day 28, *A. flavus* had the least chromium removal while the highest was recorded by *T. harzianum* at 61.48 and 96.30 % respectively.

Table 5. Percentage removal of Chromium (Cr) by different fungal isolates

Organisms/Day	% Biosorption at different days			
	D-7	D-14	D-21	D-28
A. niger	22.92	23.74	36.00	61.73
A. flavus	21.22	34.58	38.67	61.48
T. viride	32.91	46.25	66.44	94.32
T. harzianum	45.68	49.41	72.00	96.30
F. Oxysporum	31.12	39.53	48.89	78.77

3.2.4 Percentage Removal of Nickel

Table 6 showed that Day 7 recorded the lowest biosorption value (45.15 %) by*A. niger* while the highest value was noticed on day 28 (83.27 %). The lowest value by *A. flavus* was recorded on day 7 (56.29 %) while the highest value was on day 28 (92.49 %). Day 7 (67.14 %) recorded the lowest value by *T. viride* while the highest value was found noticed on 28 (95.22 %). Day 7 (72.29 %) recorded the lowest value by *T. harzianum* while the highest value was on day 28 (94.88 %). Day 7 (58.57 %) recorded the lowest value by *F. oxysporum* while the highest value was recorded on day 28 (89.42 %).Also, on day 7, *A. niger* (45.14 %)

had the least value while the highest was given by *T. harzianum* (72.29 %). On day 14, *A. niger* (45.51 %) had the least value while the highest was given by *T. harzianum* (78.02 %). On day 21, *A. niger* (58.09 %) had the least value while the highest was given by *T. viride* (86.47 %). On day 28, *A. niger* had the least nickel removal (83.27 %) while the highest was recorded by *T. viride* (95.22 %).

Table 6.Percentage removal of Nickel (Ni) by different fungal isolates

Organisms/Day	% Biosorption at different days			
	D-7	D-14	D-21	D-28
A. niger	45.14	45.51	58.09	83.27
A. flavus	56.29	63.78	76.24	92.49
T. viride	67.14	74.92	86.47	95.22
T. harzianum	72.29	78.02	82.51	94.88
F. Oxysporum	58.57	75.54	80.86	89.42

3.2.5 Percentage Removal of Zinc

A. niger recorded the minimum biosorption value on day 7 (26.16 %) while the maximum value was recorded on day 28 (81.97 %). *A. flavus* recorded the minimum value on day 7 (18.81 %) while the maximum value was recorded on day 28 (68.26 %). *T. viride* recorded the minimum value on day 7 (38.02 %) while the maximum value was recorded on day 28 (86.29 %). *T. harzianum* recorded the minimum value on day 7 (54.77 %) while the maximum value was recorded on day 28 (92.10 %). *F. oxysporum* recorded the minimum value on day 7 (30.67 %) while the maximum value was recorded on day 28 (69.75 %).In addition, on day 7, *A. flavus* (18.81 %) had the least value while the highest was given by *T. harzianum* (54.77 %). On day 14, *A. flavus* (27.55 %) had the least value while the highest was given by *T. harzianum* (63.64 %). On day 21, *F. oxysporum* (49.42 %) had the least value while the highest was given by *T. harzianum* (73.55 %). On day 28, *A. flavus* (68.26 %) had the least value while the highest was given by *T. harzianum* (92.10 %). It was observed that throughout the removal process, *Trichoderma spp.* gave the best result most especially *T. harzianum* compared with other organisms such as *A. niger*, *A. flavus*and *Fusarium oxysporum*used in the biosorption process. This can be seen in Table 7.

Table 7.Percentage removal of Zinc (Zn) by different fungal isolates

Organisms/Day	% Biosorption at different days			
	D-7	D-14	D-21	D-28
A. niger	26.16	50.71	67.34	81.97
A. flavus	18.81	27.55	53.47	68.26
T. viride	38.02	57.85	67.49	86.29
T. harzianum	54.77	63.64	73.55	92.10
F. oxysporum	30.67	42.15	49.42	69.75

3.3Biosorption Study Using Fourier-transform Infrared (FTIR) Spectroscopic Techniques

FTIR analysis was carried out to study the interaction between the fungal isolates and the contaminants in the e-waste-polluted water samples. The functional groups were used in monitoring this. Exactly 5 ml of each of the samples drawn on days 7, 14, 21, and 28 of the experimental set-ups were smeared on the attenuated total reflectance (ATR) and mounted on the machine. The smears obtained were analyzed with a Shimadzu IR Affinity-1S FTIR spectrophotometer covering a frequency of 4000-500 cm^{-1}. The machine was directed to analyze after all the parameters have been set. The result of Fourier Transform Infrared (FTIR) carried out showed that the fungal isolates used were able to cause vibration in absorption frequencies thereby leading to the disappearance and or reduction of their absorption frequencies. The peaks on the control experiment where no fungal discs were used are: 455.56 cm^{-1}, 1035.77 cm^{-1}, 1462.04 cm^{-1}, 1637.56 cm^{-1}, 2019.47 cm^{-1}, 2200.78 cm^{-1}, 2360.87 cm^{-1}, and 3327.21 cm^{-1} while that of day 28 were 432.05 cm^{-1}, 513.07 cm^{-1}, 827.46 cm^{-1}, 1001.06 cm^{-1}, 1635.64 cm^{-1}, 1975.11 cm^{-1}, 2166.06 cm^{-1}, and 3317.56 cm^{-1} as can be seen in Fig. 2 and Fig. 3 respectively.

