

Antioxidant and Cytotoxic Activities of *Pulicaria dysenterica* Methanol Extracts

ABSTRACT

Aims: The aims of the study were to analyse the polyphenols of *Pulicaria dysenterica* (L.) Bernh. methanolic extracts from aerial and underground parts, assessment of antioxidant activity and to evaluate their cytotoxicity on HeLa cells of cervical cancer.

Methodology: The total phenolic content (TPC) of extracts was determined by the Folin-Ciocalteu spectrophotometric method. The qualitative and quantitative analysis of individual polyphenolic compounds were performed by the reverse phase HPLC method. The antioxidant capacity was evaluated by both, 2,2-diphenyl-1-picrylhydrazyl radical and FRAP assay, while cytotoxicity of the extracts was assessed by MTT assay.

Results: TPC of the samples were 127.62 ± 2.22 and 244.12 ± 8.84 mg gallic acid equivalent/g extract. In the extracts chlorogenic acid in amount of 10.06 ± 0.96 and 11.32 ± 0.28 mg/g, flavonoid rutin in amount of 5.68 ± 0.13 mg/g and three caffeic acid derivatives were recorded. Extract from underground parts achieved better antioxidant activity with IC_{50} value 55.36 ± 0.75 $\mu\text{g/mL}$ and FRAP value 2411.12 ± 37.22 $\mu\text{mol Fe}^{2+}\text{g}^{-1}$ compared to the one from aerial parts. Extract from aerial parts achieved better cytotoxic activity with 50% inhibition of viability (IC_{50}) at concentration of 0.389 ± 0.07 mg/mL, against HeLa cells, compared to the extract from underground parts.

Conclusion: Analyzed *Pulicaria dysenterica* extracts contained phenolic acids and flavonoids. The extracts showed good antioxidant activity and cytotoxic properties against HeLa cells *in vitro*.

Keywords: *Pulicaria dysenterica*, polyphenols, antioxidants, cytotoxicity

1. INTRODUCTION

Pulicaria dysenterica (L.) Bernh. syn. *Inula dysenterica* L., Asteraceae, is a perennial plant, up to 100 cm height with yellow flowers, growing on damp places. It is found in South, West and Central Europe, Anatolia, Iraq, Iran, Afghanistan, Pakistan and North Africa. The aerial parts of the *Pulicaria dysenterica* are used in the treatment of diarrhea and dysentery in Iranian traditional medicine. They are also used for the treatment of dysentery in the United Kingdom. The plant has an insecticidal property, as well" [1,2]. "Phytochemical analysis of genus *Pulicaria* showed the occurrence of molecules of monoterpenes, diterpenes, sesquiterpenes, triterpenes, flavonoids and steroids. Previous studies on the chemical constituents of aerial parts of *P. dysenterica* included volatile oils, sesquiterpenes, flavonoids and caryophyllenes" [2]. "Flavonoid detected in aerial parts of *P. dysenterica* was quercetin 3-glucuronide. Other chemicals as quercetagenin 3,7-dimethyl ether, 6-hydroxykaempferol 3,4'-dimethyl ether, acetylenes were also recorded" [3,4]. „Studies on the composition of the essential oil of the aerial parts of the *P. dysenterica* from Greece and Iran confirmed well known fact that the chemical composition of the essential oil depends on various parameters such as environmental conditions. Significant difference in the composition and percentage of ingredients was recorded" [1]. A limited number of studies have been carried out concerning the chemical constituents and biological activities of the plant. In favor to its traditional use, it was shown that extracts of *P. dysenterica* aerial parts were active against

32 bacterial strain *Vibrio cholerae*, as well as against parasite *Trichomonas gallinae* in *in vitro*
33 tests [5,6]. "Cadiz-Gurrea et al. evaluated biological properties of *P. dysenterica* methanolic
34 extracts from aerial parts by *in vitro* inhibitory potential of enzymes (lipase, α -amylase, α -
35 glucosidase, tyrosinase, and cholinesterases). Extracts showed promising results for the
36 management of diabetes type II, Alzheimer's disease and skin hyperpigmentation disorders
37 and obesity. They also examined antioxidant capacity of the extracts" [2]. Furthermore, study
38 about cytotoxic effects of related *Inula species* against some tumor cell lines showed good
39 results with IC_{50} values from 17.96 μ g/mL [7]. For some species of genus *Pulicaria* cytotoxic
40 activities have been reported, as well [2].
41 However, data on the composition and activity of aerial and underground parts of this plant
42 still lack. The aims of the study were to analyse the polyphenols of *P. dysenterica* methanolic
43 extracts, assessment of antioxidant activity by different methods and to evaluate their
44 cytotoxic activities *in vitro* on HeLa cells of cervical cancer, bearing in mind broad set of
45 activities that plant polyphenols might exhibit [8,9].

46 2. MATERIAL AND METHODS

47
48 All analyses were performed using analytical grade chemicals and reagents. Folin-
49 Ciocalteu's phenol reagent, sodium carbonate, sodium acetate anhydrous, and ferric (III)
50 chloride were obtained from Merck (Germany). Acetonitrile, high performance liquid
51 chromatography (HPLC) grade and formic acid were purchased also from Merck. Methanol,
52 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical, 2,4,6-tris(2-pyridyl)-s-triazine (TPTZ), ferrous
53 (II) sulfate heptahydrate, hydrochloric acid, glacial acetic acid and HPLC-grade chlorogenic
54 acid were purchased from Sigma-Aldrich (USA). Minimum Essential Medium Eagle (MEM), 2
55 mM L-glutamine, nonessential Amino Acids, heat inactivated fetal bovine serum (HI FBS),
56 penicillin/streptomycin antibiotics and thiazolyl blue tetrazolium bromide (MTT) were
57 purchased also from Sigma-Aldrich. Rutin was obtained from Carl Roth (Germany). Water
58 for HPLC was prepared by Milli-Q Water Purification System. Double-distilled deionized
59 water or culture medium were used for solution preparations and dilutions for MTT assay.

60 2.1 Sampling Plant Material

61 Aerial and underground parts of *P. dysenterica* (PDA and PDU) were collected at the
62 mountain pass Karaula (N44°10'13.6" E18°39'07.2"), municipality Olovo, Bosnia and
63 Herzegovina, during the flowering period in July 2020. The plant material was identified
64 according to the Flora Croatica by authors and the voucher specimen (No. 20/3.1.I) was
65 deposited at the Department of Pharmacognosy, Faculty of Pharmacy, University of Tuzla
66 [10].

67 2.2 Preparation of Methanol Extracts

68 The air-dried plant material was crushed in a grinder until powder formation. The samples
69 were extracted with 98% methanol (2.5 g plant material with 50 mL of solvent) on a magnetic
70 stirrer under reflux at 50°C for 1 hour. The mixtures were filtered through a filter paper
71 (Whatman No. 1). The solvent was removed by evaporation. The dried extracts were stored
72 in the fridge at 4°C, in glass bottles for further investigations.

73 2.3 Determination of Total Phenolic Content (TPC)

74 TPC was determined by the Folin-Ciocalteu spectrophotometric method [11]. Stock solutions
75 (2 mg/mL) of extracts were prepared in methanol. One hundred μ L of an extract was mixed
76 with 7.9 mL of distilled water. Folin-Ciocalteu reagent (500 μ L) was added. After 8 min, 1.5

77 ml of 20% Na₂CO₃ was added. After 2 hours of incubation at room temperature (20°C ≤ t ≤
78 25°C), the absorbance was measured at 765 nm. Quantitative measurements were
79 performed using a standard calibration curve of different concentrations of gallic acid (20,
80 100, 200, 300, 400 and 500 mg/L) in the same way. The results for TPC are expressed in
81 galic acid equivalents (GAE) as miligram per gram of dry extract (mg GAE/g).