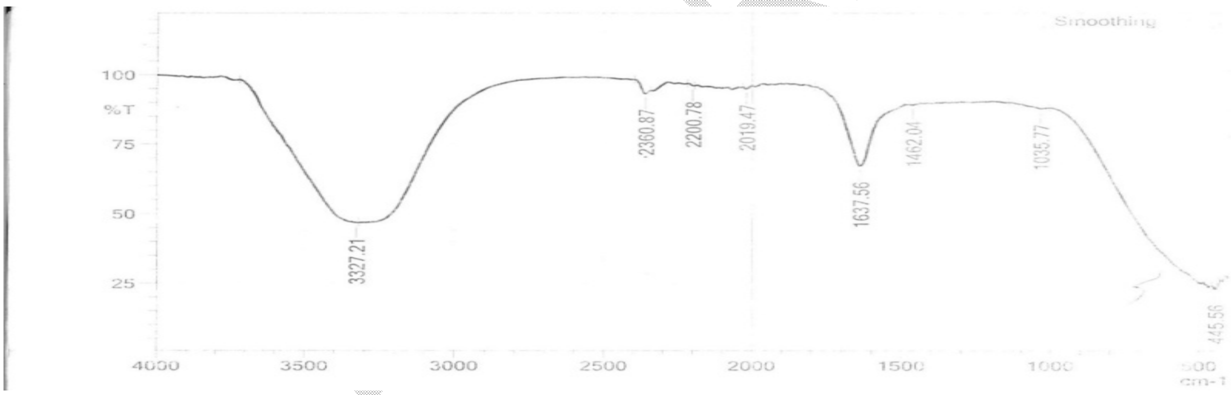

Fig. 2 FTIR of Control Experiment without fungal discs

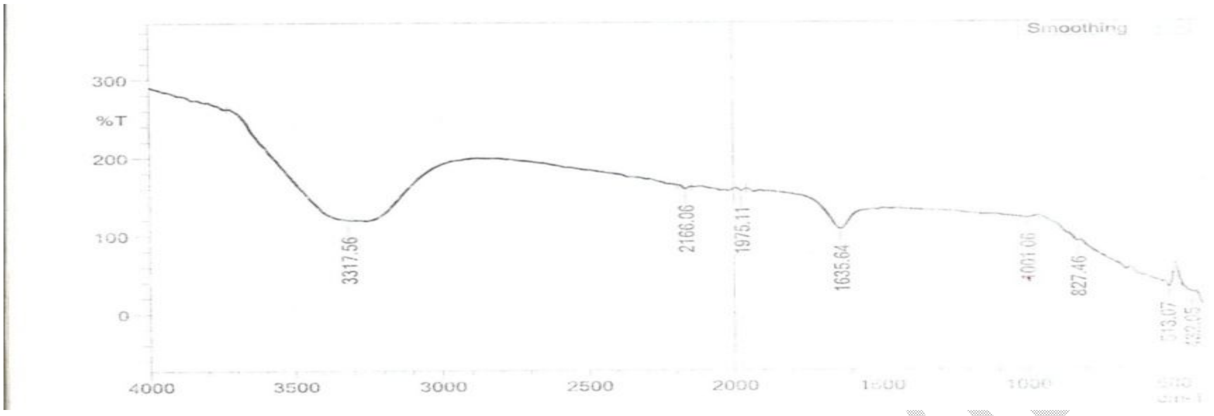

Fig. 3 FTIR at Day 28 of biosorption study with fungal discs

4.0 DISCUSSION

It was evident from this work that extracts from brewer-spent grain and honeycomb used as carbon sources were observed to greatly support *Aspergillus niger, A. flavus, Trichoderma viride, T. harzianum* and *Fusarium oxysporum* in the removal of Pb, Cd, Cr, Zn and Ni. Industrial activities such as electrical waste, technological development and so on have led to the continuous discharge of heavy metals into the environment posing a significant threat to public health worldwide. Fungi have been reported to be a far better choice for bioremediation of metal [24]. Fungi can accommodate high concentrations of heavy metal build-up without having toxic effects [25]. Hence, fungi selected for this research work have shown a unique ability to accumulate metal; they were also very easy to culture in microbiological media [26]. *Trichoderma harzianum* and *T. viride*showed higher biosorption of lead on day 28 when compared to other microorganisms which were *Aspergillus flavus*, *A. niger*and *Fusarium oxysporum*. *Aspergillus niger, A. flavus*and *F. oxysporum* showed low biosorption of lead compared to other isolates. *T. harzianum* with high lead uptake indicates that it has more binding sites on its cell wall thus acting as a biosorbent in the removal of lead from industrial effluent containing higher lead concentration [27]. Similar results have been reported for the uptake of lead by different fungi [28,29,30]