82 2.4 HPLC analysis

83 "HPLC analyses of extracts (1 mg/mL in methanol) were carried out using an Agilent 1260
84 Infinity system equipped with an Agilent 1260 Infinity Quaternary Pump, Agilent 1260 Infinity
85 Standard Autosampler, Agilent 1260 Infinity Diode Array Detector, and Agilent 1260 Infinity
86 Thermostatted Column Compartment. The separations were performed on a Merck
87 LiChroCARTR250-4 C18 reverse phase (RP) analytical column (250x4.6mm i.d., 5µm). The
88 mobile phase consisted of 0.1% formic acid in water (solvent A) and acetonitrile (solvent B).
89 The following gradient was applied: 0-15 min, linear gradient from 10% to 20% B; 15-30 min,
90 linear gradient from 20% to 30% B; 30-35 min, linear gradient from 30% to 40% B; 35-40
91 min, linear gradient from 40% to 90% B; 40-45 min, then returned to the initial conditions.
92 The injection volume was 10 µL; the flow rate was 0.8 mL/min. The detection wavelength
93 was 280 nm, 325 nm, 360 nm and the column thermostat was set at 30°C" [12]. Component
94 identification was performed comparing their retention times and UV spectra with those
95 obtained from standards. The calibration curve for chlorogenic acid was obtained by the
96 external standard method in the concentration range of 15.6-500 µg/mL (R² =0.9996,
97 y=28.93x-220.2) and the calibration curve for rutin was obtained by the external standard
98 method in the concentration range of 1-20 µg/mL.

99 2.5 Determination of Antioxidant Capacity

100 The *in vitro* antioxidant capacity of *P. dysenterica* extracts was evaluated by DPPH radical
101 scavenging assay and the ferric reducing antioxidant power (FRAP) assay.

102 2.5.1 DPPH Radical Scavenging Assay

103 Stock solutions (1 mg/mL) of extracts were prepared in methanol. The reaction mixture
104 contained 75 µL of extract solution and 75 µL of 0.3 mM DPPH solution in methanol.
105 Extracts and DPPH solution were mixed in microtiter plates and incubated in a dark place for
106 30 min at a room temperature (20°C ≤ t ≤ 25°C). A blank was measured for each sample.
107 Instead of DPPH, methanol was added to the blanks. Controls were 98% methanol plus
108 DPPH. Absorbance was measured colorimetrically at 517 nm on microtiter plate reader. The
109 DPPH scavenging activity (SA) in percentage was determined as follows:

110 $SA\% = 100 \times [(Ac - Ao) - (As - Ao)] / (Ac - Ao)$, where Ac is the absorbance of the control, Ao
111 the absorbance of the blank and As the absorbance of the sample. Rutin was used as a
112 reference substance. Results of DPPH assay are expressed as IC₅₀ (µg/mL), defined as the
113 concentration of extract required to remove 50% of free radicals [13].

114 2.5.2 FRAP Assay

115 Stock solutions (2 mg/ml) of extracts were prepared in methanol. In short, 100 µL of extracts
116 diluted with methanol were mixed with 3.0 mL of freshly prepared FRAP reagent consisting
117 of 300 mM acetate buffer (pH 3.6), 10 mM TPTZ solution in 40 mM HCl and 20 mM FeCl₃
118 (10:1:1). Samples were incubated for 30 min and absorbance measured at 593 nm on
119 spectrophotometer. A standard calibration curve has been prepared using different

120 concentrations of $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ from 100 to 1000 $\mu\text{mol/L}$ ($R^2=0.9985$, $y=0.0009x-0.0534$).
121 The results of FRAP assay are expressed as $\mu\text{mol Fe}^{2+}/\text{g}$ dry extract [14].

122 2.6 Determination of Cytotoxicity

123 2.6.1 *In vitro* culture of the cell lines

124 HeLa (cervical cancer) cell line was cultured in MEM supplemented with 2 mM glutamine,
125 1% nonessential Amino Acids, 10% HI FBS and 1% penicillin/streptomycin antibiotics. Cells
126 were maintained in humidified atmosphere containing 5% CO_2 at 37°C. For each experiment
127 cells were grown to 80% confluence in cell culture flasks.

128 2.6.2 MTT Cell Proliferation Assay

129 Cytotoxic effects of *P. dysenterica* extracts were assessed by MTT assay [15]. For each
130 experiment cells were seeded (2×10^4 cells/well) in 96 well plates and incubated overnight.
131 Next day, cells were treated with increasing final concentrations of *P. dysenterica* extracts
132 (40-4000 $\mu\text{g/mL}$) and incubated for additional 48h. After incubation the cells, MTT solution
133 0.5 mg/mL was added in each well, and plates were incubated for another 4 hours at 37°C in
134 humidified atmosphere containing 5% CO_2 . Then the medium containing MTT was removed
135 and the remaining MTT-formazan crystals were dissolved by adding 200 μL DMSO to each
136 well with continuous gentle shaking for 15 minutes. Absorbance was read using microplate
137 reader at a wavelength of 570 nm. Experiment was repeated three times and each
138 experiment was performed in triplicate. Untreated cells were used as negative control and
139 positive control were cells treated with 30% DMSO in culture medium. Samples were
140 dissolved in 10% DMSO and diluted in culture medium. The final DMSO concentration in
141 treated samples did not exceed 0.1%. Prepared stock solutions of extracts were sterilized by
142 filtration through 0.2 μm sterile syringe filters. The concentration of the extracts leading to
143 50% of viability (IC_{50}) was assessed from graph plots of the dose response curve.

144 3. RESULTS AND DISCUSSION

146 3.1 Yield and polyphenolic composition of extracts

147 The yields of the methanol extracts of *P. dysenterica* were 7.28 ± 0.20 and $9.43 \pm 0.71\%$ while
148 the TPC of the samples were 127.62 ± 2.22 and 244.12 ± 8.84 mg GAE/g extract (Table 1).
149

150 **Table 1. Yields, content of total phenols, IC_{50} and FRAP values of *P. dysenterica***
151 **methanolic extracts**

Sample	Yields (%)	TPC (mg GAE/g)	DPPH IC_{50} ($\mu\text{g/mL}^{-1}$)	FRAP [$\mu\text{mol Fe}^{2+}\text{g}^{-1}$]
PDA	7.28 ± 0.20	127.62 ± 2.22	157.06 ± 1.83	1296.67 ± 39.77
PDU	9.43 ± 0.71	244.12 ± 8.84	55.36 ± 0.75	2411.12 ± 37.22

153 Data presented as mean value \pm standard deviation (n=3),
154

155 The qualitative and quantitative analysis of individual polyphenolic compounds of the
156 extracts from aerial and underground parts of *P. dysenterica* were performed by the RP-
157 HPLC method. Obtained chromatograms are presented on Fig. 1.

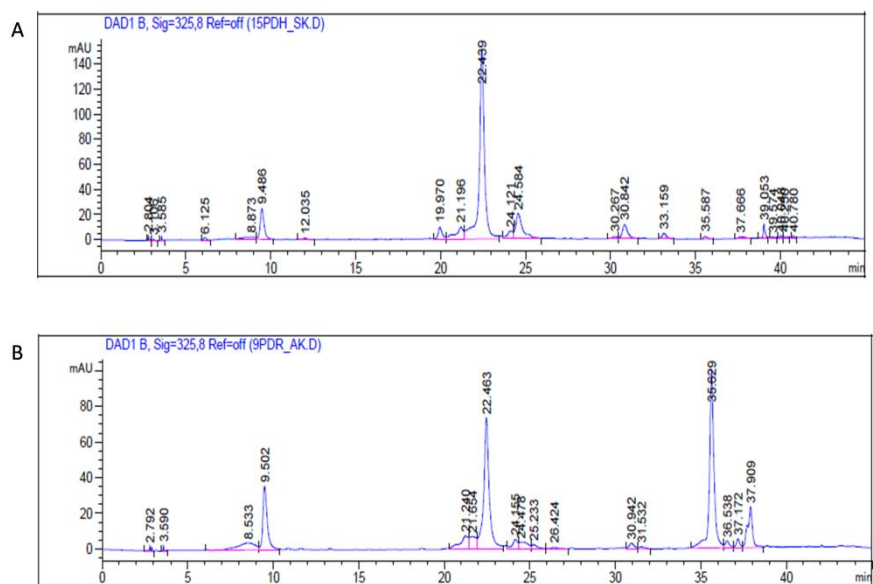


Fig. 1. HPLC chromatograms of *P. dysenterica* extract from aerial parts (A), and underground parts (B) ($\lambda = 325$ nm)

In the extract from aerial parts of *P. dysenterica* chlorogenic acid and rutin were identified, as well as one more derivative of caffeic acid. The content of rutin was 5.68 ± 0.13 mg/g. Extract from underground parts contained chlorogenic acid, three derivatives of caffeic acid, but the flavonoid rutin was not detected by the applied method. There was no big difference in chlorogenic acid content in extracts from aerial and underground parts of the plant (Table 2).