Zinc has been applied in a broad diversity of cellular processes and is obligatory for retaining macromolecule structural stability, therefore, serving as a co-factor for about 300 enzymes [31].In this study, *A. flavus* and *F. oxysporum* recorded lower biosorption levels of zinc on day twenty-eight. This reduction could be due to the toxic concentration of heavy metals by the electrical waste or an increase in a high level of tolerance and biosorption properties of *F. oxysporum* and *A. flavus*. This result agrees with the findings [32] that reported that some heavy metals' toxicity in fungi is a result of their strong binding affinities with the cell membrane which causes loss of structural integrity and impairment of cell function. *Aspergillus niger, Trichoderma viride,* and *Trichoderma harzianum* showed an important increase in the biosorption of zinc but lower compared to *Aspergillus flavus* and *Fusarium oxysporum.* Similar results have been postulated in the use of *Aspergillus niger*, *and Penicilliumchrysogenum* as biosorbent in the removal of Zinc

Nickel is recognized as a chief ecological contaminant. It has clastogenic, toxic and carcinogenic effects [33]. In this study, *Aspergillus niger,* and *Fusarium oxysporum* have lower biosorption levels for lead (Pb) compared to *Aspergillus flavus, Trichoderma viride*and *Trichoderma harzianum*. The decline observed in *Aspergillus niger* and *Fusarium oxysporum* can be due to a decrease in biosorption of the microbes from the ecological surroundings which may be due to microbe-fungi interactions of metals in fungal cell wall as postulated [34]. The report also showed that the adsorption of heavy metals is controlled by a fungi cell wall and is used for future clean-up of pollutants from the environment. Similar findings were made by researchers [19] who reported biosorption of Nickel by *Bacillus subtilis, Micrococcus luteus*and *Trichoderma harzianum*. In addition, other researchers [32] have also reported the use of *Aspergillus niger*and *A. flavus* in the biosorption of nickel. Similar results in tandem with the acceptance of Nickel by fungi have also been acknowledged in literature [35, 36]

Cadmium has been postulated by many researchers to have a renowned biological function if bio-available. Cadmium causes toxicity to cells by interacting with nucleic acids thereby producing reactive oxygen species that cause oxidative damage which leads to respiratory problems [37]. In this research, it was observed that the biosorption of cadmium by *A. niger, A. flavus* and *Fusarium oxysporum* followed the same trend all through the period of the experiment. This finding agrees with the other reports [32] that *A. flavus and A. niger* showed almost the same cadmium uptake ability during the biosorption of heavy metals. *Trichoderma viride*and *T. harzianum* recorded the highest level of biosorption of cadmium on day twenty-eight which could be due to a higher amount of cadmium uptake by the biomass compared to *Aspergillus niger, A. flavus* and *Fusarium oxysporum*due to the electrostatic interactions of the cell surface functional groups. These findings correlate with the report [26] that *T. viride*and *T. harzianum* showed higher amounts of cadmium uptake compared to other related microorganisms. Similar findings were also reported [38] that *Trichoderma* species have bioremediation effectiveness and their biosorption ability differs depending on the species. They are also known to be important fungi in decreasing Cadmium ions. Moreover, similar results which have been published about the uptake of cadmium by fungi have also been reported [39, 40, 41, 42]

Chromium is a crucial element helping the body to make use of essential elements. Extreme ingestion or exposure can lead to damage to the body. It has also been reported that *A. niger, A. flavus*and *Trichoderma viride* have been used in removing chromium from metal-contaminated wastewater [32, 43]. The use of *Bacillus cereus* in the removal of chromium from effluent has also been reported [44]. Similar findings to the uptake of chromium by fungi have also been documented [45, 46, 47, 48]. In this study, *Trichoderma viride*and *T. harzianum* showed higher biosorption compared to *Aspergillus niger, A. flavus* and *Fusarium oxysporum.*

5.0 CONCLUSION

In summary, the ability of the fungal isolates to bind and reduce heavy metals from e-waste polluted water, especially *Trichoderma harzianum* was established. It recorded a very high tolerance for biosorption of heavy metals which is an indication that it is a resourceful biological agent that can be harnessed for toxic environmental contaminants removal.

Therefore, it is essential that heavy metals be remediated from polluted water body using indigenous fungi.This can be achieved biologically using any of the fungi isolated in this work especially *Trichoderma harzianum.*

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