Table 2. Compounds of *Pulicaria dysenterica* extracts

Retention time (min)	Proposed compound	PDA extract (mg/g)	PDU extract (mg/g)
9.486	chlorogenic acid	10.06 ± 0.96	11.32 ± 0.28
19.970	rutin	5.68 ± 0.13	-
22.463	caffeic acid derivative 1	x	x
35.629	caffeic acid derivative 2	-	x
37.909	caffeic acid derivative 3	-	x

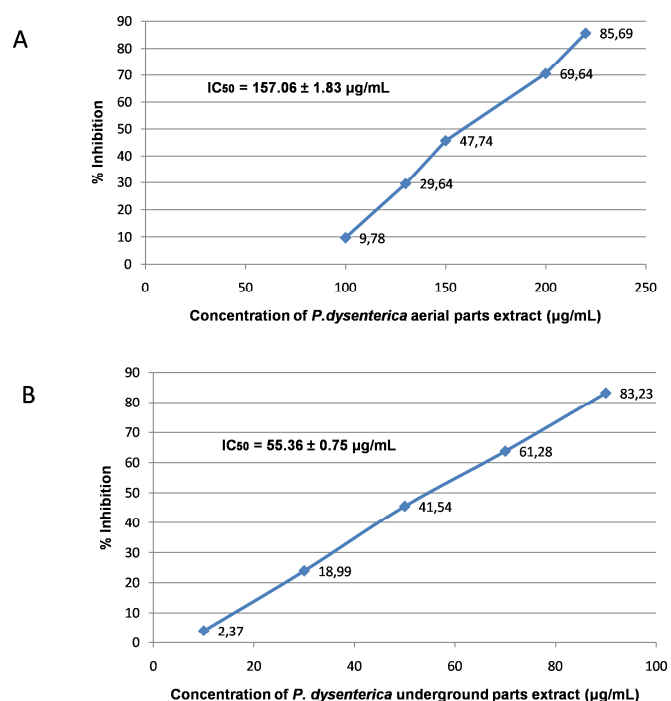
^amean value \pm standard deviation (n=3), x- presence, - not detected

In methanolic extracts from aerial parts of *P. dysenterica* from Turkey total phenolic, flavonoid and phenolic acid were detected. The phenolic content ranged from 80.62 ± 2.87 to 119.40 ± 2.67 mg GAE g^{-1} . In those extracts quinic acid, chlorogenic acid and its isomers, dicaffeoylquinic acid and its isomers, luteolin malonyl glucoside and quercetin glucoside were determined [2]. TPC in analysed extract from aerial parts is slightly higher compared to published literature data.

3.2 Results of testing antioxidant activity

"A number of studies have shown that phenolic compounds have biological activities such as antioxidant, antimicrobial, and antitumor. Thus, phenolic compounds can protect cellular

182 components against oxidative damage and therefore reduce the risk of degenerative disease
183 due to oxidative stress“ [16]. The results of DPPH and FRAP tests are shown in Table 1.
184 Graphs about the antioxidant results for DPPH assay are presented on Fig. 2.



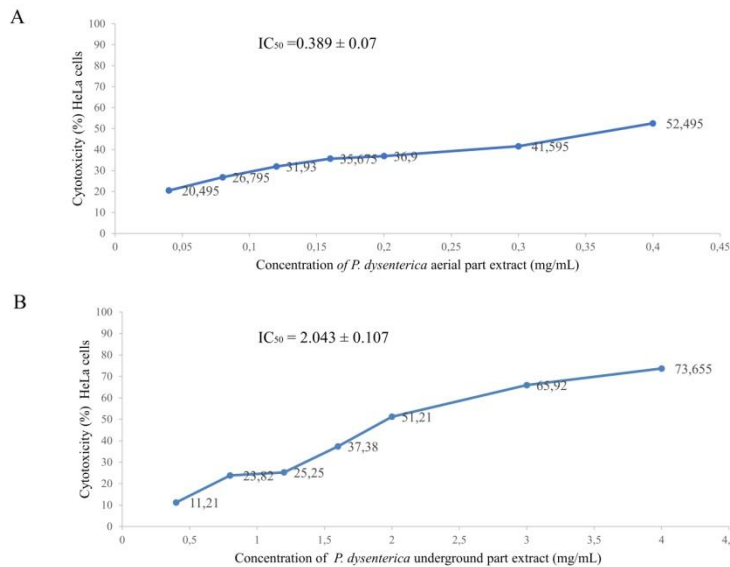
185 **Fig. 2. Antioxidant activity, assessed by DPPH of *P. dysenterica* extract from aerial**
186 **parts (A), $IC_{50}=157.06\pm 1.83 \mu\text{g/mL}$ and *P. dysenterica* extract from underground parts**
187 **(B), $IC_{50}=55.36\pm 0.75 \mu\text{g/mL}$. Values represent the mean value \pm standard deviation of**
188 **three separate experiments**

189
190
191 Higher DPPH radical scavenging activity was observed for the extract from *P. dysenterica*
192 underground parts with the IC_{50} value of $55.36\pm 0.75 \mu\text{g/mL}$ compared to the extract of aerial
193 parts. Higher antioxidant activity for the extract from *P. dysenterica* underground parts was
194 also observed with the FRAP value of $2411.12\pm 37.22 \mu\text{mol Fe}^{2+}\text{g}^{-1}$ compared to the extract
195 from aerial parts. The results obtained in this study confirm the well-known positive
196 correlation between the TPC and antioxidant capacity [17].

197 198 3.3 Results of cytotoxicity testing

199 The results of the cytotoxic activity suggested that *P. dysenterica* extracts induced a dose-
200 dependent inhibition of HeLa cell proliferation, in accordance to determined IC_{50} . The
201 measured absorbance values of extracts obtained from aerial and underground plant parts
202 were converted to percent of cell cytotoxicity with respect to negative control (Figure 1).

203



204

205 **Fig. 3. Cytotoxic effects on cancer cell line HeLa, assessed by MTT after 48 hours**
 206 **exposure to increasing concentrations of *P. dysenterica* extract from aerial parts (A),**
 207 **$IC_{50}=0.389\pm0.07$ mg/mL and *P. dysenterica* extract from underground parts (B),**
 208 **$IC_{50}=2.043\pm0.107$ mg/mL. Values represent the mean value \pm standard deviation of**
 209 **three separate experiments**

210 The IC_{50} values of methanol extracts from aerial and underground parts of *P. dysenterica*
 211 were $0.389\pm0,07$ and $2.043\pm0,107$ mg/mL respectively, against HeLa cells. For comparison,
 212 IC_{50} for curcumin against HeLa cells was 0.32 mg/mL [18]. In the previous studies methanol
 213 extract of *Pulicaria dysenterica* was tested against breast cancer cell lines and showed
 214 significant cytotoxic effects. The IC_{50} values of the extract on MCF-7 cell line was calculated
 215 as 27.05 μ g/mL [19]. To the best of our knowledge, other cytotoxic studies for *P. dysenterica*
 216 have not been published.

217 Previously, MTT assay was used to evaluate the cytotoxic effects of the 19 purified
 218 compounds isolated from the *Pulicaria insignis* against four human cancer cell lines,
 219 including HeLa. The best results against HeLa cells were obtained with flavonoids and
 220 sesquiterpene lactone [20]. *Pulicaria undulata* aerial parts extract was also evaluated for
 221 cytotoxicity against breast and hepatoma cancer. One of the flavonoids, among other
 222 isolated secondary metabolites, showed the highest activity against tested cell lines [21,22].
 223 In another study on the cytotoxicity one of the isolated pseudoguaianolide sesquiterpene
 224 from aerial parts of *P. crispa* (syn. *P. undulata*) showed cytotoxicity with $IC_{50} = 5.8\pm0.2$
 225 μ mol/mL to human bladder carcinoma cell line, EJ-138. [23].

226 Considering the presence of chlorogenic acid and rutin in the analyzed extracts, we
 227 investigated previously published data about its cytotoxic and antitumor effects on HeLa and
 228 other cancer cell lines. A study about cytotoxic activity against several cancer cell lines with
 229 water soluble green coffee bean extract was conducted. The extract was abundant with
 230 chlorogenic acid and its derivatives, and showed the highest activity on human colon cancer
 231 cells while on HeLa cells the cytotoxicity was lower with $IC_{50} = 1.4$ mg/mL [24]. Cytotoxic
 232 activity of our extract is almost four times stronger compared to the activity of extract
 233 abundant with chlorogenic acid and its derivatives. Considering that the content of

234 chlorogenic acid in our extract is about 1%, its contribution to the cytotoxicity is very small.
235 Chlorogenic acid was tested against breast cancer cell lines. Results were also low with IC_{50}
236 values ranged from $590.5 \pm 10.6 \mu\text{g/mL}$ to $1095 \pm 121.6 \mu\text{g/mL}$ [25]. "On the other side
237 chlorogenic acid isolated from *Piper betel* leaves was found to kill Bcr-Abl-positive chronic
238 myelogenous leukaemia (CML) cells *in vitro* in a dose-dependent manner without any
239 appreciable effects on Bcr-Abl-negative acute T-lymphoblastic leukemia cell line" [26,27].
240 Chlorogenic acid-induced radical oxygen species production inhibited cell viability in human
241 colon cancer cells [28]. Flavonoid rutin is demonstrated to inhibit the proliferation of breast,
242 colon, lung, and prostate cancers. It affected the cell capture and apoptosis processes,
243 reducing the number of metastatic nodules and cytotoxicity [29,30,31,32,33]. In a study
244 about cervical cancer phytotherapy, rutin showed high cytotoxic activity on HeLa cells with
245 IC_{50} value $30 \mu\text{g/mL}$ [34]. Considering the fact that currently analysed extract contains
246 approximately 0.5% of rutin, other secondary metabolites also contribute to its cytotoxicity. It
247 was recorded in several studies that effects of flavonoids were linked with their availability
248 and technological formulation. It is found that rutin in prenanoemulsion, which contributes to
249 the improvement of physical and pharmacokinetic properties of this flavonoid, improves its
250 cytotoxicity, as well [31,35].
251

252 **4. CONCLUSION**

253
254 Analyzed *Pulicaria dysenterica* extracts contained chlorogenic acid, caffeic acid derivatives
255 and flavonoid rutin. The extracts showed good antioxidant properties. Stronger antioxidant
256 activity was recorded in extract from underground then from aerial plant parts. There was
257 also a good cytotoxicity of *P. dysenterica* extracts against HeLa cells *in vitro* reported. Better
258 result was achieved for extract from aerial compared to extract from underground parts. To
259 our knowledge, this is the first report of *P. dysenterica* showing cytotoxic activity on cervical
260 cancer (HeLa) cell lines. Further research should be directed to isolation and determination
261 of the secondary metabolites, flavonoids and sesquiterpene lactones, primarily responsible
262 for the cytotoxicity.
263

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269

270 **AUTHORS' CONTRIBUTIONS**

271
272 This work was carried out in collaboration among all authors. Authors ZM and ECK designed
273 the study. ND, LK and LM contributed to samples preparation for total phenolic content
274 (TPC), HPLC analysis and antioxidant assessment. Authors ECK and EH carried out the
275 experiment for total and individual polyphenolic compounds and antioxidant assessment.
276 Authors ED and DH carried out the experiment for cytotoxicity assessment. Authors AS and
277 NS worked out majority of technical details and performed the analytic calculations and the
278 statistical analysis. Author ECK wrote the manuscript with support from author JA, MI and
279 ED. BŠK managed the literature searches and gave critical overview of the manuscript. All
280 authors read and approved the final manuscript.
281

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